

## Supplementary Methods

**Synthesis of fluorescein-lactose conjugate.** The succinimidyl ester of 5-(and-6)-carboxyfluorescein (mixed isomers, Molecular Probes) was reacted with *p*-aminophenyl- $\beta$ -D-lactopyranoside (Toronto Research Chemicals) in 0.2M sodium bicarbonate buffer pH 8.3 according to the manufacturers instructions. The product was partially purified with a SepPak C<sub>18</sub>Plus Cartridge (0.39 g, Waters) by washing with HPLC grade water (10 mL) and eluting with HPLC grade MeOH (10 mL). The lyophilized partially purified product was then subjected to a final purification by preparative HPLC using an Amide 80 column (Tosoh Bioscience) and a gradient elution (90:10 to 40:60 MeCN: H<sub>2</sub>O over 45 minutes, then 40:60 MeCN: H<sub>2</sub>O for 15 minutes). Fractions containing pure product were combined and lyophilized to yield an orange powder (40.8 mg, 89.6%). HR ESI-MS: *m/z* = 814.1960 [M + Na]<sup>+</sup> (expected for C<sub>39</sub>H<sub>37</sub>NO<sub>17</sub>Na<sup>+</sup>: *m/z* = 814.1959).

**Cst II catalyzed glycosylation reactions.** Fluorescein-lactose acceptor substrate (2 mM), *p*-nitrophenyl  $\alpha$ -sialoside (6 mM), CMP (3 mM) and Cst II (~0.5 mg/mL) were incubated for 48 hours at room temperature in 20 mM HEPES buffer (pH 7.5) containing 10 mM MgCl<sub>2</sub>. Upon completion, the reaction was purified with a SepPak C<sub>18</sub>Plus Cartridge (0.39 g, Waters) followed by preparative HPLC as described above for the fluorescein-lactose conjugate. Fractions containing the purified product were combined and lyophilized to yield an orange powder (10.1 mg, 72.0%). HR ESI-MS: *m/z* = 1081.2931 [M]<sup>+</sup> (expected for C<sub>50</sub>H<sub>53</sub>N<sub>2</sub>O<sub>25</sub>: *m/z* = 1081.2937). Owing to the complexity of the product (an unprotected trisaccharide existing as a mixture of 5- and 6-

Supplementary Material (ESI) for Chemical Communications  
This journal is (c) The Royal Society of Chemistry 2006

carboxy fluorescein regioisomers) the regiochemistry of the sialylated product could not be unambiguously deduced by  $^1\text{H}$  or  $^{13}\text{C}$  NMR spectroscopy. The regiochemistry was therefore confirmed by treatment of 5 nmol of the purified reaction product with a selective  $\alpha$ -2,3-sialidase (New England Biolabs). Under a unit defined time condition, complete reversion of the product to the fluorescein-lactose conjugate was observed (Sup. Fig. 2).

**LgtC catalyzed glycosylation reactions.** Fluorescein-lactose acceptor substrate (2 mM), 2,4-dinitrophenyl  $\beta$ -galactoside (6 mM), UDP (3 mM) and LgtC ( $\sim$ 0.5 mg/mL) were incubated for 96 hours at room temperature in 20 mM HEPES buffer (pH 7.5) containing 10 mM  $\text{MnCl}_2$ , and 10 mM DTT. The reaction mixture was partially purified with a SepPak C<sub>18</sub>Plus Cartridge (0.39 g, Waters). Due to the high rate of spontaneous hydrolysis of the alternative donor 2,4-dinitrophenyl  $\beta$ -galactoside, detailed product analysis was not possible. Presumably due to the high rates of reaction, for the positive control reaction using excess UDP Gal, tetra- and penta-saccharide products were detected by TLC (Sup. Fig. 3) and their identities were confirmed by ESI MS (data not shown). HR ESI-MS:  $m/z$  = 954.2674 [M + Na]<sup>+</sup> (expected for C<sub>45</sub>H<sub>48</sub>NO<sub>22</sub>:  $m/z$  = 954.2668).