

## **Chlorotoxin Modified Morusin-PLGA Nanoparticles for Targeted Glioblastoma Therapy**

**Srishti Agarwal**<sup>1</sup>, M. Sheikh Mohamed<sup>1</sup>, Toru Mizuki<sup>1</sup>, Toru Maekawa<sup>1</sup>, D. Sakthi Kumar<sup>1\*</sup>

<sup>1</sup>Bio-Nano Electronics Research Center, Graduate School of Interdisciplinary New Science, Toyo University, Kawagoe, Saitama, 350 - 8585, Japan,

### **SUPPLEMENTARY INFORMATION**

#### **\*Corresponding Author**

Prof. D Sakthi Kumar

Bio-Nano Electronics Research Centre

Graduate School of Interdisciplinary New Science, Toyo University,

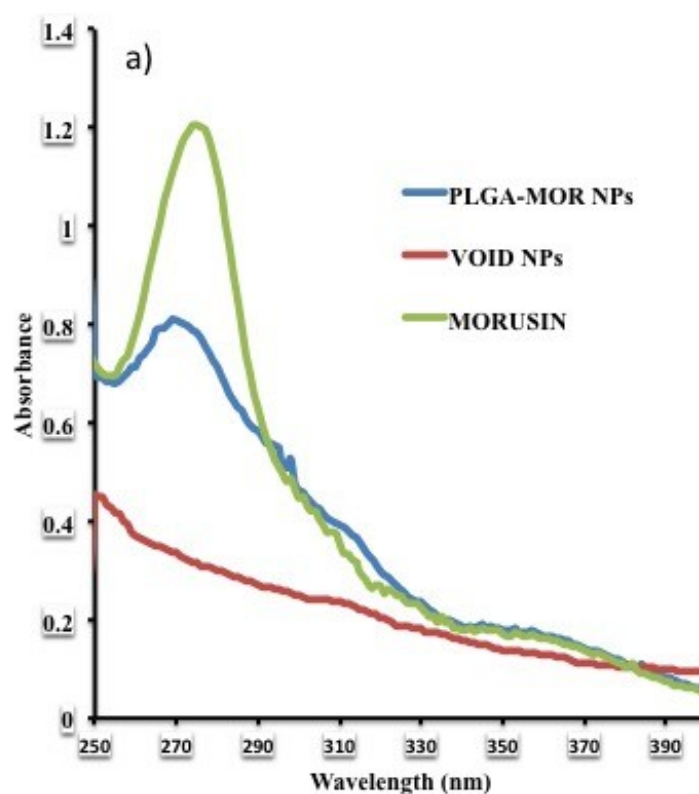
Kawagoe, Saitama-350-8585

Japan

Email: [sakthi@toyo.jp](mailto:sakthi@toyo.jp)

Phone: +81-492-39-1636/1375/1640

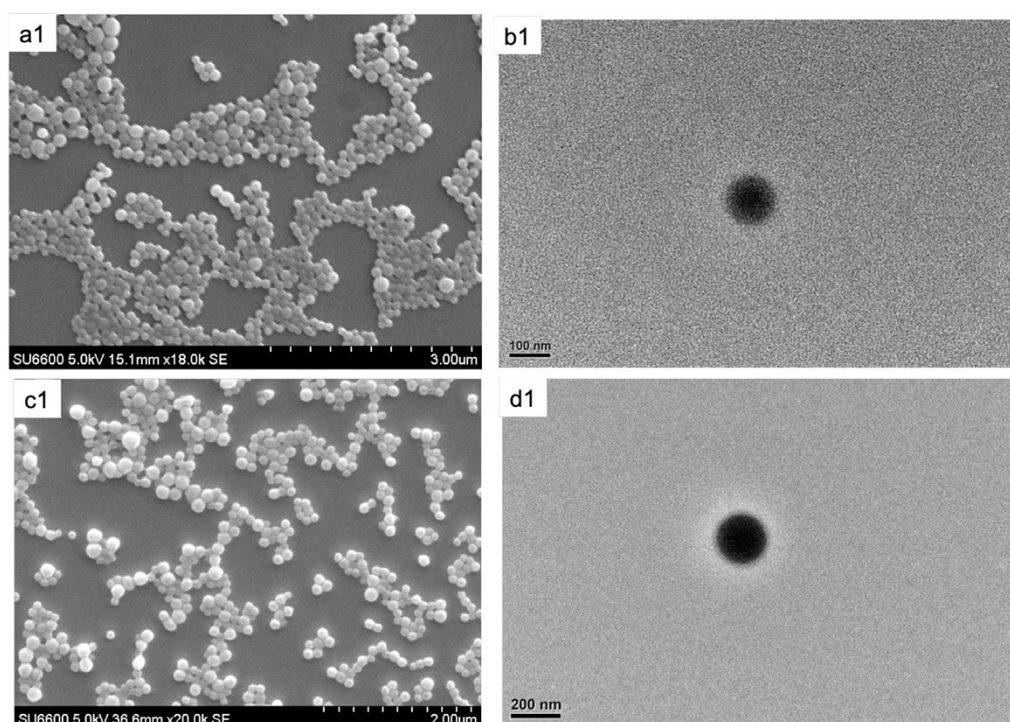
Fax: +81-366-77-1140



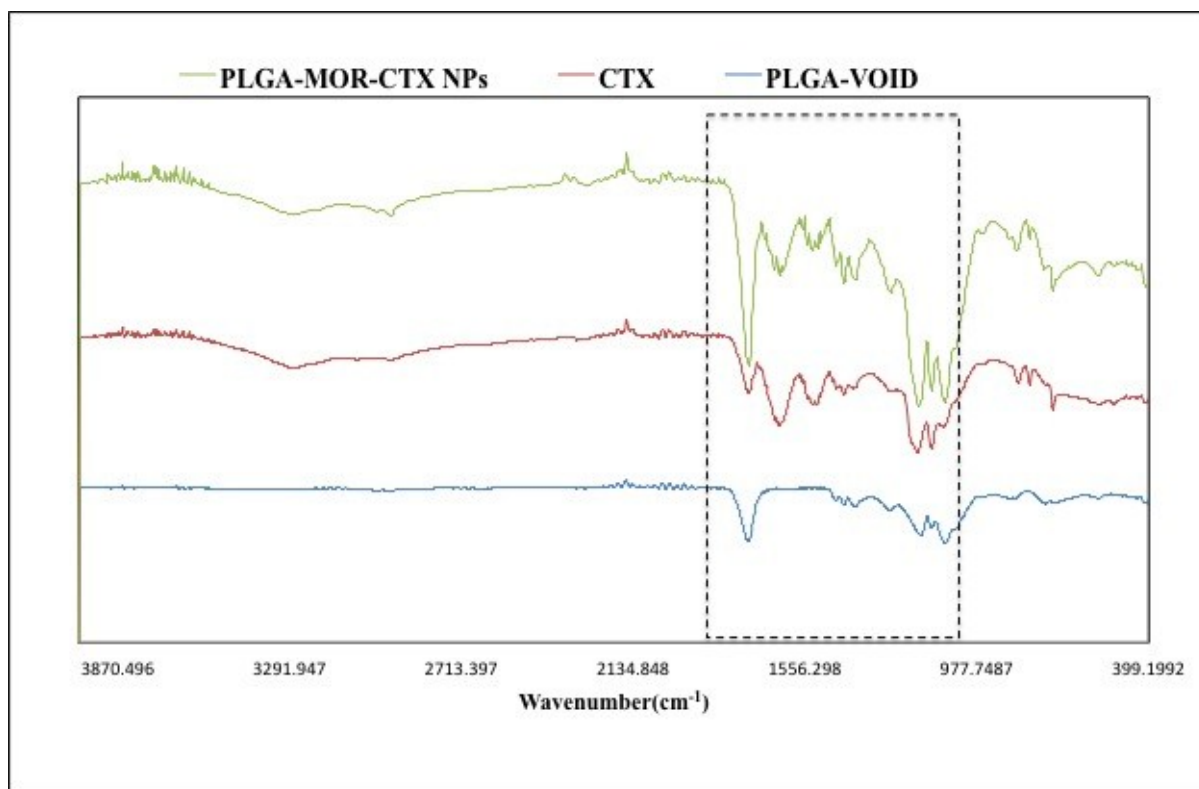
**Figure S1. Encapsulation of morusin inside PLGA-MOR NPs carried out by UV-Vis spectroscopy.** The supernatant of PLGA-MOR NPs dispersed in ethyl acetate was subjected to wavelength scan and the signature peak of morusin at 269 nm verified the presence of morusin in the NPs.

**Supplementary Table 1. Physiochemical characterization of void PLGA and PLGA-MOR NPs after a period of 1 month**

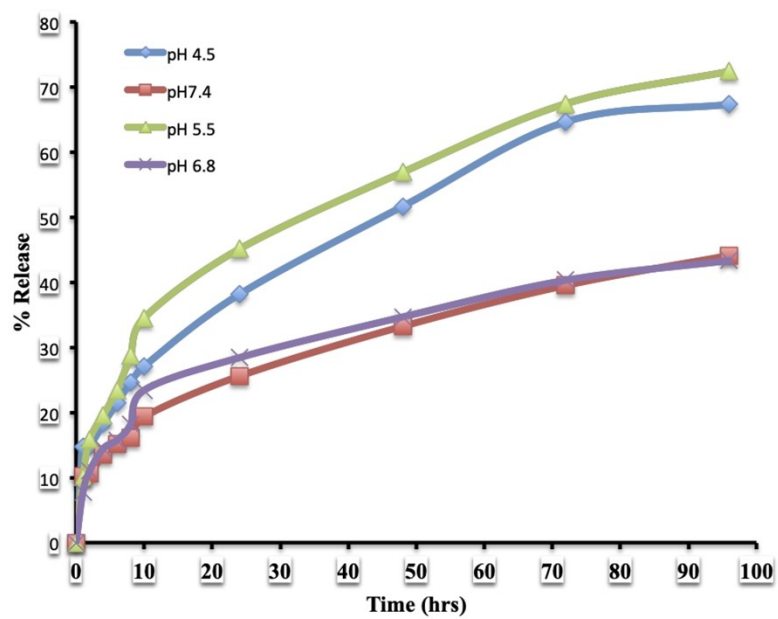
Characterization Parameter	Void PLGA NPs	PLGA-MOR NPs
Z average Diameter (nm)	175.81	242.9
Poly dispersity index (PDI)	0.125	0.209
Zeta potential (mV)	-21.7	-15.8
Entrapment Efficiency	-	97% $\pm$ 0.35



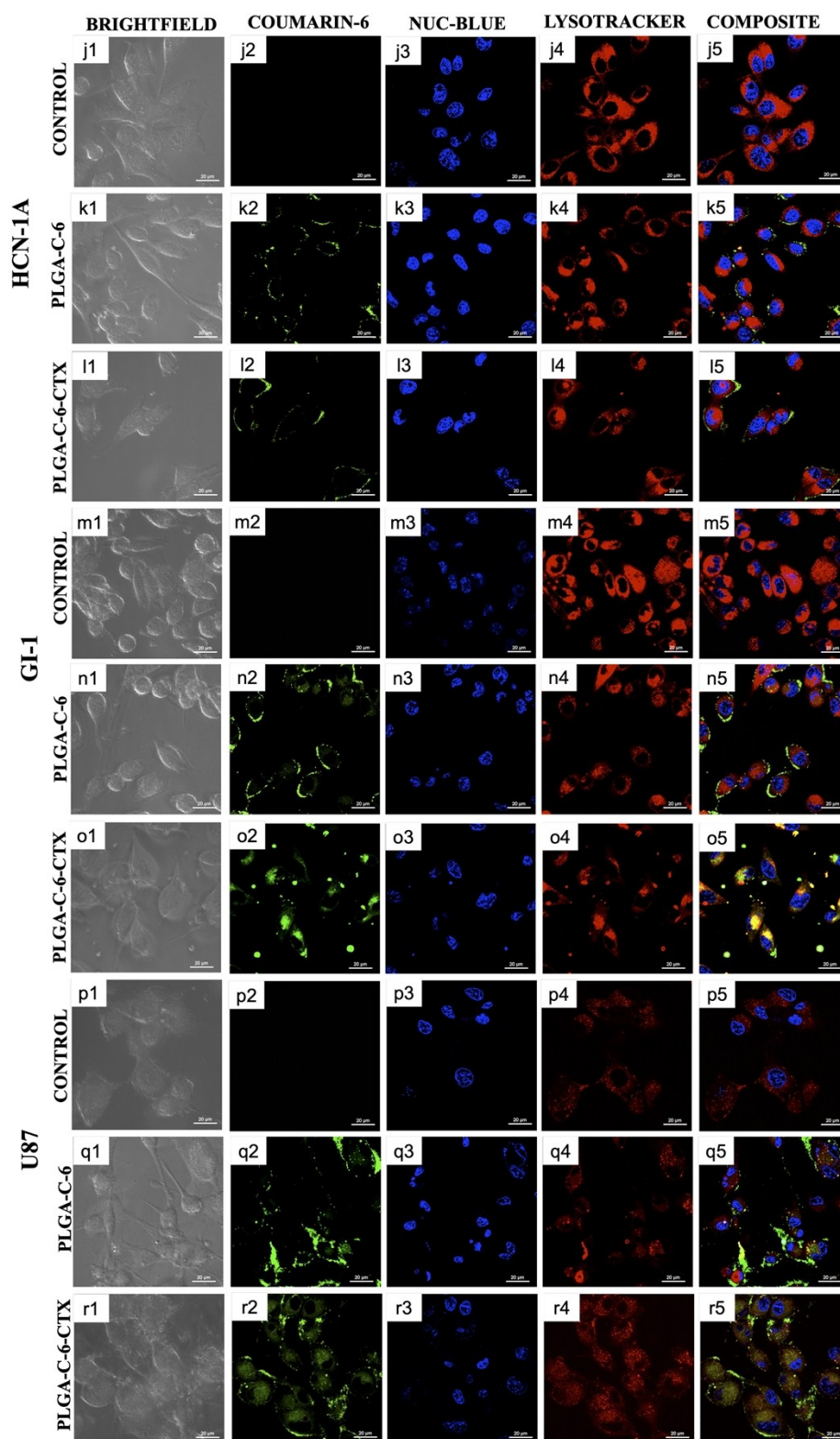
**Figure S2. Characterization of Void PLGA and PLGA-MOR NPs by Electron Microscopy:** a1, c1) SEM images of Void PLGA and PLGA-MOR NPs (At scale 3 μm & 2 μm). b, d) TEM images of Void PLGA and PLGA-MOR NPs (At scale 100 nm and 200 nm).



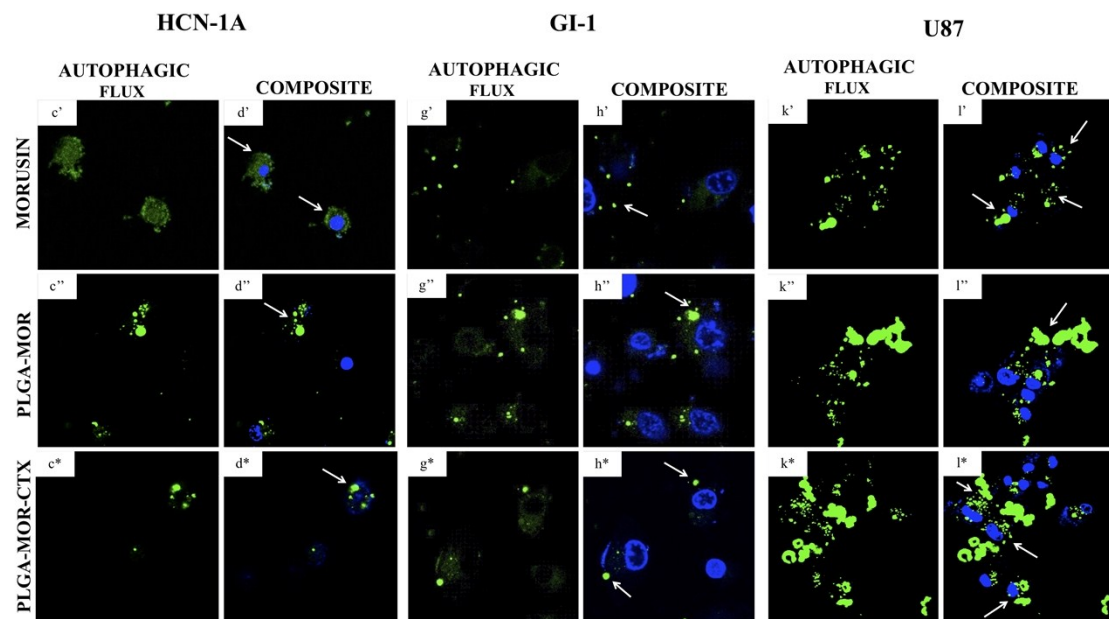
**Figure S3. ATR-FTIR analysis of free CTX, PLGA-VOID and PLGA-MOR CTX NPs confirming the conjugation of CTX to NPs.**



**Figure S4. Drug release studies.** *In vitro* drug release of morusin from PLGA-MOR-CTX NPs in pH 4.5, 5.5, 6.8 and 7.4 PBS buffer, 37°C for 96 h time period.



**Figure S5. CLSM images representing whether the excreted MMPs influence the GBM cell targeting capacity of CTX modified nanoparticles.** Nuclei were stained using NucBlue and lysosomes using lysotracker deep red to display nanoparticles co-localization. Scale bars represent 20  $\mu\text{m}$ .



**Figure S6. Magnified images for figure 12. autophagy analysis.** Autophagic flux and autophagosome formation as pointed by arrows in HCN-1A, GI-1 and U87 cells.