

Directed Evolution and Axial Chirality: Optimization of the Enantioselectivity of *Pseudomonas aeruginosa* Lipase towards the Kinetic Resolution of a Racemic Allene

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Bacterial Strains and Growth Conditions

Plasmids were transformed into *Pseudomonas aeruginosa* PABST7.1 competent cells as previously described.¹ Single colonies were picked from Luria-Bertani (LB) agar plates (supplemented with the appropriate amount of antibiotics: 100 µg/mL and tetracycline 50 µg/mL) and used to inoculate 15 mL of LB 2x liquid media in a 100 mL Erlenmeyer flask as preculture. The preculture was grown in an orbital shaker at 30°C and 300 rpm overnight.

1 mL of this preculture was used to inoculate a new erlenmeyer flask with 15 mL of LB 2x media. After 5h at 30°C and 300 rpm, lipase expression was induced by the addition of isopropyl-β-D-thiogalactopyranoside (IPTG) to give a final concentration of 0.1 mM, being incubated for 5h more. The lipase-containing supernatants were recovered by centrifuging at 8000 rpm (= 8228 x g) for 45 min and stored overnight at 4°C. (An Eppendorf Centrifuge 5804 provided with a 6x85 mL High Speed Fixed Angle Rotor, radius 11.5 cm was used).

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Reaction Conditions

100 μ L of culture supernatant, 1 mL of 100mM Tris/HCl buffer pH 7.5 and 25 μ L of substrate solution 10 mg/mL in acetonitrile were used as reaction mixture. The reactions were developed until a conversion between 15-45% had been reached. The reactions with commercial enzymes were performed in similar conditions but using 100 μ L of enzyme solution 5 mg/mL in buffer 100mM Tris/HCl buffer pH 7.5.

Extraction: 400 μ L of DCM were added to the reaction mixture and vortex for 30 s. Then 20 μ L of HCl 10% were added and stirred for additional 30 s. The solvent was transferred to a 96-glass plate for analysis by GC.

Analytical Conditions

The chiral analyses of the hydrolysis of the *p*-nitrophenyl 4-cyclohexyl-2-methylbuta-2,3-dienoate (**1**) used as substrate, were performed by GC using an IVADEX-1 chiral column (IVA analysentechnik, Meerbusch, Germany). 25m, 0,15 μ m, 0,25 mm. 160°C (20min); 20°C/min to 200°C for 50 min (total time 72 min). Carrier: Nitrogen. Press: 1.2 bar; Flow: 1.3 mL/min; 39 cm/s. Retention times: (+)-4-cyclohexyl-2-methylbuta-2,3-dienoic acid: 13.8 min, (-)-4-cyclohexyl-2-methylbuta-2,3-dienoic acid: 16.1 min, *p*-nitrophenol: 23.8 min, *p*-nitrophenyl 4-cyclohexyl-2-methylbuta-2,3-dienoate (**1**): 67 min. HPLC: Column 150 mm Chiralpak AD-RH, 4,6 mm i.D. Acetonitrile/0,1% TFA 35:65 after 40 min Gradient to 90:10 in 5 min. Retention times: (+)-4-cyclohexyl-2-methylbuta-2,3-dienoic acid: 30.0 min; (-)-4-cyclohexyl-2-methylbuta-2,3-dienoic acid: 38.1 min; *p*-nitrophenol: 9.6 min; *p*-nitrophenyl 4-cyclohexyl-2-methylbuta-2,3-dienoate (**1**): 51.8 min. The enantiomers elute in the same order as by GC.

Other lipases tested

Bacillus subtilis lipase A (expressed in *E.coli* BL21(DE3) cells using the pET22 vector as previously described,ⁱⁱ *Candida antarctica* lipase A (Roche), *Candida antarctica* lipase B (Roche), *Candida lipolytica* (Fluka), *Candida rugosa* (Sigma), Hog Pancreas Lipase (Fluka),

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Penicillium roqueforti (Fluka), Porcine Pancreas (Fluka), *Pseudomonas cepacia* (Roche), *Pseudomonas sp* (Roche), *Rhizomucor miehei* (Roche), Wheat germ lipase (Fluka).

The mutant Leu162Phe was superior to all other lipases tested for the kinetic resolution of **1**. The following observations for other lipases are interesting to note: (i) *Bacillus subtilis* lipase A wild-type reached an E value of 33 (ee = 89% (+) at 42% conversion), (ii) *Candida antarctica* lipase A gave an E value of 7.5 (ee = 62% (+) at 48 % conversion), and (iii) *Candida rugosa* lipase was the only commercial enzyme tested that showed reverse enantioselectivity in the kinetic resolution of **1** (ee = 18% (-) at 43% conversion, E = 1.6).

CAST libraries

CAST Combinatorial Active Site Saturation Test is a directed evolution strategy based on the systematic creation of focused saturation mutagenesis libraries around the substrate binding site, following specific criteria to select the amino acid residues involved.ⁱⁱⁱ

During the development of CASTing using *Pseudomonas aeruginosa* lipase (PAL), five different saturation mutagenesis libraries (NNK degeneracy)^{iv} were selected according to the combinatorial active site saturation test (CAST) criteria (Fig. 1). These libraries were tested towards a collection of bulky *p*-nitrophenyl esters with the objective to expand the substrate scope, and in parallel to enhance the enantioselectivity in the kinetic resolution of *p*-nitrophenyl 2-methyldecanoate. The wild-type enzyme gives a very low enantioselectivity (E = 1.1 (*S*)) with the latter model substrate. From the enantioselectivity point of view enzyme variants from libraries A, C and E (the only library where variants with reverse enantioselectivity were found) presented some interesting results but the mutants showing a real improvement were all from library D.

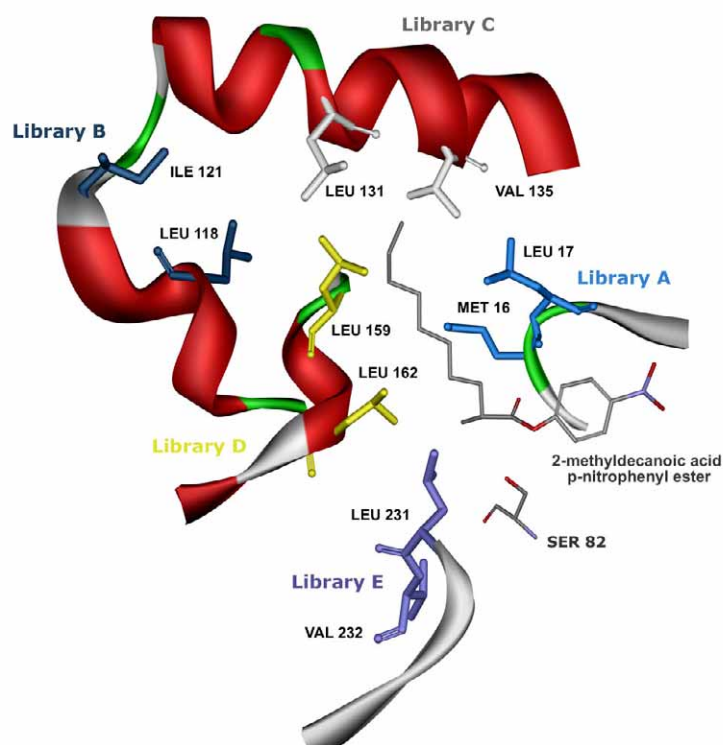


Figure 1. CAST libraries

Synthesis of 4-nitrophenyl 4-cyclohexyl-2-methyl-2,3-butadienoate (1). This substrate was prepared from the allenic acid using an esterification procedure described in ref v. The allenic acid was obtained according to ref vi.

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^{iv} The cost of the mutagenic primers can be reduced by means of the DNA degenerate code from the International Union of Biochemistry (IUB) (K = G/T, M = A/C, N = A/C/G/T, R = A/G, S = G/C, W = A/T, Y = C/T). Therefore, apart from the standard saturation mutagenesis approaches based on the use of NNK as degeneracy, the degree of saturation can be adjusted to reduce the screening effort. CASTER a small informatic tool developed in our group to help in the design of such libraries. This tool is available at <http://www.mpi-muelheim.mpg.de/reetz.html>.

^v S. Kim, J. I. Lee, Y. C. Kim, *J. Org. Chem.*, **1985**, *50*, 560-565.

^{vi} J. A. Marshall, M. A. Wolf, E. M. Wallace, *J. Org. Chem.*, **1997**, *62*, 367-371