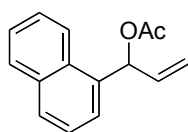


Retention of Regiochemistry of Monosubstituted Allyl Acetates in the Ruthenium Catalysed Allylic Alkylation with Malonate Anion

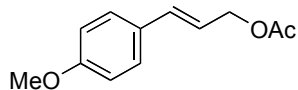
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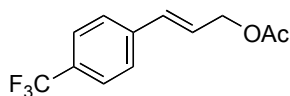
General and Materials: All manipulations were carried out under a nitrogen atmosphere. Nitrogen gas was dried by passage through P₂O₅. NMR spectra were recorded on a JEOL JNM MH-400 spectrometer (400 MHz for ¹H and 100 MHz for ¹³C) or JNM MH-500 spectrometer (500 MHz for ¹H, 125 MHz for ¹³C). Chemical shifts are reported in δ ppm referenced to an internal SiMe₄ standard for ¹H NMR. Residual chloroform (δ 77.0 for ¹³C) was used as internal reference for ¹³C NMR. ¹H and ¹³C and NMR spectra were recorded in CDCl₃ at 25 °C unless otherwise noted. [RuCl₂(*p*-cymene)]₂, PPh₃, other reagents and solvents were purchased from common commercial sources and were used without further purification. Allyl acetates **1a**,¹ **1b**,² **1c**,² **1e**,³ **1g**,² **2e**⁴ and **2f**¹² were prepared according to the literatures. Other allyl acetates **1d**,⁵ **2b**,⁶ **2c**,⁷ **2d**⁸ and **2g**⁹ were prepared by reaction of corresponding alcohols^{1,10,11} with acetic anhydride.



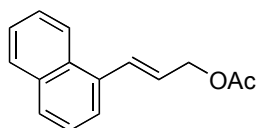
Allyl Acetate **1d**: ¹H NMR (500 MHz, CDCl₃) 2.08 (s, 3H), 5.28–5.33 (m, 2H), 6.19 (ddd, *J* = 5.5, 10.5, 16.9 Hz, 1H), 7.00 (d, *J* = 5.5 Hz, 1H), 7.45–7.55 (m, 3H), 7.59 (d, *J* = 6.8, 1H), 7.83 (d, *J* = 8.3 Hz, 1H), 7.86–7.88 (m, 1H), 8.12 (d, *J* = 8.3 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) 20.98, 73.43, 117.05, 123.61, 125.11, 125.26, 125.60, 126.17, 128.66, 128.84, 130.56, 133.75, 134.33, 135.78, 169.79.



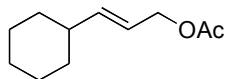
Allyl Acetate **2b**: ¹H NMR (500 MHz, CDCl₃) 2.09 (s, 3H), 3.80 (s, 3H), 4.70 (dd, *J* = 1.4 Hz, 6.4, 2H), 6.15 (dt, *J* = 6.7, 15.6 Hz, 1H), 6.59 (d, *J* = 15.6 Hz, 1H), 6.85 (m, 2H), 7.33 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) 20.94, 55.17, 65.28, 113.91, 120.75, 127.79, 128.86, 133.97, 159.51, 170.83



Allyl Acetate **2c**: ^1H NMR (500 MHz, CDCl_3) 2.13 (s, 3H), 4.76 (dd, $J = 1.25, 6.25$ Hz, 2H), 6.38 (dt, $J = 6.25, 15.5$ Hz, 1H), 6.68 (d, $J = 15.5$ Hz, 1H), 7.48 (d, $J = 8$ Hz, 2H), 7.58 (d, $J = 8.5$ Hz, 2H). ^{13}C NMR (125 MHz, CDCl_3) 20.71, 64.42, 125.43, 125.46, 126.00, 126.63, 129.66 (q, $J = 32.68$), 132.12, 139.64, 170.58.



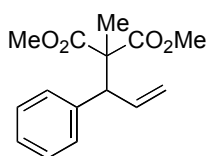
Allyl Acetate **2d**: ^1H NMR (500 MHz, CDCl_3) 2.14 (s, 3H), 4.84 (dd, $J = 1.25, 6.25$ Hz, 2H), 6.32 (dt, $J = 6.25, 15.75$ Hz, 1H), 7.42 (d, $J = 16.5$ Hz, 1H), 7.45 (d, $J = 8.00$ Hz, 1H), 7.48–7.54 (m, 2H), 7.60 (d, $J = 7.00$ Hz, 1H), 7.79 (d, $J = 8.00$ Hz, 1H), 7.84–7.86 (m, 1H), 8.19 (d, $J = 8.00$ Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) 20.74, 64.91, 123.40, 123.83, 125.31, 125.60, 125.93, 126.14, 128.14, 128.31, 130.85, 130.99, 133.33, 133.68, 170.51



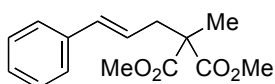
Allyl Acetate **2g**: ^1H NMR (500 MHz, CDCl_3) 1.04–1.11 (m, 2H), 1.16 (tt, $J = 3.21, 12.37$ Hz, 1H), 1.22–1.31 (m, 2H), 1.63–1.67 (m, 1H), 1.71–1.74 (m, 4H), 1.93–2.00 (m, 1H), 2.06 (s, 3H), 4.50 (d, $J = 6.4$ Hz, 2H), 5.48–5.54 (m, 1H), 5.71 (dd, $J = 6.9, 15.6$ Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) 20.99, 25.90, 26.06, 32.49, 40.28, 65.49, 121.22, 142.09, 170.82.

General Procedure for the Catalytic Allylic Alkylation of Allyl Acetates 1a-g. The reaction conditions and results are shown in Table 1. A typical procedure is given for the reaction of 1-phenyl-2-propenyl acetate (**1a**) (entry 2). To a solution of $[\text{RuCl}_2(p\text{-cymene})]_2$ (30.6 mg, 0.05 mmol), PPh_3 (26.2 mg, 0.10 mmol) in Toluene (1 mL) was added 1-phenyl-2-propenyl acetate (**1a**) (176 mg, 1.0 mmol) and dimethyl methylmalonate (219 mg, 1.5 mmol). LiHMDS (1.4 mmol, 1.4 mL of 1.0M in THF) was slowly added at 0 °C, and stirred at 60 °C for 12 h. The reaction mixture was quenched with 1N HCl (0.5 mL), then extracted with ether (3 x 2 mL). The combined organic layers were dried over MgSO_4 and concentrated *in vacuo*. The residue was chromatographed on silica gel (hexane/EtOAc = 10/1) to give 231 mg (88%) of a mixture of branch isomer **3a** and linear isomer **4a**. The ratio of **3a** and **4a** was determined to be 96 : 4 by ^1H NMR of the crude materials.

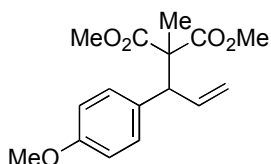
General Procedure for the Catalytic Allylic Alkylation of Allyl Acetates 2a-g. The reaction conditions and results are shown in Table 2. A typical procedure is given for the reaction of cinnamyl acetate (**2a**) (entry 3). To a solution of $[\text{RuCl}_2(p\text{-cymene})]_2$ (30.6 mg, 0.05 mmol), PPh_3 (26.2 mg, 0.10 mmol) in Toluene (1 mL) was added cinnamyl acetate (**2a**) (176 mg, 1.0 mmol) and dimethyl methylmalonate (219 mg, 1.5 mmol). NaHMDS (1.4 mmol, 1.4 mL of 1.0M in THF) was slowly added at 0 °C, and stirred at 100 °C for 12 h. The reaction mixture was quenched with 1N HCl (0.5 mL), then extracted with ether (3 x 2 mL). The combined organic layers were dried over MgSO_4 and concentrated in vacuo. The residue was chromatographed on silica gel (hexane/EtOAc = 10/1) to give 238 mg (91%) of a mixture of branch isomer **3a** and linear isomer **4a**. The ratio of **3a** and **4a** was determined to be 1 : 99 by ^1H NMR of the crude materials.



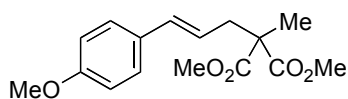
branch **3a**⁶: ^1H NMR (500 MHz, CDCl_3) 1.43 (s, 3H), 3.62 (s, 3H), 3.71 (s, 3H), 4.15 (d, $J = 8.70$ Hz, 1H), 5.09–5.15 (m, 2H), 6.32 (ddd, $J = 16.96, 10.10, 8.7$ Hz, 1H), 7.22–7.28 (m, 5H). ^{13}C NMR (125 MHz, CDCl_3) 18.37, 52.35, 54.49, 58.83, 117.75, 127.11, 128.14, 129.46, 136.84, 139.03, 171.25, 171.45.



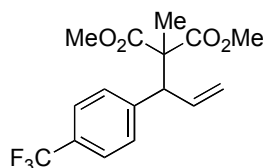
linear **4a**⁶: ^1H NMR (500 MHz, CDCl_3) 1.46 (s, 3H), 2.77 (dd, $J = 1.40, 7.33$ Hz, 2H), 3.74 (s, 6H), 6.05–6.11 (m, 1H), 6.45 (d, $J = 15.55$ Hz, 1H), 7.20–7.34 (m, 5H). ^{13}C NMR (125 MHz, CDCl_3) 20.05, 39.47, 52.53, 53.97, 124.12, 126.12, 127.39, 128.47, 134.12, 137.08, 172.32.



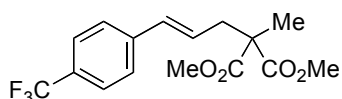
branch **3b**⁶: ^1H NMR (500 MHz, CDCl_3) 1.42 (s, 3H), 3.63 (s, 3H), 3.71 (s, 3H), 3.78 (s, 3H), 4.10 (d, $J = 8.70$ Hz, 1H), 5.07–5.13 (m, 2H), 6.28 (ddd, $J = 8.70, 8.48, 5.28$ Hz, 1H), 6.80–6.82 (m, 2H), 7.15–7.17 (m, 2H). ^{13}C NMR (125 MHz, CDCl_3) 18.23, 52.29, 53.66, 53.95, 55.06, 58.86, 113.48, 117.39, 130.43, 130.94, 137.05, 158.52, 171.27, 171.46.



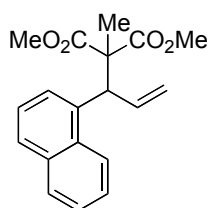
linear **4b**⁶: ¹H NMR (500 MHz, CDCl₃) 1.45 (s, 3H), 2.74 (dd, *J* = 15.0, 7.75 Hz, 2H), 3.73 (s, 6H), 3.80 (s, 3H), 5.90–5.96 (m, 1H), 6.38 (d, *J* = 15.55 Hz, 1H), 6.81–6.84 (m, 2H), 7.25–7.27 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) 19.88, 39.35, 52.37, 53.90, 55.11, 113.78, 121.65, 127.23, 129.79, 133.39, 158.96, 172.25.



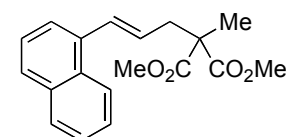
branch **3c**¹³: ¹H NMR (500 MHz, CDCl₃) 1.44 (s, 3H), 3.64 (s, 3H), 3.71 (s, 3H), 4.20 (d, *J* = 8.70 Hz, 1H), 5.09–5.19 (m, 2H), 6.30 (ddd, *J* = 16.96, 10.05, 8.70 Hz, 1H), 7.39 (d, *J* = 8.25 Hz, 2H), 7.54 (d, *J* = 8.25 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) 18.52, 52.46, 52.54, 54.28, 58.60, 118.59, 123.00, 125.01, 129.33 (d, *J*_{CF} = 32.63 Hz), 129.71, 136.01, 143.35, 170.97, 171.17



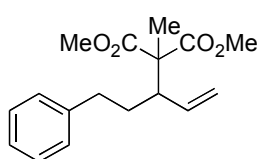
linear **4c**¹³: ¹H NMR (500 MHz, CDCl₃) 1.46 (s, 3H), 2.79 (dd, *J* = 7.33, 1.35 Hz, 2H), 3.74 (s, 6H), 6.18–6.24 (m, 1H), 6.48 (d, *J* = 16.06 Hz, 1H), 7.42 (d, *J* = 8.25 Hz, 2H), 7.54 (d, *J* = 8.25 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) 20.11, 39.45, 52.56, 53.84, 125.40, 125.43, 126.33, 127.16, 129.18 (d, *J*_{CF} = 32.31 Hz), 132.73, 140.45, 172.13



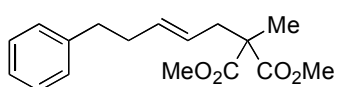
branch **3d**¹³: ¹H NMR (500 MHz, CDCl₃) 1.48 (s, 1H), 3.40 (s, 3H), 3.75 (s, 3H), 5.09–5.14 (m, 2H), 5.20 (d, *J* = 8.25 Hz, 1H), 6.42 (ddd, *J* = 16.96, 10.00, 8.25 Hz, 1H), 7.41–7.54 (m, 4H), 7.74 (dd, *J* = 7.30, 1.80 Hz, 1H), 7.83 (dd, *J* = 8.25, 1.35 Hz, 1H), 8.25 (d, *J* = 8.70 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) 19.21, 47.08, 52.26, 52.42, 59.04, 117.45, 123.26, 125.07, 125.38, 125.95, 126.17, 127.67, 128.92, 132.30, 133.97, 135.40, 137.80, 171.74, 171.76



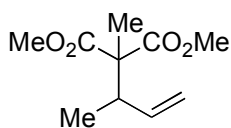
linear **4d**¹³: ¹H NMR (500 MHz, CDCl₃) 1.53 (s, 3H), 2.90 (dd, *J* = 7.55, 1.15 Hz, 2H), 3.75 (s, 6H), 6.08–6.14 (m, 1H), 7.19 (d, *J* = 15.56 Hz, 1H), 7.41–7.52 (m, 4H), 7.76 (d, *J* = 8.25 Hz, 1H), 7.83–7.85 (m, 1H), 8.05–8.07 (m, 1H). ¹³C NMR (125 MHz, CDCl₃) 20.48, 39.69, 52.58, 54.07, 65.84, 123.63, 123.98, 125.60, 125.71, 125.93, 127.52, 127.79, 128.47, 131.60, 133.51, 134.97, 172.35



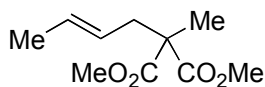
branch **3e**¹³: ¹H NMR (500 MHz, CDCl₃) 1.38 (s, 3H), 2.46-2.52 (m, 1H), 2.68-2.72 (m, 1H), 2.76 (dt, *J* = 2.05, 11.45 Hz, 1H), 3.66 (s, 3H), 3.68 (s, 3H), 5.12 (dd, *J* = 1.80, 16.96 Hz, 1H), 5.19 (dd, *J* = 1.80, 10.10 Hz, 1H), 5.64 (dt, *J* = 10.10, 16.96 Hz, 1H), 7.16-7.19 (m, 2H), 7.26-7.29 (m, 3H). ¹³C NMR (126 MHz, CDCl₃) 17.18, 31.86, 34.01, 48.85, 52.36, 57.80, 119.04, 125.76, 128.27, 128.46, 136.84, 142.04, 171.70.



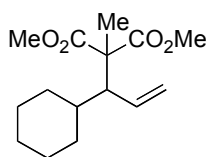
linear **4e**¹³: ¹H NMR (500 MHz, CDCl₃) 1.34 (s, 3H), 2.31 (dd, *J* = 14.68, 6.88 Hz, 2H), 2.54 (d, *J* = 7.30 Hz, 2H), 2.65 (dd, *J* = 8.25, 7.30 Hz, 2H), 3.70 (s, 6H), 5.27-5.35 (m, 1H), 5.51-5.58 (m, 1H), 7.15-7.19 (m, 3H), 7.25-7.28 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) 19.73, 34.29, 35.83, 38.93, 52.42, 53.78, 124.42, 125.76, 128.26, 128.38, 134.50, 141.72, 172.45



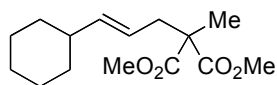
branch **3f**: ¹H NMR (500 MHz, CDCl₃) 0.98 (d, *J* = 8.00 Hz, 3H), 1.29 (s, 3H), 2.93 (six, *J* = 8.00 Hz, 1H), 3.63 (s, 3H), 3.65 (s, 3H), 4.96-5.02 (m, 2H), 5.69 (ddd, *J* = 8.00, 10.50, 17.01 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) 15.49, 16.58, 42.36, 52.34, 52.39, 57.39, 116.32, 138.74, 171.82, 171.88.



linear **4f**: ¹H NMR (500 MHz, CDCl₃) 1.37 (s, 3H), 1.64 (d, *J* = 6.00 Hz, 3H), 2.53 (d, *J* = 8.00 Hz, 2H), 3.70 (s, 6H), 5.24-5.31 (m, 1H), 5.50 (dt, *J* = 7.00, 22.00 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) 17.93, 19.66, 38.90, 52.35, 53.75, 124.73, 129.85, 172.45.



branch **3g**¹³: ¹H NMR (500 MHz, CDCl₃) 1.58 (s, 3H), 2.67 (dd, *J* = 4.00, 10.50 Hz, 1H), 3.66 (s, 3H), 3.72 (s, 3H), 5.02 (dd, *J* = 2.01, 17.50 Hz, 1H), 5.07 (dd, *J* = 2.01, 10.50 Hz, 1H), 5.66 (dt, *J* = 10.50, 17.50 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) 17.57, 54.04, 54.36, 57.92, 60.42, 118.36, 135.85, 171.95, 172.19



linear **4g**¹³: ¹H NMR (500 MHz, CDCl₃) 0.98–1.07 (m, 2H), 1.08–1.17 (m, 1H), 1.38 (s, 3H), 1.60–1.71 (m, 5H), 1.87–1.93 (m, 1H), 2.53 (d, *J* = 7.30 Hz, 2H), 3.71 (s, 6H), 5.21–5.27 (m, 1H), 5.44 (dd, *J* = 15.10, 6.85 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) 19.77, 25.92, 26.08, 32.99, 38.99, 40.73, 52.32, 53.98, 121.01, 141.55, 172.46.

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