

Supporting Material

Inhibition of *O*-GlcNAcase by a *gluco*-configured Nagstatin and a PUGNAc-imidazole Hybrid inhibitor.

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Synthesis of PUGNAc-Nagstatin hybrid

General: See reference¹ for details.

(5R,6R,7S,8S)-6,7-Bis(benzyloxy)-5-[(benzyloxy)methyl]-2,3-diiodo-5,6,7,8-tetrahydro-8-[(triisopropylsilyl)oxy]-imidazo[1,2-a]pyridine (**7**). Under Ar, a soln. of **6**² (316 mg, 0.5 mmol) in DMF (7 ml) was treated with *N*-iodosuccinimide (1.13 g, 5.0 mmol), heated to 80°, and stirred for 6 h. The brown mixture was diluted with Et₂O, washed successively with sat. NH₄Cl soln., a 5% aq. Na₂S₂O₃ soln., and H₂O, dried (Na₂SO₄), and filtered. Evaporation and FC (hexane/AcOEt 9:1) gave **7** (390 mg, 88%). Colourless viscous liquid. *R*_f (hexane/AcOEt 9:1) 0.63. $[\alpha]_{\text{D}}^{25} = -31.4$ (*c* = 1.125, CHCl₃). UV (CHCl₃): 242 (3.9). IR (CHCl₃): 3019_s, 2944_m, 2867_m, 1496_m, 1454_s, 1390_m, 1250_w, 1213_s, 1095_s, 1029_m, 882_m, 699_s. ¹H-NMR (CDCl₃, 300 MHz): 1.04–1.23 (*m*, (Me₂CH)₃Si); 3.61 (*dd*, *J* = 4.6, 9.8, CH–C(5)); 3.67 (*t*, *J* ≈ 8.4, CH'–C(5)); 3.96 (*t*, *J* ≈ 3.8, irradi. at 5.00 → *d*, *J* ≈ 3.9, H–C(7)); 4.34 (*dd*, *J* = 1.6, 4.2, irradi. at 3.96 → *d*, *J* ≈ 1.5, H–C(6)); 4.41 (*d*, *J* = 11.3, PhCH); 4.39–4.42 (*m*, H–C(5)); 4.4.3 (*d*, *J* = 12.1, PhCH); 4.54 (*d*, *J* = 11.3, PhCH); 4.55 (*d*, *J* = 12.1, PhCH); 4.59 (*d*, *J* = 10.3, PhCH); 4.63 (*d*, *J* = 12.1, PhCH); 5.00 (*d*, *J* = 3.7, irradi. at 3.96 → *s*, H–C(8)); 7.16–7.22 (*m*, 4 arom. H), 7.27–7.33 (*m*, 11 arom. H). ¹³C-NMR (CDCl₃, 75 MHz; assignment based on a HSQC spectrum): 12.69 (*d*, (Me₂CH)₃Si); 18.14, 18.20 (2*q*, (Me₂CH)₃Si); 60.46 (*d*, C(5)); 69.68 (*t*, CH₂–C(5)); 71.93, 72.94, 73.14 (3*t*, PhCH₂); 67.23 (*d*, C(8)); 72.92 (*d*, C(6)); 79.92 (*d*, C(7)); 80.20 (*s*, C(2)); 96.58 (*s*, C(3)); 127.62–128.46 (several *d*); 137.52, 137.59, 137.69 (3*s*); 150.66 (*s*, C(8a)). HR-MALDI-MS: 901.1386 (15, [M + Na]⁺, C₃₈H₄₈I₂N₂NaO₄Si⁺; calc. 901.1370), 880.1565 (28), 879.1531 (60, [M + H]⁺, C₃₈H₄₉I₂N₂O₄Si⁺; calc. 879.1551), 753.2592 (11, [M – I + 2 H]⁺, C₃₈H₅₀I₂N₂O₄Si⁺; calc. 753.2585), 706.0161

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(28), 705.0125 (87, $[M - \text{OTIPS}]^+$, $\text{C}_{29}\text{H}_{27}\text{I}_2\text{N}_2\text{O}_3^+$; calc. 705.0111), 578.1077 (100, $[M - \text{OTIPS} - \text{I}]^+$, $\text{C}_{29}\text{H}_{27}\text{IN}_2\text{O}_3^+$; calc. 578.1066). Anal. calc. for $\text{C}_{38}\text{H}_{48}\text{I}_2\text{N}_2\text{O}_4\text{Si}$ (878.70): C 51.94, H 5.51, N 3.19; found: C 52.18, H 5.64, N 3.04.

(5R,6R,7S,8S)-6,7-Bis(benzyloxy)-5-[(benzyloxy)methyl]-2-iodo-5,6,7,8-tetrahydro-8-[(triisopropylsilyl)oxy]imidazo[1,2-a]pyridine (**8**). A soln. of **7** (690 mg, 0.79 mmol) in THF (10 ml) was treated at 0° with a 1M soln. of EtMgBr in THF (1.2 ml, 1.2 mmol), stirred for 10 min, and treated with a sat. NH_4Cl soln. Extraction with AcOEt, work up, and FC (hexane/AcOEt) gave **8** (516 mg, 88%). Viscous liquid. R_f (hexane/AcOEt 4:1) 0.58. $[\alpha]_D^{25} = +22.1$ ($c = 1.2$, CHCl_3). UV (CHCl_3): 241 (3.6), 232 (3.2). IR (CHCl_3): 3031 w , 3015 m , 2945 s , 2867 s , 1602 m , 1496 m , 1454 s , 1424 m , 1363 m , 1363 m , 1261 s , 1219 s , 1097 s , 1015 s , 882 m , 809 s . $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): 0.98–1.20 (m , $(\text{Me}_2\text{CH})_3\text{Si}$); 3.71 (dd , $J = 3.1, 8.9$, H–C(6)); 3.76 (dd , $J = 5.9, 10.6$, CH–C(5)); 3.85 (dd , $J = 2.8, 10.9$, CH'–C(5)); 4.07 (t , $J = 3.4$, H–C(7)); 4.32–4.38 (m , H–C(5)); 4.39 (d , $J = 11.8$, PhCH); 4.51 (s , PhCH₂); 4.55, 4.59, 4.71 ($3d$, $J = 11.8$, 3 PhCH); 5.09 (d , $J = 3.1$, H–C(8)); 7.10 (s , H–C(3)); 7.17–7.19 (m , 2 arom. H); 7.30–7.38 (m , 13 arom. H). $^{13}\text{C-NMR}$ (CDCl_3 , 75 MHz; assignment based on HSQC spectrum): 14.76 (d , $(\text{Me}_2\text{CH})_3\text{Si}$); 18.15, 18.33 ($2q$, $(\text{Me}_2\text{CH})_3\text{Si}$); 57.26 (d , C(5)); 66.13 (d , C(8)); 67.95 (t , CH₂–C(5)); 72.44, 72.66, 73.72 ($3t$, 3 PhCH₂); 78.50 (d , C(6)); 80.90 (s , C(2)); 82.44 (d , C(7)); 123.13 (d , C(3)); 128.11–128.82 (several d); 137.41, 137.68, 137.73 ($3s$); 147.79 (s , C(8a)). HR-MALDI-MS: 775.2366 (**7**, $[M + \text{Na}]^+$, $\text{C}_{38}\text{H}_{49}\text{IN}_2\text{NaO}_4\text{Si}^+$; calc. 775.2404), 754.2595 (**38**), 753.2566 (**70**, $[M + \text{H}]^+$, $\text{C}_{38}\text{H}_{50}\text{IN}_2\text{O}_4\text{Si}^+$; calc. 753.2585), 580.1193 (**39**), 579.1151 (100, $[M - \text{OTIPS}]^+$, $\text{C}_{29}\text{H}_{28}\text{IN}_2\text{O}_3^+$; calc. 579.1145), 453.2153

(9), 452.2098 (20, $[M - \text{OTIPS} - \text{I}]^+$, $\text{C}_{29}\text{H}_{28}\text{N}_2\text{O}_3^+$; calc. 452.2100). Anal. calc. for $\text{C}_{38}\text{H}_{49}\text{IN}_2\text{O}_4\text{Si}$ (752.81): C 60.63, H 6.56, N 3.72; found: C 60.43, H 6.56, N 3.71.

(5R,6R,7S,8S)-6,7-Bis(benzyloxy)-5-[(benzyloxy)methyl]-5,6,7,8-tetrahydro-N-phenyl-8-[(triisopropylsilyl)oxy]imidazo[1,2-a]pyridine-2-carboxamide (**9**). At 0°, a soln. of **8** (134 mg, 0.18 mmol) in THF (5 ml) was treated with a 1M soln. of EtMgBr in THF (0.26 ml), stirred for 10 min, treated with PhNCO (0.22 ml, 1.8 mmol), warmed to 23° within 30 min, and stirred for 2 h. The mixture was cooled to 0°, diluted with sat. NH_4Cl soln., and extracted with Et_2O . The org. layer was washed with sat. NaHCO_3 soln. and brine, dried (NaSO_4), and evaporated. FC (hexane/AcOEt 4:1) gave **9** (108 mg, 81%). Viscous liquid. R_f (hexane/AcOEt 7:3) 0.52. IR (CHCl_3): 3433m, 3373m, 3090w, 3066m, 3020s, 2945s, 2867s, 1672s, 1596s, 1556s, 1499s, 1443m, 1363m, 1316m, 1268m, 1095s, 1028s, 883m. $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): 1.07–1.27 (m, $(\text{Me}_2\text{CH})_3\text{Si}$); 3.73 (dd, $J = 5.3, 10.6$, CH–C(5)); 3.82–3.88 (m, CH'–C(5), H–C(6)); 4.03 (t, $J = 5.0$, H–C(7)); 4.27–4.31 (m, H–C(5)); 4.41 (d, $J = 11.5$, PhCH); 4.51 (s, PhCH₂); 4.67, 4.74 (2d, $J = 11.2$, 2 PhCH); 4.78 (d, $J = 11.8$, PhCH); 5.05 (d, $J = 5.0$, H–C(8)); 7.10–7.14 (m, 2 arom. H); 7.25–7.39 (m, 16 arom. H); 7.69 (dd, $J = 1.2, 7.5$, 2 arom. H); 7.75 (s, H–C(3)); 9.03 (s, NH). $^{13}\text{C-NMR}$ (CDCl_3 , 75 MHz): 12.75 (d, $(\text{Me}_2\text{CH})_3\text{Si}$); 18.18, 18.27 (2q, $(\text{Me}_2\text{CH})_3\text{Si}$); 58.18 (d, C(5)); 67.60 (d, C(8)); 67.68 (t, CH₂–C(5)); 73.21, 73.35, 73.39 (3t, 3 PhCH₂); 76.91 (d, C(6)); 82.52 (d, C(7)); 119.34 (d, C(2) and C(6) of PhN); 120.71 (d, C(3)); 123.70 (d, C(4) of PhN); 127.80–128.96 (several d); 136.07, 136.97, 137.30, 137.55, 138.18 (5s); 145.47 (s, C(8a)); 160.72 (s, C=O). HR-MALDI-MS: 768.3746 (19, $[M + \text{Na}]^+$, $\text{C}_{45}\text{H}_{55}\text{N}_3\text{NaO}_5\text{Si}^+$; calc. 768.3809), 747.3999 (55), 746.3971 (89, $[M + \text{H}]^+$, $\text{C}_{45}\text{H}_{56}\text{N}_3\text{O}_5\text{Si}^+$; calc. 746.3989), 627.3579 (100, $[M - \text{CONHPh} + 2 \text{H}]^+$, $\text{C}_{38}\text{H}_{51}\text{N}_2\text{O}_4\text{Si}^+$;

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calc. 627.3618), 572.2496 (28, $[M - \text{OTIPS}]^+$, $\text{C}_{36}\text{H}_{34}\text{N}_3\text{O}_4^+$; calc. 572.2549), 454.2208 (30, $[M - \text{OTIPS} - \text{CONHPh} + 2 \text{H}]^+$, $\text{C}_{29}\text{H}_{30}\text{N}_2\text{O}_3^+$; calc. 454.2256).

(5R,6R,7S,8S)-6,7-Bis(benzyloxy)-5-[(benzyloxy)methyl]-5,6,7,8-tetrahydro-8-hydroxy-N-phenylimidazo[1,2-a]pyridine-2-carboxamide (**10**). A soln. of **9** (108 mg, 0.15 mmol) in THF (2 ml) was cooled to 0°, treated with 1M Bu_4NF in THF (0.3 ml, 0.3 mmol), stirred for 3 h, diluted with sat. NH_4Cl soln., and extracted with AcOEt. The combined org. layers were washed with sat. NaHCO_3 soln. and brine, dried (Na_2SO_4), and evaporated. FC (hexane/AcOEt 1:1) gave **10** (80 mg, 93%). White solid. M.p. 62–64°. R_f (hexane/AcOEt 1:1) 0.36. $[\alpha]_D^{25} = -25.0$ ($c = 0.98$, CHCl_3). UV (CHCl_3): 268 (4.3). IR (CHCl_3): 3615 m , 3496 m , 3019 s , 2975 m , 2871 w , 1672 s , 1597 s , 1561 s , 1522 s , 1498 m , 1364 w , 1327 m , 1220 s , 1099 m , 1046 m , 877 w . $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): 3.71 (dd , $J = 5.6, 10.6$, $\text{CH-C}(5)$); 3.77 (dd , $J = 4.3, 10.6$, $\text{CH'-C}(5)$); 4.03 (dd , $J = 4.8, 6.3$, irradi. at 4.85 $\rightarrow d$, $J \approx 6.3$, $\text{H-C}(7)$); 4.07 (dd , $J = 4.5, 6.3$, $\text{H-C}(6)$); 4.32 (br. q , $J \approx 4.5$, $\text{H-C}(5)$); 4.42 (d , $J = 11.5$, PhCH); 4.45–4.52 (br. s , exchange with D_2O , OH); 4.48 (d , $J = 12.1$, PhCH); 4.57, 4.70, 4.77, 4.79 ($4d$, $J = 11.5$, 4 PhCH); 4.85 (d , $J = 5.1$, irradi. at 4.03 $\rightarrow s$, $\text{H-C}(8)$); 7.09 (br. t , $J \approx 7.2$, 1 arom. H), 7.18–7.37 (m , 17 arom. H), 7.71 (dd , $J = 1.0, 7.5$, 2 arom. H); 7.78 (s , $\text{H-C}(3)$); 9.04 (s , exchange with D_2O , NH). $^{13}\text{C-NMR}$ (CDCl_3 , 75 MHz): 59.31 (d , $\text{C}(5)$); 66.91 (d , $\text{C}(8)$); 69.37 (t , $\text{CH}_2\text{-C}(5)$); 73.69, 74.16, 74.20 ($3t$, 3 PhCH_2); 76.90 (d , $\text{C}(6)$); 80.01 (d , $\text{C}(7)$); 119.34 (d , $\text{C}(2)$ and $\text{C}(6)$ of PhN); 121.91 (d , $\text{C}(3)$); 124.09 (d , $\text{C}(4)$ of PhN); 127.80–128.96 (several d); 136.97, 137.29, 137.56, 137.61, 138.38 ($5s$); 145.24 (s , $\text{C}(8a)$); 160.54 (s , C=O). HR-MALDI-MS: 612.2463 (17, $[M + \text{Na}]^+$, $\text{C}_{36}\text{H}_{35}\text{N}_3\text{NaO}_5^+$; calc. 612.2474), 591.2668 (50), 590.2641 (100, $[M + \text{H}]^+$, $\text{C}_{36}\text{H}_{36}\text{N}_3\text{O}_5^+$; calc. 590.2655), 497.2068 (18, $[M - \text{NHPh}]^+$,

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$C_{30}H_{29}N_2O_5^+$; calc. 497.2076). Anal. calc. for $C_{36}H_{35}N_3O_5$ (589.69): C 73.33, H 5.98, N 7.13; found: C 73.07, H 6.15, N 7.13.

(5R,6R,7S,8S)- and (5R,6R,7S,8R)-8-Azido-6,7-bis(benzyloxy)-5-[(benzyloxy)methyl]-5,6,7,8-tetrahydro-N-phenylimidazo[1,2-a]pyridine-2-carboxamide (**11** and **12**). A soln. of **10** (235 mg, 0.39 mmol) and diphenyl phosphoryl azide (DPPA; 0.43 ml, 1.99 mmol) in dry toluene was heated to 55°, treated dropwise within 15 min with a soln. of DBU (61 μ l, 1.99 mmol) in toluene (1 ml), stirred for 6 h, cooled to r.t., diluted with sat. NH_4Cl soln., and extracted with Et_2O . The combined org. layers were washed with brine, dried (Na_2SO_4), and evaporated. FC (hexane/AcOEt 7:3) gave **11/12** 2:3 (209 mg, 85%). R_f (hexane/AcOEt 7:3) 0.32. IR ($CHCl_3$): 3379 s , 3090 m , 3013 s , 2931 m , 2870 s , 2110 s , 1673 s , 1597 s , 1552 s , 1498 m , 1363 m , 1295 s , 1203 s , 1106 s , 1028 m , 990 m . 1H -NMR (C_6D_6 , 300 MHz, **11/12** 2:3): 3.31 (dd , $J = 5.8, 10.0, 0.4$ H), 3.36 (dd , $J = 4.1, 10.8, 0.6$ H) (CH-C(5)); 3.42–3.47 (m , CH'-C(5)); 3.56 (t , $J \approx 7.9, 0.4$ H-C(7)); 3.62–3.76 (m , 0.6 H-C(7), 0.6 H-C(5), H-C(6)); 3.90 (q , $J \approx 5.2, 0.4$ H-C(5)); 3.98 (d , $J = 12.0, 0.6$ PhCH); 4.03 (d , $J = 12.3, 0.6$ PhCH); 4.08 (d , $J = 13.1, 0.4$ PhCH); 4.09 (d , $J = 12.1, 0.6$ PhCH); 4.25 (d , $J = 12.3, 0.6$ PhCH); 4.30 (d , $J = 12.3, 0.4$ PhCH); 4.39 (d , $J = 7.4$, irradiated at 3.56 \rightarrow s , 0.4 H-C(8)); 4.42 (d , $J = 11.5, 0.4$ PhCH); 4.48 (d , $J = 11.5, 0.6$ PhCH); 4.57 (d , $J = 11.3, 0.4$ PhCH); 4.64 (d , $J = 11.0, 0.4$ PhCH); 4.67 (d , $J = 11.5, 0.4$ PhCH); 4.78 (d , $J = 3.9$, irradiated at 3.68 \rightarrow s , 0.6 H-C(8)); 6.83–6.89 (m , 1 arom. H); 7.05–7.28 (m , 17 arom. H); 7.74–7.78 (m , 2 arom. H); 7.85 (s , 0.4 H), 7.91 (s , 0.6 H) (H-C(3)); 9.31 (s , 0.6 H), 9.32 (s , 0.4 H) (NH). ^{13}C -NMR (C_6D_6 , 75 MHz, **11/12** 2:3): Signals of **11**: 58.61 (d , C(8)); 59.19 (d , C(5)); 65.76 (t , CH_2 -C(5)); 73.31, 74.56, 74.89 ($3t$, 3 PhCH $_2$); 74.76 (d , C(6)); 122.21 (d , C(3)); 141.08 (s , C(8a)); signals of **12**: 55.14 (d , C(8)); 59.27 (d , C(5));

67.68 (*t*, CH₂-C(5)); 69.72, 72.50, 73.76 (*3t*, 3 PhCH₂); 77.37 (*d*, C(6)); 123.82 (*d*, C(3)); 140.34 (*s*, C(8a)); signals of **11** and **12**: 80.35 (*d*, C(7)); 119.61 (*d*, C(2) and C(6) of PhN); 121.19 (*d*, C(4) of PhN); 127.82–128.88 (several *d*); 136.63, 136.76, 136.89, 136.93, 136.95, 137.31, 137.60, 137.91 (*8s*); 160.06 (*s*, C=O).

Transformation of 11/12 into 13 and 4. A soln. of **11/12** 3:2 (170 mg, 0.27 mmol) in AcOH (5 ml) was treated with 10% Pd/C, hydrogenated under 6 bar of H₂ for 18 h, and filtered over *Celite*. The filtrate was evaporated and co-evaporated with toluene. A soln. of the residue in pyridine (1 ml) was treated with Ac₂O (2 ml) and stirred for 24 h at r.t. Evaporation and co-evaporation with toluene gave a yellow oil (108 mg), which was dissolved in 3M NH₃ in MeOH and stirred at r.t. for 24 h. Evaporation and co-evaporation with toluene yielded crude **13/4** which was separated by reversed-phase preparative chromatography (*Lichrospher RP-18e*, 250 x 4 mm column; H₂O/MeCN 4:1) to give **13** (23 mg, 23%) and **4** (17 mg, 17%). Crystallisation from H₂O/MeCN 85:15 and drying at 10⁻⁴ Torr for 4 d gave pure samples of **13** and **4** for analysis.

Data of (5R,6R,7S,8S)-8-Acetamido-5,6,7,8-tetrahydro-6,7-dihydroxy-5-(hydroxymethyl)-N-phenylimidazo[1,2-a]pyridine-2-carboxamide (13). *R_f* (reversed phase; H₂O/MeCN 4:1) 0.47. ¹H-NMR (CD₃OD, 300 MHz): 2.11 (*s*, AcN); 3.81 (*t*, *J* = 8.7, irradi. at 4.99 → *d*, *J* ≈ 9.0, H-C(7)); 3.91 (*t*, *J* = 8.7, H-C(6)); 3.96–4.02 (*m*, H-C(5), CH-C(5)); 4.22 (*dd*, *J* = 4.0, 13.5, CH'-C(5)); 4.99 (*d*, *J* = 9.0, irradi. at 3.81 → *s*, H-C(8)); 7.12 (*tt*, *J* ≈ 0.9, 7.0, H-C(4) of PhN); 7.33 (*tdd*, *J* = 1.8, 7.5, 8.4, H-C(3) and H-C(5) of PhN); 7.69 (br. *dd*, *J* = 0.9, 8.7, H-C(2) and H-C(6) of PhN); 7.99 (*s*, H-C(3)). ¹³C-NMR (CD₃OD, 75 MHz): 22.69 (*q*, Me); 51.08 (*d*, C(8)); 61.22 (*t*, CH₂-C(5)); 62.98 (*d*, C(5)); 69.55 (*d*, C(6)); 73.64 (*d*, C(7)); 120.65 (*d*, C(2) and C(6) of PhN); 123.08 (*d*,

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H–C(3)); 124.78 (*d*, C(4) of PhN); 129.41 (*d*, C(3) and C(5) of PhN); 137.12, 139.07 (2*s*, C(1) of PhN, C(2)); 146.03 (*s*, C(8a)); 162.27 (*s*, NC=O); 173.23 (*s*, MeC=O). HR-MALDI-MS: 383.1320 (52, $[M + Na]^+$, $C_{17}H_{20}N_4NaO_5^+$; calc. 383.1331), 361.1504 (37, $[M + H]^+$, $C_{17}H_{21}N_4O_5^+$; calc. 361.1512), 343.1398 (100, $[M - OH]^+$, $C_{17}H_{19}N_4O_4^+$; calc. 343.1406), 303.0193 (24), 301.1293 (33). Anal. calc. for $C_{17}H_{20}N_4O_5 \cdot 2 H_2O$ (396.14): C 51.51, H 6.10, N 14.03; found: C 51.67, H 5.84, N 13.68.

Data of (5R,6R,7S,8R)-8-Acetamido-5,6,7,8-tetrahydro-6,7-dihydroxy-5-(hydroxymethyl)-N-phenylimidazo[1,2-a]pyridine-2-carboxamide (4). R_f (reversed phase; $H_2O/MeCN$ 4:1) 0.40. 1H -NMR (CD_3OD , 300 MHz): 2.13 (*s*, AcN); 3.96 (*dd*, $J = 6.0, 11.5$, CH–C(5)); 4.02 (*dd*, $J = 4.4, 11.8$, CH'–C(5)); 4.14 (*dd*, $J = 4.0, 6.0$, irradi. at 5.53 $\rightarrow d$, $J \approx 6.0$, H–C(7)); 4.21–4.23 (*m*, H–C(5), H–C(6)); 5.53 (*d*, $J = 3.7$, irradi. at 4.14 $\rightarrow s$, H–C(8)); 7.11 (*tt*, $J \approx 0.9, 7.5$, H–C(4) of PhN); 7.33 (br. *tt*, $J \approx 2.1, 8.3$, H–C(3) and H–C(5) of PhN); 7.70 (br. *dd*, $J = 0.9, 8.7$, H–C(2) and H–C(6) of PhN); 7.94 (*s*, H–C(3)). ^{13}C -NMR (CD_3OD , 75 MHz): 22.56 (*q*, MeC=O); 46.82 (*d*, C(8)); 63.95 (*t*, CH_2 –C(5)); 65.04 (*d*, C(5)); 69.39 (*d*, C(6)); 69.79 (*d*, C(7)); 120.67 (*d*, C(2) and C(6) of PhN); 124.48 (*d*, C(3)); 124.79 (*d*, C(4) of PhN); 129.44 (*d*, C(3) and C(5) of PhN); 136.40, 139.12 (2*s*, C(1) of Ph, C(2)); 144.77 (*s*, C(8a)); 162.37 (*s*, NC=O); 173.00 (*s*, MeC=O).

Cloning and Over-Expression

The BtGH84 ORF was truncated to produce an N-terminally tagged recombinant protein beginning at residue N23 to omit the predicted signal peptide with subsequent gene expression and protein purification as described for the native enzyme previously³.

Crystallization and Structure Solution

BtGH84 crystals were grown using hanging drop vapour diffusion from 0.6M Sodium Acetate, 13% (v/v) PEG 3500, 0.1M MES (pH 6.0) and 10% (v/v) glycerol. An additional 20% glycerol was included prior to flash freezing in liquid nitrogen. Crystals of BtGH84 in complex with PUGNAc-Imidazole hybrid inhibitor were obtained by co-crystallization with 10mM ligand. X-ray data were collected from a single crystal at 100K at the European Synchrotron Radiation Facility on beamline ID 14-2. All X-ray data were processed with HKL2000, Table 1S. A calculated Matthews' coefficient of 2.4 Å³ Da⁻¹ and a theoretical solvent content of 49% are consistent with the asymmetric unit containing one BtGH84 monomer. Structure solution was carried out using the molecular replacement technique as implemented in AMORE⁴ using BtGH84 (PDB code 2CHO) as the search model. COOT⁵ was used to make manual corrections and augmentations to the model followed by refinement using REFMAC⁶. Solvent molecules were added using COOT and checked manually. The BtGH84 structure can be traced from residue 4 of the mature protein to residue 589, with two short disordered loops between residues 20-25 and 47-54. The C-terminal domain is, however, extremely mobile in the crystal and disordered in the electron density.

Coordinates have been deposited with the Protein Databank, PDB CODE 2j47.

Kinetic Analysis of Human O-GlcNAcase

All assays were carried out in triplicate at 37 °C for 45 minutes by using a stopped assay procedure in which the enzymatic reactions (80 μ L) were quenched by the addition of a 4-fold excess (320 μ L) of quenching buffer (200 mM glycine, pH 10.75). Assays were initiated by the addition, *via* syringe, of enzyme (10 μ L), and in all cases the final pH of the resulting quenched solution was greater than 10. A time-dependent assay of the human *O*-GlcNAcase revealed that it was stable over the period of the assay: 50 mM NaH₂PO₄, 100 mM NaCl, 0.1% BSA, pH 7.4 and 50 mM citrate, 100 mM NaCl, 0.1% BSA, pH 4.25. The progress of the reaction at the end of 45 minutes was determined by measuring the extent of methyl umbelliferone liberated as determined by fluorescence measurements using a Varian CARY Eclipse Fluorescence Spectrophotometer 96-well plate system. Excitation and emission wavelengths of 368 and 450 nm were used, respectively, with 5 mm slit openings. Activity was defined as the absolute fluorescence detected for the 45 minute period. *O*-GlcNAcase was expressed fresh prior to use and was used in the inhibition assays at a concentration of 0.001 μ g/ μ L. The inhibitor was tested at six concentrations ranging from 0.00 to 18 μ M.

Kinetic Analysis of Bacterial O-GlcNAcase

Enzymatic reactions were carried out in PBS buffer (pH 7.4) and all assays were carried out by monitoring continuously at 37 °C and 400 nm the release of 4-nitrophenol from the reaction mixture. The rate of release was measured using a Cary 3E UV-VIS spectrophotometer equipped with a Peltier temperature controller. Reactions (80 μ L) were pre-heated in a 50 μ l microcuvette (Varian) for approximately 2 min followed by addition of enzyme (10 μ L) *via* syringe. Reaction velocities were determined by linear

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regression of the linear region of the reaction progress curve between the first and second minute. The bacterial *O*-GlcNAcase was expressed fresh prior to use and was used in the inhibition assays at a concentration of 0.0008 $\mu\text{g}/\mu\text{L}$. The inhibitor was tested at six concentrations ranging from 0.00 to 20 μM .

Table 1S

Data collection, phasing and refinement statistics for the structure solution of *B. thetaiotaomicron* GH84 O-GlcNAcase with PUGNAc-Imidazole hybrid Inhibitor

<i>Bt</i> GH84 with PUGNAc-Imidazole	
Data collection	
Space group	C2
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	188.4,51.3,85.5
α , β , γ (°)	90.0,100.2,90.0
Wavelength	0.933
Resolution (Å)	50-1.97 (2.04-1.97)*
<i>R</i> _{sym} or <i>R</i> _{merge}	0.083 (0.491)
<i>I</i> / σ <i>I</i>	17 (3)
Completeness (%)	100 (99)
Redundancy	4.5 (4.4)
Refinement	
Resolution (Å)	1.97
No. reflections	53713
<i>R</i> _{work} / <i>R</i> _{free}	0.19/0.23
<i>No. atoms</i>	
Protein	4674
Ligand/ion	26
Water	319
<i>B-factors</i>	
Protein	26
Ligand/ion	16
Water	35
<i>R.m.s deviations</i>	
Bond lengths (Å)	0.016
Bond angles (°)	1.5

*Highest-resolution shell is shown in parentheses.

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