Electronic supplementary information

Experimental details

Materials

D₂O (99.9%) was received from Cambridge Isotope Laboratories. HAuCl₄·3H₂O (99.99%), N-acetyl-L-cysteine (>99%) and sodium borohydride (98%) were purchased from Aldrich. Water was purified with a Milli-Q system (\geq 18 M Ω cm). All other chemicals were analysis grade and used as received.

Characterization of the particles by TEM, UV-vis and NMR

 1 H and 13 C and DEPT NMR spectra were measured on a Bruker Avance 400 spectrometer at room temperature. Solutions were prepared in D₂O at a concentration of 50 mg/ml. A relaxation delay of 5 s was used to allow adequate signal decay between pulses.

Transmission Electron Microscopy (TEM) images were recorded with a Philips C200 electron microscope operated at 200 kV. TEM samples were prepared by casting a drop of a 1 mg/ml solution in water onto a carbon-coated copper grid. UV-vis spectra of the particles in water were collected on a Cary 300 spectrometer.

IR and VCD spectroscopy

IR and vibrational circular dichroism (VCD) spectra were recorded on a Bruker PMA 50 accessory coupled to a Tensor 27 Fourier transform infrared spectrometer. A photoelastic modulator (Hinds PEM 90) set at 1/4 retardation was used to modulate the handedness of the circular polarized light. Demodulation was performed by a lock-in amplifier (SR830 DSP). An optical low-pass filter (< 1800 cm⁻¹) put before the photoelastic modulator was used to enhance the signal/noise ratio. All solutions of N-acetyl-L-cysteine and N-acetyl-L-cysteine MPNs were prepared in D₂O. NaOD was added to deprotonate the carboxylic acid group. A VCD reference spectrum of D₂O containing the same amount of NaOD was subtracted from the VCD spectrum of the dissolved molecules and MPNs, respectively. All spectra were recorded at room temperature with a resolution of 8 cm⁻¹ in a cell equipped with CaF₂ windows and a 50 μ m Teflon spacer. Both sample and reference were measured for three hours in time slices of one hour, corresponding to about 24 000 scans in total for sample and reference,

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respectively. The spectra are presented without smoothing or further data processing. More information about the experimental procedure can be found elsewhere.^{1, 2}

Density functional theory calculations

The adsorption of deprotonated N-acetyl-L-cysteine on a gold 19 cluster was investigated using the Amsterdam Density Functional (ADF) program.³ The inner electrons were frozen for Au (up to 4f), S (2p), N (1s), O (1s) and C (1s). The core was modeled by a relativistically corrected pseudopotential using the ZORA (zero order regular approximation) method. A double– ζ basis set was used. The local part of the exchange and correlation functional was modeled using a Vosko, Wilk, Nuisar parametrization of the electron gas,⁴ whereas the non-local part of the functional was modeled using the Becke correction for the exchange⁵ and the Perdew correction for the correlation.⁶ More computational details can be found elsewhere.⁷

The adsorption of deprotonated N-acetyl-L-cysteine on Au₄ clusters was studied using Gaussian03.⁸ For the gold atoms an effective core potential was used. The calculations were performed using the b3pw91^{9, 10} functional and a LanL2DZ basis set¹¹ for Au and 6-31G(d,p) basis set¹² for all other atoms. Vibrational frequencies were scaled by a factor of 0.98. IR absorption and VCD spectra were constructed from calculated dipole and rotational strengths assuming Lorentzian band shape with a half-width at half-maximum of 4 cm⁻¹.



5 nm



Figure 1: Top: Transmission electron microscopy (TEM) of N-acetyl-L-cysteine MPNs. Bottom: UV-visible spectrum of N-acetyl-L-cysteine MPNs in water (c = 0.0235 mg ml⁻¹).

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Figure 2: Top: 400 MHz ¹H spectrum of N-acetyl-L-cysteine MPNs in D₂O at 298 K. The inset shows the free ligand for comparison. The resonance at 4.8 ppm is due to water. Bottom: 100 MHz NMR of N-acetyl-L-cysteine MPNs in D₂O. (A) Distortion less enhancement by polarization transfer 135° (DEPT), (B) ¹³C NMR.

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Figure 3: Calculated conformers of deprotonated N-acetyl-L-cysteine on a Au₁₉ cluster. The structures were optimized using ADF.³ The numbers indicate the relative stability with respect to the most stable conformation a. For the calculations the structure of the Au₁₉ cluster was fixed. For computational details see the experimental part.



Figure 4: Calculated infrared spectra for the structures shown in Fig. 2 of the manuscript. Solid lines connect vibrational bands with similar character.

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