Electronic Supplementary Information

Dialysis process for the removal of surfactants to form colloidal mesoporous silica nanoparticles

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Materials

10 Hexadecyltrimethylammonium bromide (C₁₆TMABr), triethanolamine (TEA), Rhodamine 6G (R6G), and acetic acid were purchased from Wako Pure Chemical Industries, Ltd. Tetramethoxysilane (TMOS: Si(OCH₃)₄) was purchased from Kishida Chemical Ltd. Ethanol was purchased from Junsei Chemical Co., Ltd.

15 Experimental

1) Preparation of colloidal mesostructured silica (CMSS)

TEA (0.420 g) and 2.00 g of C_{16} TMABr was added to 240 ml of water and the solution was stirred for 30 min at 80 °C. Then 11 mmol of TMOS was added to the C_{16} TMABr solution and stirred vigorously for 2 h at 80 °C. Finally, the colloid was filtered with a filtering paper (No. 5). The molar 20 ratio of the precursor solution was 1 TMOS : 0.50 C_{16} TMABr : 0.25 TEA : 1200 H₂O.

2) Removal of surfactants from CMSS

In the next step, 50 ml of the colloid was transferred into a dialysis membrane tube composed of cellulose (Molecular Weight Cut Off = 12000-14000) and was dialyzed for 24 h against 250 ml of the mixture containing 2M acetic acid and ethanol (1:1, v/v) to remove C₁₆TMABr and this process was

25 repeated three times. Then the tube, which contained colloidal mesoporous silica (CMPS), was immersed in water to remove acetic acid / EtOH in the tube, and the process was repeated three times. CMPS was filtered through a 0.45 μm regenerated cellulose syringe filter.

3) Adsorption of Rhodamine 6G into CMPS

A tube containing 10 ml of CMPS was immersed into a R6G solution (40 ml, 50 mg/l) and the 30 solution was stirred to accelerate the transfer of the dye for 12 h at room temperature. Finally, the concentrations of R6G inside and outside the tube were determined by the absorbance at 526 nm.

Characterization

Dynamic light scattering (DLS) measurements were conducted with a HORIBA LB-550 Dynamic 35 Light Scattering Nanoparticle Size Analyzer at 17 °C. Zeta-potential measurements were conducted with an OTSUKA ELECTRONICS ELSZ-1 at 20 °C. UV-vis spectra were obtained by using a SHIMADZU UV-2500PC spectrophotometer. Thermogravimetry differential thermal analysis (TG-DTA) measurements were carried out with a RIGAKU Thermo Plus 2 instrument under a dry air flow at a heating rate of 10 °C min⁻¹ up to 800 °C. CHN elemental analysis was performed with a

40 Perkin-Elmer, 2400 Series II. The Nitrogen gas adsorption-desorption measurement were performed by an Quantachrome Instruments Autosorb-2 instrument at 77K. Samples were preheated at 120 °C for 3 h under 1×10^{-2} Torr. The Brunauer–Emmett–Teller (BET) surface area was calculated from the adsorption data in the relative pressure range from 0.05 to 0.30. The pore size distribution was evaluated using the non-local density functional theory (NLDFT). SEM and bright field STEM studies were carried out on HITACHI S-5500 microscope operated at 30kV. SEM samples, CMSS or CMPS were dropped and dried carbon-coated micro-grid (Okenshoji Co.). TEM studies were carried out on JEOL JEM-2010 microscope operated at 200kV. TEM samples, CMSS or CMPS were dropped and dried on a carbon-coated micro-grid (Okenshoji Co.). SAXS 2D-images and patterns of the samples 5 were obtained on a RIGAKU Nano-Viewer. The samples for the SAXS measurement were used as colloidal state. The particle diameter distributions were calculated based on the scattering profiles by

colloidal state. The particle diameter distributions were calculated based on the scattering profiles by using Nano-solverTM. The powder XRD patterns of the dried samples were obtained on a Mac Science M03XHF22 diffractometer with Mn-filtered Fe K α radiation.





Fig. S1 Transmittance spectra of CMSS and CMPS.



Fig. S2 (a) N₂ sorption isotherm, BET surface area, and pore volume of d-CMPS, (b) Pore size distribution of d-CMPS obtained by NLDFT method, and (c) XRD pattern of d-CMPS. 5



Fig. S3 (left) Small-angle X-ray scattering (SAXS) 2D-images of CMSS and CMPS. (right) SAXS 5 patterns of CMSS and CMPS. (inset) Calculated particle diameter distributions based on the scattering profiles by using Nano-solverTM.



Fig. S4 TEM images of CMSS. a) low magnification image (scale bar: 200 nm) and b) high 5 magnification image of the square in the image (a) (the size of the white circle is 20 nm in diameter).





Fig. S5 Visual appearance of (a) CMSS (C_{16} TMABr / Si = 0.50) and (b) aggregated CMSS 5 (C_{16} TMABr / Si = 0.13).





Fig. S6 SEM images of CMPS. (a) Arrow indicates the presence of "barrel-like" mesostructures (scale 5 bar: 100 nm) and (b) Magnified image of the area indicated by an arrow in the image (a) (scale bar: 20 nm).