Electronic Supporting Information

Promising Use of Sm(III)–{1,2-propanediamine-*N*,*N*,*N*',*N*'-tetra(α,α-dideuterioacetate)} Complex for Direct Simultaneous NMR Determination of Absolute Configurations of Each α-Amino Acids in Peptide Hydrolysate Mixtures

Kenji Omata, ^a Mika Fujioka, ^a Kuninobu Kabuto, ^a Yoichi Sasaki^b

^aDepartment of Chemistry, Graduate School of Science, Tohoku University, 6-3, Aoba, Aramaki, Aoba-ku, Sendai, Japan; E-mail: omata@he.tohoku.ac.jp ^bCatalysis Research Center, Hokkaido University, Kita-ku, Sapporo, Japan; E-mail: ysasaki@sci.hokudai.ac.jp

Materials and methods:

Preparation of (R)- and (S)-propylenediamine-N,N,N',N'-tetra(α,α -diduterioacetate) [(R)and (S)-H₄pdta-d₈]: The deuterated ligands were prepared from (R)- and (S)-propylenediamine and bromoacetic acid-d₃ (Aldrich) in the same way as described in the literature for the undeuterated ligands. See R. M. Wing, K. P. Callahan, *Inorg. Chem.* **1969**, *8*, 2303. Deuterium contents of (R)- and (S)-H₄pdta-d₈ was evaluated to be > 99 % by NMR.

(*R*)-*H*₄*pdta*-d₈ (recryst. from H₂O): Mp. 188 °C (decomp.); $[\alpha]_D^{23}$ –46.4 (*c* 0.45, H₂O); ¹H NMR (400 MHz, D₂O) δ 1.10 (3H, d, *J* = 6.6 Hz), 2.96 (1H, dd, *J* = 14.7, 12.2 Hz), 3.14 (1H, dd, *J* = 14.7, 3.3 Hz), 3.59 (1H, m); ¹³C NMR (100 MHz, D₂O) δ 10.88 (CH₃), 53 (CD₂, br), 55.5 (CD₂, br), 55.78 (CH₂), 58.06 (CH), 171.84 (CO₂H), 173.51 (CO₂H); Anal. Found: C, 38.83; N, 8.02. Calcd for C₁₁H₁₀D₈N₂O₈·*n*H₂O (*n* = 1.3): C, 39.02; N 8.27.

(*S*)- H_4pdta - d_8 (recryst. from H₂O): $[\alpha]_D^{23}$ +47.4 (c 0.50, H₂O): Anal. Found: C, 39.70; N, 8.48. Calcd for C₁₁H₁₀D₈N₂O₈·nH₂O (n = 1): C, 39.75; N, 8.43.

Preparation of of $Na[Sm-{(R)-pdta-d_8}]'$ and $Na[Sm-{(S)-pdta-d_8}]'$ (Na[(R)-3]) and Na[(S)-3]):

Na[(R)-3] and Na[(S)-3] were prepared from Sm_2O_3 and the corresponding pdta-d₈ ligands in a similar way for the preparation of undeuterated Eu-pdta. See C. Kabuto, K. Kabuto, Y. Sasaki, T. Nishiyama, K. Umakoshi, *J. Chem. Soc.*, *Chem. Commun.* **1993**, 381.

Na[(*R*)-**3**]: Mp. >290, $[\alpha]_{D}^{25.5}$ -3.1 (*c* 0.99, H₂O, pH 8.98), $[\alpha]_{D}$ is very dependent on temperature and pH; HRMS(ESI, negative) Calcd for C₁₁H₆D₈N₂O₈Sm (M – Na⁺)⁻: 462.0455, Found 462.0460. Anal. Calcd for C₁₁H₆D₈N₂NaO₈Sm·H₂O: C, 26.34; N, 5.58. Found: C, 26.31; N, 5.52.

Na[(*S*)-**3**]: $[\alpha]_D^{25.5}$ +3.5 (*c* 0.99, H₂O, pH 9.07); Anal. Calcd for C₁₁H₆D₈N₂NaO₈Sm·H₂O: C, 26.34; N, 5.58. Found: C, 26.16; H, 3.35; N, 5.378.

X-ray crystallographic analysis of Na [rac-3]:

The structure of Na[rac-3] was confirmed by X-ray crystallographic analysis on racemic 3 prepared by mixing equal amounts of Na[(R)-3] and Na[(S)-3]. See C. Kabuto, K. Kabuto, Y. Sasaki, T. Nishiyama, K. Umakoshi, *J. Chem. Soc., Chem. Commun.* **1993**, 381.

Data collected on a Rigaku Saturn CCD diffractometer using MoK α radiation and refined by least-squares method on F^2 with the SHELXH-97 program (G. M. Sheldrick, SHELX-97,

Program for the Solution of Crystal Structures, Universität Göttingen, **1997**). Crystallographic data have been deposited with Cambridge Crystallograpic Data Centre as supplementary publication no. CCDC 651161. The data can be obtained free charge from The Cambridge Crystallographic Data Centre via <u>www.ccdc.cam.ac.uk/data</u> request/cif.

Crystal data: $[Sm(C_{11}H_{14}O_8N_4)(H_2O)_2]_6Na_6^+ \cdot 30H_2O; M_r=3610.42$, monoclinic, space group $P2_1/n$ (No. 14); crystal sizes = 0.4x0.2x0.15mm³, a = 14.701(1) b = 36.584(17), c = 25.677(1)Å, $\beta = 90.134(3)^\circ$, V=13809(11) Å³; Z = 4, $\rho_{calad} = 1.736 \text{ g/cm}^3$, μ (Mo-K α) = 2.645 mm⁻¹, $\lambda = 0.71073$ Å, F(000) = 7224, ω -scan mode, $\theta_{min-max} = 3.0-27.0^\circ$, T = 173K, total data = 158740, unique data = 31008 ($R_{int} = 0.075$), 1680 parameters, $R_1(19482$ data with $I > 2\sigma(I)$) = 0.0511 and $wR_2(\text{all reflections}) = 0.1728$, GOF = 1.000, $\rho_{max} = 3.40 \text{ eÅ}^3$, ρ_{min} =-2.10 eÅ³. Non-hydrogen atoms in the cationic part and Na atoms were refined anisotropically, and H atoms were fixed. Of which 30 independent guest water molecules, 26 oxygen atoms were refined anisotropically (occupancy = 1.0) and 4 atoms isotropically (occupancy = 0.5), where no H atoms were applied.

NMR shift study on amino acid mixtures:

A mixture, consisted of equimolar $2\sim10$ amino acids (0.096 mmol in total), was dissolved in 1.6 ml of D₂O. After pH of the solution was adjusted to 10.1 using D₂O solutions of NaOD and DCl (0.1-0.01 M), 0.6 ml portions of the solution were taken in two separated sample tubes. To one of the two tubes, aliquots of 0.36 M D₂O solution (pH adjusted to 8.0) of (*R*)-**1** were added successively. By the same procedure, D₂O solution of (*S*)-**1** was added to the other. NMR spectra were determined on each sample solution.

Hydrolysis of [*D-Ala*², *Met*⁵]*-enkephalin*: Commercially available [D-Ala², Met⁵]*-enkephalin* acetic acid salt (Peptide Institute Inc. (Japan), 25 mg, 0.038 mmol) was dissolved in 10 ml of 5.7 M HCl and was heated at 110 °C for 24 h in a sealed tube. After the solution was concentrated to dryness under reduced pressure, the residue was dissolved in 1.6 ml of D₂O. After adjusting pH to 10 with NaOD-D₂O solution, the NMR shift study was conducted at 400 MHz.

A typical procedure for assigning the NMR signals measured for amino acid mixtures.

1. Assignment of each ¹³C signal of mixtures by comparison with the NMR spectra of typical amino acids



in D_2O pH adjusted around 10.

 ^{13}C NMR spectra of single amino acids in D₂O (pH ~10).

Dependence of the chemical shifts on the concentration and induced shift on the ¹³C signals of substrate amino acids by the Sm reagent were sufficiently small therefore comparison of the ¹³C NMR spectrum of the mixture with those of single amino acids is useful to assign the signals observed for mixture.

 13 C NMR spectrum of an amino acid mixture containing L-Ala, D-Val, L-Ser, D-Asn, L-Glu, and L-4-hydroxyproline (entry 4 in table 1) in D₂O (pH ~10). Signals marked with circles can be assigned as those of uncommon amino acid (L-4-hydroxyproline).

2. Assignment of ¹H signal from C-H correlation.



C-H COSY spectrum of an amino acid mixture containing L-Ala, D-Val, L-Ser, D-Asn, L-Glu, and L-4-hydroxyproline (entry 4 in table 1) in D_2O (pH ~10).

Each ¹H signal can be assigned on the basis of C-H correlation spectrum.

- 3. Confirmation of the assignment of 1H signals by H-H correlation spectrum.
- A. H-H COSY of spectrum of a six amino acid mixturte (Table 1, entry 4).



B. TOCSY spectrum of a eight amino acid mixturte in the presence of Sm{(S)-pdta-d8} (Table 1, entry 3).





Figure S1. Partial ¹H NMR spectra of the mixture of eight amino acids (L-Ala, D-Val, D-Met, D-Phe, L-Ser, L-Glu, L-Asn, L-His) in the presence of enantiomeric **3**. ¹H NMR (400 MHz, D_2O), pH 10.1, [Sm] = 6.0 mM, concentration of each amino acid = 7.5 mM. Upper: in the presence of (*R*)-**3**, Lower: in the presence of (*S*)-**3**.