Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B.

A new strategy to improve the water solubility of organic fluorescent probe using silicon nanodots and fabricate a two-photon SiNDs-ANPA-N₃ for visualizing hydrogen sulfide in living cells and onion

tissues

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Reagents and Apparatus

All chemicals were of analytical grade and used without further purification. Phorbol-12-myristate-13-acetate (PMA) obtained from Sigma. was Carboxymethoxylamine hemihydrochloride (AOA) and 4-Bromo-1,8-naphthalic anhydride (98%) were purchased from Annaiji (Shanghai, China). Nhydroxysulfosuccinimide sodium salt (Sulfo-NHS) and (3-Aminopropyl) trimethoxysilane (APTMS) (97%) were purchased from Aladdin. 1-(3-Dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC·HCl) was purchased from Shanghai Medpep Co., Ltd. Dulbecco's modified Eagle's medium (DMEM) was purchased from HyClone (Waltham, MA, U.S.A.). Fetal bovine serum (FBS) was taken from Tianhang Biological Technology (Zhejiang, China) and 3-(4,5-dimethyl-2thiazoyl)-2,5-diphenyl tetrazolium bromide (MTT) was purchased from Genview (U.S.A). Physiological buffer solution (PBS) consisted of 8.00 g/L NaCl, 0.20 g/L KCl, 0.20 g/L KH₂PO₄, and 2.78 g/L Na₂HPO₄·12H₂O (pH 7.4). Sodium phosphate buffer solution was prepared by mixing 0.10 M H₃PO₄ solution with 0.10 M Na₃PO₄ solution and adjusted to the required pH values. Water used throughout the experiments was Milli-Q ultrapure water (Millipore= $18.25 \text{ M}\Omega \cdot \text{cm}$).

UV-vis absorption spectra were acquired with an UV-2600 spectrophotometer (Daojin, Tokyo, Japan) and fluorescence spectra were obtained with a RF-6000 spectrophotometer (Shimadzu, Tokyo, Japan). TEM image was obtained with a JEM-2100 (HR) transmission electron microscope by dropping an aqueous SiNDs-ANPA-N₃ fluorescent probe solution and SiNDs solution onto ultrathin carbon support film (300 meshes) with the excess solvent evaporated. FTIR spectra over the range 800-4000 cm⁻¹ were performed on IS10Fourier transform infrared spectrometer. ¹H NMR spectra were recorded on AVANCE III HD at 400 MHz (Bruker, Switzerland). High-resolution mass spectrometry (HR-MS) was performed on a LTQ-Orbitrap Elite (Thermo Fisher Scientific, USA) mass spectrometer. The confocal microscopy fluorescence images were obtained with a 60 × oil (NA = 0.55) objective lens by UltraVIEW VoX confocal microscope (PerkinElmer, America) and NOL-LSM 710 (Carl Zeiss 710, Germany) and analyzed by EZ-C1 software. Onion tissue images were recorded by Olympus inverted fluorescence microscope IX71 equipped with a cooled CCD camera (Olympus DP72).



Fig. S1. The ¹H NMR (400 MHz, DMSO-d6) spectrum of ANPA-N₃. δ 8.44 (d, J = 7.3 Hz, 1H), 8.36 (d, J = 8.0 Hz, 1H), 8.31 (d, J = 8.4 Hz, 1H), 7.79 (t, J = 7.4 Hz, 1H), 7.65 (d, J = 8.0 Hz, 1H), 4.22 (t, J = 7.8 Hz, 2H), 2.58 (t, J = 7.8, 2H).



Fig. S2. The ¹³C NMR (400 MHz, DMSO-d6) spectrum of ANPA-N₃. δ 173.0, 163.5, 163.0, 143.2, 132.0, 131.9, 128.8, 128.6, 127.7, 123.8, 122.4, 118.4, 116.2, 36.2, 32.7.



Fig. S3. The HRMS of ANPA-N₃: Calcd. for C₁₅H₁₀N₄O₄Na (M+Na)⁺ 333.0594, found 333.0579.



Fig. S4. Different amounts of ANPA-N₃ (a, 0.025 mmol; b, 0.05 mmol; c, 0.075 mmol; d, 0.1 mmol; e, 0.2 mmol; f, 0.3 mmol; g, 0.4 mmol; h, 0.5mmol) coupled onto the SiNDs.



Fig. S5. Fluorescence spectra of SiNDS-ANPA-N₃ (0.075 mg/mL) response to 200 μ M NaHS in PBS buffer (pH 7.4). The SiNDS-ANPA-N₃ were prepared by 200 mg SiNDs with different amounts of ANPA-N₃ (a, 0.025 mmol; b, 0.05 mmol; c, 0.075 mmol; d, 0.1 mmol; e, 0.2 mmol; f, 0.3 mmol; g, 0.4 mmol; h, 0.5 mmol). Excitation was at 430 nm; slit widths of excitation and emission, 3 nm/3 nm.

Calculation of the amount of the ANPA-N₃ in the stock solution

SiNDs and ANPA-N₃ produce the characteristic absorption band at 337 nm (Fig. S6B) and 360 nm (Fig. S6A), respectively. Since the SiNDs-ANPA-N₃ exhibits characteristic absorption band at 360 nm which overlaps with the absorption of SiNDs; while upon reacting with NaHS, SiNDs-ANPA-N₃ turns into SiNDs-ANPA-NH₂ and displays characteristic absorption at 430 nm which will not overlap with the absorption of SiNDs (Fig. S6B); hence to more accurately determine the amount of the amount of the ANPA-N₃ in the stock solution, the SiNDs-ANPA-N₃ solution and ANPA-N₃ solution were added excessive amount of NaHS, so as to transform the SiNDs-ANPA-N₃ into SiNDs-ANPA-N₃ and ANPA-N₃ into ANPA-NH₂.¹



Fig. S6. (A) Absorption spectra of ANPA-N₃ (red) and ANPA-N₃ solution was added excessive amount of NaHS then transformed to ANPA-NH₂ (blue) in PBS buffer (pH 7.4). (B) Absorption spectra of SiNDs (green), SiNDs-ANPA-N₃ (black) and SiNDs-ANPA-N₃ solution was added excessive amount of NaHS then transformed to SiNDs-

ANPA-NH₂ (Red) in PBS buffer (pH 7.4). (C) Standard calibration curve of known concentrations of ANPA-NH₂ [y = 0.0102x + 0.0007, R² = 0.9995]. (D) Standard calibration curve of known concentrations of ANPA-N₃ [y = 0.0079x + 0.0132, R² = 0.9994].

References

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Fig. S7. (A) TEM images of label-free SiNDs. (B) The TEM diameter distribution.



Fig. S8. Two-photon excited fluorescence spectrum of SiNDs-ANPA-N₃ (0.075 mg/mL) towards 200 μ M NaHS in PBS buffer (pH 7.4). Excitation was at 853 nm; slit widths of excitation and emission, 3 nm/3 nm.



Fig. S9. Time-dependent fluorescence intensity changes of SiNDs-ANPA-N₃ (0.075 mg/mL) in PBS buffer (pH 7.4) in the presence of 50 μ M, 100 μ M, 200 μ M NaHS. Excitation was at 430 nm; slit widths of excitation and emission, 3 nm/3 nm.



Fig. S10. One-photon confocal microscopy images of HeLa cells incubated with SiNDs-ANPA-N₃ (3 mg/mL) upon 405 nm excitation for different times (A, 0.5 h; B, 1.0 h; C, 1.5 h; D, 2.0 h). Then treatment amount of 100 μ M NaHS. E, The average fluorescence intensity of the above images. Scale bar: 20 μ m.



Fig. S11. One-photon confocal microscopy images of HeLa cells incubated with SiNDs-ANPA-N₃ (3 mg/mL) upon 405 nm excitation in the absence or presence of stimulant or scavenger. A-C, the cells incubated with only SiNDs-ANPA-N₃ for 1 h; D-F, the cells incubated with PMA (2 μ L, 1 μ g/mL) for 1 h then treated with SiNDs-ANPA-N₃ for 1 h; G-I, cells incubated with Cys (400 μ M) for 1 h and then treated with SiNDs-ANPA-N₃ for 1 h. Scale bar: 20 μ m.



Fig. S12. One-photon confocal microscopy images of MCF-7 cells incubated with SiNDs-ANPA-N₃ (3 mg/mL) upon 405 nm excitation in the absence or presence of stimulants and scavengers. A-C, the cells incubated with only SiNDs-ANPA-N₃ for 1 h; D-F, the cells incubated with PMA (2 μ L, 1 μ g/mL) for 1 h then treated with SiNDs-ANPA-N₃ for 1 h; G-I, the cells incubated with Cys (400 μ M) for 1 h and then treated with SiNDs-ANPA-N₃ for 1 h. Scale bar: 20 μ m.



Fig. S13. Two-photon confocal microscopy images of HL-7702 cells incubated with SiNDs-ANPA-N₃ (3 mg/mL) upon 853 nm excitation in the absence or presence of stimulants and scavengers. A-C, the cells incubated with only SiNDs-ANPA-N₃ for 1h; D-F, the cells incubated with PMA (2 μ L, 1 μ g/mL) for 1 h then treated with SiNDs-ANPA-N₃ for 1 h; G-I, the cells incubated with Cys (400 μ M) for 1 h and then treated with SiNDs-ANPA-N₃ for 1 h. Scale bar: 20 μ m