

Supporting Information

Glycosyl Tricyclic Orthoesters as Versatile Intermediates for the Preparation of Glycosyl Phosphate Building Blocks

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General Information

All chemicals used were reagent grade and used as supplied except where noted. All reactions were performed in oven-dried glassware under an inert atmosphere unless noted otherwise. Reagent grade dichloromethane (CH_2Cl_2) was passed through activated neutral alumina column prior to use. Reagent grade N,N-dimethylformamide (DMF) and methanol (MeOH) were dried over activated molecular sieves prior to use. Pyridine, triethylamine and acetonitrile were distilled over CaH_2 prior to use. Analytical thin layer chromatography (TLC) was performed on Merck silica gel 60 F_{254} plates (0.25mm). Compounds were visualized by UV irradiation or dipping the plate in a cerium sulfate-ammonium molybdate (CAM) solution. Flash column chromatography was carried out using forced flow of the indicated solvent on Fluka Kieselgel 60 (230-400 mesh).

^1H , ^{13}C and ^{31}P NMR spectra were recorded on a Varian Mercury 300 (300 MHz), Varian Gemini 300 (300 MHz), Bruker DRX500 (500 MHz) in CDCl_3 with chemical shifts referenced to internal standards CDCl_3 (7.26 ppm ^1H , 77.0 ppm ^{13}C) unless otherwise stated. ^{31}P spectra are reported in δ value relative to H_3PO_4 (0.0 ppm) as an external reference. Splitting patterns are indicated as s, singlet; d, doublet; t, triplet; q, quartet; brs, broad singlet for ^1H NMR data. High-resolution mass spectral (HRMS) analyses were performed by the MS-service at the Laboratory for Organic Chemistry (LOC) at ETH Zürich. High-resolution MALDI and ESI mass spectra were run on an IonSpec Ultra instrument. IR spectra were recorded on a Perkin-Elmer 1600 FTIR spectrometer. Optical rotations were measured using a Perkin-Elmer 241 polarimeter. Elemental analyses were performed by the Microanalytical Laboratory of the LOC, ETH Zürich. The

automated synthesis was performed on an ABI 431A peptide synthesizer with a custom-made jacketed glass reaction vessel.

Preparation of Glycosyl Tricyclic Orthoester via Acid-Catalyzed Intramolecular Trans-orthoesterification of Glycosyl 1,2-Orthoester

3,4-*O*-Benzyl α -D-mannopyranose 1,2,6-orthobenzoate (**5a**):

From mannosyl 1,2-orthoester **2a**. To a solution of **2a** [1] (195 mg, 0.65 mmol) in CH₃CN (4 mL) in the presence of activated 4Å MS (220 mg) was added CSA (35 mg, 0.15 mmol) at room temperature. The mixture was stirred for 12 h when additional amount of CSA (15 mg) was added. Further stirred for 12 h, the reaction was quenched by the addition of Et₃N (0.15 mL). Filtration through a pad of Celite, followed by removal of the solvents gave the crude tricyclic orthoester. This crude material was dissolved in DMF (4.5 mL), added BnBr (0.24 mL, 2.0 mmol) and NaH (48 mg, 2.0 mmol) at 0 °C. The mixture was gradually warmed up to room temperature during 12 h. Excess NaH was quenched by the addition of MeOH. Typical aqueous workup, followed by silica gel column chromatography gave **5a** (244 mg, 85%) as a white solid.

From mannosyl 1,2-orthoester **2b**. To a solution of **2b** [2] (440 mg, 1.34 mmol) in CH₃CN (8 mL) in the presence of activated 4Å MS (220 mg) was added CSA (35 mg, 0.15 mmol) at room temperature. The mixture was stirred for 12 h before quenched with Et₃N (0.13 mL). Filtration through a pad of Celite, followed by removal of the solvents gave the crude tricyclic orthoester. This crude material was dissolved in DMF (7.5 mL), added BnBr (0.5 mL, 4.20 mmol) and NaH (90 mg, 3.9 mmol) at 0 °C. The mixture was gradually warmed up to room temperature and the reaction completed after 3 h. Excess NaH was quenched by the addition of MeOH. Typical aqueous workup, followed by silica gel column chromatography gave **5a** (578 mg, 95%) as a white solid. *R*_f 0.40 (Hexanes/EtOAc = 4 : 1); [α]_D²⁵ = +2.1 (*c* = 1.2, CHCl₃); m.p.=110–111 °C; ¹H NMR (300 MHz, CDCl₃) δ 3.74 (dd, *J* = 12.9, 3.6 Hz, 1H), 3.77 (dd, *J* = 7.5, 2.4 Hz, 1H), 4.18 (d, *J* = 12.9 Hz, 1H), 4.21 (dd, *J* = 7.5, 1.3 Hz, 1H), 4.24 (dd, *J* = 3.6, 1.3 Hz 1H), 4.64 (dd, *J* = 7.5, 1.3 Hz, 1H), 4.66 (d, *J* = 12.0 Hz, 1H), 4.80 (d, *J* = 12.0 Hz, 1H), 4.84 (app s, 2H), 5.86 (d, *J* = 5.7 Hz, 1H), 7.25-7.44 (m, 13H), 7.62-7.65 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 70.6,

72.4, 72.6, 73.2, 74.5, 77.4, 79.3, 99.7, 121.9, 126.0, 127.8, 127.9, 128.0, 128.4, 128.5, 129.3, 137.1, 138.0; Anal. Calcd for C₂₇H₂₆O₆; C, 72.63; H, 5.87; Found: C, 72.48; H, 5.86.

3,4-*O*-Benzyl α -D-mannopyranose 1,2,6-orthoacetate (5b**):**

To a solution of **2d** [3] (500 mg, 1.91 mmol) in CH₃CN (10 mL) in the presence of activated 4Å MS (300 mg) was added CSA (44 mg, 0.19 mmol) at room temperature. The mixture was stirred for 12 h before quenched with Et₃N (0.20 mL). Filtration through a pad of Celite, followed by removal of the solvents gave the crude tricyclic orthoester. This crude material was dissolved in DMF (10 mL), added BnBr (0.68 mL, 5.70 mmol) and NaH (137 mg, 5.7 mmol, 60% in mineral oil) at 0 °C. The mixture was gradually warmed up to room temperature and the reaction completed after 3 h. Excess NaH was quenched by the addition of MeOH. Typical aqueous workup, followed by silica gel column chromatography gave **5b** (700 mg, 95%) as a colorless syrup. R_f 0.28 (Hexanes/EtOAc = 4 : 1); [α]_D²⁵ = -8.1 (*c* = 7.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.65 (s, 3H), 3.52 (dd, *J* = 13.0, 3.3 Hz, 1H), 3.66 (dd, *J* = 7.5, 2.4 Hz, 1H), 3.98 (app d, *J* = 13.0 Hz, 1H), 4.04 (dd, *J* = 7.5, 1.2 Hz, 1H), 4.06 (dd, *J* = 3.3, 1.2 Hz 1H), 4.45 (dd, *J* = 6.0, 2.4 Hz, 1H), 4.60 (d, *J* = 12.0 Hz, 1H), 4.76 (d, *J* = 12.0 Hz, 1H), 4.80 (app s, 2H), 5.68 (d, *J* = 6.0 Hz, 1H), 7.25-7.42 (m, 10H); ¹³C NMR (75 MHz, CDCl₃) δ 23.8, 69.8, 72.4, 73.1, 73.8, 77.2, 79.1, 99.4, 122.9, 127.7, 127.8, 127.9, 128.3, 128.4, 137.9, 138.0; HRMS-MALDI (*m/z*): [M+Na]⁺ calcd for C₂₂H₂₄O₆, 407.1471; Found: 407.1459.

3-*O*-Benzyl α -D-xylopyranose 1,2,4-orthoacetate (5c**):**

To a solution of **2e** [4] (348 mg, 1.50 mmol) in CH₃CN (40 mL) was added CSA (42 mg, 0.18 mmol) at room temperature. The mixture was stirred for 12 h before quenched with Et₃N (0.15 mL). Filtration through a pad of Celite, followed by removal of the solvents gave the crude tricyclic orthoester. Two-third portion of this crude material was dissolved in DMF (3 mL), added BnBr (0.13 mL, 1.1 mmol) and NaH (32 mg, 1.3 mmol, 60% in mineral oil) at 0 °C. The mixture was gradually warmed up to room temperature and the reaction completed after 5 h. Excess NaH was quenched by the addition of MeOH. Typical aqueous workup, followed by silica gel column chromatography gave **5c** (194 mg, 73%) as a syrup. R_f 0.53 (CH₂Cl₂/EtOAc = 1 : 1). [α]_D²⁵ = +44.0 (*c* = 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.63 (s, 3H), 3.91–3.96 (m,

1H), 4.12–4.19 (m, 3H), 4.35–4.39 (m, 1H), 4.58–4.71 (m, 2H), 5.75 (d, $J = 4.8$ Hz, 1 H), 7.28–7.40 (m, 5H); ^{13}C NMR (75 MHz, CDCl_3) δ 20.3, 65.5, 69.9, 71.4, 72.2, 72.3, 97.9, 118.2, 127.7 (2x), 128.1, 128.5 (2x), 137.4; HRMS-MALDI (m/z): $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{14}\text{H}_{16}\text{O}_5$, 287.0889; Found: 287.0880.

3-*O*-Benzyl α -D-xylopyranose 1,2,4-orthobenzoate (5d**):**

To a solution of **2f** [5] (2.65 g, 9.89 mmol) in CH_3CN (50 mL) was added silica gel (10 g, Fluka Kieselgel 50), and the mixture was stirred at room temperature for 12 h. The reaction mixture was filtered through a short pad of Celite and the solvent was removed in vacuo. Purification by silica gel chromatography gave tricyclic orthoester α -D-xylopyranose 1,2,4-orthobenzoate (1.33 g, 60%) as a colorless solid. R_f 0.31 ($\text{CH}_2\text{Cl}_2/\text{EtOAc} = 1 : 1$). $[\alpha]_D^{rt} = +71.5$ ($c = 1.0$, CHCl_3); m.p. = 145–147 °C; ^1H NMR (300 MHz, CDCl_3) δ 2.34 (d, $J = 10.2$ Hz, 1H), 4.24–4.29 (m, 1H), 4.28 (dd, $J = 12.0$, 4.5 Hz, 1H), 4.44 (app d, $J = 12.0$ Hz, 1H), 4.49 (td, $J = 4.5$, 2.0 Hz, 1H), 4.57 (app dt, $J = 4.5$, 2.0 Hz, 1H), 5.96 (d, $J = 4.8$ Hz, 1H), 7.36–7.44 (m, 3H), 7.66–7.70 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 63.8, 65.9, 73.1, 74.5, 98.0, 117.7, 125.9, 128.1, 129.7, 133.5; Anal. Calcd for $\text{C}_{12}\text{H}_{12}\text{O}_5$; C, 61.02; H, 5.12; Found: C, 60.98; H, 5.05.

To a solution of α -D-xylopyranose 1,2,4-orthobenzoate (777 mg, 3.29 mmol) in DMF (12 mL) was added BnBr (0.78 mL, 6.58 mmol) and NaH (158 mg, 6.58 mmol, 60% in mineral oil) at 0 °C. The mixture was allowed to warm up to room temperature gradually and stirred overnight. Excess NaH was quenched by the addition of MeOH. Typical aqueous workup, followed by silica gel column chromatography gave **5d** (1.11 g, quant.) as a white solid. R_f 0.49 (Hexanes/EtOAc = 4 : 1). $[\alpha]_D^{rt} = +43.8$ ($c = 1.4$, CHCl_3); m.p. = 54–55 °C; ^1H NMR (300 MHz, CDCl_3) δ 4.10–4.12 (m, 1H), 4.27 (dd, $J = 12.0$, 6.9 Hz, 1H), 4.34 (dd, $J = 12.0$, 1.0 Hz, 1H), 4.38 (td, $J = 4.2$, 2.4 Hz, 1H), 4.58 (dt, $J = 4.8$, 2.0 Hz, 1H), 4.67 (d, $J = 12.1$ Hz, 1H), 4.74 (d, $J = 12.1$ Hz, 1H), 5.94 (d, $J = 5.1$ Hz, 1H), 7.33–7.42 (m, 8H), 7.65–7.68 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 65.7, 70.0, 72.2, 72.4, 72.7, 98.1, 117.3, 127.8, 128.1, 128.2, 128.6, 129.8, 133.7, 137.4; Anal. Calcd for $\text{C}_{19}\text{H}_{18}\text{O}_5$; C, 69.93; H, 5.56; Found: C, 69.66; H, 5.57.

3,6-Di-*O*-benzyl α -D-glucopyranose 1,2,4-orthoacetate (5e**):**

To a solution of **2g** [6] (89.9 mg, 0.381 mmol) in acetonitrile (1.9 mL) was added 4Å AW 300 MS, and the mixture was stirred at 40 °C for 14 h, then at 60 °C for 3 h. The reaction mixture was

filtered through a short pad of Celite and the solvent was removed in vacuo. In a separate flask, NaH (116 mg, 60% in mineral oil) was washed with hexane twice and DMF (0.45 mL) was added. To this suspension of NaH in DMF was added the crude tricyclic orthoester mixture in DMF (1 mL) at 0°C, and BnBr (345 μ L, 2.9 mmol) was added dropwise. The reaction mixture was stirred at room temperature for 13 h, and H₂O was added to quench the reaction. After extraction with diethyl ether, combined organic layer was washed with brine, and dried over magnesium sulfate. Filtration, evaporation, and purification by silica gel chromatography gave **5e** as a colorless syrup (73.0 mg, 50%). $[\alpha]_D^{25} = +21.1$ ($c = 0.82$, CHCl₃) ¹H NMR (300 MHz, CDCl₃) δ 1.63 (s, 3H), 3.72 (dd, $J = 9.6, 7.8$ Hz, 1H), 3.81 (dd, $J = 9.6, 6.6$ Hz, 1H), 3.97 (dt, $J = 4.5$ Hz, 1H), 4.26 (dd, $J = 4.5, 2.1$ Hz, 1H), 4.40 (dt, $J = 4.8, 2.1$ Hz, 1H), 4.48 (d, $J = 12.3$ Hz, 1H), 4.53 (brs, 2H), 4.63 (d, $J = 12.0$ Hz, 1H), 4.65 (t, $J = 6.9$ Hz, 1H), 5.80 (d, $J = 4.8$ Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 20.4, 69.9, 71.1, 71.4, 72.0, 72.3, 73.2, 75.8, 97.8, 118.8, 127.5, 127.6, 127.7, 128.0, 128.3, 128.5, 137.2, 137.9; HRMS-MALDI (m/z): $[M+Na]^+$ Calcd for C₂₂H₂₄O₆, 407.1471; Found: 407.1467.

α -D-Glucopyranose 1,2,4-orthoacetate

α -D-Glucopyranose 1,2,4-orthoacetate can be purified with silica gel chromatography, before being subjected to benzylation conditions. Spectral data are as follows.

$[\alpha]_D^{25} = +110$ ($c = 0.5$, MeOH); m.p. = 119-120 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.65 (s, 3H), 3.86 (dd, $J = 12.0, 3.3$ Hz, 1H), 4.02 (dd, $J = 12.0, 4.2$ Hz, 1H), 4.17 (d, $J = 3.3$ Hz, 1H), 4.28 (dd, $J = 4.5, 2.1$ Hz, 1H), 4.38 (dt, $J = 4.8, 2.1$ Hz, 1H), 4.59 (t, $J = 3.6$ Hz, 1H), 5.85 (d, $J = 4.8$ Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 20.3, 62.3, 63.8, 74.2, 75.1, 76.7, 97.6, 119.0; HRMS-ESI (m/z): $[M]^-$ calcd for C₈H₁₁O₆, 203.0556; Found: 203.0559.

Opening of Glycosyl Tricyclic Orthoester with Dibutyl Phosphate: General Procedure

To a solution of suitably protected glycosyl tricyclic orthoester (1.0 equiv) in either CH₂Cl₂ or CH₃CN was added freshly activated 4Å molecular sieves. The mixture was stirred for 15 min stirring at room temperature, before added dibutyl phosphate (6 equiv.) in one portion. After completion of the reaction (TLC analysis), the reaction was cooled to 0°C and triethylamine (10 equiv) was added. The solution was warmed to room temperature and filtered off through a pad

of Et₃N-deactivated silica gel. The resulting mixture was purified by flash silica column chromatography.

Dibutyl-(2-*O*-benzoyl-3,4-di-*O*-benzyl- α -D-mannopyranosyl) phosphate (6a):

General procedure with mannosyl tricyclic orthoester **5a** (51 mg, 0.11 mmol), dibutyl phosphate (136 μ L, 0.68 mmol), 4Å MS (30 mg), CH₂Cl₂ (1.5 mL), room temperature, 24 h, gave **6a** (62 mg, 83%) as a colorless syrup. *R*_f 0.32 (Hexanes/EtOAc = 1:1); [α]_D^{r.t.} = +0.44 (*c* = 2.5, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.92-0.98 (m, 6H), 1.35-1.49 (m, 4H), 1.63-1.73 (m, 4H), 2.16 (t, *J* = 6.9 Hz, 1H), 3.81-3.92 (m, 2H), 3.94-4.16 (m, 7H), 4.60 (d, *J* = 11.7 Hz, 1H), 4.68 (d, *J* = 10.8 Hz, 1H), 4.80 (d, *J* = 11.7 Hz, 1H), 4.92 (d, *J* = 10.8 Hz, 1H), 5.42 (app t, *J* = 2.4 Hz, 1H), 5.77 (dd, *J* = 6.3, 2.0 Hz, 1H), 7.24-7.37 (m, 10H), 7.45-7.51 (m, 2H), 7.58-7.64 (m, 1H), 8.06-8.09 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 13.7, 18.8, 32.3, 61.6, 68.1, 68.6, 71.9, 73.3, 73.9, 75.4, 77.3, 95.6, 127.7, 127.8, 127.9, 128.0, 128.3, 128.4, 128.5, 129.4, 129.9, 133.4, 137.6, 137.9, 165.1; ³¹P NMR (121MHz, CDCl₃): δ -2.19. HRMS-MALDI (*m/z*): [M+Na]⁺ Calcd for C₃₅H₄₅O₁₀P, 679.2643; Found: 679.2639.

Dibutyl-(2-*O*-acetyl-3,4-di-*O*-benzyl- α -D-mannopyranosyl) phosphate (6b):

General procedure with microwave irradiation at 120 °C, using mannose tricyclic orthoester **5b** (41 mg, 0.11 mmol), 4Å MS (50 mg), dibutyl phosphate (0.38 mL, 0.64 mmol), CH₃CN (4 mL), 5 min, gave **6b** as a colorless oil (38.2 mg, 60%), *R*_f 0.18 (Hexanes/EtOAc = 1:1); [α]_D^{r.t.} = +22.7 (*c* = 1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.88–0.99 (m, 6H), 1.32–1.48 (m, 4H), 1.59–1.72 (m, 4H), 2.01 (brs, 1H), 2.15 (s, 3H), 3.73–3.91 (m, 4H), 3.95–4.13 (m, 5H), 4.48–4.77 (m, 3H), 4.88–4.95 (m, 1H), 5.42 (dd, *J* = 3.0, 2.4 Hz, 1H), 5.62 (dd, *J* = 6.2, 1.8 Hz, 1H), 7.27–7.38 (m, 10H); ¹³C NMR (75 MHz, CDCl₃) δ 13.7, 18.7, 21.0, 32.2, 32.3, 61.6, 68.0, 68.1, 68.3, 72.0, 73.4, 73.7, 75.3, 77.1, 95.5, 95.6, 127.7 (2x), 127.9 (2x), 128.0 (2x), 128.3 (2x), 128.4 (2x), 137.5, 137.9, 169.7; ³¹P NMR (121MHz, CDCl₃): δ -2.55; IR (film) ν_{max} 3372, 2961, 2875, 1748, 1456, 1368, 1233, 1029, 956 cm⁻¹; HRMS-MALDI (*m/z*): [M+Na]⁺ Calcd for C₃₀H₄₃O₁₀P, 617.2486; Found: 617.2484.

Dibutyl-(2-*O*-acetyl-3-*O*-benzyl- α -D-xylopyranosyl) phosphate (6c):

General procedure with xylose tricyclic orthoester **5c** (51 mg, 0.19 mmol), dibutyl phosphate (0.23 mL, 1.16 mmol), 4Å MS (30 mg), CH₂Cl₂ (1 mL), room temperature, 24 h, gave **6c** as a colorless syrup (69 mg, 65%). R_f 0.22 (Hexanes/EtOAc = 1:1); $[\alpha]_D^{r.t.} = +63.2$ (*c* = 1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.89–1.00 (m, 6H), 1.32–1.49 (m, 4H), 1.60–1.74 (m, 4H), 2.05 (s, 3H), 2.32 (brs, 1H), 3.68–3.88 (m, 4H), 3.97–4.17 (m, 4H), 4.67–4.90 (m, 3H), 5.73 (dd, *J* = 6.6, 3.3 Hz, 1H), 7.23–7.41 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ 13.5, 13.5, 18.5, 18.5, 20.7, 32.0, 32.1, 63.0, 67.7, 67.8, 69.1, 72.2, 72.3, 74.9, 79.1, 94.5, 94.5, 127.6 (2x), 127.9, 128.5 (2x), 138.1, 169.9; ³¹P NMR (121MHz, CDCl₃): δ –2.07; HRMS-MALDI (*m/z*): [M+Na]⁺ Calcd for C₂₂H₃₅O₉, 497.1911; Found: 497.1918.

Dibutyl-(2-*O*-benzoyl-3-*O*-benzyl-α-D-xylopyranosyl) phosphate (6d):

General procedure with xylosyl tricyclic orthoester **5d** (56 mg, 0.17 mmol), dibutyl phosphate (204 μL, 1.03 mmol), 4Å MS (30 mg), CH₂Cl₂ (2.0 mL), room temperature, 24 h gave **6d** (82 mg, 90%) as a colorless solid. R_f 0.51 (Hexanes/EtOAc = 1:2); m.p. = 103–104 °C; $[\alpha]_D^{r.t.} = +80.7$ (*c* = 2.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.79–0.88 (m, 6H), 1.20–1.32 (m, 4H), 1.42–1.56 (m, 4H), 2.69 (s, 1H), 3.80–4.00 (m, 8H), 4.73 (d, *J* = 11.7 Hz, 1H), 4.85 (d, *J* = 11.7 Hz, 1H), 5.11 (app dt, *J* = 9.6, 3.3 Hz, 1H), 5.86 (dd, *J* = 6.9, 3.3 Hz, 1H), 7.23–7.27 (m, 5H), 7.42–7.46 (m, 2H), 7.56–7.61 (m, 1H), 8.03–8.06 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 13.7, 18.6, 32.2, 63.2, 68.0, 69.6, 73.1, 75.4, 79.5, 94.9, 128.1, 128.2, 128.7, 129.5, 130.0, 133.7, 138.1, 165.9; ³¹P NMR (121MHz, CDCl₃): δ –1.83; Anal. Calcd for C₂₇H₃₇O₉P; C, 60.44; H, 6.95; Found: C, 60.55; H, 6.75.

Dibutyl-(2-*O*-acetyl-3,4-di-*O*-benzyl-β-D-glucopyranosyl) phosphate (6e):

General procedure with glucosyl tricyclic orthoester **5e** (46 mg, 0.12 mmol), dibutyl phosphate (142 μL, 0.72 mmol), 4Å MS (23 mg), CH₃CN (1.2 mL), room temperature, 5 h gave **6e** (69 mg, 90%) as a colorless syrup. $[\alpha]_D^{r.t.} = +61.0$ (*c* = 1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.84–0.93 (m, 6H), 1.25–1.42 (m, 4H), 1.53–1.64 (m, 4H), 1.97 (s, 3H), 3.51–3.59 (m, 2H), 3.63–3.79 (m, 3H), 3.89–4.06 (m, 4H), 4.51 (d, *J* = 12.0 Hz, 1H), 4.56 (d, *J* = 12.0 Hz, 1H), 4.69 (d, *J* = 12.0 Hz, 1H), 4.79 (d, *J* = 12.0 Hz, 1H), 5.03 (app t, *J* = 9.0 Hz, 1H), 5.18 (app t, *J* = 7.2 Hz, 1H), 7.24–7.41 (m, 10H). ¹³C NMR (75 MHz, CDCl₃) δ 13.6, 18.7, 20.9, 32.1, 68.0, 69.6, 71.3, 72.7, 73.6, 74.6, 75.0, 81.8, 96.5, 127.5, 127.7, 128.3, 128.4, 137.5, 138.0, 169.3; ³¹P NMR (121MHz,

CDCl₃): δ -2.59; HRMS-MALDI (m/z): [M+Na]⁺ Calcd for C₃₀H₄₃O₁₀P, 617.2492; Found: 617.2500.

Dibutyl 2-*O*-benzoyl-3,4-di-*O*-benzyl-6-*O*-(9-fluorenylmethoxycarbonyl)- α -D-mannopyranosyl phosphate (4a):

Following the general procedure for the selective opening of glycosyl tricyclic orthoester, mannosyl orthoester **5a** (131 mg, 0.29 mmol) was treated with dibutyl phosphate (291 μ L, 1.47 mmol) in CH₂Cl₂ (3.0 mL) at room temperature for 24 h. To this mixture at 0 °C was added pyridine (0.47 mL, 5.8 mmol) and FmocCl (150 mg, 0.58 mmol). The mixture was stirred for 2 h before subjected to typical aqueous workup. The crude material was purified by silica gel column chromatography to give the target compound **4a** (236 mg, 85%) as a syrup. R_f 0.55 (Hexanes/EtOAc = 2 : 1). $[\alpha]_D^{25} = +6.4$ ($c = 1.8$, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 0.92-0.98 (m, 6H), 1.36-1.50 (m, 4H), 1.65-1.73 (m, 4H), 4.00-4.21 (m, 7H), 4.27 (t, $J = 7.5$ Hz, 1H), 4.37-4.48 (m, 4H), 4.62 (d, $J = 11.1$ Hz, 1H), 4.64 (d, $J = 10.8$ Hz, 1H), 4.84 (d, $J = 11.1$ Hz, 1H), 4.94 (d, $J = 10.8$ Hz, 1H), 5.72 (app t, $J = 2.4$ Hz, 1H), 5.80 (dd, $J = 6.3, 1.8$ Hz, 1H), 7.26-7.45 (m, 15H), 7.52-7.65 (m, 3H), 7.78 (d, $J = 7.2$ Hz, 2H), 8.13 (d, $J = 7.2$ Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 13.7, 18.9, 32.4, 46.8, 66.4, 68.1, 68.3, 70.1, 71.5, 71.8, 73.1, 75.4, 77.3, 95.4, 120.0, 125.1, 127.2, 127.8-128.4, 129.4, 129.9, 133.4, 137.4, 137.6, 141.2, 143.1, 143.3, 155.0, 165.1; ³¹P NMR (121 MHz, CDCl₃) δ -2.23; Anal. Calcd for C₅₀H₅₅O₁₂P; C, 68.33; H, 6.31; Found: C, 68.23; H, 6.21.

Multigram synthesis of building block 5a from D-mannose (Scheme 3):

A solution of benzoyl chloride (193 mL, 1.665 mol) in pyridine (600 mL) was cooled to 0 °C in an ice-water bath. D-mannose (50 g, 0.278 mol) was added to the cooled reaction solution in small portions over a period of 30 min. The reaction mixture was allowed to warm to room temperature and the solution was kept stirring at room temperature for 12 h. The solvent was removed *in vacuo* and the residual material was extracted by EtOAc and 1 N HCl (aq.). The combined organic layer was washed with saturated NaHCO₃ (aq.) and concentrated *in vacuo*. The

crude product was recrystallized in hot EtOH to yield the perbenzoylated D-mannose as a white solid.

Perbenzoylated D-mannopyranoside (65.5 g, 99.32 mmol) was treated with acetic anhydride (14 mL, 150 mmol) and a solution of HBr in AcOH (100 mL of 33% solution, 579 mmol). The reaction was stirred at room temperature for 24 h, then poured in ice-cold water in a 500 mL separatory funnel, and extracted by CH₂Cl₂. The combined organic layer was concentrated *in vacuo* at room temperature to afford 1-bromo-2,3,4,6-*O*-benzoyl- α -D-mannopyranoside. The product obtained from this step was directly treated with 2,6-lutidine (46.27 mL, 397 mmol) and allyl alcohol (135 mL, 1986 mmol). The reaction was stirred at room temperature for 20 h and concentrated *in vacuo*. The crude product was co-evaporated with toluene (3x), dried under high vacuum for 12 h, dissolved in CH₂Cl₂, washed with water, and concentrated *in vacuo* to obtain a semi-solid product which was used in the next step without further purifications.

The following reactions were carried out in 15 to 50 mmol scales. The protected bicyclic orthoester crude product (ca 50 mmol) was dissolved in 1:1 THF/MeOH and treated with a freshly prepared solution of NaOMe in MeOH (metal Na in MeOH, 0.05 eq.) at reflux temperature for 8 h. The reaction mixture was concentrated *in vacuo*, re-dissolved in CH₂Cl₂ and filtered to a silica gel plug to remove methyl benzoate. Without further purifications the triol-bicyclic orthoester crude product was dissolved in CH₃CN and treated with camphor sulfonic acid (0.15 eq.) and 4Å molecular sieves (1x the mass of the crude product) at room temperature for 24 h. The reaction mixture was neutralized by addition of Et₃N and filtered to remove 4Å molecular sieves and concentrated *in vacuo*. The tricyclic orthoester crude product was dissolved in DMF and cooled to 0 °C before BnBr (3 eq.) and NaH (3 eq.) were added to the solution consecutively. The reaction mixture was allowed to warm to room temperature and kept stirring at room temperature for 12 h. The reaction mixture was extracted with Et₂O and water and washed with brine. The combined organic layer was concentrated *in vacuo*, and recrystallized in EtOAc and hexanes to obtain the title compound as a white solid (70% - 80%).

Automated Modules:

Module A: The resin is washed 6 times with THF for 15 sec. each.

Module B: The resin is washed with DCM for 15 sec. followed by hexanes. Repeated 6 times.

Module C: The resin is washed 6 times with DCM for 15 sec. each.

Module D: The building block (5 eq., 0.125 mmol in 1.0 mL DCM) is delivered to the reaction vessel containing the resin. The mixture is allowed to cool for 3 min. (with vortex for 30 sec. followed by standing for 30 sec.). TMSOTf (5 eq., 0.125 mmol, in 1.0 mL DCM) is added to the reaction vessel with vortex in two portions, with 2 min. interval. The reaction mixture is then left for 45 min. (with vortex for 30 sec. followed by standing for 30 sec.). After that time, the solution is drained and the resin is washed once with DCM.

Module E: The resin is submitted to piperidine (20% v/v in DMF, 2 mL) for 5 min. (with vortex for 30 sec., followed by standing for 30 sec.). After that time, the solution is drained and the resin is submitted to the same conditions twice.

Module F: The resin is washed 6 times with acetic acid (0.2 M in DCM) for 15 sec. each.

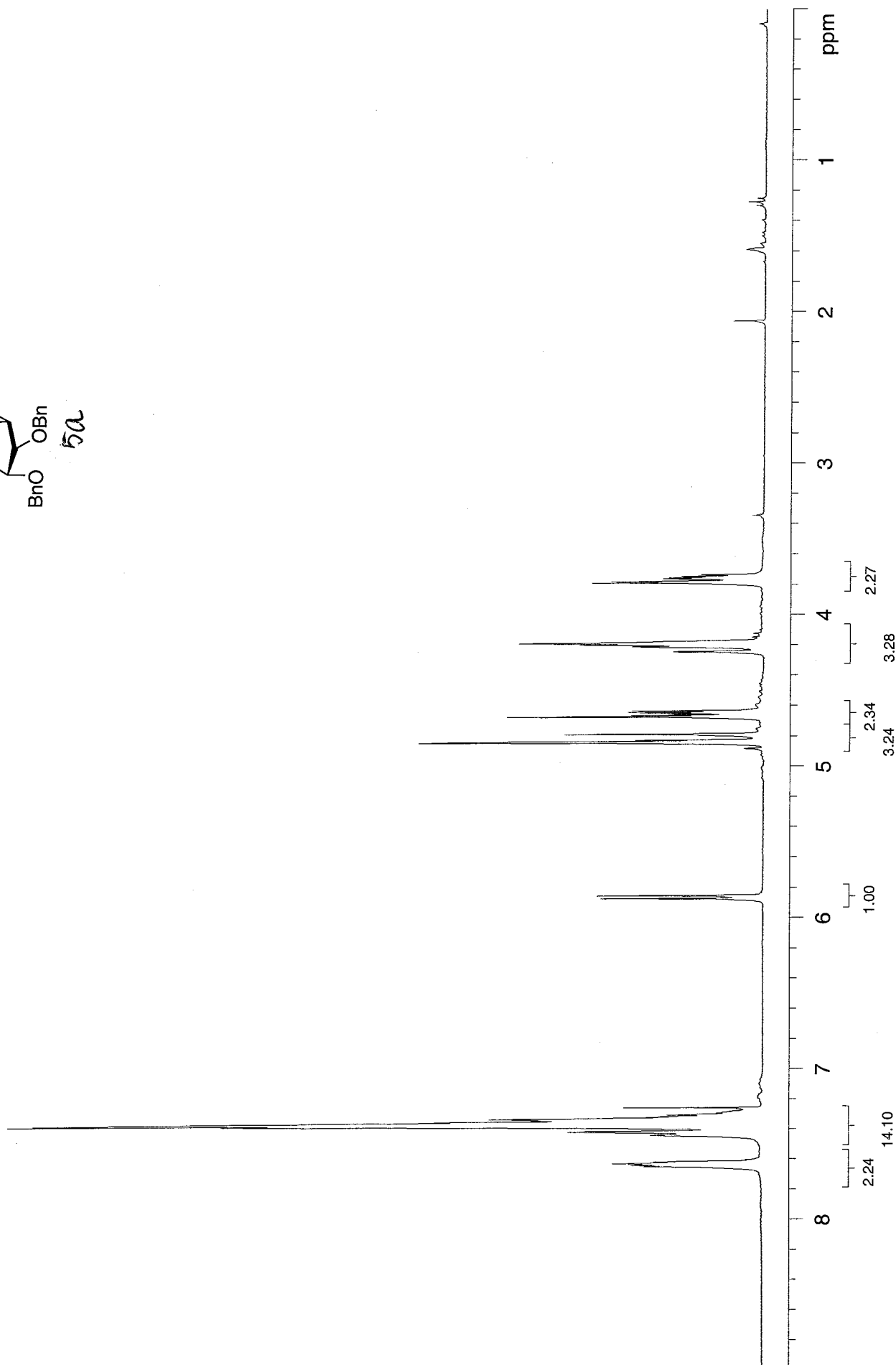
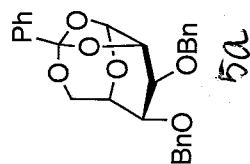
Automated synthesis of α (1-6) hexamannoside 7:

The octenediol resin (104 mg, 26 μ mol) was loaded in the reaction vessel and the modules A, B, and C were performed at room temperature. The reaction vessel was cooled to -15°C and modules D and C were performed. The reaction vessel was warmed to room temperature and modules A, C, E, C, F, A, B, and C were performed to complete the first cycle. This cycle was executed six times to furnish the resin-bound hexasaccharide. The resin was washed manually with alternating DCM and Methanol and dried under high vacuum overnight. The resin was swelled in 2 mL DCM and treated with 2 mg of Grubb's first generation catalyst. The flask was put under an atmosphere of ethylene and gently stirred overnight. The resin was washed six times with 2 mL of DCM. The washing solution was concentrated and loaded on a silica gel column. Elution with 20-30% ethyl acetate/hexanes furnished the desired hexamannoside **7** (57.5 mg, 80%). $[\alpha]_D^{25} = +44.8$ ($c = 0.83$, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.18-8.07 (m, 12H), 7.55-7.47 (m, 18H), 7.35-7.09 (m, 60H), 5.85-5.72 (m, 6H), 5.65-5.64 (m, 1H), 5.11-4.70 (m, 19H), 4.58 (t, $J=10.5$ Hz, 2H), 4.46-4.32 (m, 9H), 4.13-3.38 (m, 34H), 2.18-2.06 (m, 2H), 1.69-1.57 (m, 2H), ; ^{13}C NMR (75 MHz, CDCl_3) δ 165.9, 165.6, 165.6, 165.5, 138.6, 138.5, 138.4, 138.0, 137.9, 137.7, 137.6, 137.6, 137.6, 133.3, 130.0, 129.9, 128.7, 128.6, 128.5, 128.4, 128.2, 128.2,

128.2, 128.1, 128.1, 127.7, 127.7, 127.4, 127.4, 127.3, 127.2, 127.2, 115.1, 98.5, 98.5, 98.4, 98.3, 98.2, 98.0, 78.7, 78.4, 78.3, 78.2, 78.0, 77.7, 77.3, 75.2, 75.1, 75.1, 74.2, 74.0, 73.8, 73.7, 72.1, 71.7, 71.4, 71.2, 71.0, 70.8, 69.1, 68.6, 68.4, 67.4, 66.1, 65.8, 65.5, 61.9, 30.3, 28.6; HRMS-MALDI (m/z): $[M+Na]^+$ Calcd for $C_{167}H_{166}O_{37}$, 2786.100; Found: 2786.092.

Reference:

- [1] Hölemann, A.; Stocker, B. L.; Seeberger, P. H. *J. Org. Chem.* **2006**, *71*, 8071-8088.
- [2] Kwon, Y. U.; Soucy, R. L.; Snyder, D. A.; Seeberger, P. H. *Chem. Eur. J.* **2005**, *11*, 2493-2504.
- [3] (a) Paulsen, H.; Helpap, B. *Carbohydr. Res.* **1991**, *216*, 289-313; (b) Lindhorst, T. K. *J. Carbohydr. Chem.* **1997**, *16*, 237-243.
- [4] Bochkov, A. F.; Voznyi, Y. V.; Chernetskii, V. N.; Dashunin, V. M.; Rodionov, A. V. *Izv. Akad. Nauk SSSR, Ser. Khim.* **1975**, 420-423.
- [5] Bochkov, A. F.; Obruchnikov, I. V.; Kochetkov, N. K. *Izv. Akad. Nauk SSSR, Ser. Khim.* **1971**, 1291-1295.
- [6] (a) Bakinovskii, L. V.; Bairamova, N. E.; Tsvetkov, Yu. E.; Betaneli, V. I. *Carbohydr. Res.* **1981**, *98*, 181-93; (b) Du, Y.; Zhang, M.; Kong, F. *J. Chem. Soc., Perkin Trans I* **2001**, 2289-2293.



XL12-195-columned

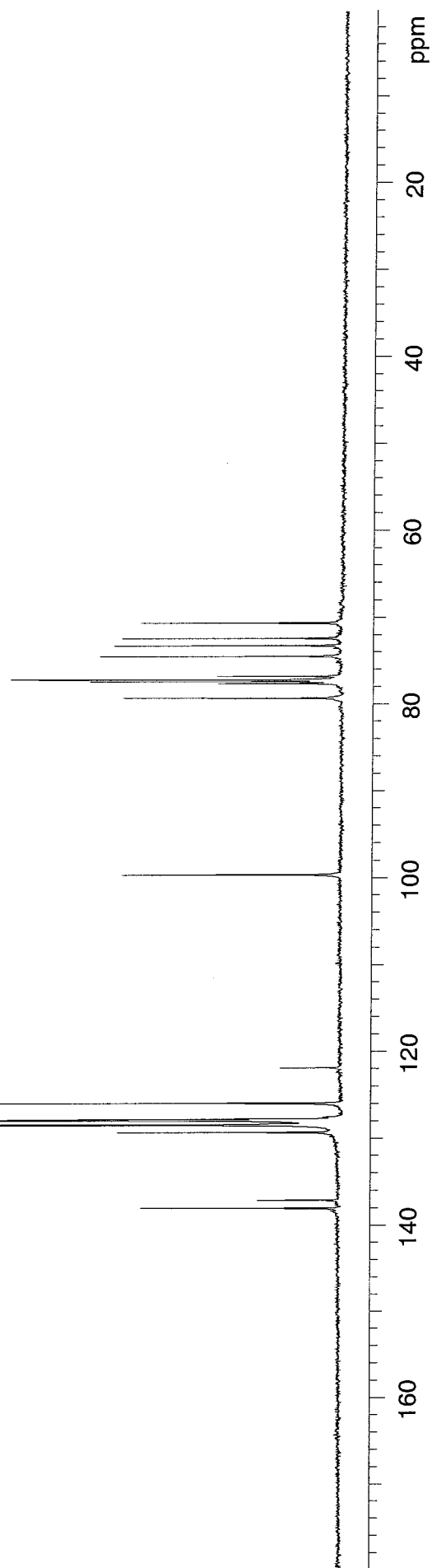
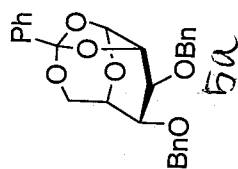
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Pulse Sequence: s2pul

¹³C OBSERVE

File: CARBON

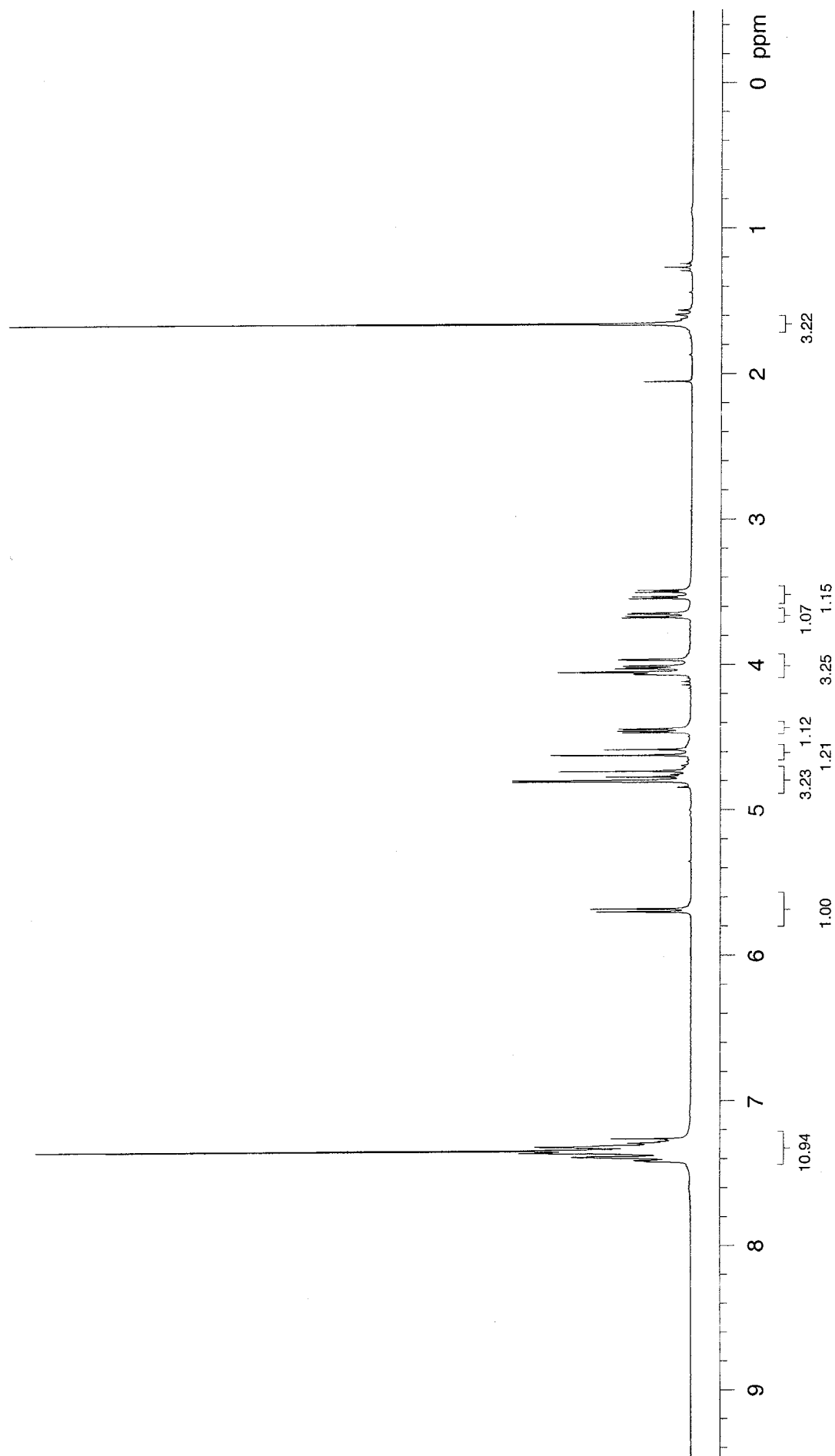
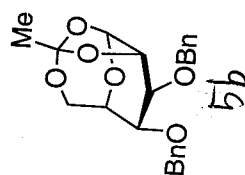
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XL13-199-pure1H

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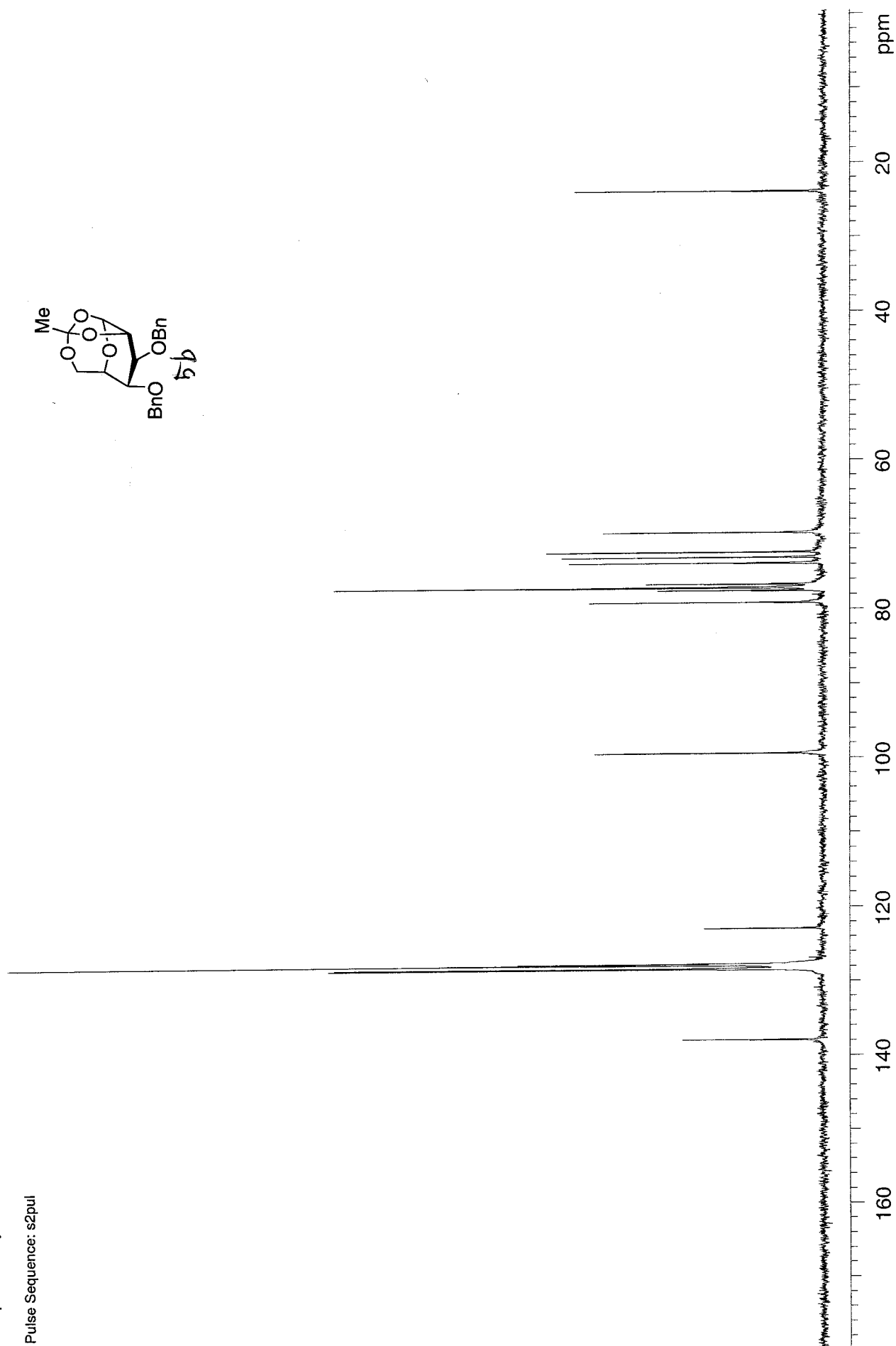
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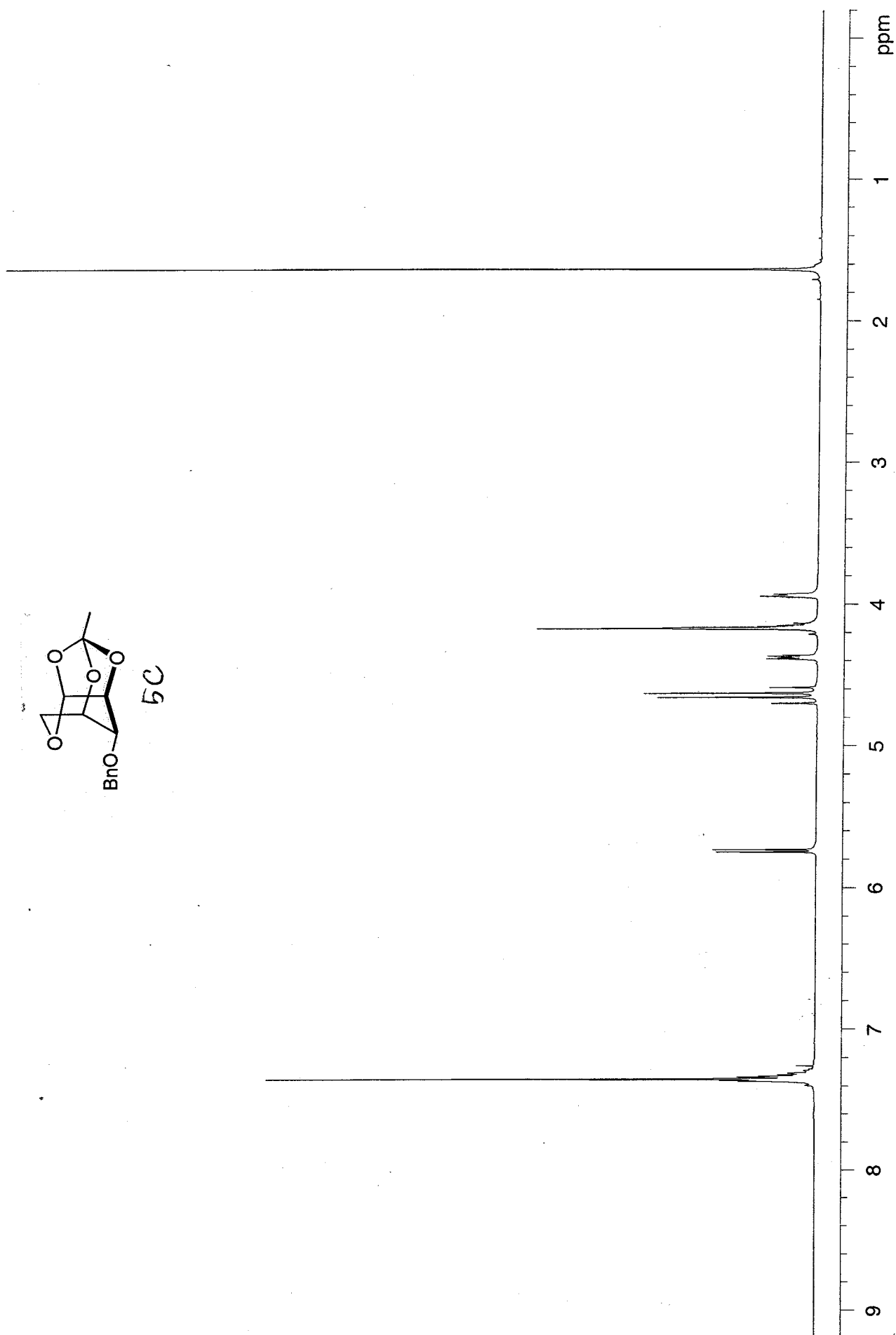
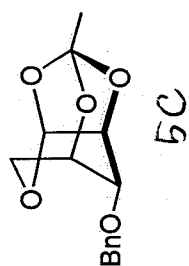


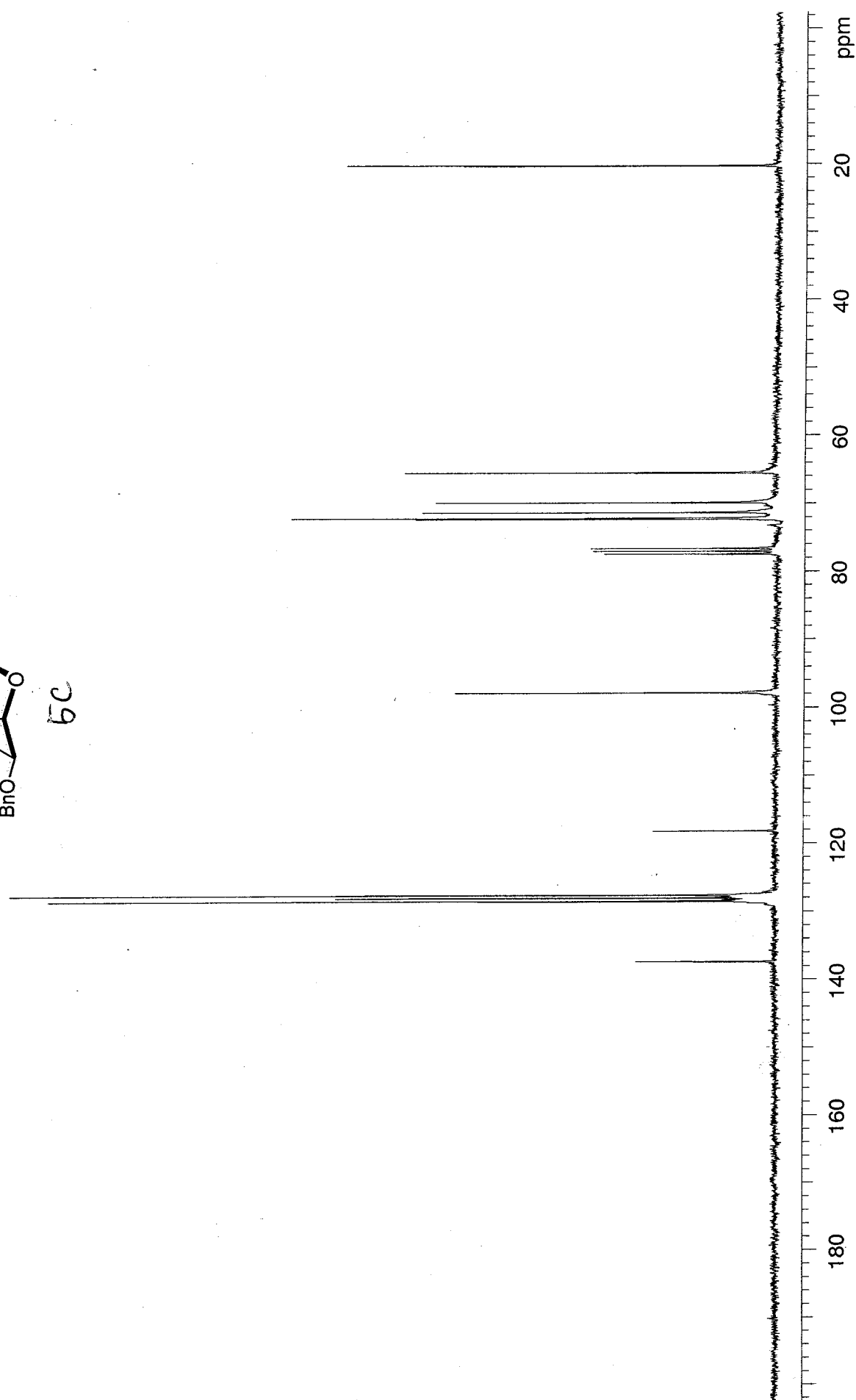
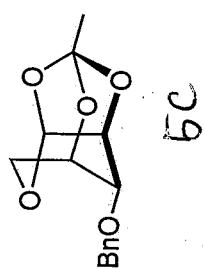
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Sample directory: we

Pulse Sequence: s2pul





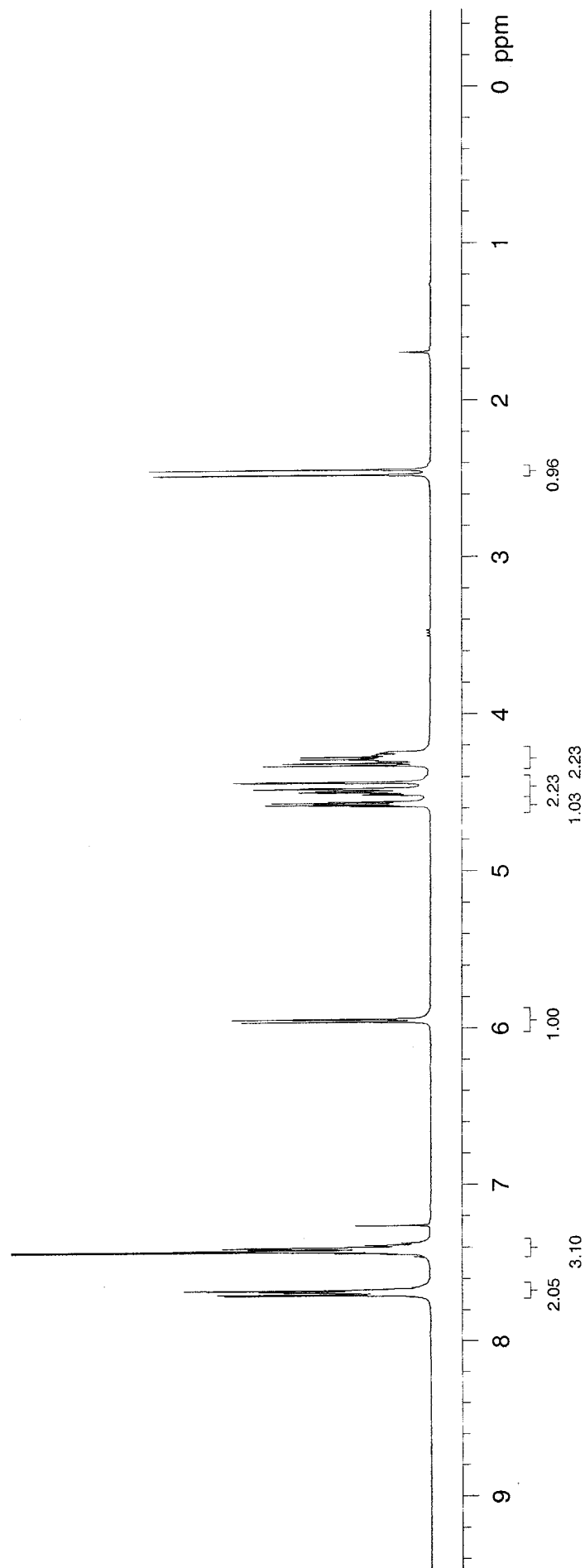




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Pulse Sequence: s2pul

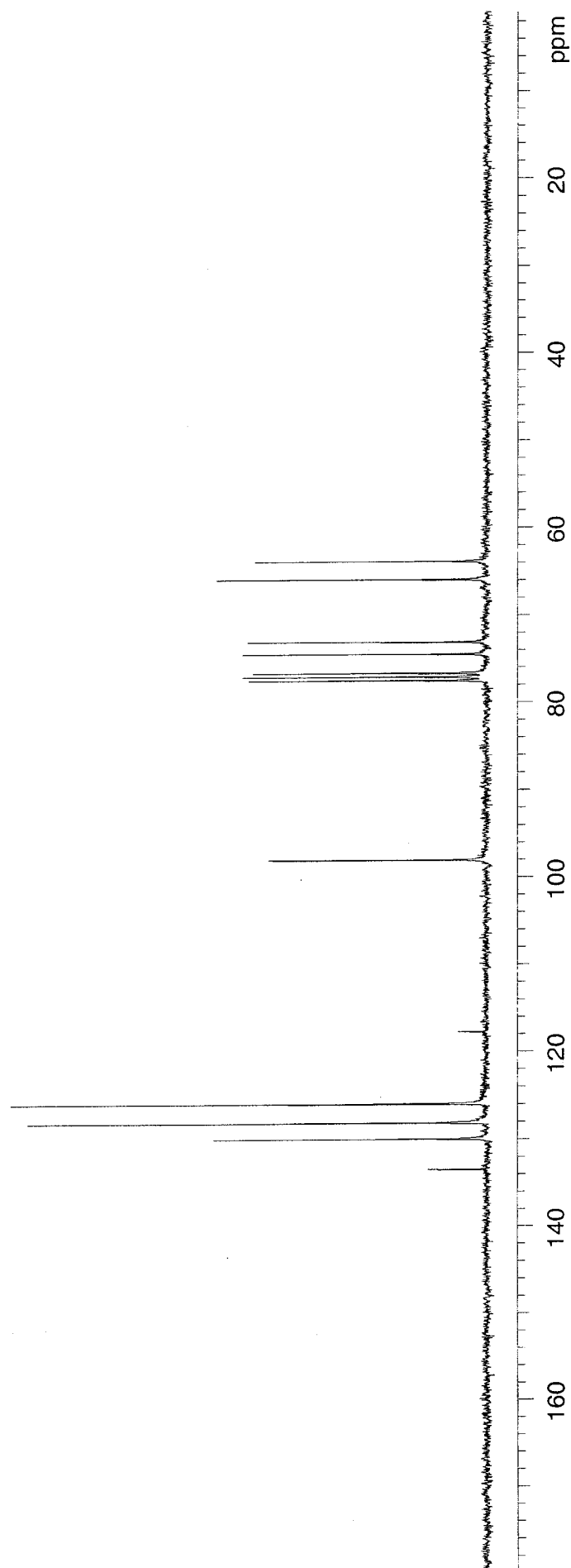




¹³C OBSERVE

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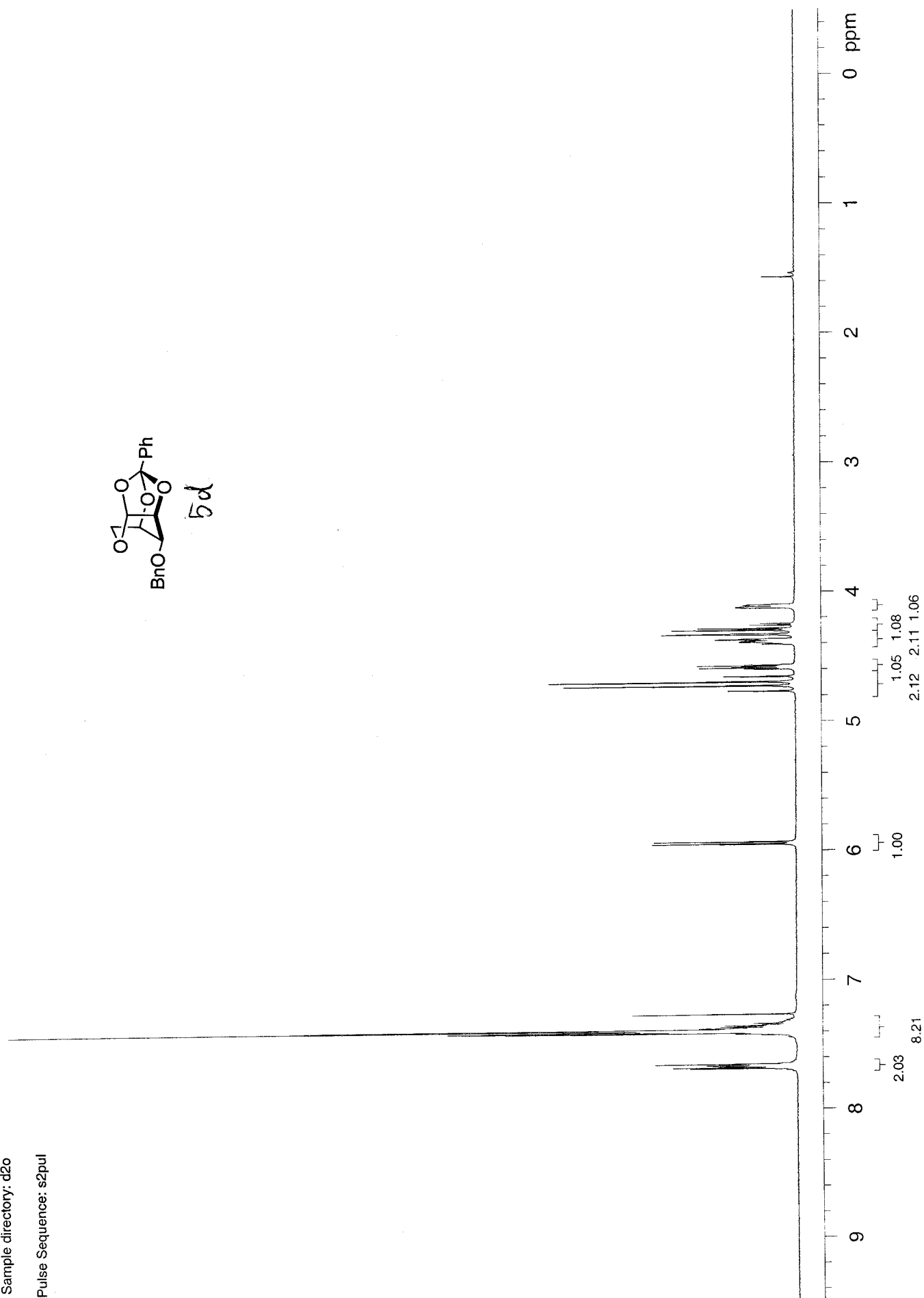
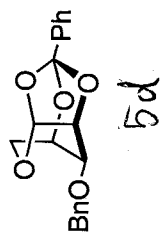
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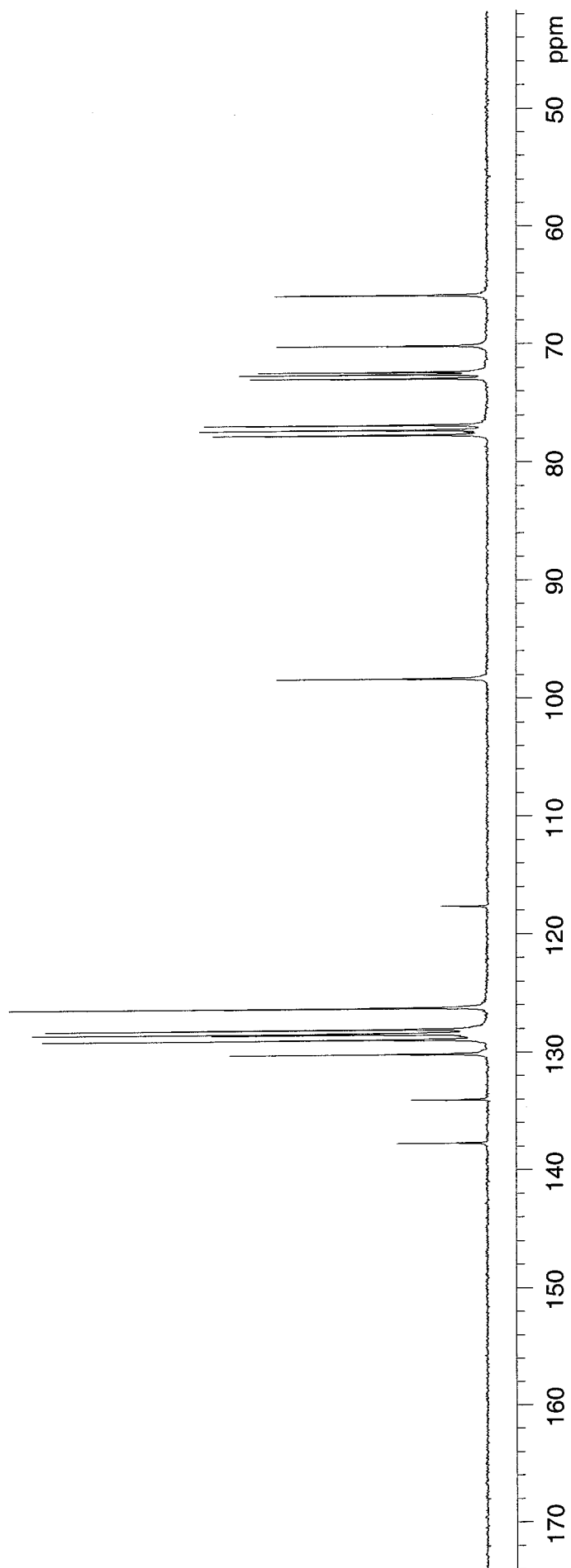
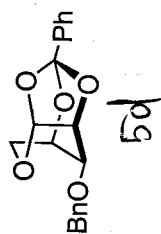
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Pulse Sequence: s2pul



¹³C OBSERVE

Pulse Sequence: s2pul



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STANDARD 1H OBSERVE

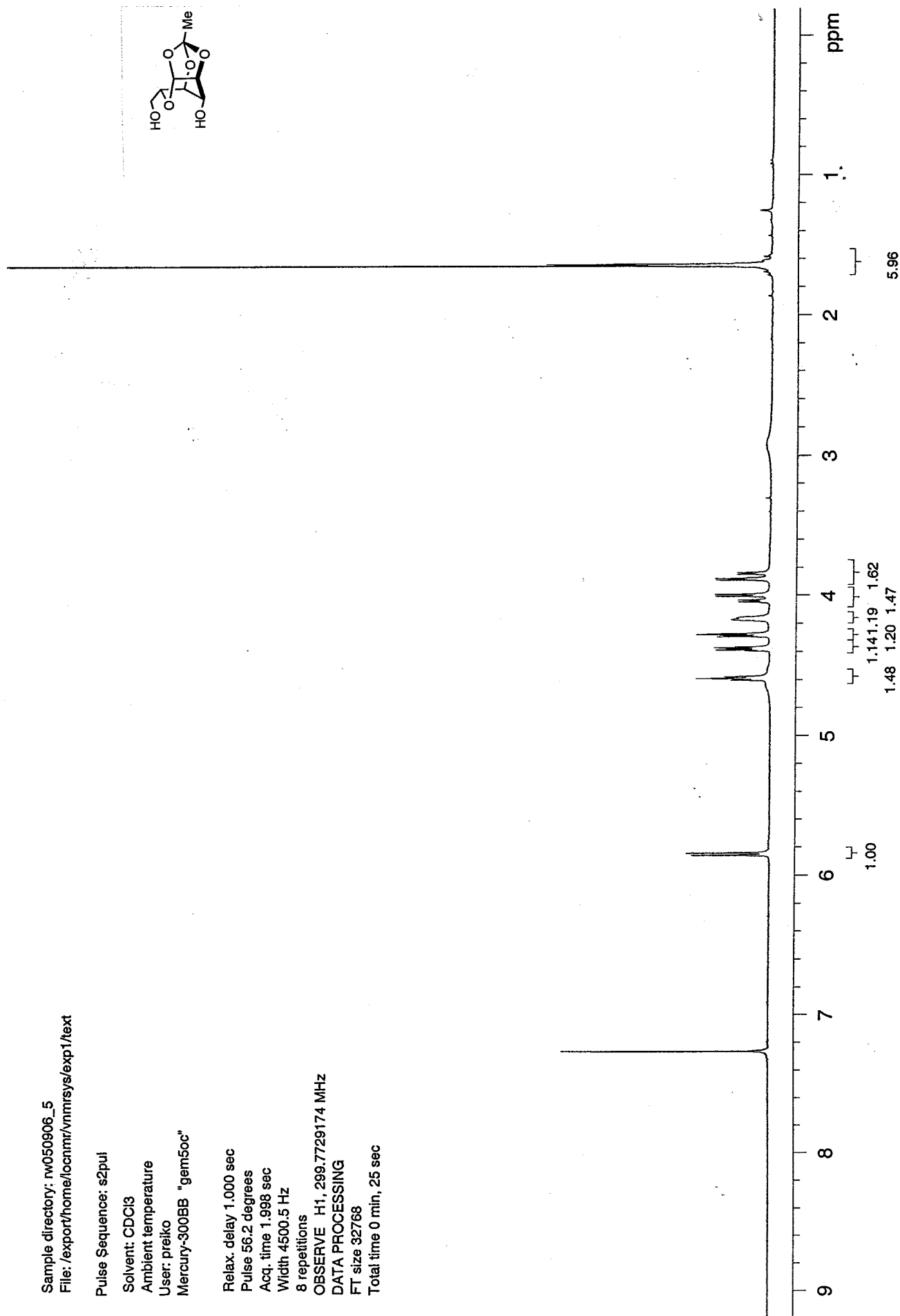
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File: /export/home/locnmr/vnmrSYS/exp1/text

Pulse Sequence: s2pul

Solvent: CDCl3
Ambient temperature
User: preiko
Mercury-300BB "gem5oc"

Relax. delay 1.000 sec
Pulse 56.2 degrees
Acq. time 1.998 sec
Width 4500.5 Hz
8 repetitions

OBSERVE H1, 299.7729174 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 25 sec



LOC ETHZ NMR Mercury-vx 300MHz Nr.6 09/15/06 19:46:33 USER:preiko GROUP:seeber SAMPLE:rw150906_1C

13C OBSERVE

Pulse Sequence: s2pul

Solvent: CDCl₃

Ambient temperature

User: preiko

File: rw150906_1C

UNITYplus-300 "nmroc"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 20000.0 Hz

512 repetitions

OBSERVE C13, 75.4911544 MHz

DECOUPLE H1, 300.2242455 MHz

Power 35 dB

continuously on

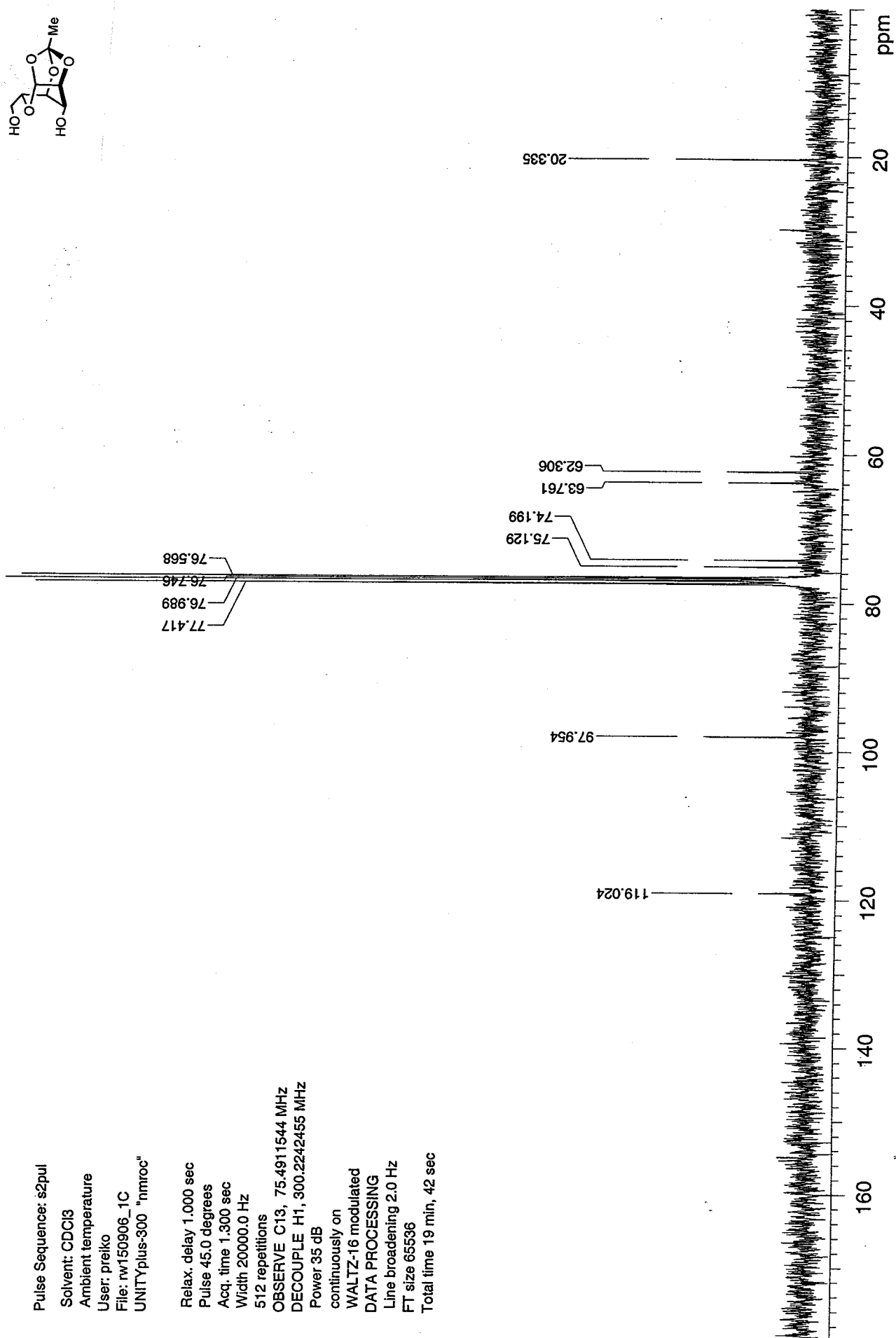
WALTZ-16 modulated

DATA PROCESSING

Line broadening 2.0 Hz

FT size 65536

Total time 19 min, 42 sec



LOC ETHZ NMR Mercury-vx 300MHz Nr.5 09/07/06 15:33:40 USER:preiko GROUP:seeber SAMPLE:rw070906_5

STANDARD 1H OBSERVE

Sample directory: rw070906_5
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Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

User: preiko

Mercury-300BB "gem5oc"

Relax. delay 1.000 sec

Pulse 56.2 degrees

Acq. time 1.998 sec

Width 4500.5 Hz

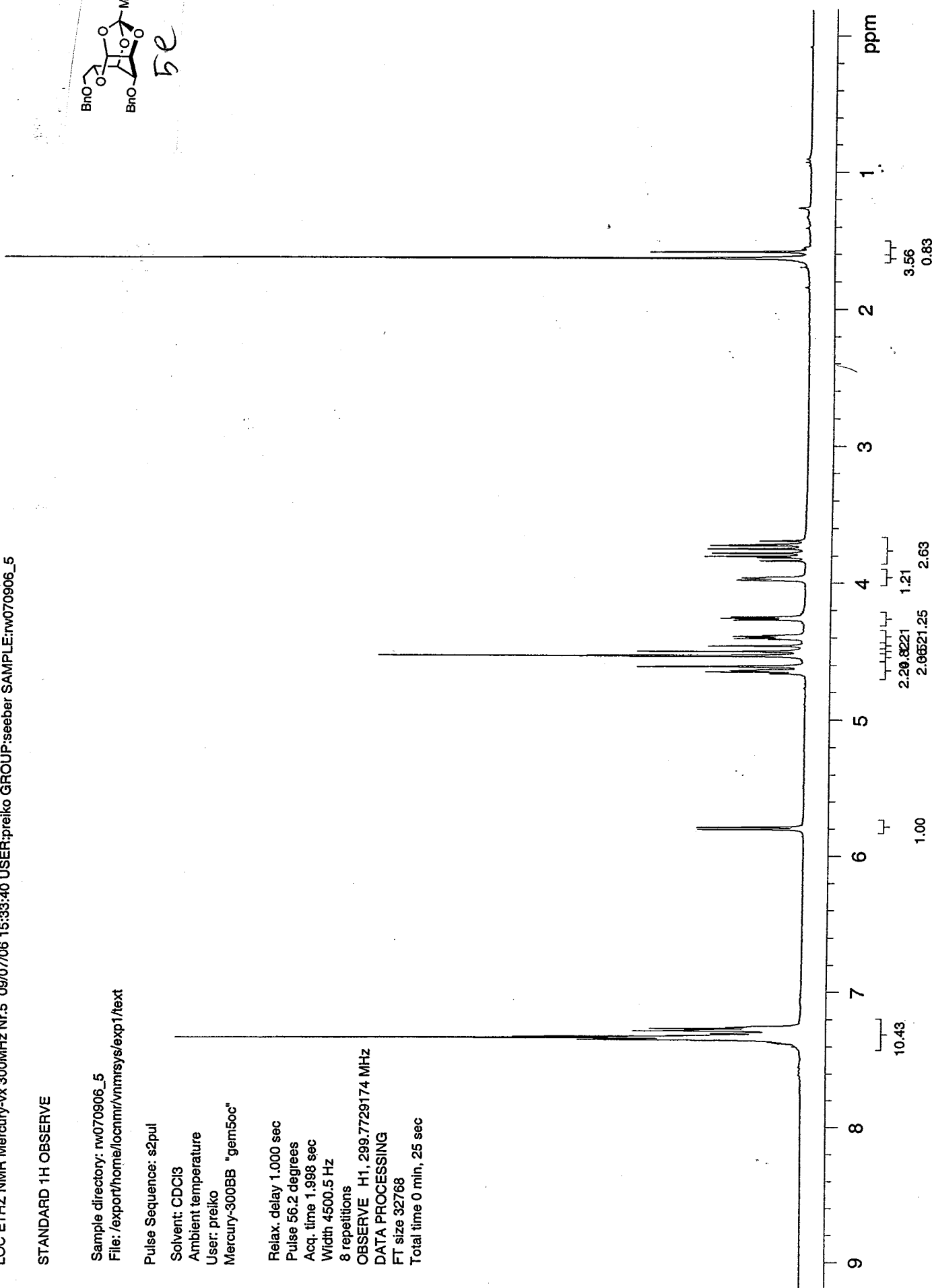
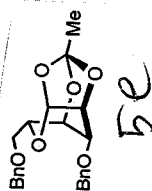
8 repetitions

OBSERVE H1, 299.7729174 MHz

DATA PROCESSING

FT size 32768

Total time 0 min, 25 sec



LOC ETHZ NMR Mercury-vx 300MHz Nr.6 09/14/06 12:51:35 USER:preiko GROUP:seeber SAMPLE:rw140906_1

13C OBSERVE

File: CARBON

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

User: preiko

Mercury-300BB "gem6oc"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 20000.0 Hz

128 repetitions

OBSERVE C13, 75.4911544 MHz

DECOUPLE H1, 300.2242455 MHz

Power 35 dB

continuously on

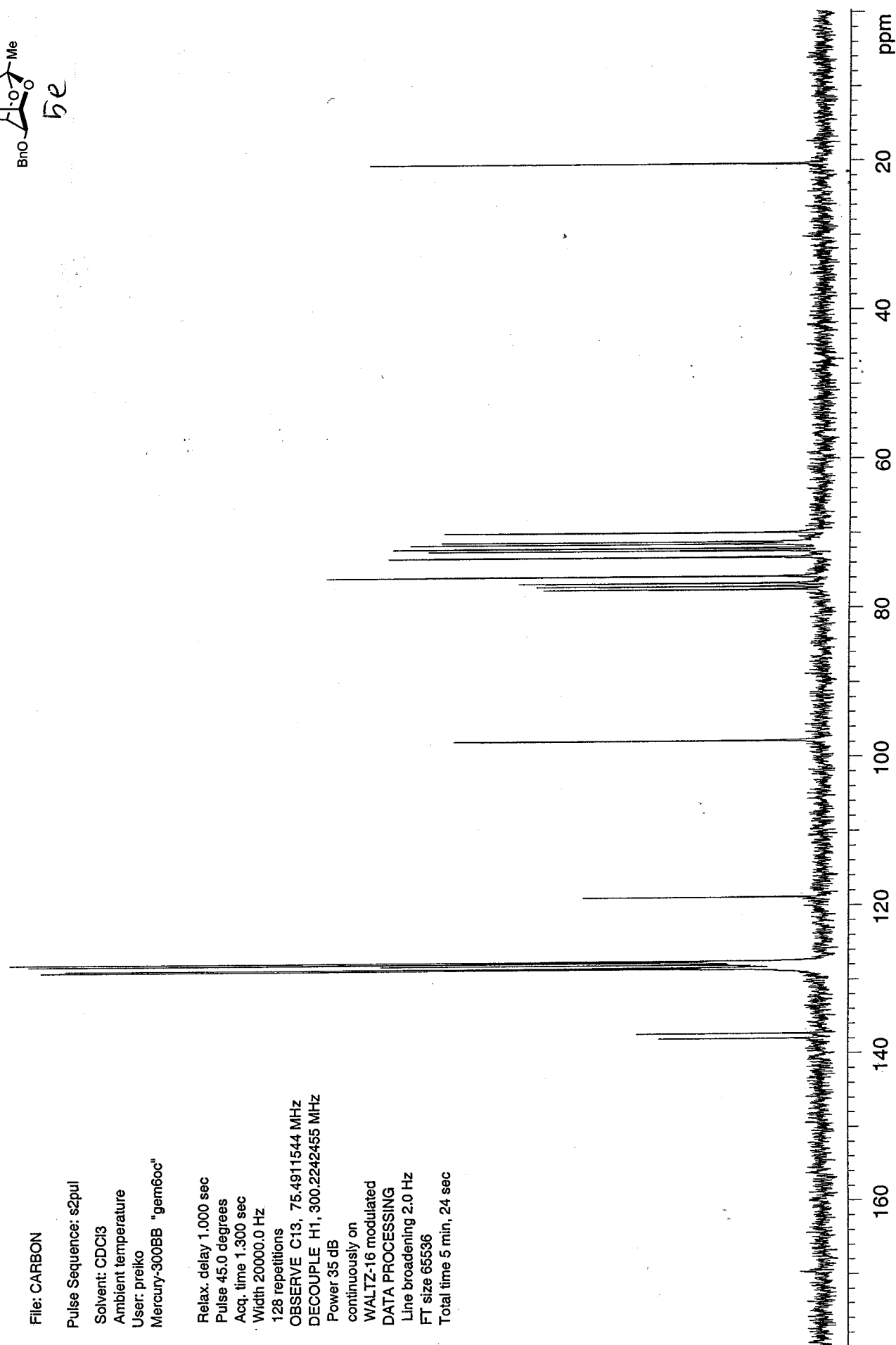
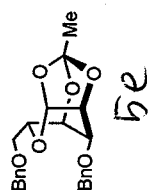
WALTZ-16 modulated

DATA PROCESSING

Line broadening 2.0 Hz

FT size 65536

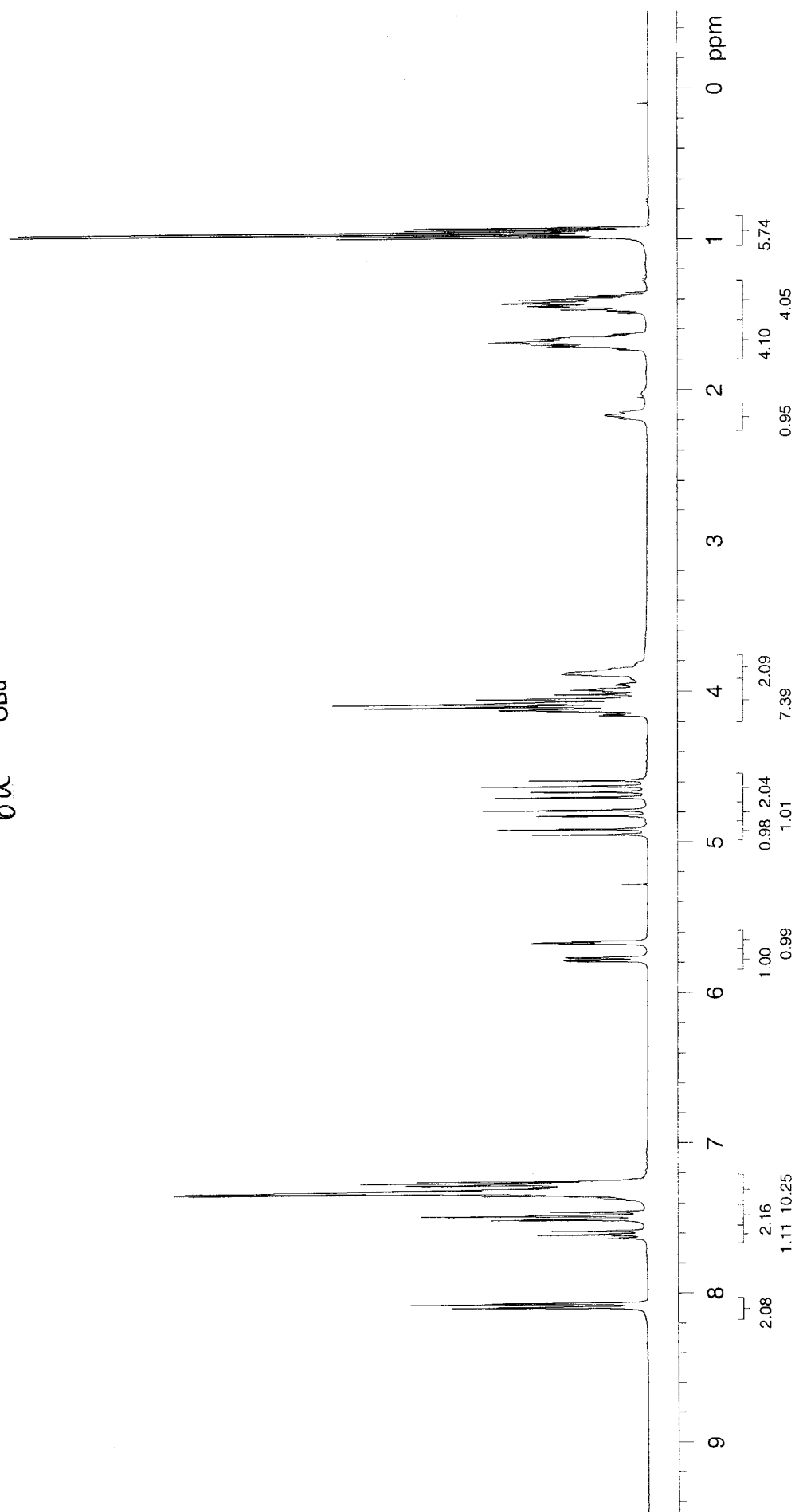
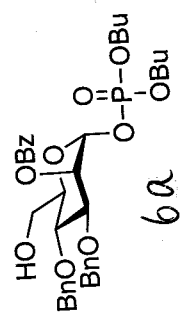
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XL13-196-pure1H

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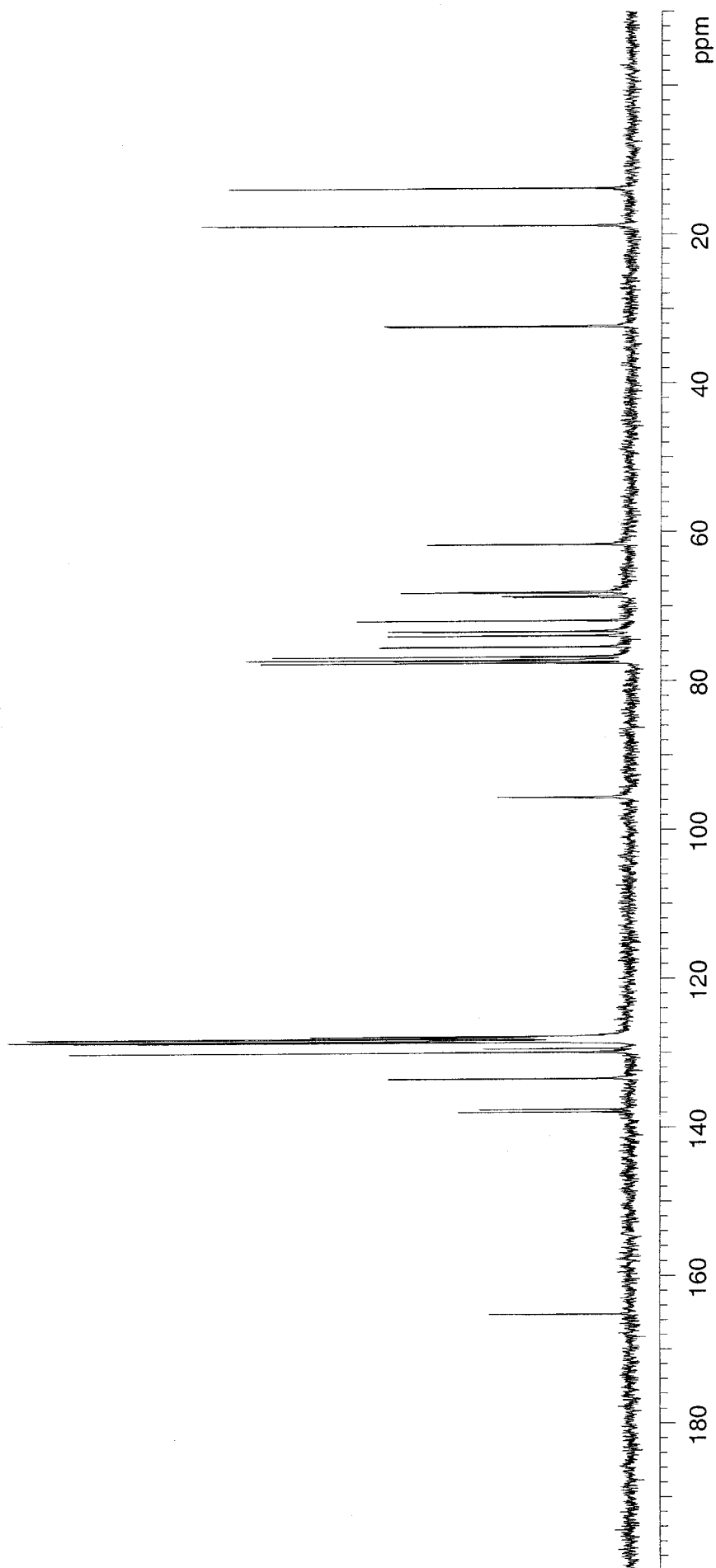
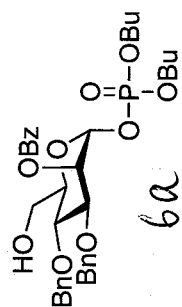
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XL13-196-pure13C

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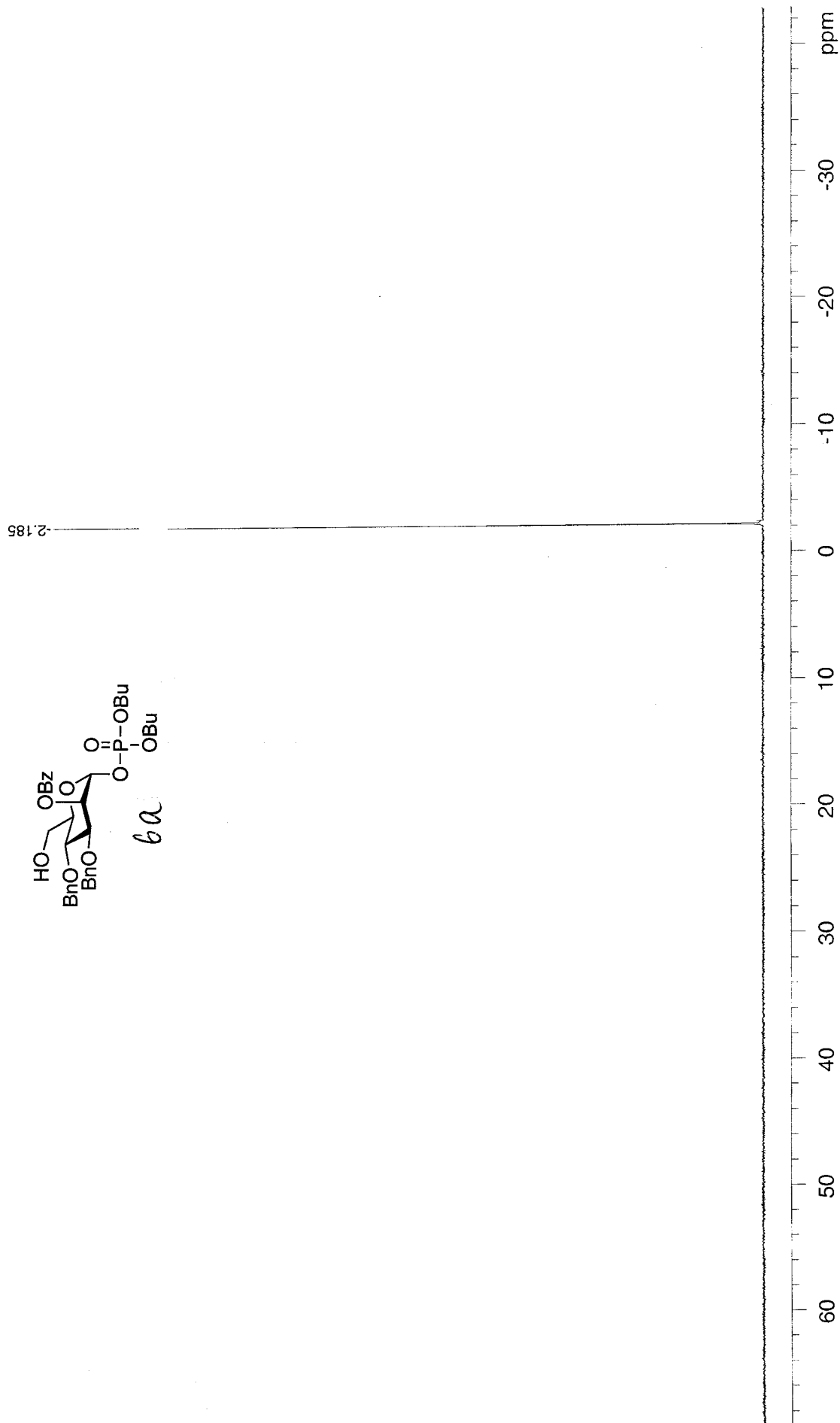
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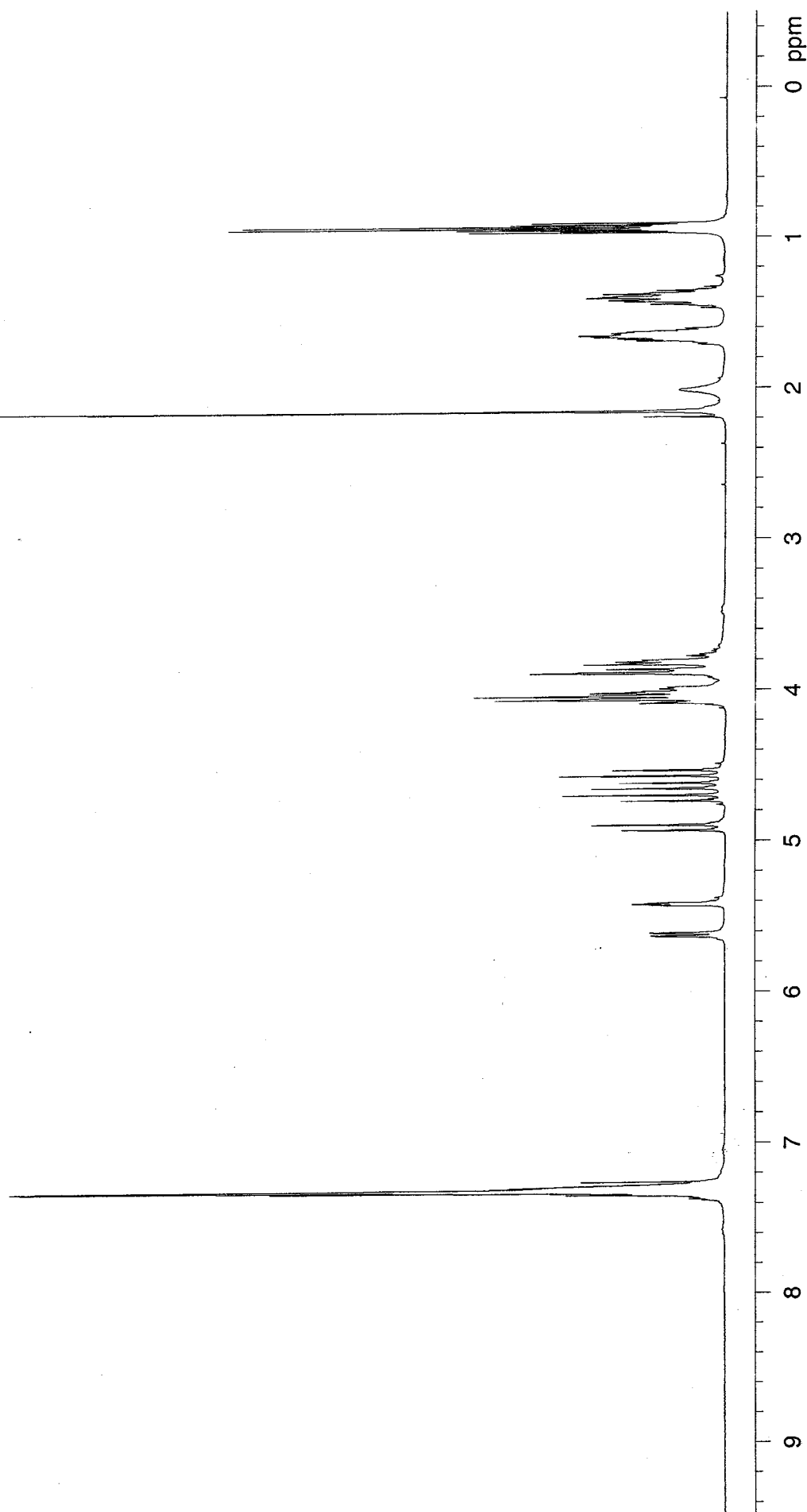
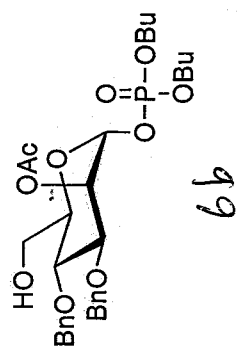


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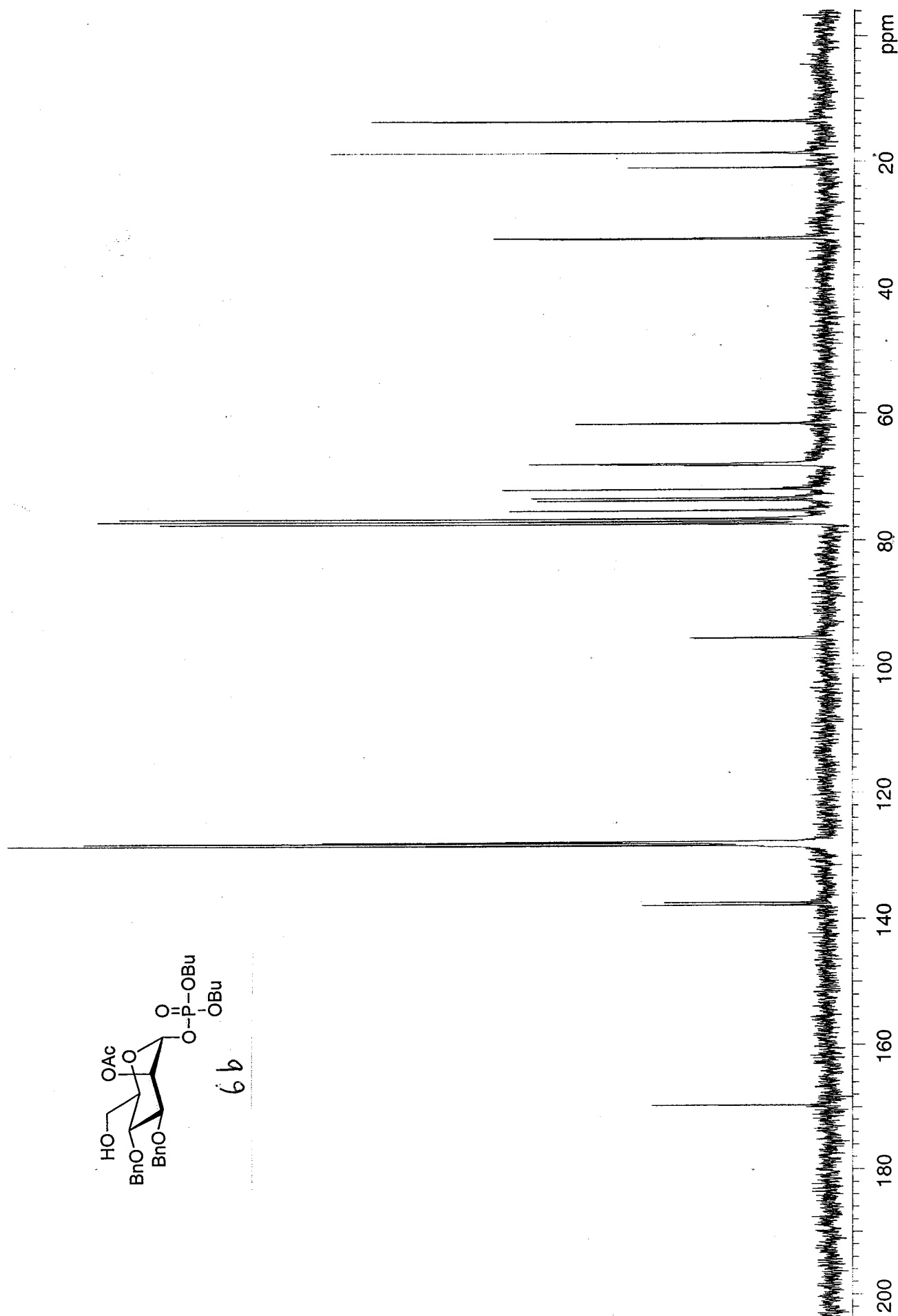
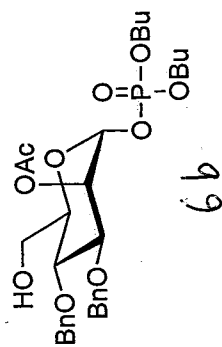
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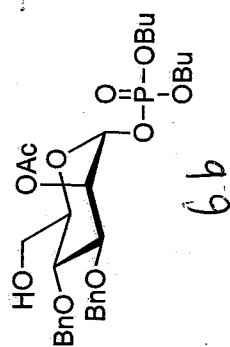




File: CARBON

Pulse Sequence: s2pul





6b

ppm

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-60

-40

-20

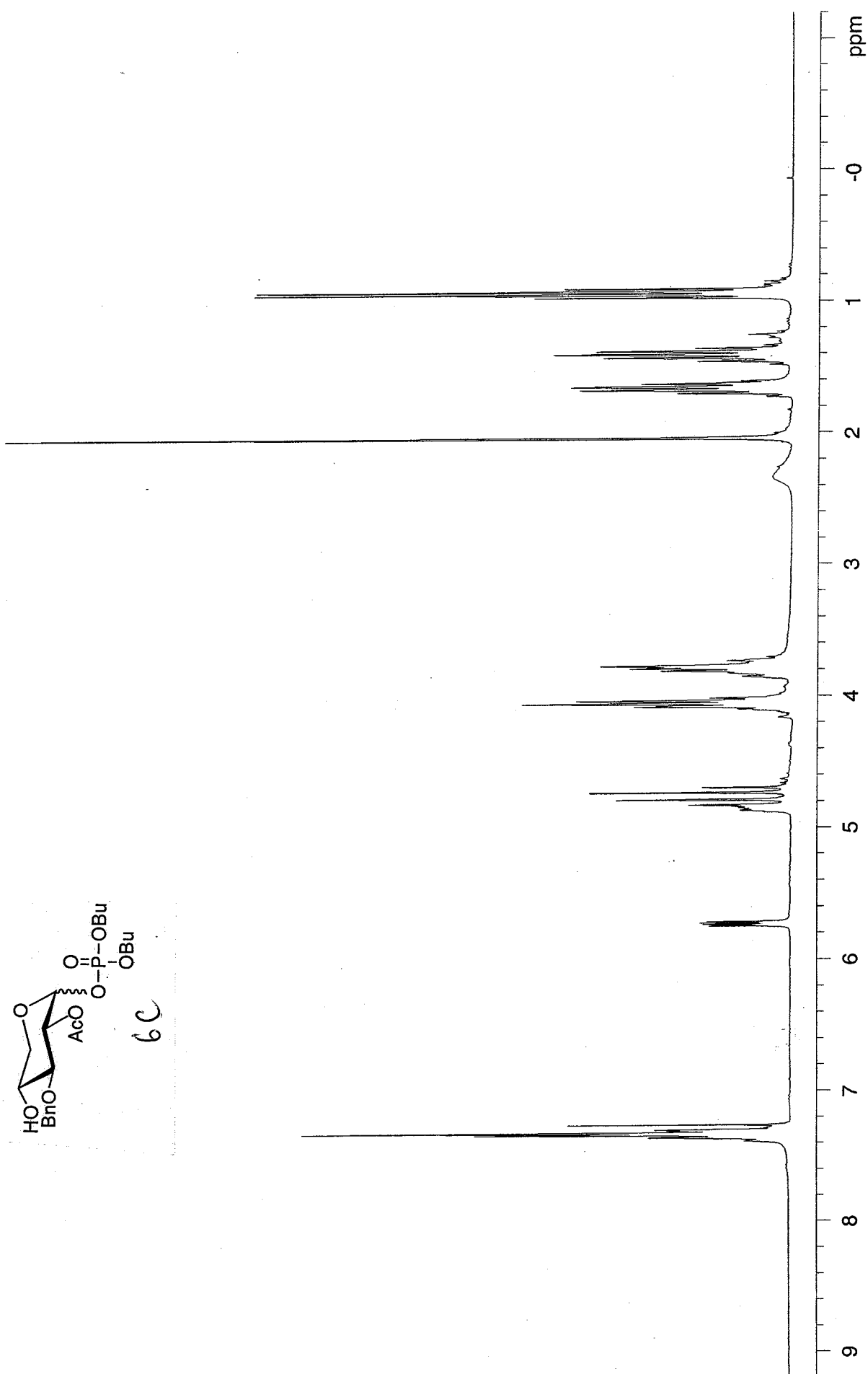
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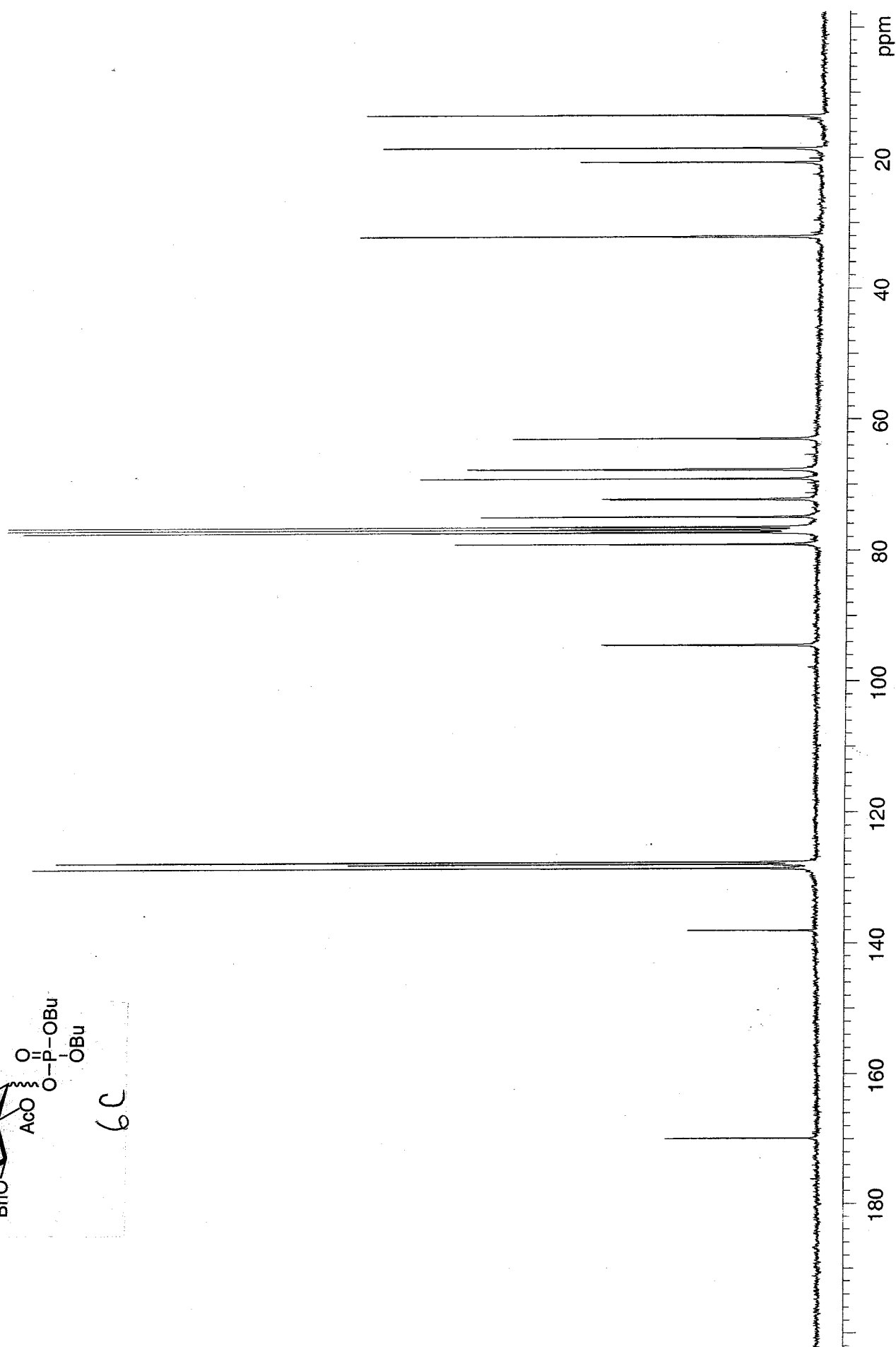
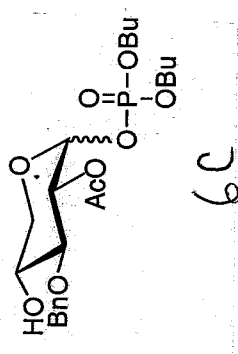
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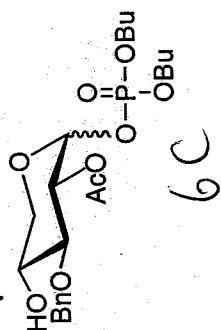
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60

80







ppm

-80

-60

-40

-20

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20

40

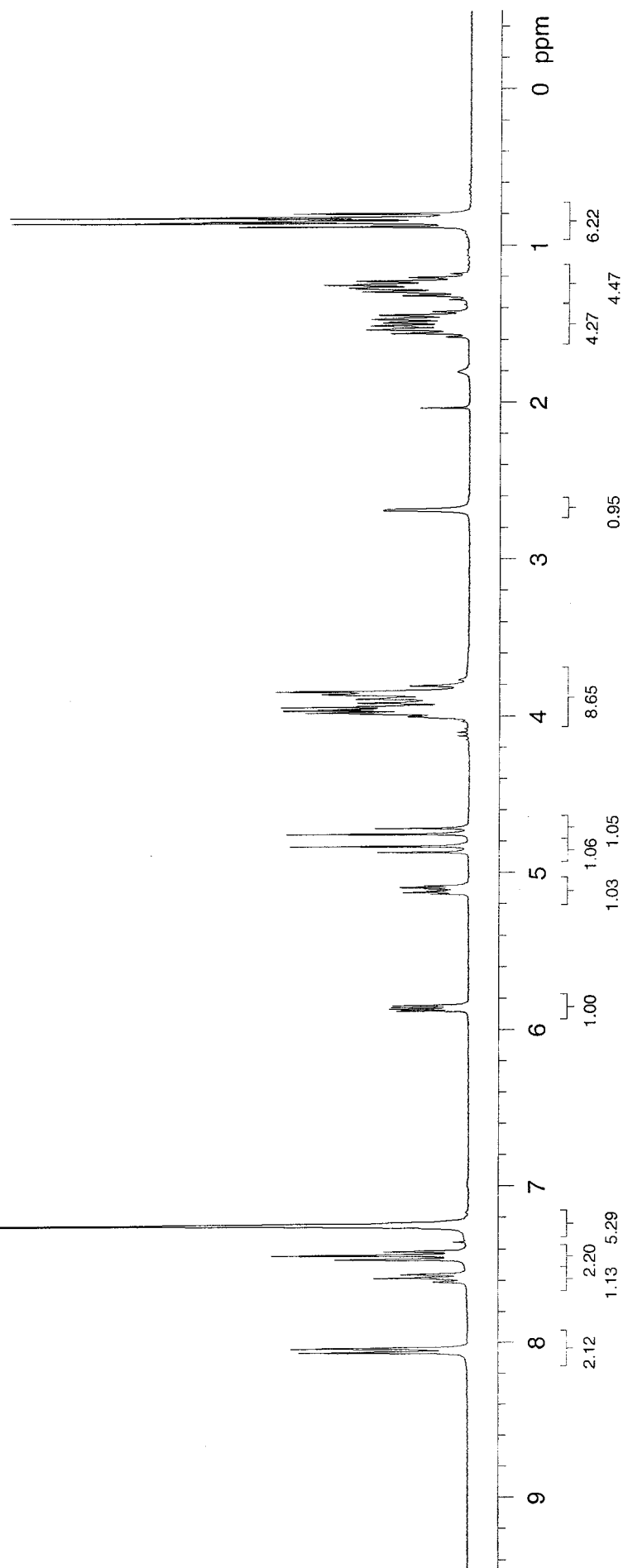
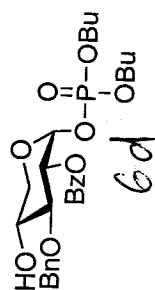
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80

XL13-34-1H

File: PROTON

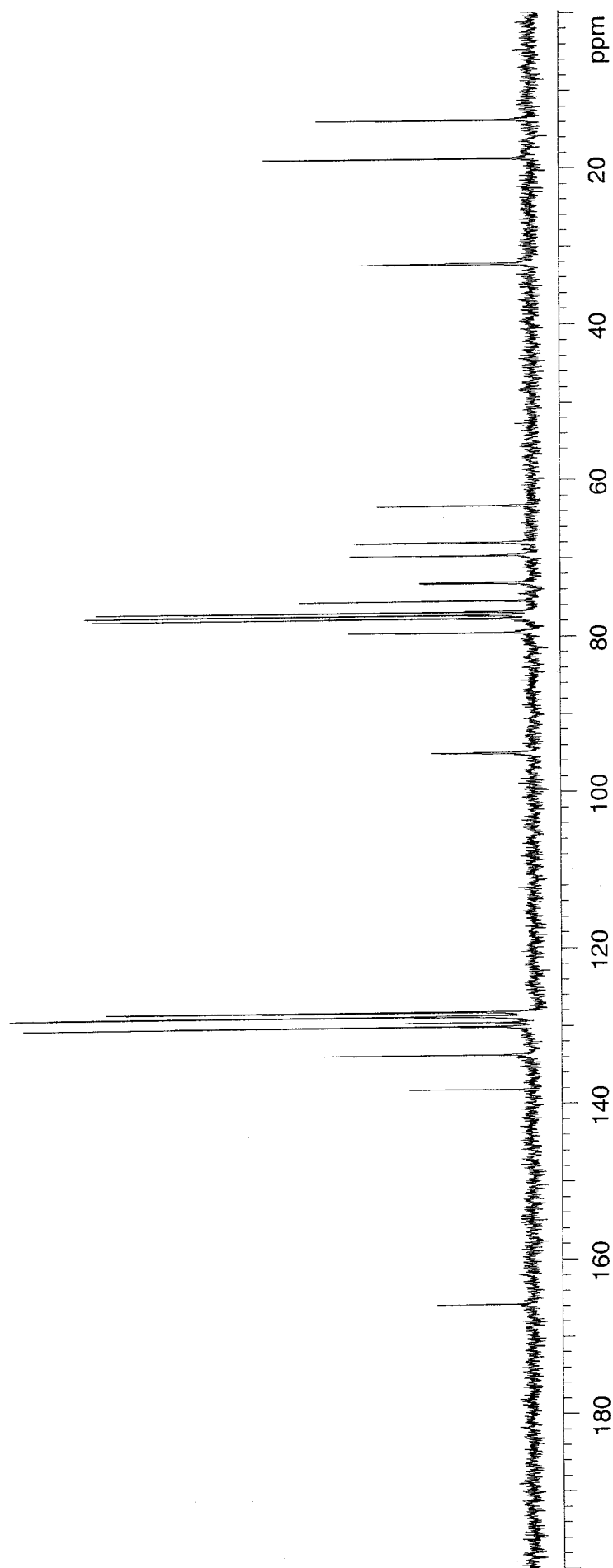
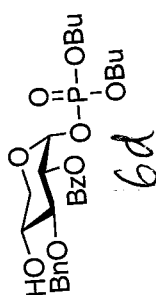
Pulse Sequence: s2pul



¹³C OBSERVE

Sample directory: we

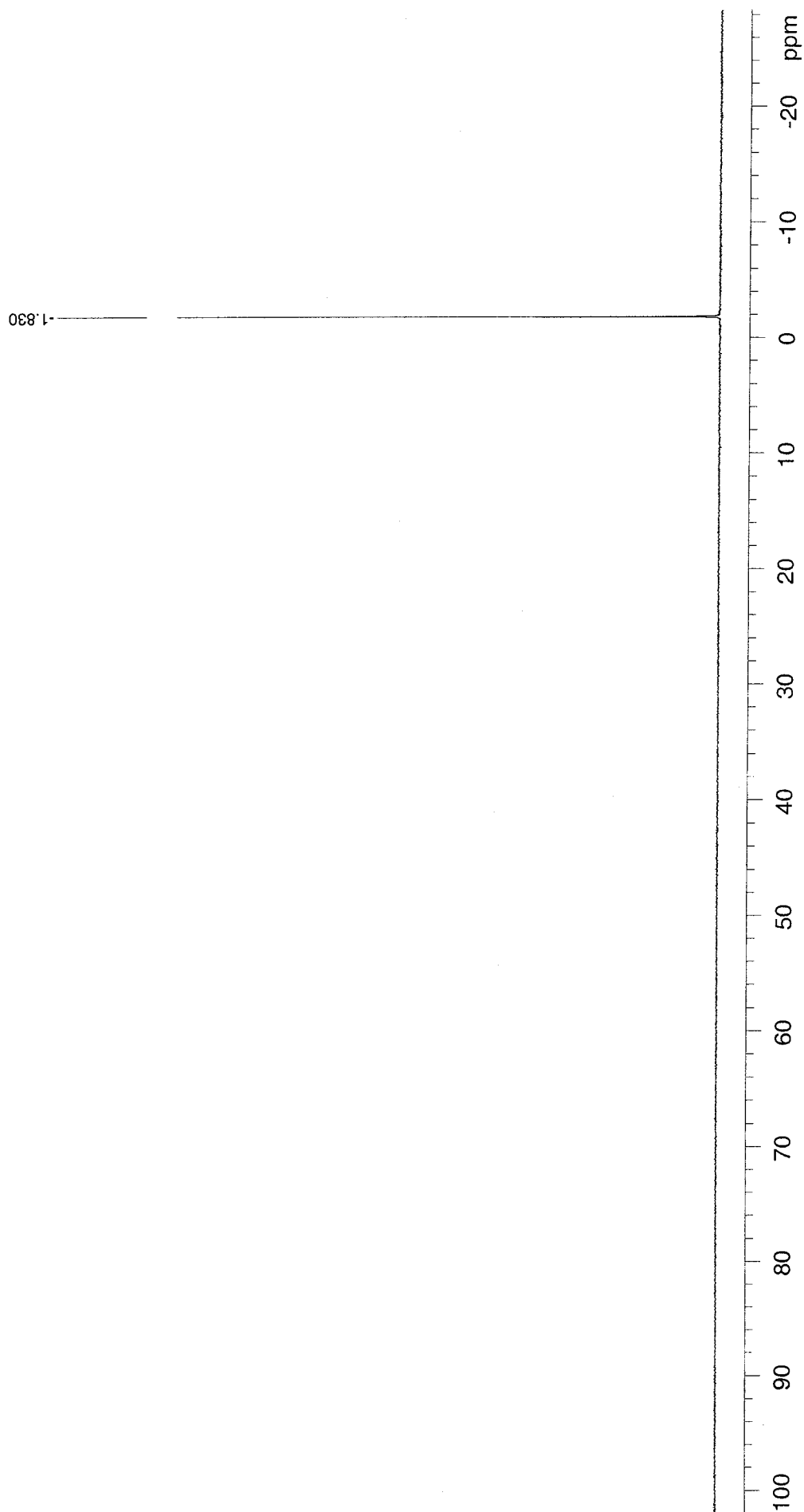
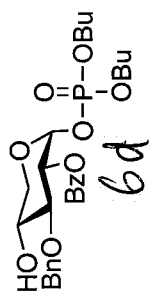
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XL13-34-31P

File: PHOSPHORUS

Pulse Sequence: s2pul



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STANDARD 1H OBSERVE

Sample directory: rw301006_4

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

User: preiko

File: rw301006_4H

Mercury-200 "nmr"

Pulse 30.0 degrees

Acq. time 3.138 sec

Width 5099.4 Hz

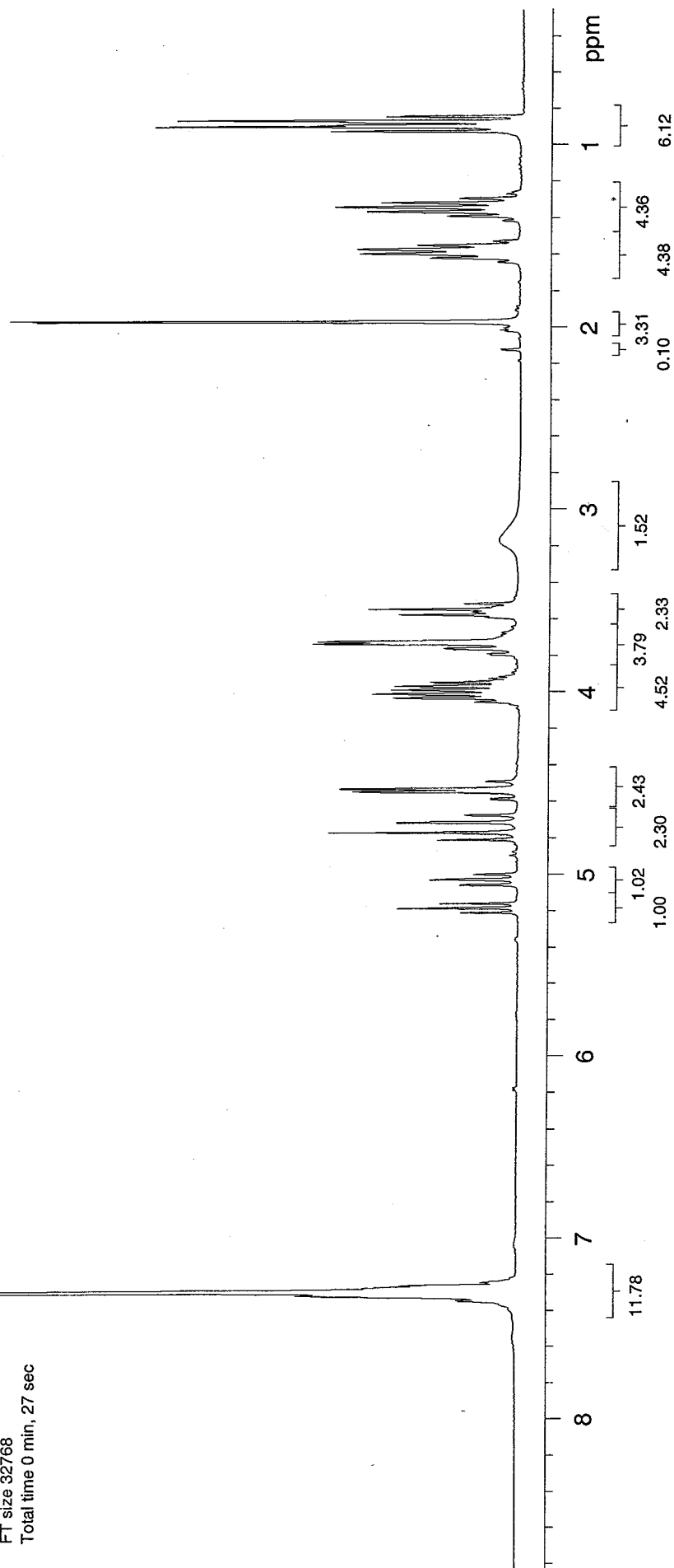
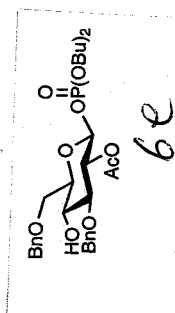
8 repetitions

OBSERVE H1, 300.2230602 MHz

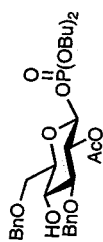
DATA PROCESSING

FT size 32768

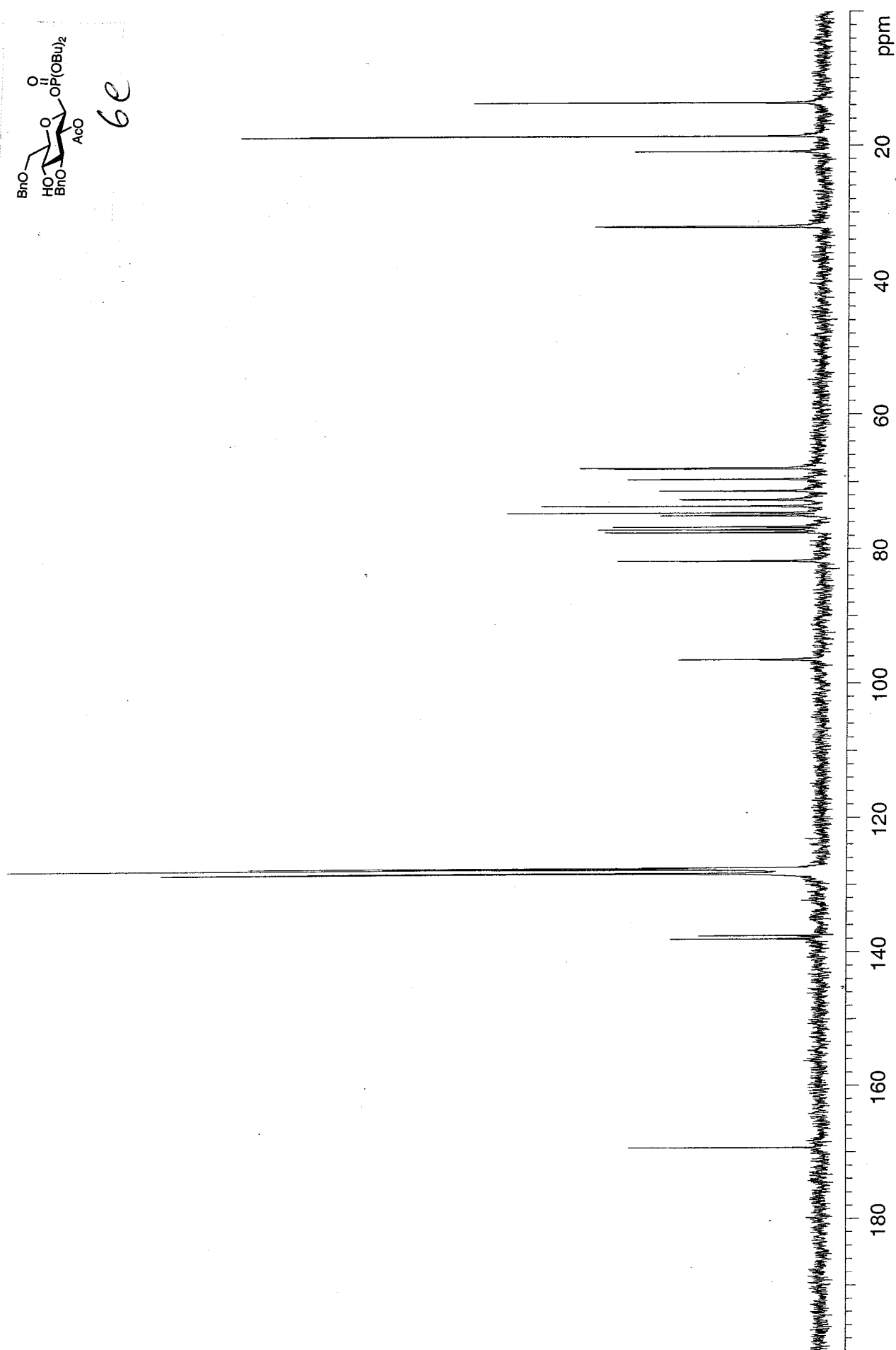
Total time 0 min, 27 sec



LOC ETHZ NMR Mercury-vx 300MHz Nr.6 03/27/07 11:01:38 USER:preiko GROUP:seeber SAMPLE:rw301006_4



6e



LOC ETHZ NMR Mercury-vx 300MHz Nr.6 10/30/06 18:05:51 USER:preiko GROUP:seeber SAMPLE:rw301006_4

P-31 STANDARD PARAMETERS
PHOSPHATE REGION

Pulse Sequence: s2pul

Solvent: CDCl₃

Ambient temperature

User: preiko

File: rw301006_4P

Mercury-300BB "gem6oc"

Pulse 45.0 degrees

Acq. time 1.002 sec

Width 31948.9 Hz

64 repetitions

OBSERVE P31, 121.5324811 MHz

DECOUPLE H1, 300.2242455 MHz

Power 35 dB

continuously on

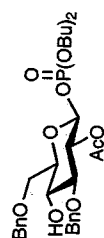
WALTZ-16 modulated

DATA PROCESSING

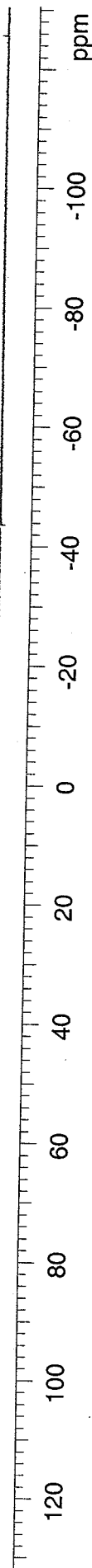
Line broadening 1.0 Hz

FT size 65536

Total time 1 min, 22 sec



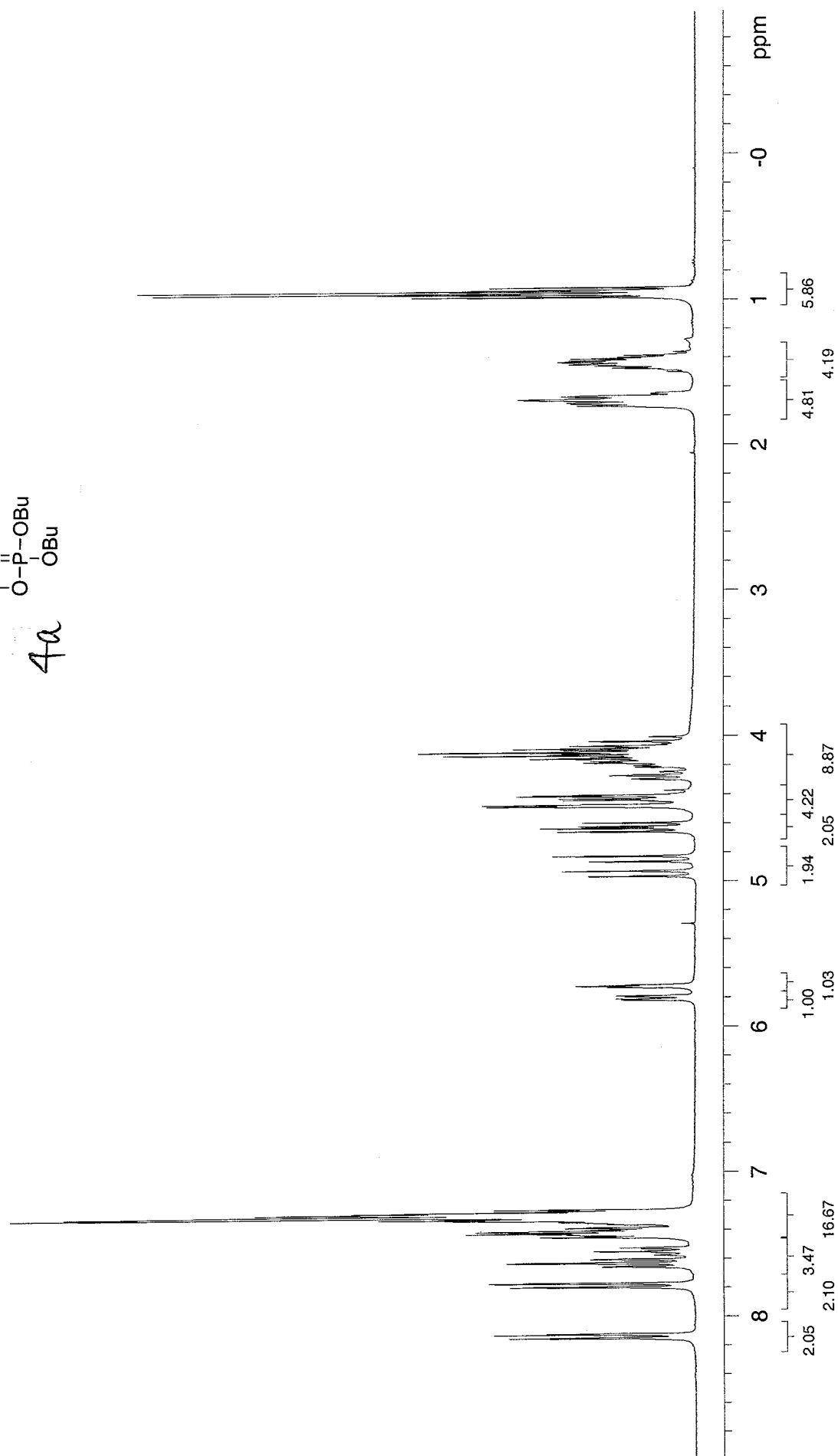
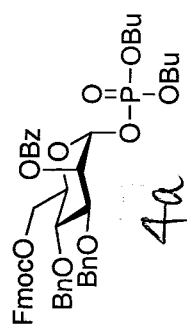
-2.587



XL13-6-pure 1H

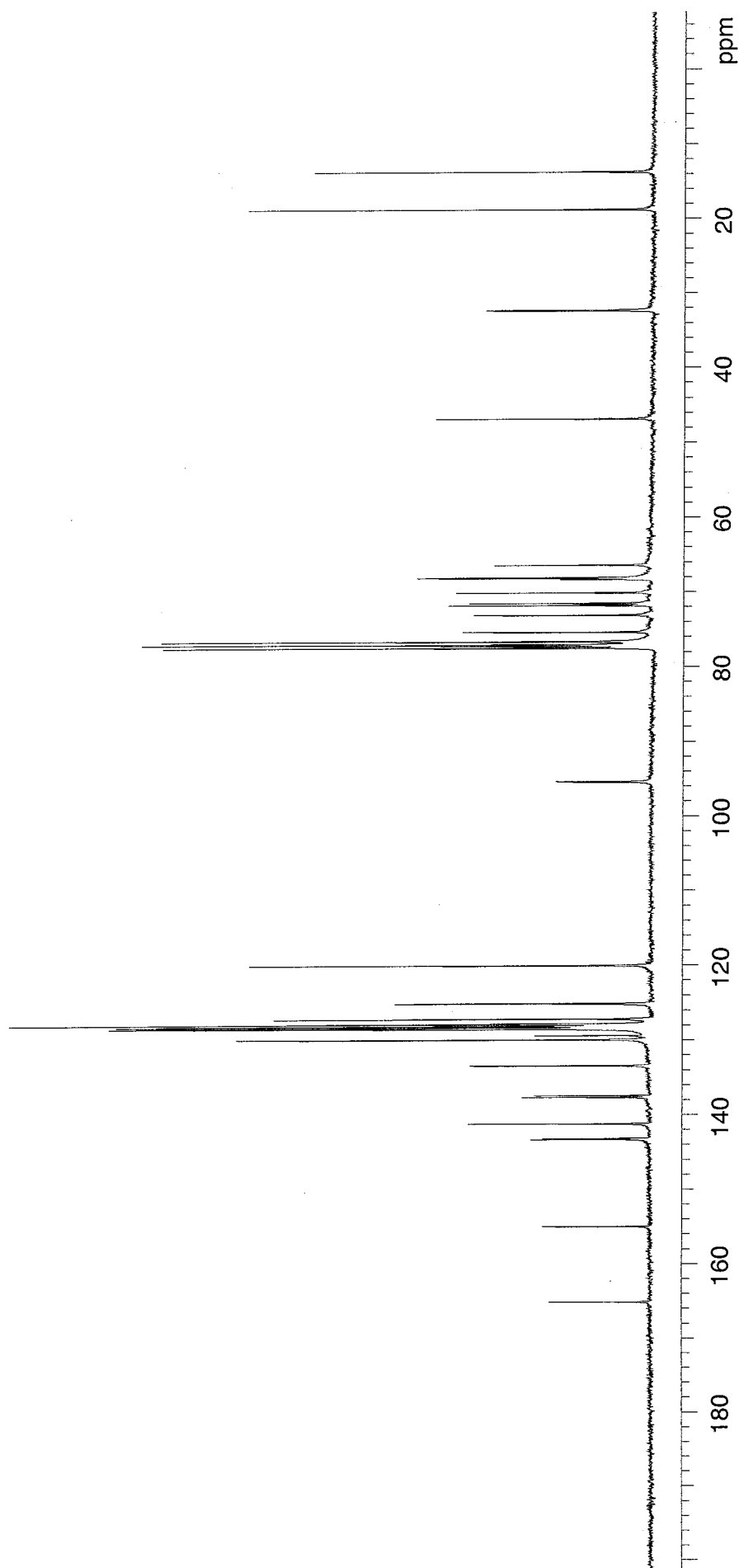
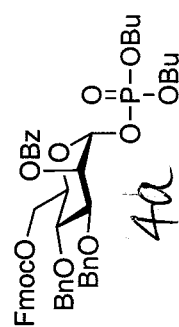
File: PROTON

Pulse Sequence: s2pul



XL13-6-13C

Pulse Sequence: s2pul



XL13-6-pure 31P

File: PHOSPHORUS

Pulse Sequence: s2pul

