

*Supporting Information for*

*Ratiometric Fluorescence Detection of a Tag Fused Protein  
Using the Dual-Emission Artificial Molecular Probe*

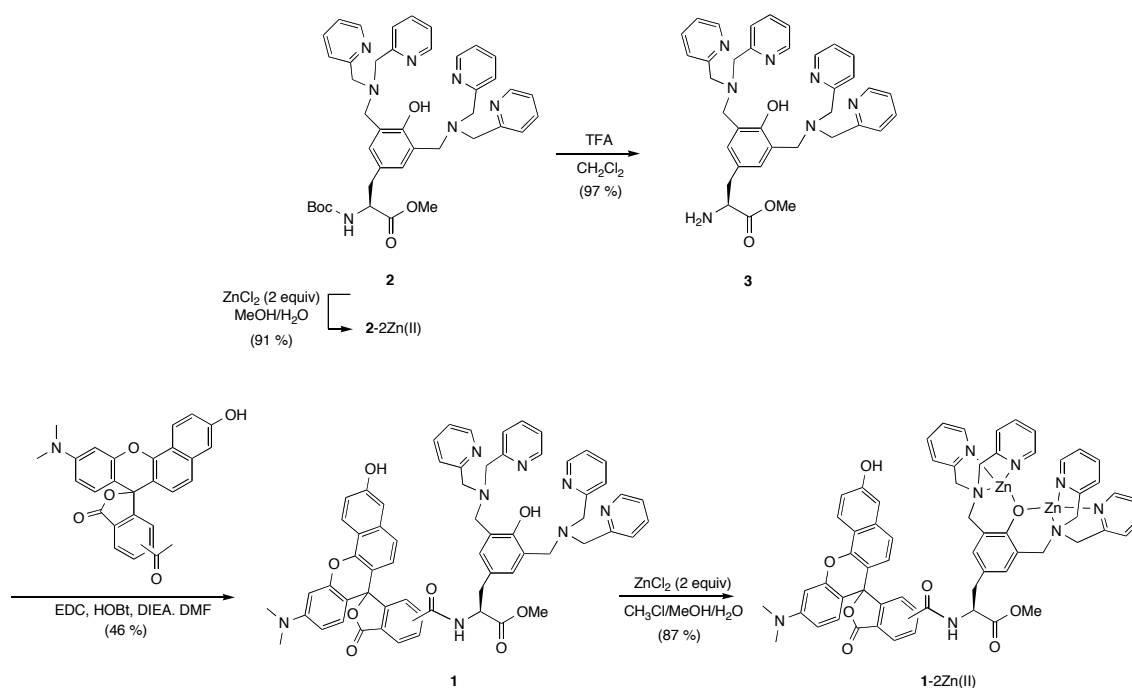
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## General methods

Unless otherwise noted, all chemical reagents were purchased from commercial suppliers and used without further purification.  $^1\text{H-NMR}$  spectra were recorded using a JNM-EX400 (JEOL, 400 MHz) spectrometer and the chemical shifts ( $\delta$ ppm) are referenced to the respective solvent. FAB mass spectra were recorded using a QP5050A (Shimadzu). Fluorescent spectra and Absorption spectra were recorded on a Perkin-Elmer LS55 spectrometer and U-2550 (Shimadzu), respectively. Reverse phase HPLC was conducted with a Lachrom (Hitachi) with C18 columns.

## Synthesis and Compound Characterizations



Scheme S1 Synthesis of 1-2Zn(II).

The details of synthesis of **2**, 2-2Zn(II) and **3** were described previously.<sup>8</sup>

**SNARF-DpaTyr (1)** A mixture of **3** (20 mg, 0.033 mmol), carboxy SNARF (mixture of 5- and 6- carboxylate mixture) (15 mg, 0.033 mmol), HOBT  $\cdot$   $\text{H}_2\text{O}$  (7.5 mg, 0.050 mmol), EDC  $\cdot$  HCl (9.4 mg, 0.050 mmol) and *N,N'*-diisopropylethylamine (30 mL, 0.17 mmol) in dry DMF (2 mL) was stirred at room temperature for 6 h. The mixture

was poured into water and extracted with ethyl acetate. The organic layer was washed with brine and dried over Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvent in vacuo, the residue was dissolved in CH<sub>3</sub>Cl (containing a small amount of MeOH) and the solution was diluted with Hexane to form precipitate. The precipitate was filtered and washed with Hexane to give **1** (16 mg, 46%) as a purple powder. **1** was used in the next reaction without further purification. Due to assignment difficulty, <sup>1</sup>H NMR of **1** was measured after isolation of each isomer by reverse phase HPLC.

<sup>1</sup>H NMR (400 MHz; CD<sub>3</sub>OD; solvent)

isomer-1 : δ<sub>H</sub> 3.00 – 3.06 (2H, m, CH<sub>2</sub>), 3.43 (6H, s, NMe), 3.76 (3H, s, Me), 4.15 – 4.28 (12H, m, CH<sub>2</sub>), 6.91 – 6.96 (1H, m, Ar-H), 7.24 – 7.29 (5H, m, Ar-H), 7.37 – 7.44 (12H, m, Ar-H), 7.56 – 7.61 (2H, m, Ar-H), 7.82 (4H, td, *J* = 8.0, 1.6 Hz, Ar-H), 8.07 – 8.12 (1H, m, Ar-H), 8.57 – 8.58 (5H, m, Ar-H), 8.83 (1H, d, *J* = 9.2 Hz, Ar-H).

isomer-2 : δ<sub>H</sub> 2.82 – 2.97 (2H, m, CH<sub>2</sub>), 3.42 (6H, s, NMe), 3.70 (3H, d, *J* = 4.0, Me), 4.00 – 4.20 (12H, m, CH<sub>2</sub>), 6.58 (1H, dd, *J* = 7.2, 1.2 Hz, Ar-H), 6.87 – 7.00 (3H, m, Ar-H), 7.07 – 7.16 (5H, m, Ar-H), 7.22 – 7.31 (3H, m, Ar-H), 7.36 – 7.44 (10H, m, Ar-H), 7.55 (1H, t, *J* = 8.8 Hz, Ar-H), 7.66 (1H, dd, *J* = 14.2, 1.2 Hz, Ar-H), 7.82 (4H, qd, *J* = 7.8, 1.6 Hz, Ar-H), 7.98 – 8.05 (2H, m, Ar-H), 8.25 – 8.31 (1H, m, Ar-H), 8.53 – 8.63 (4H, m, Ar-H), 8.81 (1H, d, *J* = 8.8 Hz, Ar-H).

FAB-HRMS *m/e* calcd for [M + H]<sup>+</sup> 1053.4299, found 1053.4312.

**SNARF-DpaTyr(Zn) (1-2Zn(II))** To a solution of **1** (mixture of the isomers 7.7 mg, 7.3 μmol) in distilled MeOH / CHCl<sub>3</sub> (1.5 mL / 0.5 mL) was added aqueous solution of ZnCl<sub>2</sub> (100 mM; 136 mL, 0.014 mmol), and the solution was stirred at room temperature for 1 h. The solution was concentrated by evaporation, and then lyophilized under vacuum. The obtained solid was suspended in ethyl acetate, filtered, and dried in vacuo to give **1-2Zn(II)** (7.9 mg, 87%) as a purple powder.

FAB-HRMS *m/e* calcd for [SNARF-DpaTyr + 2Zn + 2Cl]<sup>+</sup> 1249.2103, found 1249.2106.

### Preparation of D4 peptide (Boc-DDDD-NH<sub>2</sub>)

The Boc-DDDD-NH<sub>2</sub> was synthesized by the standard Boc chemistry in solution phase from Boc-Asp(Obzl)-OH and H-ASP(Obzl)-CONH<sub>2</sub>. Detailed synthetic procedure and compound characterization was described previously.<sup>8</sup>

### Preparations of the tag-fused RNases

The tagged RNases were constructed by the self-assembly of a S-protein (purchased from Aldrich) with a S-peptide<sup>S1</sup> tethering a D4-tag (DDDD) or His<sub>6</sub>-tag (HHHHHH) at its N-terminus, which was synthesized by the automated peptide synthesizer and purified by reverse-phase HPLC. Detailed synthetic procedures and compound characterizations were described previously.<sup>8</sup>

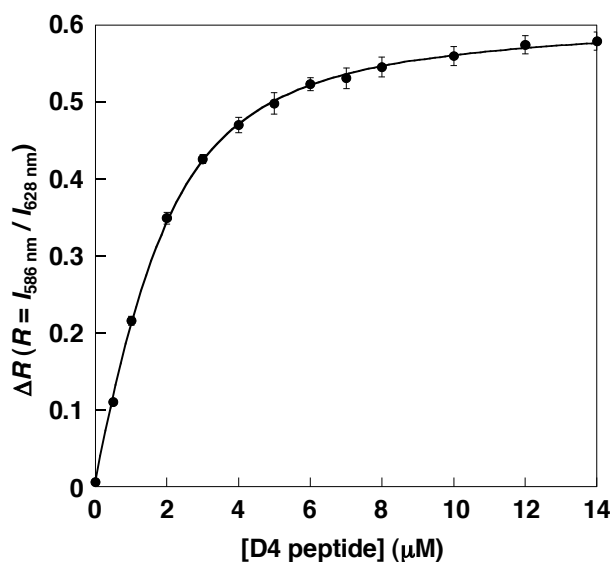


Fig. S1 Fluorescence titration curve of the emission intensity ratio  $R$  ( $R = I_{586 \text{ nm}} / I_{628 \text{ nm}}$ ) of 1-2Zn(II) (2 μM) upon addition of D4 peptide in 50 mM HEPES buffer, pH 7.2, at 25 °C.

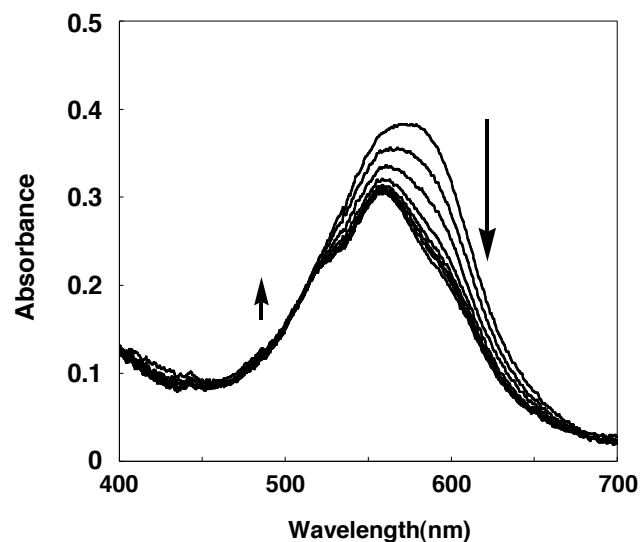


Fig. S2 Absorption spectral change of 1-2Zn(II) (2  $\mu$ M) upon the addition of D4 peptide : [D4 peptide] = 0, 0.5, 1, 2, 3, 4, 5, 6, 7, 8, 10, 12 and 14  $\mu$ M in 50 mM HEPES buffer, pH 7.2, at 25  $^{\circ}$ C.

### References

- S1. (a) P. R. Connelly, R. Varadarajan, J. M. Sturtevant and F. M. Richards, *Biochemistry*, 1990, **29**, 6108-6114. (b) I. Hamachi, R. Eboshi, J. Watanabe and S. Shinkai, *J. Am. Chem. Soc.*, 2000, **122**, 4530-4531.