Supporting Information

Biotinylated and fluorophore-incorporated polymeric mixed micelles

for tumor cell-specific turn-on fluorescence imaging of Al³⁺

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mPEG-Dye b



The structural characterization of compound 1

¹H NMR (CDCl₃, 400 MHz): δ 11.486 (s, H_a), 9.706 (s, H_b), 7.43 (d, H_c), 6.545 (d, H_d), 6.416 (s, H_e), 4.003 (t, 2 H_f), 1.793 (t, 2 H_g), 1.585(d, 2 H_h), 1.447-1.259 (m, 24H_i), 0.88 (d, 3 H_i).



Figure S2. ¹H NMR spectrum of compound 1 in CDCl₃.

The structural characterization of compound 2

¹H NMR (DMSO-d₆, 400 MHz):δ7.90 (d, 2H_a), 7.054 (d, 2H_b), 4.157 (t, 2H_c), 3.792 (s,

3H_d), 3.738(t, 2H_e), 3.486 -3.421 (m, 176 H_f), 3.247 (s, 3 H_g).



Figure S3. ¹H NMR spectrum of compound 2 in deuterated DMSO.

¹H NMR (CDCl₃, 400 MHz): δ 7.907 (d, 2H_a), 6.866 (d, 2H_b), 4.117 (t, 2H_c), 3.81 (s, 3H_d), 3.746 (t, 2H_e), 3.646 -3.487 (m, 176 H_f), 3.313 (s, 3 H_g).



Figure S4. ¹H NMR spectrum of compound 2 in CDCl₃.

The structural characterization of compound 3

¹H NMR (DMSO-d₆, 400 MHz): δ 9.622 (t, H_a), 7.799 (d, 2H_b), 6.997 (d, 2H_c), 4.157 (d, 2H_d), 4.134 (t, 2H_e), 3.742(t, 2H_f), 3.505 -3.43 (m, 176 H_g), 3.235 (s, 3 H_h), 1.234 (s, H₂O).



Figure S5. ¹H NMR spectrum of compound 3 in deuterated DMSO.

¹H NMR (CDCl₃, 400 MHz): 7.762 (d, 2H_b), 6.942 (d, 2H_c), 4.163 (d, 2H_e), 3.846 (t, 2H_f), 3.689 -3.523 (m, 176 H_g), 3.355 (s, 3 H_h), 1.228 (s, H₂O).



The structural characterization of the polymer mPEG-Dye

Isomer a: ¹H NMR (DMSO-d₆,400 MHz): δ 11.888 (s, H₁), 8.514 (s, H₂), 7.916 (d, 2H₃), 7.398(d, H₄), 7.099(d, 2H₅), 6.517(m, H₆, H₇),4.18(t, 2H₈),3.973(t, 2H₉), 3.771 (t,2H₁₀), 3.582-3.501 (m, 176H₁₁), 3.232 (s, 3H₁₂), 1.693 (m,2H₁₃), 1.40 -1.232 (m, 26H₁₄), 0.845 (t, 3H₁₅)

Isomer b: ¹H NMR (DMSO-d₆,400 MHz): δ11.689 (s, H₁), 8.319 (s, H₂), 7.892(d, 2H₃), 7.369(d, H₄), 7.070(d, 2H₅), 6.468(m, H₆, H₇), 4.18(t, 2H₈), 3.973(t, 2H₉), 3.771 (t, 2H₁₀), 3.582-3.501 (m, 176H₁₁), 3.232 (s, 3H₁₂), 1.693 (m, 2H₁₃), 1.40 -1.232 (m, 26H₁₄), 0.845 (t, 3H₁₅)



Figure S7. ¹H NMR spectrum of polymer mPEG-Dye in deuterated DMSO. ¹H NMR (CDCl₃,400 MHz): δ8.409 (s, H₂), 7.821(d, 2H₃), 7.303(d, H₄), 6.945(d, 2H₅), 6.513(m, H₆, H₇), 4.178(t, 2H₈), 3.937(t, 2H₉), 3.861 (t, 2H₁₀), 3.702-3.537 (m, 176H₁₁), 3.366 (s, 3H₁₂), 1.738 (m, 2H₁₃), 1.429 -1.241 (m, 26H₁₄), 0.862 (t, 3H₁₅)



Figure S8. ¹H NMR spectrum of polymer mPEG-Dye in CDCl₃.



Figure S9. (a) Benesi-Hildebrand analysis of the emission changes for the complexation between mPEG-Dye and Al^{3+} . (b) Linear fluorescence enhancement of mPEG-Dye (20 μ M) response to Al^{3+} concentration (0-10 μ M).



Figure S10. Job's plots for determining the stoichiometry of mPEG-Dye (M) and Al³⁺ in H₂O. The total con centration of mPEG-Dye and Al³⁺ was 20 μ M (λ_{ex} = 408 nm, λ_{em} = 467 nm).



Figure S11. ¹H NMR of mPEG-Dye (10 mM) in DMSO-d₆ upon addition of various amount of Al^{3+} cation.

Table S1. Physicochemical properties of two micelles								
Sample	TEM particle size(nm)	Hydrodynamic particle size(nm)	PDI					
mPEG-Dye	15±3	18±0.05	0.213±0.01					
mPEG-Dye-Biotin	18±3	21±0.04	0.201 ± 0.01					



Figure S12. Fluorescence spectra of mPEG-Dye (20 μ M) alone, mPEG-Dye-Biotin (20 μ M) alone, mPEG-Dye (20 μ M) with Al³⁺ (200 μ M) and mPEG-Dye-Biotin (20 μ M) with Al³⁺ (200 μ M) in H₂O.



Figure S13. (a) Benesi-Hildebrand analysis of the emission changes for the complexation between mPEG-Dye-Biotin and Al^{3+} . (b) Linear fluorescence enhancement of mPEG-Dye-Biotin (20 μ M) response to Al^{3+} concentration (0-10 μ M).



Figure S14. Cytotoxicity of mPEG-Dye-Biotin and mPEG-Dye against HeLa cells. Data are presented as \pm SD (n=4).

Table 52 Comparison of some senili base probes for Al ⁵ detection	Table	e S2 Co	omparison	of some	schiff base	probes	for Al ³⁻	⁺ detection
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Probe	Testing media	Association	Detection	Imaging
		Constant	limit (M)	Al ⁵
		(\mathbf{M}^{*})		in live
Dragant work	A (1120)15	0.5 × 104	2.02×10^{-8}	Vas
(mDEC Due Dietin)	Aqueous	9.3×10^{-5}	$2.02 \times 10^{\circ}$	res
(IIIPEG-Dye-Blouin)	Solution	4.00×104	2.7×10^{-8}	Na
Diarylethene-based	Acetomume	4.90×10^{-1}	2.7×10^{-6}	INO
nuorescentchemosensor (10) ⁴	M. (1	4 72 104	1 24 10 5	NT
Nitrogen neterocycle	Methanol	4.72×10^{4}	1.24×10^{-5}	NO
platformbased fluorescence				
chemosensor(L) ²	CH CN	5 44 104	2 1 10-7	NT
Fluorene-based Schiff-base	CH ₃ CN	5.44×10^{4}	3.1×10^{-7}	No
fluorescent chemosensor(F3) ³		0.4.4 1.02	5 10-7	Ът
Tetra(3-[benzoylhydrazone]-	DMSO: H_2O	9.44×10^{2}	5×10^{-7}	No
methyl-4-hydroxyphenyl) ethene	(9:1, v/v)			
$(IV)^4$		0.66 1.05	1 7	
Oligothiophene-phenylamine-	THF: H_2O	8.66×10^{3}	1.77×10^{-7}	No
based fluorescence sensor (3TP) ⁵	(7:3, v/v)			
2-amino-4,5-imidazoledicarbo-	MeOH: H_2O	2.7×10^{3}	3.44×10^{-6}	No
nitrile and 8-hydroxy-julolidine-9-	(5:15, v/v)			
carboxaldehyde(1) ⁶				
Fluorescence chemosensor based	C ₂ H ₅ OH:H ₂ O	7.03×10^{3}	1.1×10^{-7}	No
on rhodamine and azobenzene	(4:1, v/v)			
moieties(L) ⁷				
Schiff base fluorescence probe	EtOH:H ₂ O	6.74×10^{3}	1.07×10^{-6}	Yes
(STH) ⁸	(9:1, v/v)			
Unsymmetrical azine derivative	DMSO: H_2O :	2.47×10^{4}	1.65×10^{-7}	Yes
(NDEA) ⁹	MeOH			
	(0.1:1.9:8.0,			
	v/v)			
Schiff base-type fluorescent	Aqueous	7.80×10^{4}	0.17×10^{-6}	Yes
chemosensor ¹⁰	solution			

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