

## Internal electron transfer in multi-site redox enzymes is accessed by laser excitation of thiouredopyrene-3,6,8-trisulfonate (TUPS)

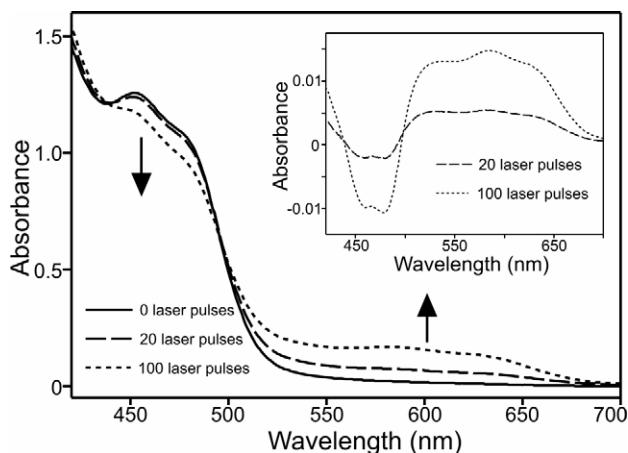
Derren J. Heyes<sup>a</sup>, Anne-Marie Quinn<sup>a</sup>, Paul M. Cullis<sup>b</sup> Michael Lee<sup>b</sup>, Andrew W. Munro<sup>a</sup>, and Nigel S. Scrutton<sup>a\*</sup>

\*Faculty of Life Sciences, Manchester Interdisciplinary Biocentre, University of Manchester, 131 Princess St., Manchester, M1 7DN, UK

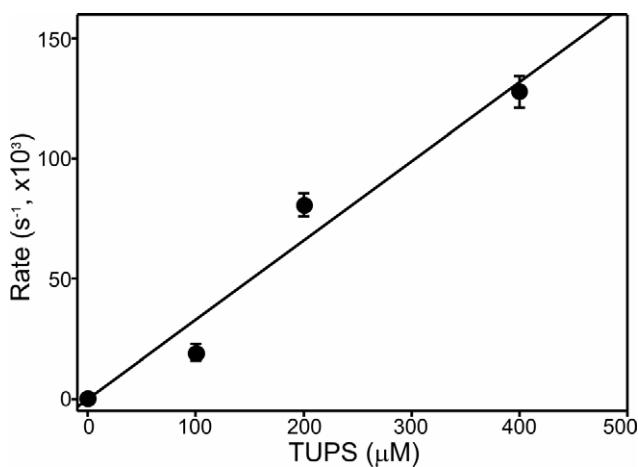
<sup>†</sup>Department of Chemistry, University of Leicester, Leicester, LE1 7RH, UK

### Supplementary figures

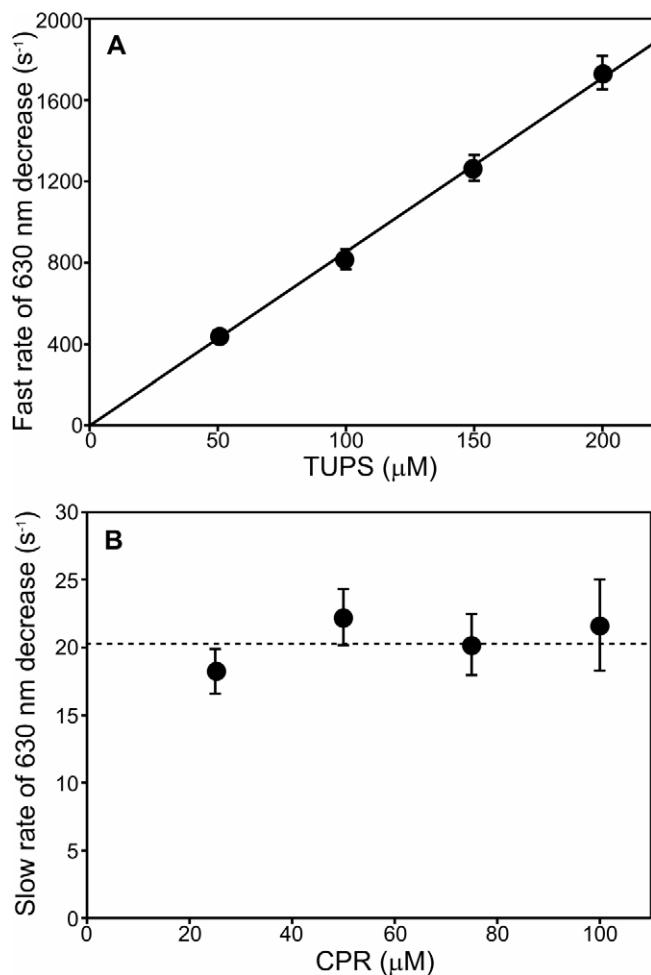
**Figure S1** Absorbance spectra of 50  $\mu$ M CPR in the presence of 200  $\mu$ M TUPS before laser photoexcitation and after excitation with 20 and 100 laser pulses at 355 nm. The inset shows the absorbance difference spectra, using a non-excited sample as a blank.



**Figure S2** The rate of electron transfer from photoexcited TUPS to CPR measured at a range of TUPS concentrations. The error bars were calculated from the average of at least 5 transients.



**Figure S3.** Dependence of slower kinetic phases on the concentration of TUPS and CPR. (A) The rate of the initial phase of decrease in absorbance at 630 nm measured over a range of TUPS concentrations. The error bars were calculated from the average of at least 5 transients. (B) The rate of the slower phase of decrease in absorbance at 630 nm measured over a range of CPR concentrations. The error bars were calculated from the average of at least 5 transients.



**Figure S4** Absorbance spectra of TUPS-nNOS reductase sample after covalent attachment to the protein as described in the *Experimental* section. The TUPS and nNOS reductase absorption peaks are indicated.

