Electronic Supplementary Information

Ligand Amplification in a Dynamic Combinatorial Glycopeptide Library

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S1. Mass Spectrometry

Electrospray ionization-mass spectrometric (ESI-MS) analysis of DCLs was conducted on a Micromass LCT time-of-flight mass spectrometer coupled to a Waters Alliance 2790 HPLC. Samples were loop injected into the MS using the LC autosampler. The ESI capillary and cone voltages were optimized at 2.8 kV and 50 V respectively. DCL samples were quenched by the addition of formic acid (4 %), and diluted 20-fold in MeOH before injection into the MS. 'Native' ESI-MS of WGA was performed on a Q-TOFmicro quadrupole-time of flight mass spectrometer. The standard Micromass source was replaced with an Advion BioSciences NanoMateTM chip-based nano-ESI source. Protein samples were sprayed from 10 mM NH₄OAc (pH 7) using a chip nozzle voltage of 1.69 kV, and a cone voltage of 100 V. Collisional cooling of ions was achieved by partially closing a valve on the rotary vacuum pump, leading to an increased pressure in the intermediate vacuum region of the mass spectrometer. CsI was used for calibration.

Denaturing ESI-MS analysis of WGA revealed the presence of two major polypeptide chains with masses of 17086.6 and 17175.8 \pm 2 Da, accounting for the multiple signals seen for the dimer in Fig. 3 in the main text.

S2. Experimental

bis-N-Butoxycarbonyl-L-glycinyl-L-cysteine methylester 3

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Diisoprpylamine (1 mL) was added to a solution of *bis*-L-cysteine methylester (0.39 g, 1.4 mmol), 1,3-dicyclohexylcarbodiimide (0.55 g, 2.8 mmol), hydroxybenztriazole (0.36 g, 2.8 mmol) and *N*-Butoxycarbonyl-L-glycine (0.50 g, 2.8 mmol) in DCM (50 mL) and the reaction mixture was stirred at room temperature under Argon. After 24 h, t.l.c. (ethyl acetate:methanol, 5:1) showed the formation of a product (R_f 0.4). The reaction mixture was filtered, diluted with DCM (40 mL) and washed with water (2 x 30 mL). The organic layers was dried (MgSO₄), filtered and concentrated *in vacuo* to yield bis-*N*-butoxycarbonyl-L-glycinyl-L-cysteine methylester (0.49g, 60%) as a white amorphous solid; $[\alpha]_D^{21}$ +35.5 (*c*, 1.0 in CHCl₃); δ_H (400 MHz, CDCl₃) 1.46 (18H, s, (CH₃)₃), 3.20 (4H, m, CH₂-Cys), 4.70-4.74 (4H, m, α CH₂-Gly), 3.79 (6H, s, OMe), 4.86 (2H, m, α H-Cys), 5.51 (2H, m, NH-Gly), 7.23 (2H, d *J*_{NH, α H-Cys} 7.4 Hz, NH-Cys); δ_C (500 MHz, CD₃OD) 25.39 (t, CH₂(Cys)), 28.15 (q, C(<u>CH₃</u>)₃), 157.87 (s, NC(O)O), 170.49 (s, C(O) Gly), 170.99 (s, C(O) Cys); *m/z* (ESI⁺) 605 (M+Na⁺, 100%); HRMS (ESI⁺) calcd for C₂₂H₃₉N₄O₁₀S₂ 583.2108. Found 583.2120.

N-Butoxycarbonyl-L-glycinyl-L-cysteine methylester 4



Tributyl phosphine (245 mL, 1.0 mmol) was added to a solution of bis-*N*-butoxycarbonyl-L-glycinyl-L-cysteine methylester (246 mg, 0.83 mmol) in THF (10 mL) and methanol (2 mL). After 40 min, t.l.c. (1:1, petrol:ethyl acetate) indicated formation of a product (R_f 0.3) with complete consumption of the starting material (R_f 0.0). The reaction mixture was concentrated *in vacuo* and the residue was purified by flash column chromatography to afford *N*-butoxycarbonyl-L-glycinyl-L-cysteine methylester (496 mg, quantitative) as a foam; [α]_D²¹ +33.9 (*c*, 1.1 in CHCl₃); v_{max}

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(thin film) 3320 (br, SH), 1744, 1674 (s, C=O) cm⁻¹; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.35 (9H, s, (CH₃)₃), 1.47 (1H, at, *J* 8.8 Hz, SH), 2.87-2.91 (2H, m, CH₂-Cys), 3.68 (3H, s, OMe), 3.76 (2H, d, *J* 4.9 Hz, CH₂-Gly), 4.75-4.79 (1H, m, α H-Cys), 5.69 (1H, m, NH-Gly), 7.17 (1H, d *J* 6.4 Hz, NH-Cys); $\delta_{\rm C}$ (100 MHz, CDCl₃) 26.6 (t, CH₂-Cys), 28.2 (q, CH₃), 52.6 (q, OMe), 53.7 (d, α C), 80.0 (s, <u>C</u>(CH₃)₃), 156.2, 169.8, 170.4 (3 x s, 3 x C=O); *m/z* (ESI⁺) 315 (M+Na⁺,100), 293 (M+H⁺, 70 %); HRMS (ESI⁺) calcd. for C₁₁H₂₁N₂O₅S (M+H⁺) 293.1171. Found 293.1174.

N-Butoxycarbonyl-glycinyl-L-cysteinyl-(S-1-β-D-glucopyranosyl disulfide) methylester G



A solution of *N*-butoxycarbonyl-glycinyl-L-cysteine methylester (45 mg, 0.13 mmol) in methanol (8 mL) was added dropwise to a solution of phenyl 1-selenenylsulfide-β-D-glucopyranoside (100 mg, 0.3 mmol) and triethylamine (15 µL, 0.15 mmol) in methanol (12 mL) and the resulting solution was stirred at room temperature. After 30 min, t.l.c. (ethyl acetate:methanol, 9:1) indicated the formation of a major product (R_f 0.4). The reaction mixture was concentrated *in vacuo* and the residue was purified by flash column chromatography (ethyl acetate:methanol, 9:1) to yield *N*butoxycarbonyl-glycinyl-L-cysteinyl-(S-1-β-D-glucopyranosyl disulfide) methylester (32 mg, 36 %) as a white amorphous solid; $[\alpha]_D^{21}$ -101.2 (*c*, 1.2 in MeOH); v_{max} (thin film) 3346 (br, OH, NH), 1740, 1675 (s, C=O) cm⁻¹; δ_H (400 MHz, CD₃OD) 1.48 (9H, s, C(CH₃)₃), 3.08 (1H, dd, *J*_{CH,αH} 8.0 Hz, *J*_{CH,CH}⁻ 13.9 Hz, C<u>H</u>H⁻-Cys), 3.32-3.38 (5H, m, CH<u>H</u>[']-Cys, H-4, H-5, CH₂-Gly), 3.43 (1H, at, *J* 8.8 Hz, H-3), 3.53 (1H, at, *J* 9.0 Hz, H-2), 3.71 (1H, dd, *J*_{5,6} 5.5 Hz, *J*_{6,6}⁻ 12.0 Hz, H-6), 3.76 (3H, s, OMe), 3.91 (1H, dd, *J*_{5,6}⁻ 1.8 Hz, H-6'), 4.38 (1H, d, *J*_{1,2} 9.4 Hz, H-1), 4.96 (1H, m, αH); δ_C (100 MHz, CD₃OD) 27.3 (q, CH₃), 4.01 (t, CH₂-Cys), 43.1 (t, CH₂-Gly), 51.6 (q, OMe), 61.5 (t, C-6), 69.9, 81.1 (2 x d, C-4, C-5), 71.0 (d, C-2), 78.0 (d, C-3), 80.0 (s, $\underline{C}(CH_3)_3$), 90.2 (d, C-1), 171.1 (s, C=O); *m/z* (ESI⁺) 509 (M+Na⁺, 100 %); HRMS calcd. for C₁₇H₃₀N₂O₁₀S₂Na 509.1240. Found 509.1245.

S3. Library formation

Libraries consisted of 100µg each of *N*-butoxycarbonyl-glycinyl-L-cysteinyl-(S-2acetamino-2-deoxy-1- β -D-glucopyranosyl disulfide) methylester and *N*butoxycarbonyl-L-cysteinyl-(*S*-2-acetamino-2-deoxy-1- β -D-glucopyranosyl disulfide)-L-threonine methylester. Where DTT was used 5µg was added to the library. Solvents were water, Buffer: a 10mM solution of ammonium acetate adjusted to pH 7.5 by addition of ammonia and a 4% by volume solution of formic acid. 10 µL samples were taken and quenched in 100 µL of 4% formic acid solution before analysis. Wheat Germ Agglutinin (100 µg of a 5 mg/mL solution) was added after 78 h.