

Enantioselective Allylation of Aldehydes Catalyzed by Chiral Indium(III) Complexes Immobilized in Ionic Liquid

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Supporting Information

General

Experiments involving moisture and/or air sensitive components were performed in oven-dried glassware. Commercial solvents and reagents were used without further purification with the following exceptions: Hexane, dichloromethane, ethyl acetate were fractionally distilled. Aldehydes were distilled before used. Azeotropic drying of starting materials or reagents was performed by the addition of the stated amount of anhydrous tetrahydrofuran, ensued by azeotropic removal of tetrahydrofuran with traces of moisture in vacuo followed by subsequent purging with nitrogen.

Analytical thin layer chromatography (TLC) was performed using Merck 60 F₂₅₄ precoated silica gel plate (0.2 mm thickness). Subsequent to elution, plates were visualized using UV radiation (254 nm) on Spectroline Model ENF-24061/F 254 nm. Further visualization was possible by staining with basic solution of potassium permanganate or acidic solution of ceric molybdate, followed by heating on a hot plate. Flash chromatography was performed using Merck silica gel 60 with freshly distilled solvents. Columns were typically packed as slurry and equilibrated with the appropriate solvent system prior to use.

Infrared spectra were recorded on a Bio-Rad FTS 165 FTIR spectrometer. Liquid samples were examined as film between NaCl salt plates.

Proton nuclear magnetic resonance spectra (¹H NMR) were recorded on a Bruker Avance DPX 300 and Bruker AMX 500 spectrophotometer (CDCl₃ as solvent). Chemical shifts for ¹H NMR spectra are reported as δ in units of parts per million (ppm) downfield from SiMe₄ (δ 0.0) and relative to the signal of chloroform-d (δ 7.2600, singlet). Multiplicities were given as: s (singlet); d (doublet); t (triplet); q (quartet); dd (doublets of doublet); ddd (doublets of doublets of doublet); dddd (doublets of doublets of doublets of doublet); dt (doublets of triplet); or m (multiplets). The number of protons (n) for a given resonance is indicated by nH. Coupling constants are reported as a J value in Hz. Carbon nuclear magnetic resonance spectra (¹³C NMR) are reported as δ in units of parts per million (ppm) downfield from SiMe₄ (δ 0.0) and relative to the signal of chloroform-d (δ 77.03, triplet). The proportion of diastereomers and geometric isomers was determined from the integration of ¹H NMR and ¹³C NMR spectra.

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Mass spectral analyses were carried out on a VG 7035 micromass mass spectrophotometer at a source temperature of 200 °C and at an ion current of 70 eV. Mass spectral data were reported in units of mass to charge (m/z) and % intensity.

Experimental Section

Representative procedure for asymmetric allylation of aldehydes: Preparation of (R)-1-phenylbut-3-en-1-ol

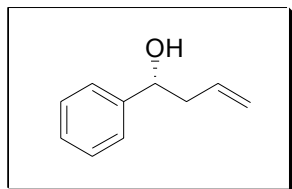
To an oven dried 5mL round-bottom flask equipped with a magnetic stirring bar was added In(OTf)₃ (16.9 mg, 0.03 mmol, 0.2 equiv.) and 4Å molecular sieve (120 mg). The solid was azeotropically dried with anhydrous tetrahydrofuran twice (2 mL x 2) prior to the addition of [hmim]PF₆ (0.5 mL) and dichloromethane (0.5 mL). PYBOX (**1**) (13 mg, 0.033 mmol, 0.22 equiv.) was added and the mixture was stirred under nitrogen at room temperature for 2 hours to afford a white suspension. A mixture of benzaldehyde (15 µL, 0.15 mmol, 1 equiv.) and TMSCl (23 µL, 0.18 mmol, 1.2 equiv.) in dichloromethane (0.2 mL) was added to the resulting suspension and stirred for 10 minutes. The mixture was then cooled to -60 °C for 15 minutes followed by addition of allyltributylstannane (57 µL, 0.18 mmol, 1.2 equiv.). The reaction mixture was stirred at -60 °C for 30 hours, then dichloromethane was removed *in vacuo*; the reaction mixture was extracted with dry hexane (5x5 mL). The combined hexane was treated with saturated sodium bicarbonate solution at room temperature for 30 min., washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residual crude product was purified via silica gel chromatography to afford the homoallylic alcohol as colorless oil (72% yield and 88% ee).

Recycling of chiral catalytic system for preparation of (R)-1-phenylbut-3-en-1-ol

After washing the reaction mixture with dry hexane (5x5 mL), the catalytic system was azeotropically dried with anhydrous tetrahydrofuran twice (2 mL x 2). A second run was performed under identical reaction conditions and resulted in the formation of product in 76% yield and 87% ee.

Characterization of tertiary homoallylic alcohols in Table 2

(*R*)-1-phenylbut-3-en-1-ol



Selectivity: 88 % ee

R_f = 0.38 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 7.37-7.27 (m, 5H), 5.89-5.75 (m, 1H), 5.20-5.13 (m, 2H), 4.75 (t, J = 5.6 Hz, 1H), 2.54-2.49 (m, 2H), 2.20 (br, 1H)

^{13}C NMR (75.4 MHz, CDCl_3): δ 143.9, 134.5, 128.4, 127.6, 125.8, 118.4, 73.3, 43.8

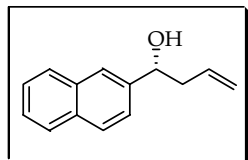
FTIR (neat): 3468, 2932, 1707, 1642, 1494, 1452, 1051, 999, 916, 758, 701 cm^{-1} .

HRMS Calcd for $\text{C}_{10}\text{H}_{12}\text{O}$ [M^+]: 148.0888. Found: 148.0899.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiracel OD column (Hexane: *i*-propanol 98:2, 1.0 mL/min: t_1 = 10.35 min for *R* enantiomer, t_2 = 13.86 min for *S* enantiomer).

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(*R*)-1-Naphthalen-2-yl-but-3-en-1-ol



Selectivity: 91 % ee

R_f = 0.40 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 7.81-7.85 (m, 4H), 7.45-7.50 (m, 3H), 5.77-5.91 (m, 1H), 5.13-5.22 (m, 2H), 4.91 (t, J = 6.4 Hz, 1H), 2.53-2.68 (m, 2H), 2.14 (br, 1H)

^{13}C NMR (75.4 MHz, CDCl_3): δ 141.2, 134.3, 133.2, 132.9, 128.1, 127.9, 127.6, 126.0, 125.7, 124.4, 123.9, 118.4, 73.3, 43.6

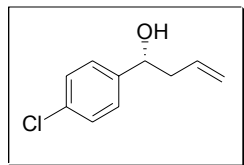
FTIR (neat): 3380cm^{-1} .

HRMS Calcd for $\text{C}_{14}\text{H}_{14}\text{O}$ [M^+]: 198.1047. Found: 198.1054.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiracel OD-H column (Hexane: i-propanol 98:2, 1 mL/min: t_1 = 40.11min for *S* enantiomer, t_2 = 46.68min for *R* enantiomer).

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(*R*)-1-(4-chlorophenyl)but-3-en-1-ol



Selectivity: 94 % ee

R_f = 0.5 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 7.31 (d, J = 2.7Hz, 2H), 7.12 (d, J = 2.8Hz, 2H), 5.84-5.70 (m, 1H), 5.19-5.11 (m, 2H), 4.70-4.65 (m, 1H), 2.55-2.37 (m, 2H), 2.11(br, 1H).

^{13}C NMR (75.4 MHz, CDCl_3): δ 146.88, 134.21, 133.84, 129.58, 127.51, 125.94, 123.88, 118.74, 72.45, 43.64.

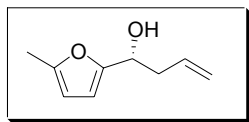
FTIR (neat): 3367, 2905, 1573, 1432, 1196 cm^{-1} .

HRMS Calcd for $\text{C}_{10}\text{H}_{11}\text{ClO}$ [M^+]: 182.0498. Found: 182.0502.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiracel ASH column (Hexane: *i*-propanol 98:2, 1 mL/min: t_1 = 8.72min for the *R* enantiomer, t_2 = 9.34 min for the *S* enantiomer).

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(*R*)-1-(5-methyl-furan-2-yl)-but-3-en-1-ol



Selectivity: 92 % ee

R_f = 0.5 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 6.10(d, J = 3.2Hz, 1H), 5.90-5.75(m, 2H), 5.22-5.11(m, 1H), 4.67(q, J = 6.4Hz, 1H), 2.63-2.58(m, 2H), 2.28(d, J = 0.8Hz, 3H), 2.01(d, J = 5.3Hz, 1H).

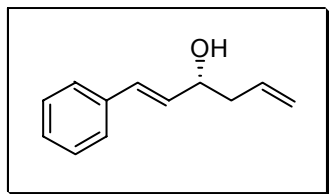
^{13}C NMR (75.4 MHz, CDCl_3): δ 154.11, 151.62, 133.92, 118.19, 106.78, 105.94, 66.79, 39.90, 13.38.

FTIR (neat): 3388, 2921, 1580, 1438 cm^{-1} .

HRMS Calcd for $\text{C}_9\text{H}_{12}\text{O}_2$ [M^+]: 152.0837. Found: 152.0839.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiracel OJ column (Hexane: i-propanol 95:5, 1 mL/min: t_1 =5.95min for the *R* enantiomer, t_2 =7.93 min for the *S* enantiomer.

(R)-1-phenylhexa-1,5-dien-3-ol



Selectivity: 91 % ee

R_f = 0.40 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 7.21-7.40 (m, 5H), 6.60 (d, J = 15.9 Hz, 1H), 6.25 (dd, J = 15.9, 6.3 Hz, 1H), 5.79-5.93 (m, 1H), 5.15-5.21 (m, 2H), 4.35-4.37 (m, 1H), 2.33-2.48 (m, 2H), 1.80 (br, 1H)

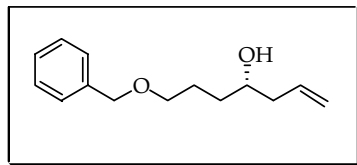
^{13}C NMR (75.4 MHz, CDCl_3): δ 136.6, 134.0, 131.5, 130.3, 128.5, 127.6, 126.4, 118.4, 71.6, 42.0.

FTIR (neat): 3414 cm^{-1} .

HRMS Calcd for $\text{C}_{12}\text{H}_{14}\text{O}$ [M^+]: 174.1045. Found: 174.1040.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiralcel OD column (Hexane: i-propanol 98:2, 1.0 mL/min: t_1 = 14.69 min for *R* enantiomer, t_2 = 26.58 min for *S* enantiomer).

(S)-7-(benzyloxy)hept-1-en-4-ol



Selectivity: 92 % ee

R_f = 0.38 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 7.39-7.28 (m, 5H), 5.90-5.77 (m, 1H), 5.17-5.09 (m, 2H), 4.52 (s, 2H), 3.71-3.61 (m, 1H), 3.52 (t, J = 5.9 Hz, 2H), 2.32-2.13 (m, 2H), 1.78-1.63 (m, 2H), 1.53-1.41 (m, 2H)

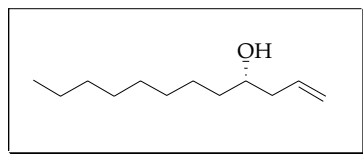
^{13}C NMR (75.4 MHz, CDCl_3): δ 138.2, 135.0, 128.4, 127.7, 127.6, 117.7, 73.0, 70.6, 70.4, 42.0, 34.0, 26.2

FTIR (neat): 3451, 2928, 2862, 1641, 1452, 1097, 1026, 998, 915, 740, 699 cm^{-1} .

HRMS Calcd for $\text{C}_{14}\text{H}_{20}\text{O}_2$ [M^+]: 220.1463. Found: 220.1465.

The enantiomeric excess was determined by HPLC analysis employing a Daicel Chiracel OB column (Hexane: i-propanol 99:1, 1.0 mL/min: t_1 = 22.24 min for the *R* enantiomer, t_2 = 28.95 min for the *S* enantiomer).

(S)-dodec-1-en-4-ol



Selectivity: 86 % ee

R_f = 0.53 (4:1 hexane/ethyl acetate)

^1H NMR (300 MHz, CDCl_3): δ 5.90-5.76 (m, 1H), 5.17-5.11 (m, 2H), 3.64 (m, 1H), 2.36 (m, 2H), 1.48-1.43 (m, 2H), 1.33-1.25 (m, 12H), 0.88 (t, J = 6.3 Hz, 3H)

^{13}C NMR (75.4 MHz, CDCl_3): δ 134.9, 118.0, 70.7, 41.9, 36.8, 31.9, 29.7, 29.3, 25.7, 22.7, 14.1.

FTIR (neat): 3557, 2924, 2855, 1642, 1464, 995, 913 cm^{-1} .

HRMS Calcd for $\text{C}_{12}\text{H}_{24}\text{O}$ [M^+]: 184.1827. Found: 184.1830.

Chiral resolution using R-(+)- α -trifluoromethyl- α -methoxy-phenylacetic acid (Mosher acid). The enantiomeric excess was found to be 86 % by 500 MHz ^1H NMR analysis of its Mosher derivative at δ 2.40 for the *R* enantiomer and 2.33 for the *S* enantiomer.