## **Electronic Supplementary Information**

Engineering FeCo alloy@N-doped carbon layers by directly pyrolyzing Prussian blue analogue: new peroxidase mimetic for chemiluminescence glucose biosensing

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## General procedure for CL detection

Fig. S1 shows a diagram of the flow injection CL detection system, Two peristaltic pumps were used to deliver the solutions, in which one delivers FeCo@NC suspension and water carrier stream, the other does luminol solution and H<sub>2</sub>O<sub>2</sub> (or sample solution). The produced CL signal was monitored by a PMT (at a working voltage of -600 V) and recorded by a computer. The data was processed by REMAX software. The net CL intensity ( $\Delta I=I-I_0$ , where I and  $I_0$  were the CL intensity of sample and blank solutions, respectively) versus the concentration of target was used for calibration. At each target concentration, the injection was repeated for at least three times, and the average CL signal was recorded.



**Fig. S1**. Schematic diagram of the FIA (flow injection analysis)–CL system. A: FeCo@NC suspension; B: water carrier stream; C: luminol solution; D:  $H_2O_2$  or sample solution; PMT: photomultiplier tube, PC: personal computer.



Fig. S2. SEM images of (A) Fe<sup>III</sup>-Co PBA and (B) FeCo@NC-600.



Fig. S3. FT-IR spectra of Fe<sup>III</sup>-Co PBA and FeCo@NC-600.



Fig. S4. XPS survey spectra for FeCo@NC-600.



Fig. S5. EDS spectrum of FeCo@NC-600.



Fig. S6. Ramen spectrum of the FeCo@NC-X obtained at different carbonization temperature in the 400 °C to 900 °C range.



Fig. S7. TG-DTG curves of Fe<sup>III</sup>-Co PBA in  $N_2$  atmosphere.



**Fig. S8.** N<sub>2</sub> adsorption-desorption isotherms of Fe<sup>III</sup>-Co PBA and FeCo@NC-600. Inset: Pore size distribution curves of Fe<sup>III</sup>-Co PBA and FeCo@NC-600.



**Fig. S9.** The UV-visible absorption spectra of (A) TMB $-H_2O_2$ , (B) ABTS $-H_2O_2$  and (C) OPD $-H_2O_2$  systems in the presence of different catalysts. Reaction conditions: Concentration of TMB, ABTS and OPD: 0.5 mM; 0.1 mM  $H_2O_2$ ; Concentration of FeCo@NC-600, Fe@NC and Co@NC): 10 mg/L; pH 3.5 (0.2 M HAc-NaAc); 20 min reaction at room temperature.



**Fig. S10.** (A) Amperometric response of bare GCE (glassy carbon electrodes), Fe@NC modified GCE, Co@NC modified GCE and FeCo@NC-600 modified GCE in 0.1 M PBS buffer (pH 7.4) at applied potential of -0.4 V upon successive addition of 1.0 M H<sub>2</sub>O<sub>2</sub> at time intervals of 40 s. (B) The relative activity of FeCo@NC-600 with different radical scavengers. SOD concentration: 50 U/mL; 100 U/mL and 200 U/mL; AA concentration: 0.04 mM; 0.1 mM and 0.2 mM; Thiourea concentration: 0.5 mM; 1 mM and 2.5 mM. (C) ESR spectra of •OH radicals in the H<sub>2</sub>O<sub>2</sub>-DMSO system with and without FeCo@NC-600. (D) ESR spectra of  $\cdot$ O<sub>2</sub><sup>-</sup> radicals in the H<sub>2</sub>O<sub>2</sub>-DMPO system with and without FeCo@NC-600.



Fig. S11. Effect of carbonization temperature on the activity of the resultant FeCo@NC-X.



**Fig. S12.** CL kinetic curves of luminol-H<sub>2</sub>O<sub>2</sub>-FeCo@NC-600 system in O<sub>2</sub>-saturated (red), air-saturated (blue) and N<sub>2</sub>-saturated solutions (black), respectively.



**Fig. S13.** (A) Effect of pH of luminol: 10  $\mu$ M luminol, 0.5  $\mu$ M H<sub>2</sub>O<sub>2</sub>, 10 mg L<sup>-1</sup> FeCo@NC-600. (B) Effect of luminol concentration: 0.1 M Na<sub>2</sub>CO<sub>3</sub>-NaHCO<sub>3</sub> buffer solution (pH 11.3), 0.5  $\mu$ M H<sub>2</sub>O<sub>2</sub>, 10 mg L<sup>-1</sup> FeCo@NC-600. (C) Effect of FeCo@NC-600 concentration: 10  $\mu$ M luminol in 0.1 M Na<sub>2</sub>CO<sub>3</sub>-NaHCO<sub>3</sub> buffer solution (pH 11.3), 1.0  $\mu$ M H<sub>2</sub>O<sub>2</sub>. Error bars represent the standard deviation for three replicates detection. (D) The response curve of H<sub>2</sub>O<sub>2</sub> under the optimized conditions. Inset: the linear relationship between net CL intensity and concentration of H<sub>2</sub>O<sub>2</sub>. Error bars were acquired from three replicates determination.



**Fig. S14.** Selectivity of the proposed method for  $H_2O_2$  detection in the present of potential interfering substances. Blank: 1  $\mu$ M  $H_2O_2$ . The concentrations of tested substances were 1 mM for leucine, glycine, alanie, L-phenylalanine and glutanmic acid, 0.5 mM for proline, methionine, serine, arginine and glutamine, 10  $\mu$ M for tryptophane, 5  $\mu$ M for cysteine, glutanthione and homocysteine, 1  $\mu$ M for uric acid and ascorbic acid, 100 mg/L for HSA (human serum albumin) and 10 mg/L for IgG (immunoglobulin G), respectively. The error bars represents standard deviations based on three independent measurements.



Fig. S15. XRD spectra of the synthesized FeCo NPs.



**Fig. S16.** Variation of catalytic activity of the (A) FeCo NPs and (B) FeCo@NC-600 with storage time.

Table S1. Atomic concentration (%) of C, N, O, Fe, and Co in FeCo@NC-600.

Element	At % from EDS	At % from XPS
С	36.41	71.87
N	2.61	7.89
0	5.99	12.38
Fe	29.15	4.49
Со	25.84	3.37

Samples	Surface area $(m^2/g)$	Total pore volume (cm <sup>3</sup> /g)
Fe <sup>III</sup> -Co PBA	42.84	0.12
FeCo@NC-600	37.63	0.16

Table S2. The surface area and pore volume of Fe<sup>III</sup>-Co PBA and FeCo@NC-600.

Table S3. Atomic concentration (%) of N in FeCo@NC-X.

Samples	At % of N
FeCo@NC-500	11.7
FeCo@NC-600	7.89
FeCo@NC-700	6.66
FeCo@NC-800	2.82
FeCo@NC-900	1.93

Table S4. Recoveries of 1  $\mu$ M H<sub>2</sub>O<sub>2</sub> in the presence of foreign species.

Coexisting species	Concentration (mg/L)	Recovery (%)	Coexisting species	Concentration (mg/L)	Recovery (%)
Na <sup>+</sup>	23	101.90±2.19	Al <sup>3+</sup>	1.08	99.60±1.30
$\mathrm{K}^+$	31.2	100.00±3.23	Cl-	35.5	101.90±2.19
$\mathrm{NH_{4}^{+}}$	7.2	103.25±0.11	Br	1.6	$100.14 \pm 0.72$
Ca <sup>2+</sup>	16	105.38±0.57	NO <sub>3</sub> -	24.8	96.71±0.38
$Mg^{2+}$	4.8	96.71±0.38	$SO_4^{2-}$	48	$103.29 \pm 3.08$
$Zn^{2+}$	0.33	99.18±2.04	NO <sub>2</sub> -	0.46	102.19±0.61
$Pb^{2+}$	4.1	102.52±1.84	SO <sub>3</sub> <sup>2-</sup>	0.8	94.21±0.80
Ni <sup>2+</sup>	0.3	8.67±1.73	HPO4 <sup>2-</sup>	1.92	$100.01 \pm 2.05$
Cr <sup>3+</sup>	52*	98.31±1.13	H <sub>2</sub> PO <sub>4</sub> -	9.7	103.92±2.36
$Ba^{2+}$	1.37	95.84±1.65	PO <sub>4</sub> <sup>3-</sup>	0.48	99.80±0.43

\*  $\mu g L^{-1}$ 

 Table S5. Comparison of the linear ranges and detection limits for glucose afforded

System	Linear range (M)	LOD (M)	Refs
ECL-GOD/AuNPs/PANi	1.0×10 <sup>-7</sup> -1.0×10 <sup>-4</sup>	5.0×10 <sup>-8</sup>	[5]
CL-MOF-235/β-CD	1.0×10 <sup>-8</sup> -3.0×10 <sup>-6</sup>	1.0×10 <sup>-8</sup>	[7]
CL-MIL-53(Fe)	1.0×10 <sup>-7</sup> -1.0×10 <sup>-5</sup>	5.0×10 <sup>-8</sup>	[8]
CL-gold NPs	8.5×10-7-1.0×10-4	4.3×10-7	[49]
CL-Co-Fe LDHs	5.0×10 <sup>-8</sup> -2.0×10 <sup>-5</sup>	2.0×10 <sup>-8</sup>	[50]
CL-carbon nitride quantum dots	5.0×10-7-5.0×10-5	1.0×10 <sup>-7</sup>	[51]
CL-GOD/GNPs/CNTs	2.25×10-6-1.75×10-4	1.0×10 <sup>-6</sup>	[52]
CL-CF-CoFe <sub>2</sub> O <sub>4</sub> NPs	5.0×10 <sup>-8</sup> -1.0×10 <sup>-5</sup>	1.0×10 <sup>-8</sup>	[53]
CL-CoFe <sub>2</sub> O <sub>4</sub> MNPs	1.0×10 <sup>-7</sup> -1.0×10 <sup>-5</sup>	2.4×10 <sup>-8</sup>	[54]
CL-WS <sub>2</sub> NS and AgNCs	3.0×10 <sup>-8</sup> -2.0×10 <sup>-5</sup>	1.3×10 <sup>-8</sup>	[55]
CL-Hemin@HKUST-1	7.5×10 <sup>-6</sup> -7.5×10 <sup>-4</sup>	7.5×10 <sup>-6</sup>	[56]
CL-FeCo@NC-600	1.0×10 <sup>-8</sup> -1.0×10 <sup>-5</sup>	8.5×10 <sup>-9</sup>	This work

by the proposed and other CL methods.