Design of a novel G-quenched molecular beacon: A simple and efficient strategy for DNA sequence analysis

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Figure S1: Structures of the MB2-5 and MB7

General experimental:

Oligonucleotide synthesis and characterisation: All reagents for DNA synthesis were purchased from Glen Research. **MB1** was synthesized by automated DNA synthesizer. The fluorophores were post-synthetically incorporated into **MB1** to get desired molecular beacons, **MB2**, **3**, and **4**. As a target loop strands, we synthesised unmodified ODN **11**, a fully matched sequence, and ODN **8-10**, one-base-mismatched sequences at the central position of the loop sequence. ODNs were purified by reverse phase HPLC on a CHEMCO-BOND 5-ODS-H column (10 x 150 mm, elution with 50 mM ammonium formate buffer (AF), pH 7.0, linear gradient over 50 min from 3% to 40% acetonitrile at a flow rate 2.0 ml/min). ODNs containing modified nucleotides were fully digested with calf intestine alkaline phosphatase (50 U/mL), snake venom phosphodiesterase (0.15 U/mL), and P1 nuclease (50 U/mL) at 37 °C for 3 h. Digested solutions were analysed by HPLC on a CHEMCO-BOND 5-ODS-H column (4.6 x 150 mm), elution with a solvent mixture of 50 mM ammonium formate buffer (AF), pH 7.0, linear gradient over 60 min from 3% to 50% acetonitrile at a flow rate 1.0

mL/min). The concentration of each ODNs was determined by comparing peak areas with standard solution containing dA, dC, dG, and dT at a concentration of 0.1 mM. Mass spectra of ODNs purified by HPLC were determined with a MALDI-TOF mass spectroscopy, Shimadzu, AXIMA-LNR.

ODNs		MALDI-TOF	MALDI-TOF
		Mass cald. for	Mass found
1	MB1	7870.25	7870.35
2	MB2	8101.52	8101.04
3	MB3	8073.42	8073.87
4	MB4	8090.47	8090.94
5	MB5	8230.57	8230.74
5	MB7	9323.34	9323.74

 Table S1: MALDI-TOF Mass Spectral data for the ODNs

Synthesis of modified G-quenched molecular beacons (MBs): The fluorophores were post-synthetically incorporated into MB1 to get desired molecular beacons, MB2, 3, and 4. Thus, active esters (1.0 mg) were dissolved in a small amount of dry DMF (20 μ L) and added to the 5'-amino modified MBs (20 μ L) in a total volume of 150 μ L of 1.0M sodium NaHCO₃ and incubated for 8 hours at 37 °C. Purification and the characterization of the products were performed according to the standard process as described above.

Melting temperature (T_m) measurements: All T_m s of the ODNs (2.5 µM, final duplex concentration) were taken in 50 mM sodium phosphate buffers (pH 7.0) containing 100 mM sodium chloride. Absorbance *vs* temperature profiles were measured at 260 nm using a Shimadzu UV-2550 spectrophotometer equipped with a Peltier temperature controller using 1 cm path length cell. The absorbance of the samples was monitored at 260 nm from 4 °C to 90 °C with a heating rate of 1 °C/min. From these profiles, first derivatives were calculated to determine T_m values.

UV absorption measurements: ODN solutions were prepared as described in Tm measurement experiment. Absorption spectra were obtained using a Shimadzu UV-2550 spectrophotometer at room temperature using 1 cm path length cell.

Fluorescence measurements: ODN solutions were prepared as described in $T_{\rm m}$ measurement experiment. Fluorescence spectra were obtained using a Shimadzu RF-5300PC spectrophotometer at 25°C using 1cm path length cell. The excitation and the emission bandwidth was 1.5 nm.





MB1 with or without target ODNs

Figure S2: UV-visible spectra and thermal melting curve of the probe MB1 alone (MB) and with their targets (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]



MB2 with or without target ODNs

MB4 with or without target ODNs

Figure S3: UV-visible spectra of probes **MB2** and **MB4** alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]





MB4 with or without target ODNs

Figure S4: Thermal denaturation curves of probes **MB2** and **MB4** alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]



MB2 with or without target ODNs

MB4 with or without target ODNs

Figure S5: Fluorescence excitation spectra of probes MB2 and MB4 alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]





MB5 with or without target ODNs

Figure S6: UV-visible spectra of probes MB3 and MB5 alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]





MB5 with or without target ODNs

Figure S7: Thermal denaturation curves of probes MB3 and MB5 alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]





MB5 with or without target ODNs

Figure S8: Fluorescence excitation spectra of probes MB3 and MB5 alone (MBs) and with their target (ODN 8-11) [2.5 μ M, 50mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT]



Wavelength (nm)

MB7 with or without target ODNs

Figure S9: UV, thermal denaturation curves and fluorescence excitation spectra of probe MB7 alone (MBs) and with its target (ODN 8-11) [2.5 μ M, 50 mM sodium phosphate, 0.1 M sodium chloride, pH 7.0, RT].



Figure S10: Structures of the MB8 and MB9



Figure S11: Fluorescene excitation spectra of hairpin **MB8** (a) and **MB9** (b) [(2.5 μ M) and the different duplexes formed by hybridization with ODN 8-11 (2.5 μ M, 50 mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT)]. Excitation wavelength was 403 nm. "MB" denotes the hairpin states.



Figure S12: Fluorescence spectra of hairpin **MB8** (a) and **MB9** (b) [(2.5 μ M) and the different duplexes formed by hybridization with ODN 8-11 (2.5 μ M, 50 mM sodium phosphate, 0.1M sodium chloride, pH 7.0, RT)]. Excitation wavelength was 403 nm . "MB" denotes the hairpin states.