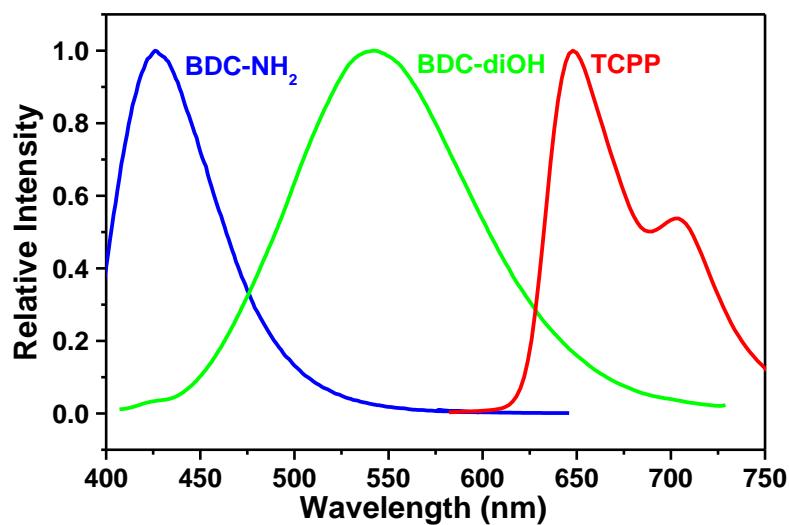


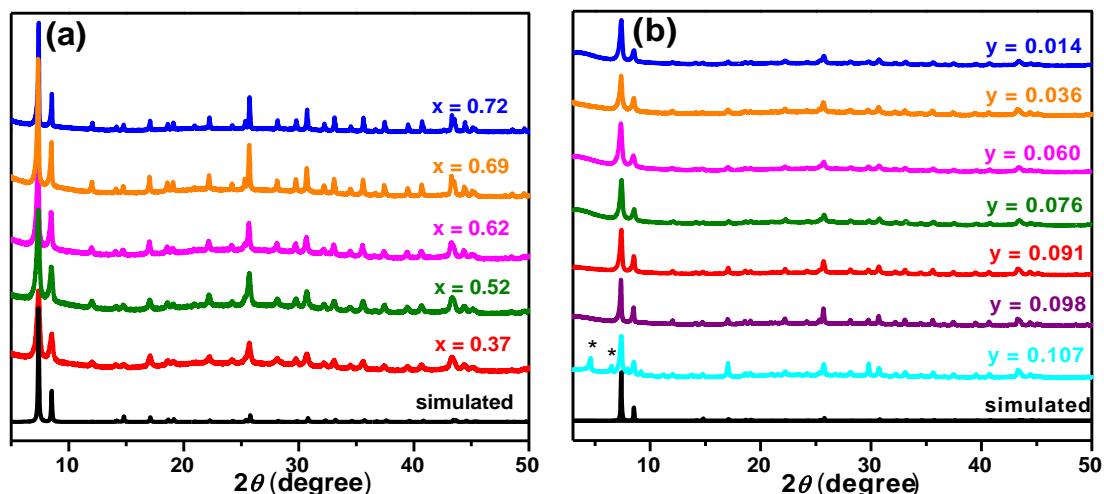
## **Electronic Supplementary Information**

### **White-light-emitting single MOF sensor based array for berberine homologues discrimination**

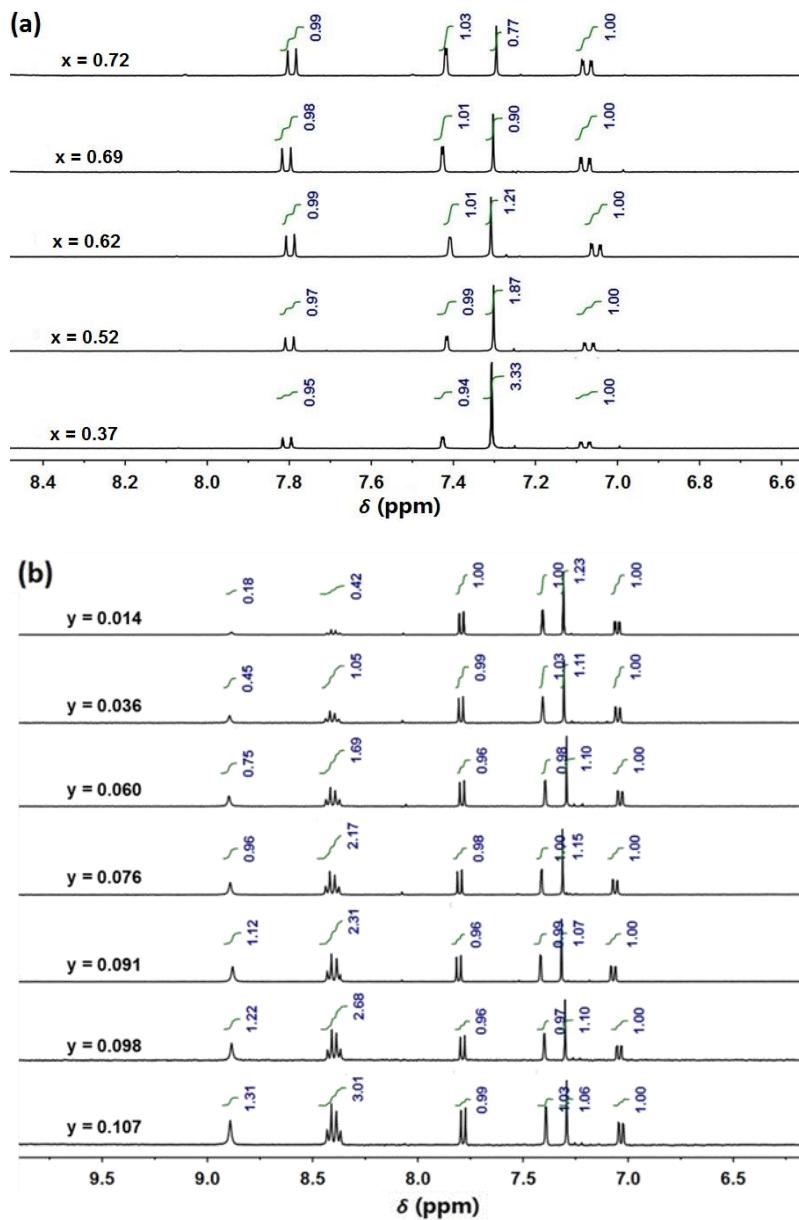
Qian Wang, Qiao Liu, Xiao-Meng Du, Bo Zhao, Yue Li\* and Wen-Juan Ruan\*



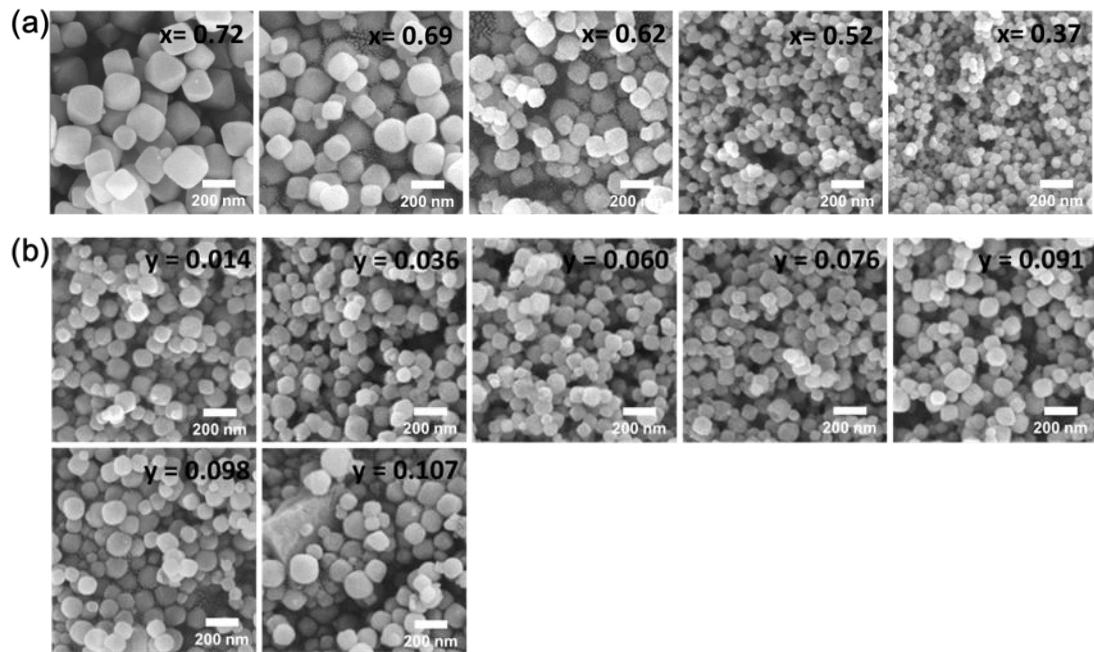
**Fig. S1** Fluorescent spectra of free  $\text{BDC-NH}_2$ ,  $\text{BDC-diOH}$  and  $\text{TCPP}$  ligands,  $\lambda_{\text{em}} = 388 \text{ nm}$ .



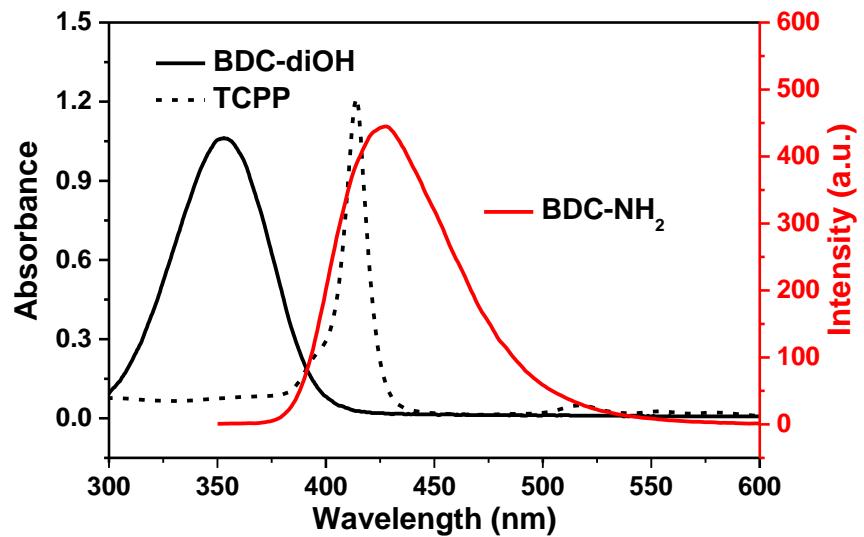
**Fig. S2** PXRD patterns of (a)  $\text{UiO-66-(NH}_2\text{)}_x(\text{diOH})_{1-x}$  and (b)  $\text{TCPP}_y \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  samples. The diffraction peaks of impurity phase are marked by “\*”.



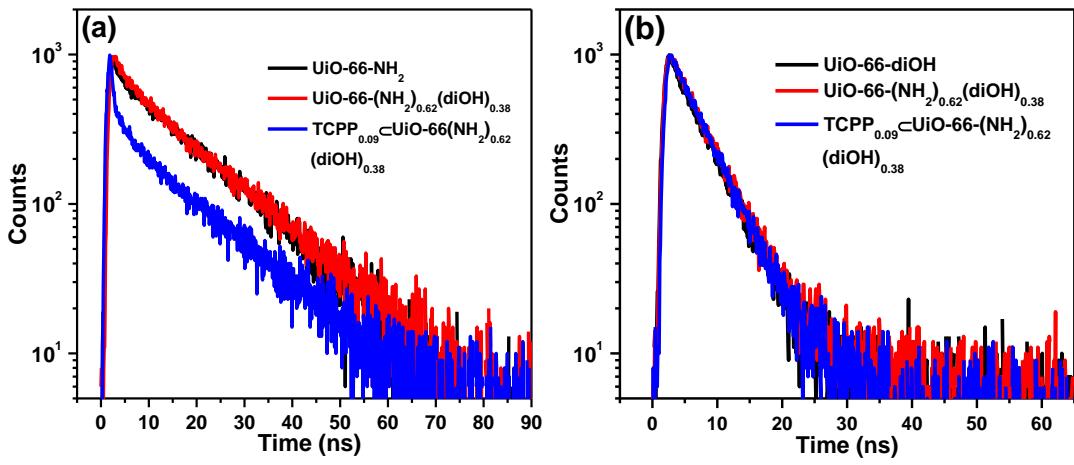
**Fig. S3**  $^1\text{H}$  NMR spectra of digested solutions of (a)  $\text{UiO-66-(NH}_2\text{)}_x(\text{diOH})_{1-x}$  and (b)  $\text{TCPP}_y \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  with dilute HF in  $\text{DMSO-d}_6$ . BDC- $\text{NH}_2$   $\delta$ : 7.08 (d,  $J = 8.0$  Hz, 1H), 7.42 (s, 1H), 7.78 (d,  $J = 7.6$  Hz, 1H). BDC-diOH  $\delta$ : 7.30 (s, 2H).  $\delta$ : 8.40 (q, 16H), 8.90 (s, 8H).



**Fig. S4** SEM images of (a)  $\text{UiO-66-(NH}_2\text{)}_x(\text{diOH})_{1-x}$  and (b)  $\text{TCPP}_y \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  samples.

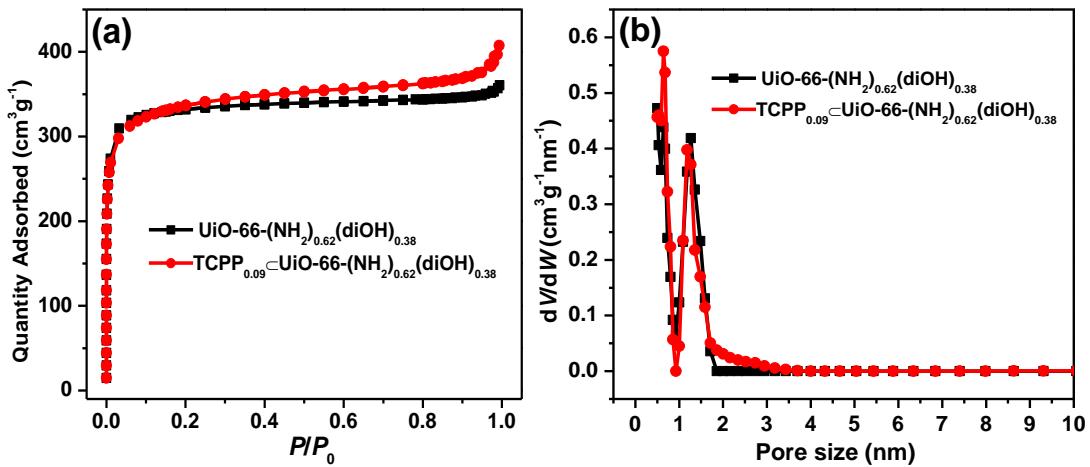


**Fig. S5** Comparison of the emission spectrum of  $\text{BDC-NH}_2$  with the absorption spectra of  $\text{BDC-diOH}$  and  $\text{TCPP}$ .



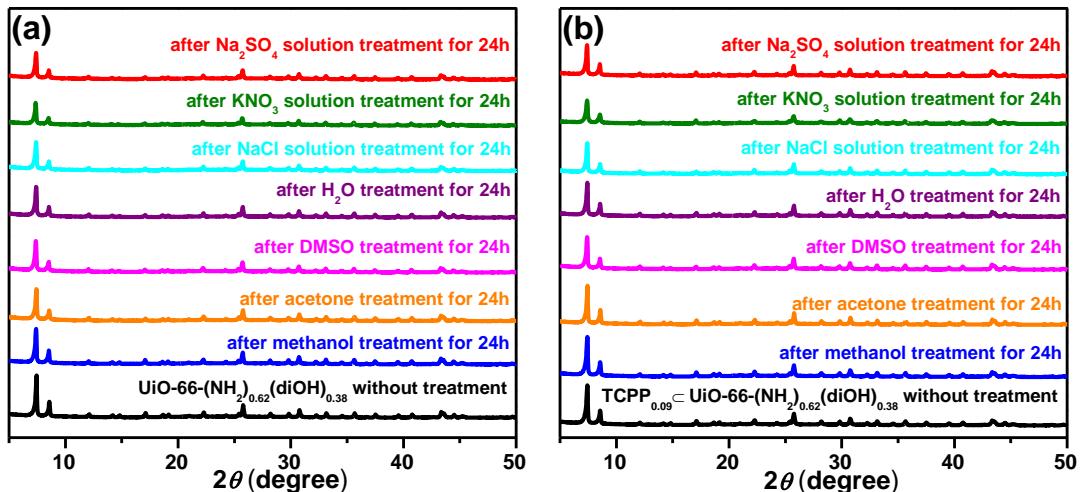
**Fig. S6** Time-resolved decays of the emission bands at (a) 440 and (b) 536 nm.

The emission band at 440 nm comes from the fluorescence of BDC-NH<sub>2</sub> ligand. The decays of this band in UiO-66-NH<sub>2</sub> and UiO-66-(NH<sub>2</sub>)<sub>0.62</sub>(diOH)<sub>0.38</sub> could be satisfactorily fitted to single-exponential kinetics with the lifetimes of 15.6 and 15.7 ns, respectively. After the incorporation of TCPP, this band presented a biexponential decay with a major term with longer lifetime ( $\tau_1 = 14.9$  ns, 90.5%) and a minor subnanosecond term ( $\tau_2 = 0.8$  ns, 9.5%). The average lifetime of these two terms was 13.6 ns. The emission band at 536 nm comes from BDC-diOH. The decay kinetics of this component was single-exponential and gave nearly the same lifetime in UiO-66-diOH (4.7 ns), UiO-66-(NH<sub>2</sub>)<sub>0.62</sub>(diOH)<sub>0.38</sub> (4.7 ns) and TCPP<sub>0.09</sub><UiO-66-(NH<sub>2</sub>)<sub>0.62</sub>(diOH)<sub>0.38</sub> (4.8 ns).

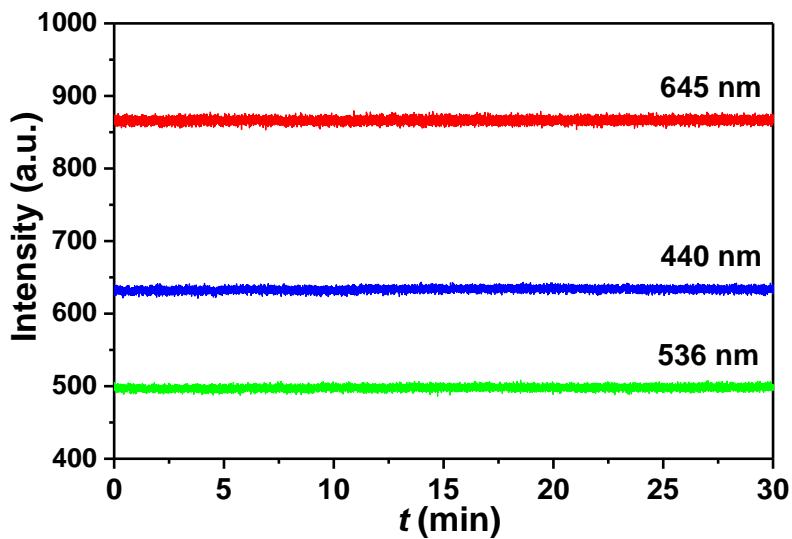


**Fig. S7** (a)  $\text{N}_2$  adsorption isotherms (at 77 K) and (b) BJH pore-size distribution curves of  $\text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  and  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$ .

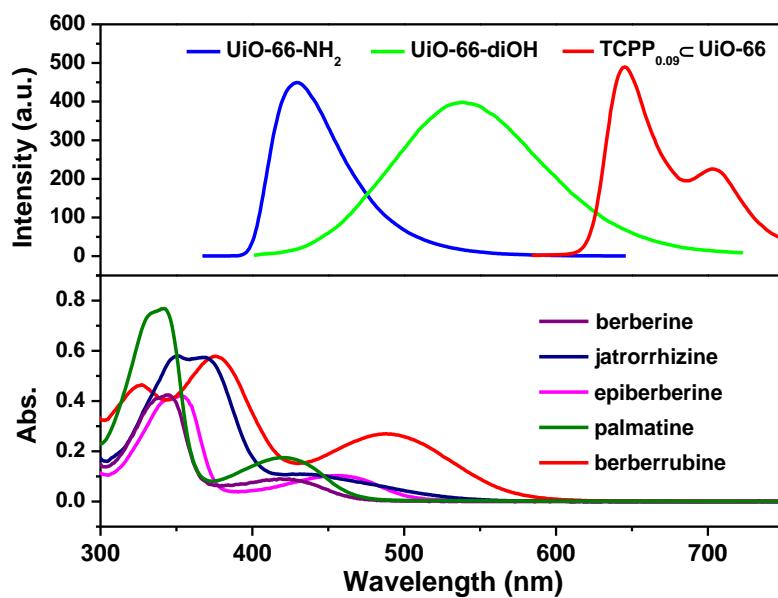
With above isotherms, the BET surface areas of  $\text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  and  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  were calculated to be  $1084$  and  $1045 \text{ m}^2 \text{ g}^{-1}$ , respectively, and their pore volumes were  $0.409$  and  $0.414 \text{ cm}^3 \text{ g}^{-1}$ . The apertures of these two samples are in the range of  $0.5\text{--}2$  nm, so their pores belong to micropore.



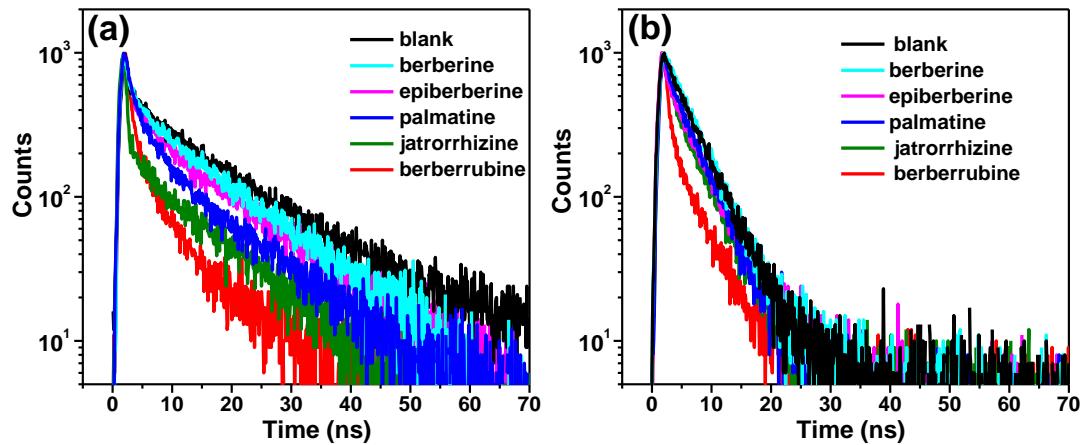
**Fig. S8** PXRD patterns of the  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  samples immersed in different organic solvents and saturated salt solutions.



**Fig. S9** Temporal change of the fluorescent intensities at 440, 536 and 645 nm with prolonging measurement of  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  suspension,  $\lambda_{\text{ex}} = 388 \text{ nm}$ .

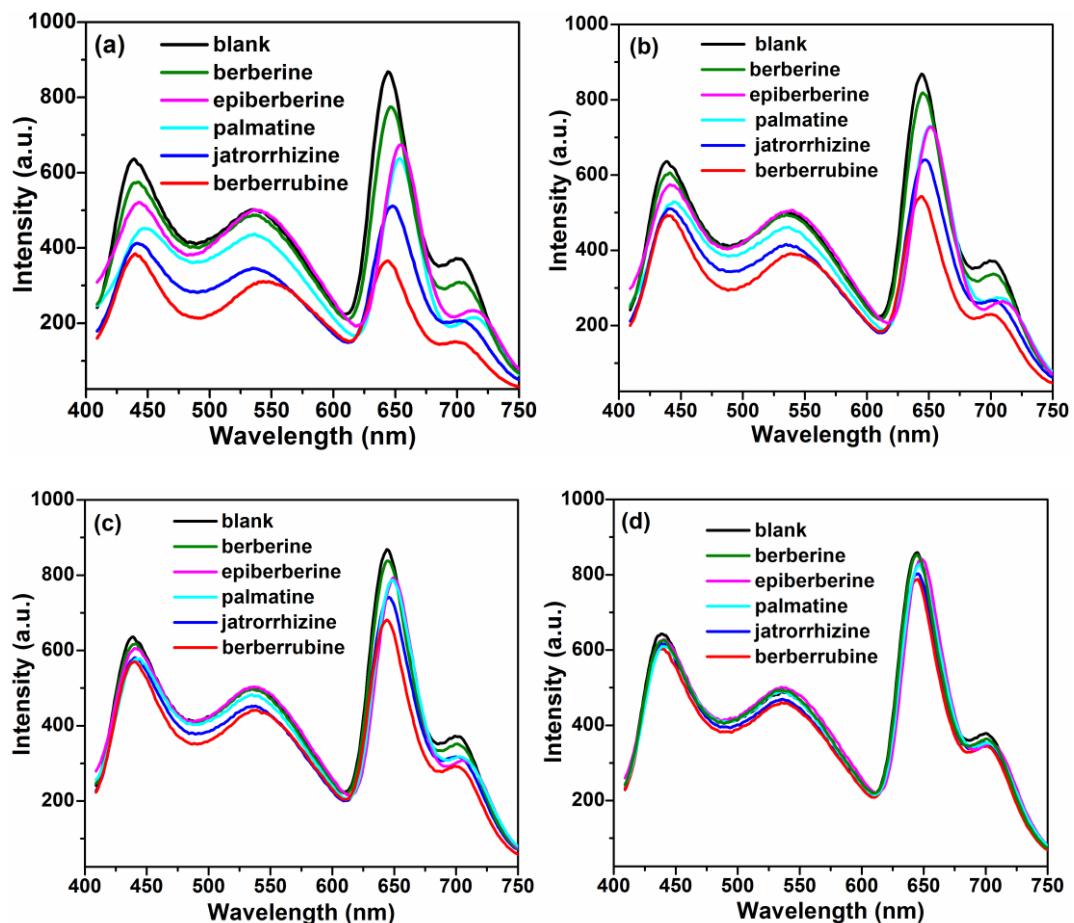


**Fig. S10** Comparison of the emission spectra of luminescent ligands in MOF backbones with the absorption spectra of berberine homologues.

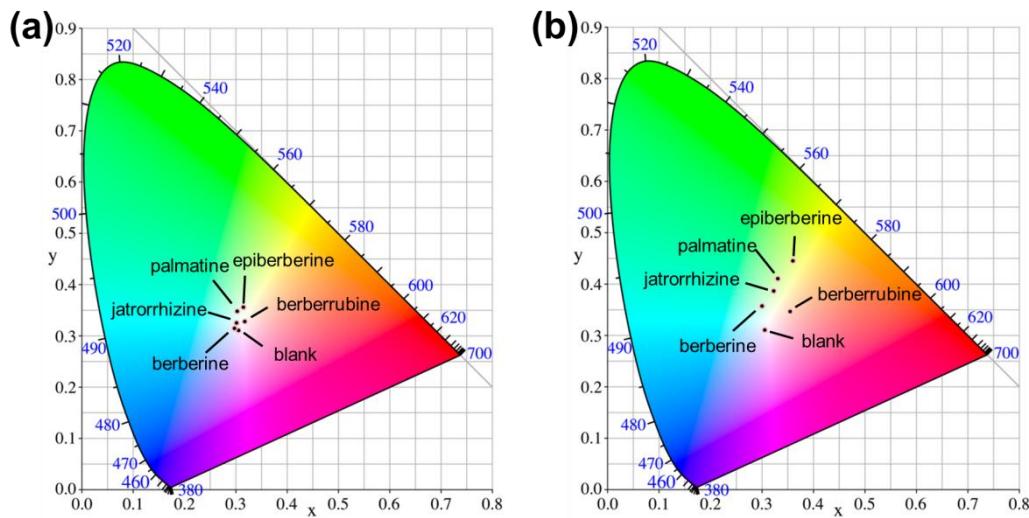


**Fig. S11** Time-resolved decays of the emission bands at (a) 440 and (b) 536 nm upon the exposure of  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  to berberine homologues.

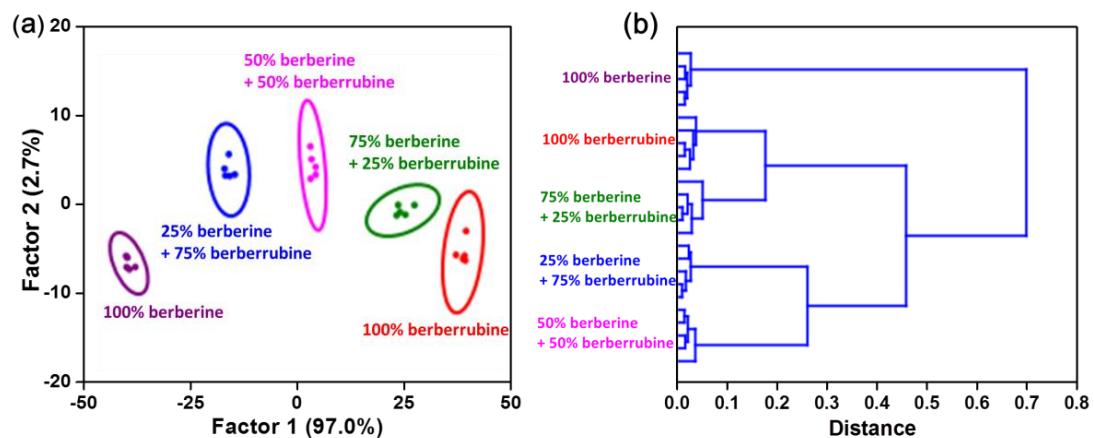
After the addition of berberine, epiberberine, palmatine, jatrorrhizine and berberrubine, the lifetime at 440 nm was shortened from the original value of 13.76 ns to 13.32, 12.66, 11.65, 10.10 and 7.28 ns, respectively. The lifetime of the emission band at 536 nm was nearly unchanged from the original value of 4.84 ns to 4.92, 4.72, 4.44, 4.26 and 3.58 ns, respectively.



**Fig. S12** Fluorescent spectra of  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  ( $50 \text{ mg L}^{-1}$ ) in the presence of berberine homologues with the concentrations of (a) 20, (b) 10, (c) 5 and (d) 2  $\mu\text{M}$ .



**Fig. S13** CIE chromaticity diagrams of  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  in the presence of (a) 50 and (b) 200  $\mu\text{M}$  berberine homologues.



**Fig. S14** (a) LDA and (b) HCA plots for  $\text{TCPP}_{0.09} \subset \text{UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  against the mixtures of berberine and berberrubine,  $C_{\text{berberine}} + C_{\text{berberrubine}} = 50 \mu\text{M}$ .

**Table S1** Compositions and CIE coordinates of synthesized MOF samples

Addition amount in synthesis (mmol)			Molar ratio of BDC-NH <sub>2</sub> : BDC-diOH : TCPP	CIE coordinates
H <sub>2</sub> BDC-NH <sub>2</sub>	H <sub>2</sub> BDC-diOH	H <sub>4</sub> TCPP	in MOFs	
0.150	0	0	1 : 0 : 0	(0.1540, 0.0564)
0.125	0.025	0	0.72 : 0.28 : 0	(0.1998, 0.1792)
0.120	0.030	0	0.69 : 0.31 : 0	(0.2187, 0.2211)
0.112	0.038	0	0.62 : 0.38 : 0	(0.2470, 0.2836)
0.100	0.050	0	0.52 : 0.48 : 0	(0.2850, 0.3662)
0.075	0.075	0	0.37 : 0.63 : 0	(0.3041, 0.4164)
0	0.150	0	0 : 1 : 0	(0.3492, 0.5197)
0.112	0.038	0.003	0.62 : 0.38 : 0.014	(0.2595, 0.2965)
0.112	0.038	0.006	0.64 : 0.36 : 0.036	(0.2686, 0.3019)
0.112	0.038	0.009	0.65 : 0.35 : 0.060	(0.2754, 0.3033)
0.112	0.038	0.012	0.63 : 0.37 : 0.076	(0.2858, 0.3052)
0.112	0.038	0.015	0.65 : 0.35 : 0.091	(0.3012, 0.3063)
0.112	0.038	0.018	0.65 : 0.35 : 0.098	(0.3349, 0.3135)
0.112	0.038	0.021	0.65 : 0.35 : 0.107	—

**Table S2** Spectral overlap integrals of the emission of UiO-66-NH<sub>2</sub> with the absorptions of berberine homologues

berberine	epiberberine	palmatine	jatrorrhizine	berberrubine
<i>J</i> <sup>a</sup> 3.74×10 <sup>13</sup>	5.67×10 <sup>13</sup>	7.28×10 <sup>13</sup>	7.41×10 <sup>13</sup>	1.65×10 <sup>14</sup>

<sup>a</sup> The spectral overlap integral (*J*) is calculated with the equation of<sup>1</sup>

$$J = \int \bar{f}_D(\lambda) \varepsilon_A(\lambda) \lambda^4 d\lambda$$

where  $\bar{f}_D$  is the donor (UiO-66-NH<sub>2</sub>) emission spectrum normalized to an area of 1, and  $\varepsilon_A$  is the acceptor (berberine homologue) molar extinction coefficient obtained from the absorption spectrum.

**Table S3** Training matrix of the fluorescent response ( $1 - I/I_0$ ) patterns of  $\text{TCPP}_{0.09}\text{-UiO-66-(NH}_2\text{)}_{0.62}(\text{diOH})_{0.38}$  ( $50 \text{ mg L}^{-1}$ ) against alkaloids ( $50 \mu\text{M}$ )

Analyte	440 nm	536 nm	645 nm
berberine	0.173	0.052	0.202
	0.185	0.068	0.219
	0.175	0.013	0.212
	0.168	0.062	0.213
	0.165	0.053	0.219
jatrorrhizine	0.608	0.546	0.657
	0.611	0.540	0.661
	0.598	0.541	0.653
	0.619	0.549	0.669
	0.602	0.532	0.660
epiberberine	0.331	-0.015	0.502
	0.344	-0.010	0.527
	0.330	-0.007	0.514
	0.320	-0.009	0.524
	0.339	-0.013	0.526
palmatine	0.528	0.244	0.533
	0.513	0.243	0.525
	0.509	0.259	0.532
	0.520	0.256	0.550
	0.533	0.276	0.509
berberrubine	0.681	0.662	0.808
	0.660	0.673	0.815
	0.672	0.672	0.837
	0.694	0.661	0.798
	0.671	0.670	0.818
arecoline	0.019	0.017	0.019
	0.018	0.009	0.020
	0.028	0.018	0.016
	0.025	0.015	0.026
	0.020	0.011	0.019
theophylline	0.021	0.018	0.020
	0.022	0.015	0.028
	0.022	0.024	0.017
	0.026	0.014	0.018
	0.025	0.021	0.017
reserpine	0.009	0.014	0.012
	0.011	0.014	0.021
	0.010	0.017	0.015
	0.013	0.011	0.019
	0.008	0.011	0.010
quinine	0.035	0.031	0.041

0.031	0.032	0.054
0.035	0.028	0.039
0.034	0.029	0.045
0.032	0.034	0.048

**Table S4** Jackknifed classification matrix of LDA in the differentiation of berberine homologues

Actual	Predicted					Total
	berberine	jatrorrhizine	epiberberine	palmatine	berberrubine	
berberine	5	0	0	0	0	5
jatrorrhizine	0	5	0	0	0	5
epiberberine	0	0	5	0	0	5
palmatine	0	0	0	5	0	5
berberrubine	0	0	0	0	5	5
Total	5	5	5	5	5	25

**Table S5** Identification of 15 berberine homologue samples with unknown concentrations

Sample	Actual	Concentration (mM)		Fluorescent response ( $1 - I/I_0$ )			Predicted
		Added	Measured	440 nm	536 nm	645 nm	
1	berberine	0.80	0.73	0.165	0.052	0.184	berberine
2	jatrorrhizine	0.80	0.76	0.605	0.528	0.657	jatrorrhizine
3	epiberberine	0.80	0.72	0.301	-0.012	0.525	epiberberine
4	palmatine	0.80	0.84	0.491	0.232	0.557	palmatine
5	berberrubine	0.80	0.75	0.646	0.623	0.835	berberrubine
6	berberine	1.00	0.93	0.163	0.055	0.202	berberine
7	jatrorrhizine	1.00	0.98	0.646	0.512	0.695	jatrorrhizine
8	epiberberine	1.00	0.87	0.295	-0.013	0.498	epiberberine
9	palmatine	1.00	0.94	0.524	0.216	0.525	palmatine
10	berberrubine	1.00	1.06	0.683	0.649	0.854	berberrubine
11	berberine	1.20	1.10	0.152	0.059	0.214	berberine
12	jatrorrhizine	1.20	1.29	0.605	0.489	0.667	jatrorrhizine
13	epiberberine	1.20	1.08	0.293	-0.017	0.512	epiberberine
14	palmatine	1.20	1.32	0.564	0.287	0.608	palmatine
15	berberrubine	1.20	1.14	0.625	0.616	0.785	berberrubine

## **References**

1. L. Yuan, W. Lin, K. Zheng and S. Zhu, *Acc. Chem. Res.*, 2013, **46**, 1462–1473.