Electronic Supplementary Information

Tripodal-Oxazoline-Based Homochiral Coordination Cages with Internal Binding Sites

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1. General Methods

All commercial reagents are of ACS reagent grade and used as supplied. DMF and CH₂Cl₂ were dried over 4 Å molecular sieves and CaH, respectively, and distilled before use. Metal precursors were prepared following the established procedures.¹⁻³ Column chromatography was carried out on Merck silica gel 60 (230-400 mesh). Melting points were obtained with an electrothermal capillary apparatus and were uncorrected. Optical rotations were measured using Rudolph Research Autopol III digital polarimeter using a sodium lamp (D line, 589 nm) and are reported in degrees with concentration in unit of 10 mg mL⁻¹. ¹H and ¹³C spectra were recorded using a Bruker AMX 300 or DRX 500 spectrometer and all chemical shifts were reported as δ in parts per million (ppm) downfield from tetramethylsilane ($\delta = 0.0$). ³¹P NMR spectra were recorded using a Bruker XP200 (81 MHz) spectrometer, and all chemical shifts were reported in ppm relative to external 80% H₃PO₄ at 0.00 ppm. ¹⁹F NMR spectra were recorded using a Buruker XP200 (188.3 MHz) spectrometer, and all chemical shifts were reported relative to external CFCl₃ at 0.00 ppm. Mass spectral analysis was recorded on Jeol JMS-AX505WA and is reported in units of mass to charge (m/z). CSI mass spectra were recorded on a QSTAR XL Hybrid LC/MS/MS System. Elemental analyses and HRMS were performed by the Korea Basic Science Institute at Kyung-Pook Branch Analytical Laboratory.

Reference:

- 1. S. Fallis, G. K. Anderson and N. P. Rath, Organometallics, 1991, 10, 3180-3184.
- P. J. Stang, D. H. Cao, S. Saito and A. M. Arif, J. Am. Chem. Soc. 1995, 117, 6273-6283.
- 3. M. Fujita, J. Yazaki and K. Ogura, J. Am. Chem. Soc. 1990, 112, 5645-5647.
- 2. Syntheses and Characterization of compounds 1a and 1b.



The Synthesis of intermediates **6** has been followed by the procedure reported in the reference 4 in the main paper. More specifically, compounds **5** and **6** have been used in the synthesis of other tripodal receptors, and their characterization data are included in a submitted paper.

2-[(3,5-Bis{[4-(3-hydroxyphenyl)-4,5-dihyrooxazol-2-yl]methyl}-2,4,6-triethyl)phenyl]methyl-4-(3-hydroxyphenyl)-4,5-dihydrooxazole-4-yl]phenoxy}methyl-4pyridine (1b). NaH (350 mg, 14.55 mmol) was added in small portions into a solution of 6b (1.0 g, 1.46 mmol) in dry DMF (20 mL). After hydrogen gas evolution is subsided, the mixture was stirred at room temperature for 30 min. A solution of 4bromomethylpyridine (1.84 g, 7.27 mmol) in CH₂Cl₂ (100 mL) was introduced into the reaction mixture using a syringe. The resulting mixture was stirred at room temperature for 24 h, and it was poured into an Erlenmeyer flask containing a mixture of water/dichloromethane. The organic layer was separated, washed with brine, dried over anhydrous MgSO₄, and concentrated to dryness. The residue was purified by column chromatography (MeOH:EtOAc, 3:7) to afford the desired pyridyl-tris(oxazoline) **1b** as white powders (0.75 g, 54%). (4-Bromomethylpyridine was freshly prepared before use by the following method: A saturated aqueous Na₂CO₃ solution was added to a stirred solution of 4-bromomethylpyridine HBr (1.84 g, 7.27 mmol) in distilled water (50 ml) at 0 °C, at such a rate to maintain the temperature <5 °C and to reach pH ~7. Then, 4-bromomethylpyridine was extracted with dichloromethane, dried over anhydrous MgSO₄, and filtered. The filtrate (100 ml) was used without further purification.) **1b**: mp 109-101 °C; $[\alpha]^{21}_{D} = +2.5$ (c = 1.0, CH₂Cl₂); ¹H NMR (300 MHz, CD₂Cl₂) δ 8.59 (d, J = 5.7 6H), 7.36 (d, J = 5.6 6H), 7.24 (dd, J = 7.9, 7.5 3H), 6.88-6.84 (m, 9H), 5.13 (dd, J = 9.1, 8.4 3H), 5.07 (s, 6H), 4.60 (dd, J = 9.4, 8.6 3H), 4.03 (dd, J = 8.4, 7.9 3H), 3.85 (s, 6H), 2.94 (q, J = 7.5 6H), 1.24 (t, J = 7.5 9H); ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.9, 159.0, 150.5, 146.6, 145.1, 142.6, 130.7, 130.2, 121.9, 120.0, 114.0, 113.5, 75.1, 70.0, 68.6, 29.1, 23.9, 14.9; MS (FAB) *m/z* (rel. intensity) 962 (M+1, 100), 871 (12), 735 (8); HRMS (FAB) cald. for C₆₀H₆₁O₆N₆ 961.4653, found 961.4650.



2-[(3,5-Bis{[4-(3-hydroxyphenyl)-4,5-dihyrooxazol-2-yl]methyl}-2,4,6-trimethyl)phenyl]methyl-4-(3-hydroxyphenyl)-4,5-dihydrooxazole-4-yl]phenoxy}methyl-4-

pyridine (1a). This compound was synthesized similarly as above starting from compound **5a**. The crude product was purified by column chromatography (MeOH:EtOAc, 3:7) to afford 1a in 63% yield as white solids: mp 92-94 °C; ¹H NMR (300 MHz, acetone-d₆) δ 8.48 (d, *J* = 5.3 6H), 7.44 (d, *J* = 4.8 6H), 7.20 (dd, *J* = 8.3, 7.6 3H), 6.89-6.79 (m, 9H), 5.06 (s, 9H), 4.59 (dd, *J* = 9.4, 8.6 3H), 3.99 (dd, *J* = 8.7, 8.1 3H), 3.86 (s, 6H), 2.45 (s, 9H); ¹³C NMR (75 MHz, acetone-d₆) δ 170.3, 160.2, 150.4, 149.5, 145.6, 137.3, 132.0, 131.2, 123.4, 120.8, 115.2, 114.4, 76.4, 70.1, 69.1, 30.8, 17.8; MS (FAB) *m/z* (rel. intensity) 919 (M+1, 100), 846 (41); HRMS (FAB) cald. for C₅₇H₅₄N₆O₆ 918.4105, found 918.4112.



Cage 2b. This complex was assembled by mixing tripodal oxazoline 1b with *trans*- $Pd(OTf)_2(PEt_3)_2$ in a molar ratio of 2:3 in CH_2Cl_2 at room temperature for 5 min. Removal of the solvent in vacuo afforded the desired cage in a quantitative yield. The

peak at 1.2 ppm in the ¹H NMR spectrum is owing to the presence of "free" Et_3P because a slight excess amount of the palladium source was used:



Fig. S1 ³¹P NMR spectra obtained for a mixture of **1b** and Pd(PEt₃)₂(OTf)₂ in CD₂Cl₂ at 25 $^{\circ}$ C with different molar ratios: (a) at a molar ratio of 2:3; (b) when more than 1.5 equiv of the Pd source was added to the solution (a).







(a)



Fig. S2 (a) CSI-MS spectrum for the cage **2b**, (b) an enlarged part of the m/z range 3500-4100, (c) an enlarged part of the m/z range 1000-1800. Experimental conditions: acceleration voltage, 5.0 kV; needle voltage, 2.8 kV; orifice voltage, 202 V; ring lens voltage, 254 V; spray temperature, 20 °C; resolution: 1000; flow rate, 1.0 mL/h; solvent: CH₂Cl₂.



Fig. S3 A selected region of the VT-NMR experimental data for the cage **2b** (from the bottom, taken at -40, -20, -5, 10, 25, 40 $^{\circ}$ C)

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Fig. S4 The disassembly experiment of the cage **2b** in the presence of Et₃N: (a) cage **2b** as single component, (b) partial disassembly into its free ligand **1b** after addition of 0.7 equiv of Et₃N, (c) complete disassembly into **1b** in the presence of excess Et₃N (10 equiv) at 300K. (\blacksquare : cage, \square : ligand, \blacktriangle : solvent, \triangle : Et₃N)



Fig. S5 Self-assembly of **1a** and [Pd(OTf)₂(PEt₃)₂], monitored by ¹H NMR in acetone-d₆ at 25 °C: (a) free **1a**; (b) a mixture of [**1a**]/[Pd(OTf)₂(PEt₃)₂] in a ratio of 2/1; (c) in a ratio of 1/1; (d) in a ratio of 2/3.



Fig. S6 ¹H NMR spectral change of the cage **2b** upon gradual addition of $NH_4 \cdot PF_6$ in CD_2Cl_2 at 25 °C: (a) free cage **2b**, (b) 0.5 equiv. of the guest added, (c) 1.0 equiv. of the guest added, (d) 2.0 equiv. the guest added (Note: a small amount of acetone was used to dissolve the guest).



Fig. S7 ¹H NMR spectral change of cage **2b** upon addition of enantiomeric and racemic α -phenylethylammonium perchlorate (The guest methyl group indicated in the box is enlarged and shown at the right side): (a) [**2b**]/[(*R*)-guest] = 1/2; (b) [**2b**]/[(*S*)-guest] = 1/2; (c) [**2b**]/[racemic guest] = 1/4.