Electronic supplementary information (ESI) For

A Photosensitive {Ru-NO}⁶ Nitrosyl bearing Dansyl Chromophore: Novel NO Donor with a Fluorometric On/Off Switch

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Experimental Section

Chemical Reagents and Syntheses. Dansyl-chloride, imidazole, CsCO₃, and other starting materials were obtained from Sigma-Aldrich Chemical Co. except for AgBF₄, which was obtained from Alfa-Aesar. All solvents were purchased from Fisher Sciences and distilled prior to use: Et₂O from Na; MeCN and CH₂Cl₂ from CaH₂. NanoPure H₂O was used to prepare all aqueous solutions. The ligands H₂Me₂bpb and dansyl-imidazole (ds-Im) were synthesized as previously described.

 $[(Me_2bpb)Ru(NO)(Ds-im)]BF_4$ (1). A batch of 0.140 g (0.277 mmol) of [(Me₂bpb)Ru(NO)Cl] was stirred in 40 mL of MeCN and treated with 1 equiv of AgBF₄ (0.054 g, 0.277 mmol). Upon heating to reflux temperature, the greenish-brown solution generated a beige precipitate was observed (AgCl) and the solution held at this temperature for 12 h. Next, 3 equiv of dansyl-imidazole (0.250 g, 0.831 mmol) in 5 mL of MeCN were added to the hot solution and continue refluxing for only 1 h. Upon cooling, the resulting orange-red solution was concentrated to ~ 5 mL, and stored at -20°C overnight to precipitate trace amounts of AgCl and unreacted starting material (chloride adduct). After filtration, the solvent was removed in vacuo, and the residue dissolved in ~5 mL of CH₂Cl₂. Addition of Et₂O and subsequent cooling to -20 °C overnight affords the product as a pure greenish-red solid. The nitrosyl-dye conjugate 1 was washed with several small portions of Et_2O and dried in vacuo. Yield: 60 mg (26%). Anal. calcd. for C₃₅H₃₁N₈O₅SRuBF₄ (1): C 48.65, H 3.62, N 12.98; found: C 48.50, H 3.68, N 12.95. Selected IR frequencies (KBr disk, cm^{-1}): 1868 (s, v_{NO}), 1639 (vs, v_{CO}), 1597 (vs), 1566 (w), 1483 (s), 1376 (m), 1354 (m), 1184 (m), 1170 (w), 1155 (w), 1070 (vs), 795 (m), 762 (m), 685 (m), 634 (s), 586 (m), 487 (w). Electronic absorption spectrum in MeCN, λ_{max} in nm (ϵ in M⁻¹ cm⁻¹): 385 (4 950), 300 (9 530), 272 (15 790). ¹H NMR in CD₃CN, δ from TMS: 9.01 (d 2H), 8.68 (d 1H), 8.40 (s 2H), 8.35 (d 1H), 8.23 (t 2H), 8.00 (d 2H), 7.88 (t 2H), 7.79 (s 1H), 7.60 (t 1H), 7.48 (m 2H), 7.29 (d 1H), 7.26 (s 1H), 6.56 (s 1H), 2.89 (s 6H).

 $[(Me_2bpb)Ru(NO)(im)]BF_4$ (2). A slurry of 0.175 g (0.342 mmol) of [(Me₂bpb)Ru(NO)Cl] in 20 mL MeCN was mixed with a 5 mL solution of 0.067 g (0.342 mmol) of AgBF₄ in MeCN. This green-brown solution was refluxed for 16 hrs, which resulted in a turbid green-red solution. A 5 mL solution of MeCN containing 3 equiv of imidazole (0.070 g, 1.026 mmol) was added and reflux continued for several hrs. The solution was cooled to ambient temperature and filtered over Celite to remove AgCl. The dark red solution was concentrated to ~10 mL and placed at -20 °C overnight to precipitate trace impurities. The next day, 5 mL of Et₂O was added to the filtrate and the solution kept at -20 °C for several days to afford 2 as a dark red crystalline solid. Yield: 143 mg (68 %). Anal. calcd. for C₂₃H₂₀N₇O₃RuBF₄ (**2**): C 43.83, H 3.20, N 15.56; found: C 44.50, H 3.08, N 14.95. Single crystals suitable for X-ray diffraction were obtained from an MeCN: $Et_2O(1:1)$ mixture of **2** at ambient temperature, which afforded dark red blocks. Selected IR frequencies (KBr disk, cm⁻¹): 1872 (vs, v_{NO}), 1642 (s, v_{CO}), 1590 (vs), 1483 (m), 1400 (w), 1388 (w), 1360 (m), 1074 (s), 1031 (s), 756 (m), 684 (m), 661 (m). Electronic absorption spectrum in MeCN, λ_{max} in nm (ϵ in M⁻¹ cm⁻¹): 550 (500), 394 (7 160), 302 (13 110), 277 (20 330). ¹H NMR in CD₃CN, δ from TMS: 9.14 (d 2H), 8.45 (s 2H), 8.32 (m 2H), 8.18 (d 2H), 7.92 (m 2H), 7.38 (s 1H), 6.84 (s 1H), 6.50 (s 1H), 2.33 (s 6H).

Physical Measurements. Electronic absorption spectra were recorded on a Varian Cary 50 spectrophotometer, while fluorescence emission spectra were monitored using a Perkin-Elmer LS50B Fluorescence/Luminescence Spectrometer. IR spectra were obtained on a Perkin-Elmer Spectrum One FTIR spectrometer, while 1H-NMR spectra were scanned using Varian 600 MHz instrumentation. X-band EPR spectra of photolyzed ($\lambda \ge 400$ nm, 0.4 W) solutions of 1 and 2 were obtained using a Bruker 500 ELEXSYS spectrometer at 125 K, and concaminant photorelease of NO was monitored using the *in*NO Nitric Oxide Monitoring System (Innovative Instruments, Inc.) fitted with the *amino*-2000 electrode.

Photolysis Experiments. Quantitative photolyses of **1** and **2** in MeCN or aqueous solutions with 400 nm monochromatic light were performed using an Apex Illuminator

(150 W Xenon lamp) equipped with a Cornerstone 130 1/8M monochromator, while qualitative samples (EPR, biological studies) were illuminated with an IL 410 Illumination System from Electro-FiberOptics Corp (halogen lamp) equipped with a $\lambda \ge$ 400 nm cutoff filter (measured light intensity = 0.4 W).

Cell Culture and Staining Protocols. The breast cancer cell line (MDA-MB-231) was a kind gift from Dr. Mina Bissell. Cell growth media and FBS were obtained from Gibco or Gemini bioproducts, respectively. Microscope chamber slides were purchased from Nunc Lab-Tek, while reagents for preparing the PBS supplemented with 1 mM Ca²⁺/Mg²⁺ (PBS-Ca/Mg) were procured from Sigma-Aldrich. Briefly, cells were grown in DMEM/5% FBS supplemented with penicillin/streptomycin. Cells at $\sim 70\%$ confluence in 8-well microscope chamber slides were washed 3 times with PBS-Ca/Mg and fixed with 4% paraformaldehyde (PFA) for 20 min at room temperature. The cells were then treated with a solution containing 4 µL of 10 mM stock solution of 1 in MeCN diluted with 196 µL of PBS-Ca/Mg. After 1 h incubation at 37°C, cells were washed 3 times with PBS-Ca/Mg. Light-exposed samples were illuminated with 1 min of visible light ($\lambda \ge 400$ nm, ~0.3 W measured intensity) or kept in the dark (control). Lastly, all samples were stained with 10 µM DAPI for 15 min (to visualize nuclei) and washed with PBS-Ca/Mg and H₂O. The slides were mounted with Fluoromount-G (Southern Biotech) under glass cover slips, and sealed with nail varnish. Fluorescence microscopy images were visualized with an Axiovert 200 fluorescence microscope (Zeiss) coupled to an AttoArc 2 HBO 100 W light source (Zeiss), and captured by a CCD camera controlled by SPOT Advanced software v4.0.9. Exposure times were chosen to best reflect the image as observed through microscope lens: blue (DAPI): 500 ms; Green (nitrosyl-dye conjugate 1): 3-7 s.

DAPI = 4'-6-Diamidino-2-phenylindole; FBS= Fetal bovine serum; PBS= Phosphate buffer saline.



Fig. S1. ¹H-NMR spectrum in CD₃CN of the ligands and metal complexes utilized in this work (from top to bottom): (*i*) imidozole only, (im); (*ii*) the non-fluorescent nitrosyl $[(Me_2bpb)Ru(NO)(im)]BF_4$ (**2**), (*iii*) the unbound ligand dansyl-Imidazole (Ds-im); (*iv*) the nitrosyl-fluorophore conjugate $[(Me_2bpb)Ru(NO)(Ds-im)]BF_4$ (**1**). Instrument parameters: 600 MHz Varian spectrometer; 298 K.



Fig. S2. Thermal ellipsoid plot (50% probability level) of $[(Me_2bpb)Ru(NO)(im)]^+$, the cation of **2**. Selected bond distances and bond angles: Ru–N1 = 1.9944(16); Ru–N2 = 1.9938(15); Ru–N3 = 2.1212(16); Ru–N4 = 2.1242(17); Ru–N5 = 2.0910(16); Ru–N6 = 1.7577(17); Ru–N6–O3 = 174.38(17). The structure has ca. 6% disorder in the orientation of the complex, but only the disordered position of the Ru was included in the model. This disorder was indicated by diffuse scattering in the diffraction pattern. The BF₄ group is disordered into two orientations and also involves a partially occupied molecule of CH₃CN. Hydrogen atoms were not added into the structure factor calculation for the latter molecule of CH₃CN.

Spectroscopic properties of 2:

The IR spectrum of **2** ($v_{NO} = 1872 \text{ cm}^{-1}$; $v_{CO} = 1642 \text{ cm}^{-1}$) is nearly identical to that of **1** with the notable exception of the absence of characteristic v_{SO} stretches near 1180 cm⁻¹ (as observed in **1**). The ¹H-NMR spectrum of **2** confirms the bound imidazole moiety, whose symmetry (in CD₃CN solution) is clearly broken upon binding the metal center (see Figure S1). X-ray quality crystals of **2** were grown at room temperature using a dilute solution in MeCN/Et₂O mixture (1:1). The nearly linear Ru–N–O moiety

 $(174.38(17)^{\circ})$, and its bond distances (Ru–N(O) = 1.7577(17) Å; N–O = 1.143(2) Å) are typical of photoactive {Ru-NO}⁶ nitrosyls. The bond distances from the ruthenium center to the carboxamido-*N* donors (Ru–N1 = 1.9938(15) Å) are similar to those observed in other nitrosyls derived from deprotonated bpb^{2–} ligands (reference 16b of the text). The Ru–N_{py} distances (Ru–N3 = 2.1212(16) Å) are also within the expected range.



Fig. S3. X-band EPR spectrum of a photolyzed solution of **1** in $H_2O/glycerol$ (3:1) glass (125 K). The g values of the primary signals are indicated. Instrument settings: microwave frequency, 9.4 GHz; microwave power, 1 mW; frequency modulation, 100 kHz; modulation amplitude, 2 G.

Table S1. Crystal data and structure solution parameters for [(Me₂bpb)Ru(NO)(im)]BF₄, complex **2**. Diffraction data was collected on a Bruker Apex 2 diffractometer, and the crystal structure solved using direct methods in SHELXS-97 software package (Sheldrick, 1997). Additional structural parameters are available in the cif file in ESI.

empirical formula FW crystal color crystal size (mm) T (K) wavelength (Å) crystal system space group a (Å) b (Å) c (Å) α (deg) β (deg) γ (deg) γ (deg) V (Å ³) Z d_{calc} (g/cm ³) μ (mm ⁻¹) Reflections collected Independent reflections GOF ^a on F^2 final R indices $[I > 2\sigma(I)]$	$C_{23.75}H_{21.25}N_{7.38}O_{3}RuBF_{4}$ 645.86 dark red block 0.33 × 0.27 × 0.16 90(2) 0.7103 triclinic $P\bar{t}$ 8.9006(4) 10.4858(5) 14.2498(7) 102.537(3) 104.160(3) 93.973(3) 1248.21(10) 2 1.718 0.702 24557 8239 [R(int) = 0.0190] 1.085 RI = 0.0330 w $R2 = 0.0882$
$\begin{bmatrix} I > 2\sigma(I) \end{bmatrix}$ <i>R</i> indices ^b	wR2 = 0.0882 R1 = 0.0371
all data ^c	wR2 = 0.0924

^a GOF = $[\Sigma[w(F_o^2 - F_c^2)^2]/M - N)]^{1/2}$ (M = number of reflections, N = number of parameters refined). ^b R1 = $\Sigma||F_o|| - |F_c|| / \Sigma |F_o|$; ^c wR2 = $[\Sigma[w(F_o^2 - F_c^2)^2]/\Sigma[w(F_o^2)^2]]^{1/2}$