

Electronic Supplementary Information

Direct Catalytic Asymmetric Aldol Reaction of β -Keto Esters with Formaldehyde Promoted by a Dinuclear Ni₂-Schiff Base Complex

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Experimental Section

General:

Infrared (IR) spectra were recorded on a JASCO FT/IR 410 Fourier transform infrared spectrophotometer. NMR spectra were recorded on JEOL JNM-LA500, ECX500, or ECA500 spectrometers, operating at 500 MHz for ¹H NMR and 125.65 MHz for ¹³C NMR. Chemical shifts in CDCl₃ were reported in the scale relative to tetramethylsilane (0 ppm) for ¹H NMR. For ¹³C NMR, chemical shifts were reported in the scale relative to CHCl₃ (77.0 ppm) as an internal reference. Column chromatography was performed with silica gel Merck 60 (230-400 mesh ASTM). Optical rotations were measured on a JASCO P-1010 polarimeter. ESI mass spectra were measured on Waters micromass ZQ (for LRMS) and FAB mass spectra (for HRMS) were measured on a JEOL JMS-700 spectrometer. The enantiomeric excess (ee) was determined by HPLC analysis. HPLC was performed on JASCO HPLC systems consisting of the following: pump, PU-2080 plus; detector, UV-2075 plus, measured at 280 nm; column, DAICEL CHIRALPAK AS-H or AD-H; mobile phase, hexane-2-propanol. Diisopropylether (*i*Pr₂O) was distilled from sodium benzophenone ketyl. Other reagents were purified by the usual methods. Formaldehyde solution (37% w/w in H₂O) and paraformaldehyde were purchased from Aldich. β -Keto esters^[S1, S2] were prepared by following the same procedures as described in literatures.

References

[S1] C. Palomo, M. Oiarbide, J. M. García, P. Bañuelos, J. M. Odriozola, J. Razkin, A. Linden, *Org. Lett.* **2008**, *10*, 2637.

[S2] T. B. Poulsen, L. Bernardi, M. Bell, K. A. Jørgensen, *Angew. Chem., Int. Ed.* **2006**, *45*, 6551.

Preparation of Ni₂-Schiff Base 1 Complex:

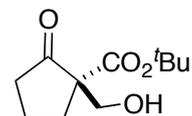
To a solution of (*R,R*)-Schiff base ligand **1** (400 mg, 0.76 mmol) in EtOH (7.6 mL), was added Ni(OAc)₂•4H₂O (378 mg, 1.52 mmol), and the mixture was stirred for 12 h under reflux. After cooling down to room temperature, H₂O (1.0 mL) was added to the mixture. The precipitate (Ni₂/Schiff base **1** complex) was collected by filtration. Then, the solid was washed with hexane and EtOH. The solid was dried under reduced pressure at 50 °C to afford the Ni₂-Schiff base **1** complex (417 mg) as a dark yellow solid. The complex was used for the asymmetric reaction without further purification, and was stored under Ar at room temperature. Catalytic activity did not change for 5 months.

General Procedure for Catalytic Asymmetric Hydroxymethylation of β-Keto Esters Using a Ni₂-Schiff Base 1 Catalyst:

To a stirred solution of the Ni₂/Schiff base **1** catalyst (0.096 mg, 0.15 μmol) in *i*Pr₂O (7.5 mL) was added β-keto ester **2a** (26.2 mg, 0.15 mmol). To the mixture at 40 °C was added 37% aqueous formaldehyde solution (21.7 mg, 0.165 mmol), and the resulting suspension was stirred for 1 h. The mixture was diluted with diethyl ether, and the precipitate was removed by filtration through a pad of Celite. After the filtrate was concentrated under reduced pressure, the residue was purified by silica gel flash column chromatography (hexane/ethyl acetate = 3/1) to give the desired product **3a** (30.2 mg, 94% yield) as a colorless oil.

(*S*)-*tert*-butyl 1-(hydroxymethyl)-2-oxocyclopentanecarboxylate (**3a**)

colorless oil; IR (neat) ν 3460, 2973, 1748, 1721 cm⁻¹; ¹H NMR (CDCl₃) δ 1.45 (s, 9H), 1.93-2.11 (m, 2H), 2.14-2.20 (m, 1H), 2.25-2.36 (m, 2H), 2.42-2.50 (m, 1H), 2.74 (dd, *J* = 4.6, 9.0, 1H), 3.75 (dd, *J* = 9.0, 12.0, 1H), 3.85 (dd, *J* = 4.6, 12.0, 1H); ¹³C NMR (CDCl₃) δ 19.7, 27.9, 31.3, 38.4, 62.2, 63.8, 82.6, 170.8, 215.4; LRMS(ESI): *m/z* 237 [M+Na]⁺; HRMS (FAB): *m/z* calculated for C₁₁H₁₈O₄Cs⁺ [M+Cs]⁺: 347.0260, found: 347.0272, [α]_D^{16.0} +6.3 (*c* 1.00, CHCl₃); HPLC (DAICEL CHIRALPAK AS-H, hexane/2-propanol

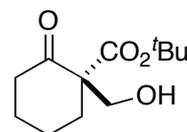


= 95/5, flow 0.5 mL/min, detection at 280 nm) t_R 20.4 min (major) and 25.2 min (minor).

(S)-tert-butyl 1-(hydroxymethyl)-2-oxocyclohexanecarboxylate (3b)

colorless oil; IR (neat) ν 3463, 2978, 1709 cm^{-1} ; ^1H NMR (CDCl_3)

δ 1.49 (s, 9H), 1.49-1.63 (m, 3H), 1.78-1.82 (m, 1H), 2.02-2.08 (m, 1H), 2.27-2.32 (m, 1H), 2.41-2.47 (m, 1H), 2.60-2.68 (m, 1H), 2.85

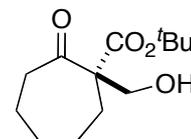


(dd, $J = 5.3, 9.7$ Hz, 1H), 3.68 (dd, $J = 9.7, 11.5$ Hz, 1H), 3.78 (dd, $J = 5.3, 11.5$ Hz, 1H); ^{13}C NMR (CDCl_3) δ 22.0, 26.9, 27.9, 32.9, 41.0, 63.1, 66.7, 82.7, 170.3, 211.3; LRMS(ESI): m/z 251 $[\text{M}+\text{Na}]^+$; HRMS (FAB): m/z calculated for $\text{C}_{12}\text{H}_{20}\text{O}_4\text{Cs}^+$ $[\text{M}+\text{Cs}]^+$: 361.0416, found: 361.0406; $[\alpha]_D^{19.0} +5.2$ (c 1.04, CHCl_3), HPLC (DAICEL CHIRALPAK AS-H, hexane/2-propanol = 95/5, flow 0.5 mL/min, detection at 280 nm) t_R 14.3 min (major) and 17.2 min (minor).

(S)-tert-butyl 1-(hydroxymethyl)-2-oxocycloheptanecarboxylate (3c)

colorless oil; IR (neat) ν 3535, 2976, 2932, 2862, 1701 cm^{-1} ; ^1H NMR

(CDCl_3) δ 1.48 (s, 9H), 1.58-1.64 (m, 4H), 1.74-1.84 (m, 3H), 1.91-1.96 (m, 1H), 2.49- 2.54 (m, 1H), 2.71-2.76 (m, 1H), 3.04 (dd, $J =$

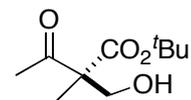


4.9, 9.7 Hz, 1H), 3.68 (dd, $J = 9.7, 11.4$ Hz, 1H), 3.94 (dd, $J = 4.9, 11.4$ Hz, 1H); ^{13}C NMR (CDCl_3) δ 25.2, 25.6, 27.9, 30.1, 30.9, 43.0, 64.4, 67.3, 82.6, 171.3, 212.6; ESI-MS m/z 265 $[\text{M}+\text{Na}]^+$; HRMS calcd. for $\text{C}_{13}\text{H}_{22}\text{O}_4\text{Cs}$ $[\text{M}+\text{Cs}]^+$: 375.0573, found 375.0561; $[\alpha]_D^{18.0} +4.6$ (c 1.23, CHCl_3); HPLC (DAICEL CHIRALPAK AD-H, hexane/2-propanol = 90/10, flow 0.5 mL/min, detection at 280 nm) t_R 13.1 min (major) and 16.1 min (minor).

(S)-tert-butyl 2-(hydroxymethyl)-2-methyl-3-oxobutanoate (3d)

colorless oil; IR (neat) ν 3419, 2979, 1735, 1715 cm^{-1} ; ^1H NMR

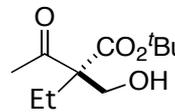
(CDCl_3) δ 1.35 (s, 3H), 1.48 (s, 9H), 2.23 (s, 3H), 2.73 (dd, $J = 6.9, 8.0$



Hz), 3.76 (dd, $J = 8.0, 11.5$ Hz), 3.85 (dd, $J = 6.9, 11.5$); ^{13}C NMR (CDCl_3) δ 17.2, 27.0, 27.9, 61.7, 66.6, 82.6, 171.5, 207.0; ESI-MS m/z 225 $[\text{M}+\text{Na}]^+$; HRMS calcd. for $\text{C}_{10}\text{H}_{18}\text{O}_4\text{Cs}$ $[\text{M}+\text{Cs}]^+$: 335.0260, found 335.0250; $[\alpha]_D^{19.0} +4.6$ (c 1.03, CHCl_3); HPLC (DAICEL CHIRALPAK AD-H, hexane/2-propanol = 90/10, flow 0.5 mL/min, detection at 280 nm) t_R 13.2 min (major) and 15.1 min (minor).

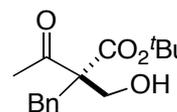
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colorless oil; $^1\text{H NMR}$ (CDCl_3) δ 0.90 (t, $J = 8.0$ Hz, 3H), 1.49 (s, 9H), 1.81-1.89 (m, 1H), 1.92-2.00 (m, 1H), 2.23 (s, 3H), 2.47 (dd, $J = 6.0, 8.0$ Hz, 1H), 3.85 (dd, $J = 8.0, 12.0$ Hz, 1H), 3.96 (dd, $J = 6.0, 12.0$ Hz, 1H); $^{13}\text{C NMR}$ (CDCl_3) δ 8.8, 24.3, 27.5, 27.9, 63.9, 65.9, 82.5, 171.1, 206.8; ESI-MS m/z 239 $[\text{M}+\text{Na}]^+$; HRMS calcd. for $\text{C}_{11}\text{H}_{20}\text{O}_4$ $[\text{M}+\text{Cs}]^+$: 349.0416, found 349.0408; $[\alpha]_{\text{D}}^{26.0} +5.3$ (c 1.11, CHCl_3); HPLC (DAICEL CHIRALPAK AS-H, hexane/2-propanol = 90/10, flow 0.5 mL/min, detection at 280 nm) t_{R} 12.3 min (major) and 13.1 min (minor).



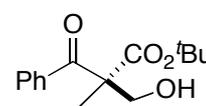
(S)-tert-butyl 2-benzyl-2-(hydroxymethyl)-3-oxobutanoate (3f)

colorless oil; IR (neat) ν 3463, 2978, 2938, 1709 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) δ 1.46 (s, 9H), 2.24 (dd, $J = 5.5, 7.5$, 1H), 2.27 (s, 3H), 3.18 (d, $J = 14.0$ Hz, 1H), 3.25 (d, $J = 14.0$, 1H), 3.79 (dd, $J = 7.5, 12.0$ Hz, 1H), 3.93 (dd, $J = 5.5, 12.0$ Hz, 1H), 7.18-7.29 (m, 5H); $^{13}\text{C NMR}$ (CDCl_3) δ 27.87, 27.92, 36.5, 63.7, 66.7, 82.9, 127.0, 128.3, 130.1, 135.9, 170.2, 205.6; ESI-MS m/z 301 $[\text{M}+\text{Na}]^+$; HRMS calcd. for $\text{C}_{16}\text{H}_{22}\text{O}_4$ $[\text{M}+\text{Cs}]^+$: 411.0573, found 411.0580; $[\alpha]_{\text{D}}^{26.0} +2.6$ (c 1.23, CHCl_3); HPLC (DAICEL CHIRALPAK AD-H, hexane/2-propanol = 9/1, flow 0.5 mL/min, detection at 254 nm) t_{R} 15.0 min (major) and 18.6 min (minor).



(S)-tert-butyl 2-benzyl-2-(hydroxymethyl)-3-oxo-3-phenylpropanoate (3g)

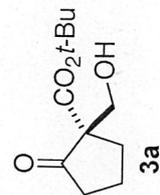
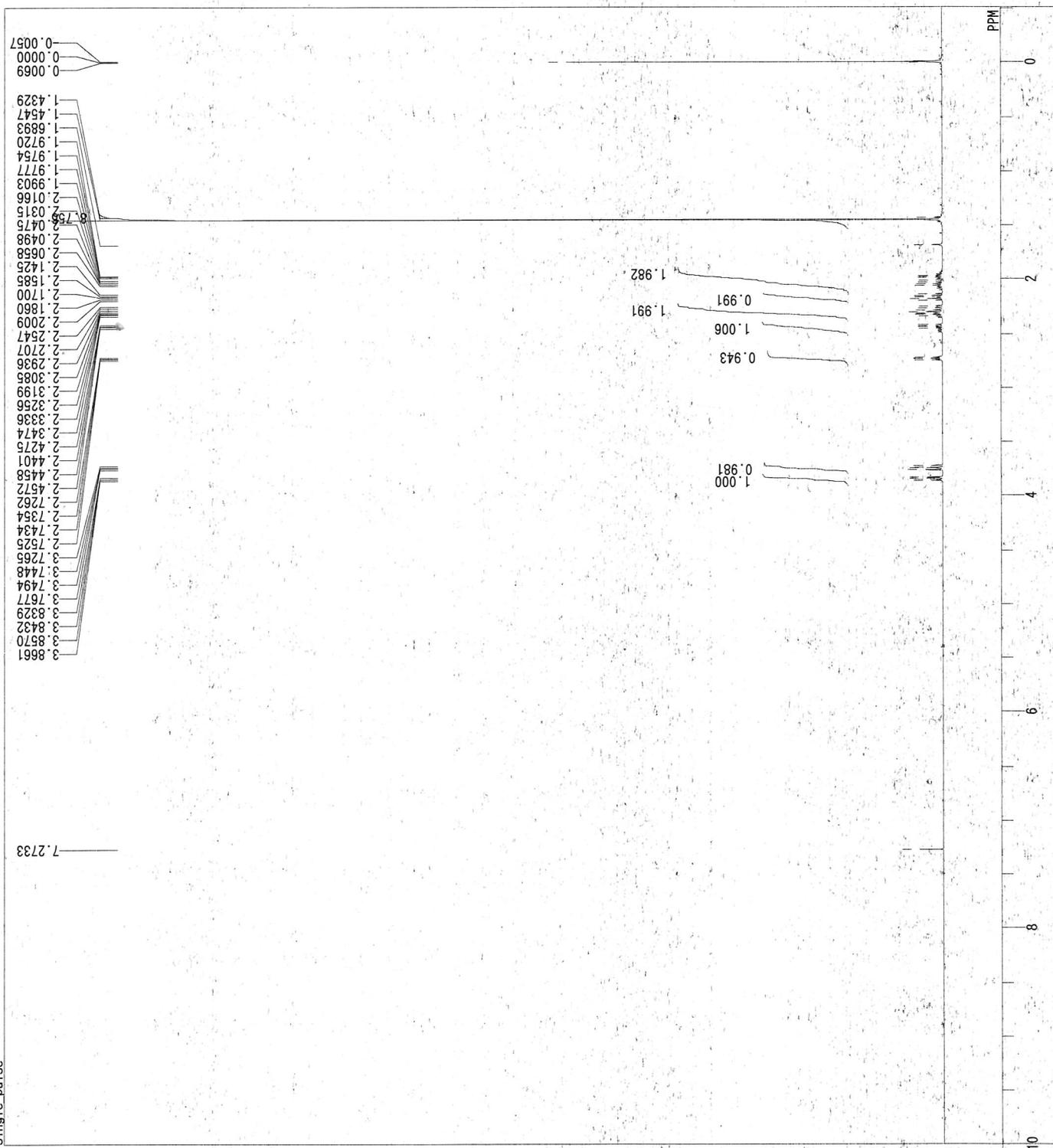
colorless solid; mp 66-70 $^{\circ}\text{C}$; IR (neat) ν 3487, 2976, 2932, 1718, 1679 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) δ 1.32 (s, 9H), 1.58 (s, 3H), 2.79 (dd, $J = 6.9, 6.9$, 1H), 3.93 (d, $J = 6.9, 11.5$ Hz, 1H), 4.01 (dd, $J = 6.9, 11.5$ Hz, 1H), 7.42-7.46 (m, 2H), 7.53-7.57 (m, 1H), 7.86-7.89 (m, 2H); $^{13}\text{C NMR}$ (CDCl_3) δ 18.6, 27.6, 59.6, 68.0, 82.6, 128.4, 128.8, 133.0, 135.4, 171.8, 199.3; ESI-MS m/z 287 $[\text{M}+\text{Na}]^+$; HRMS calcd. for $\text{C}_{15}\text{H}_{20}\text{O}_4$ $[\text{M}+\text{Cs}]^+$: 397.0416, found 397.0414; $[\alpha]_{\text{D}}^{13.0} +2.6$ (c 1.00, CHCl_3); HPLC (DAICEL CHIRALPAK AD-H, hexane/2-propanol = 9/1, flow 0.5 mL/min, detection at 254 nm) t_{R} 13.9 min (major) and 15.3 min (minor).



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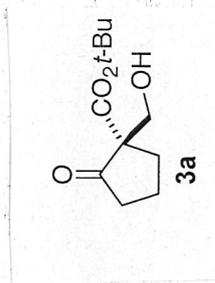
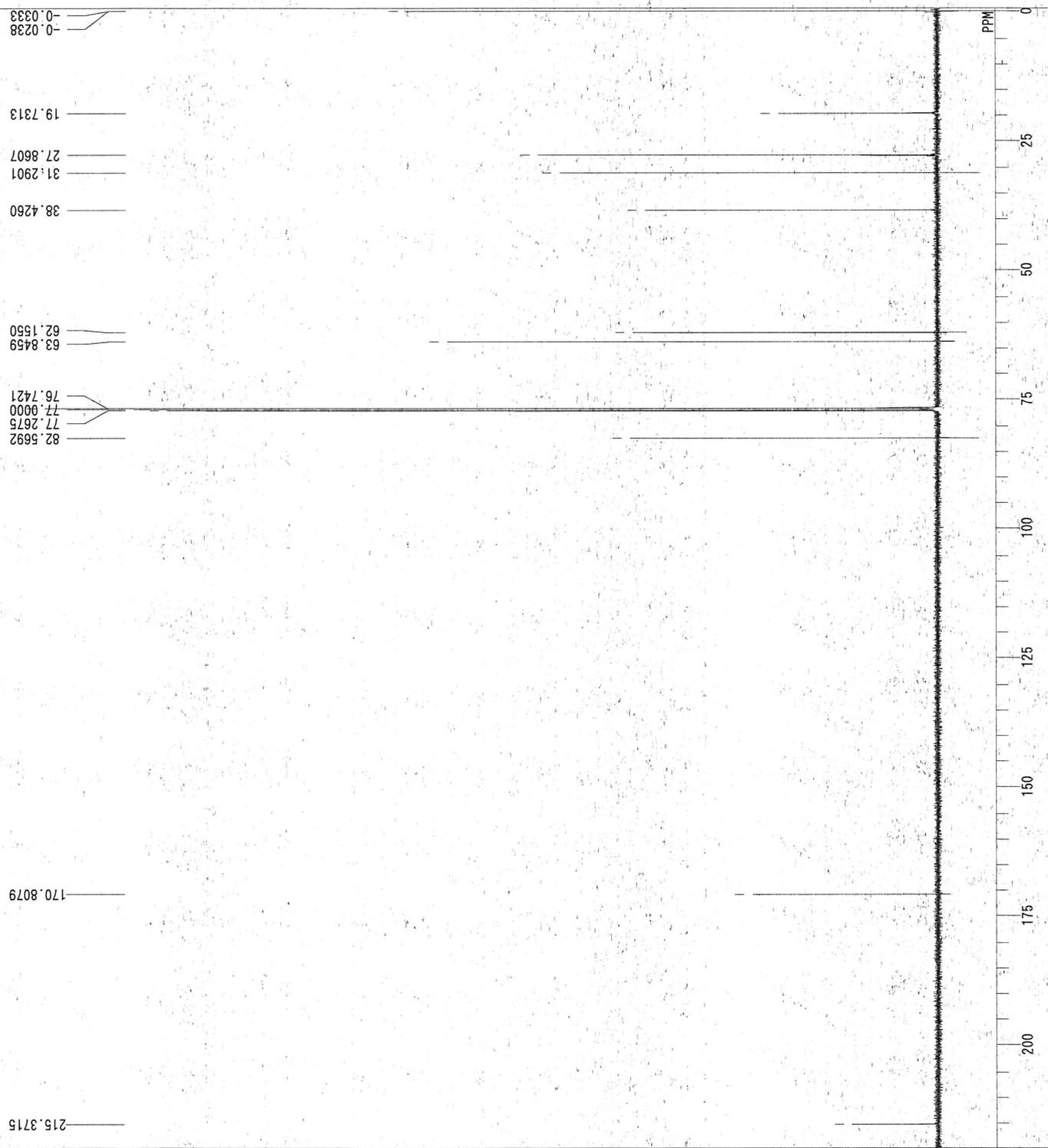
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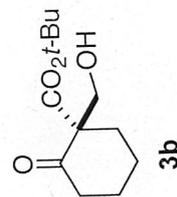
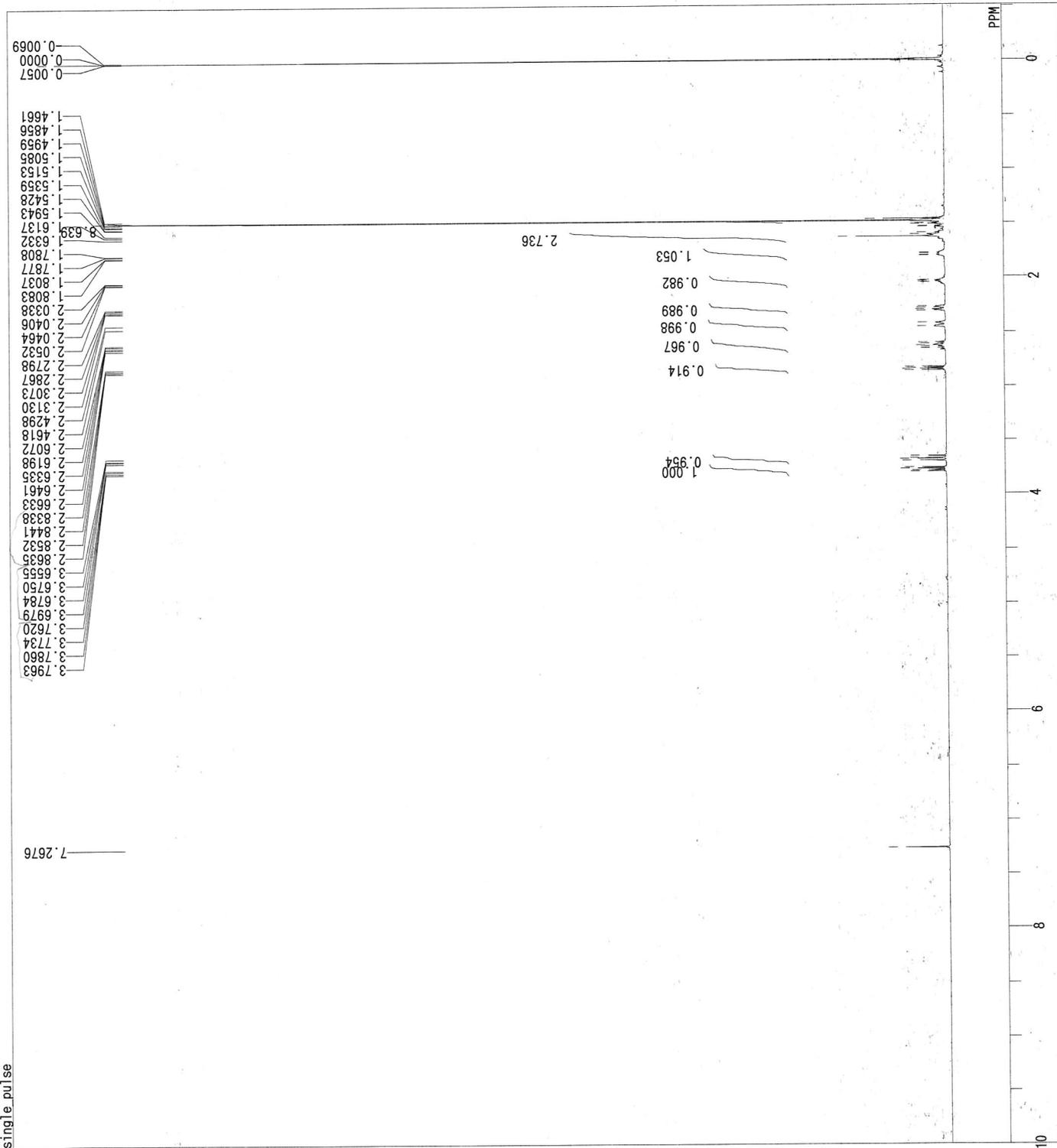
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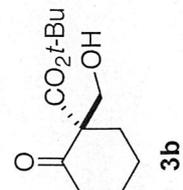
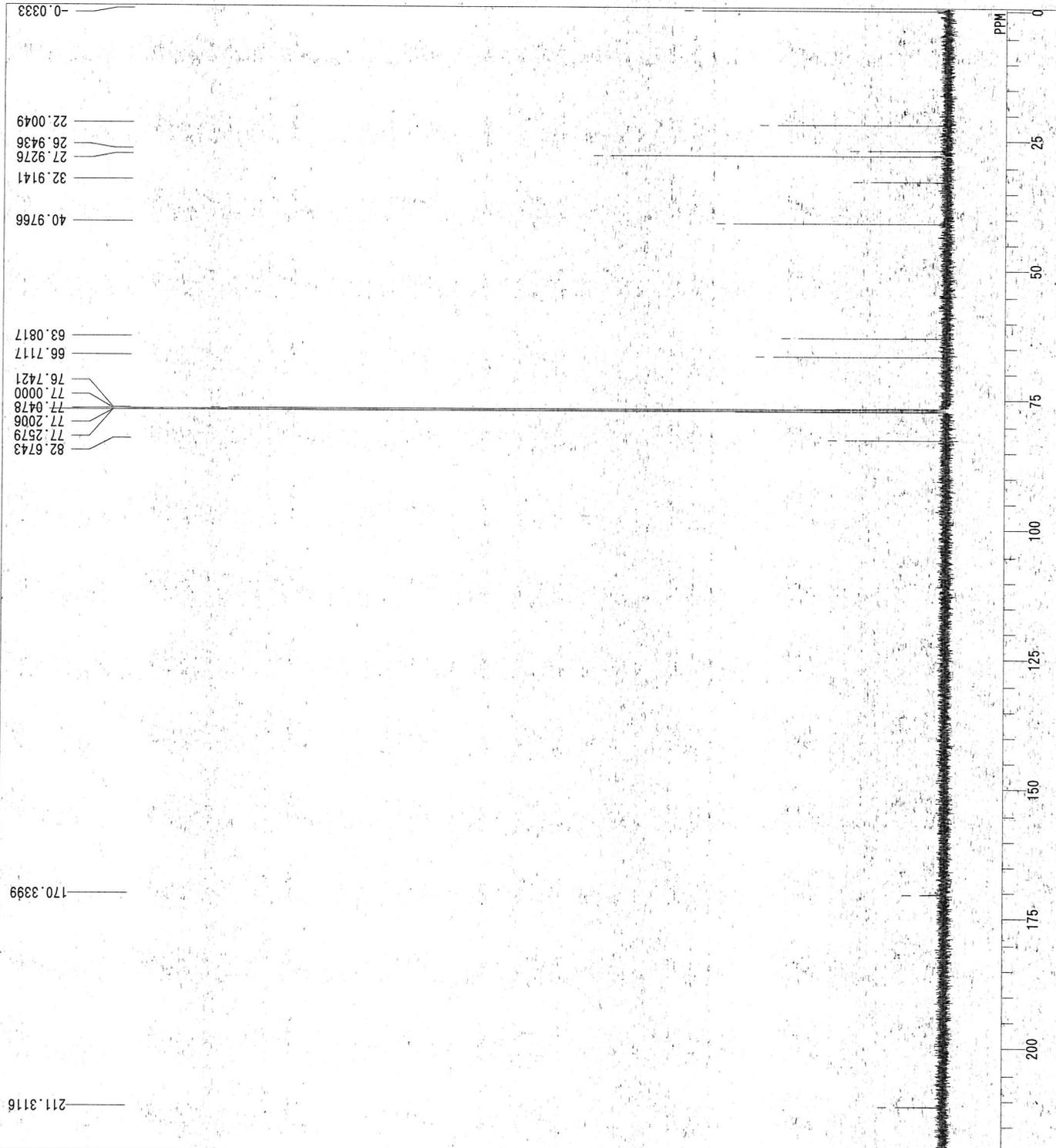


3b

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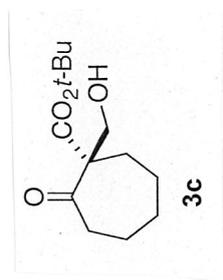
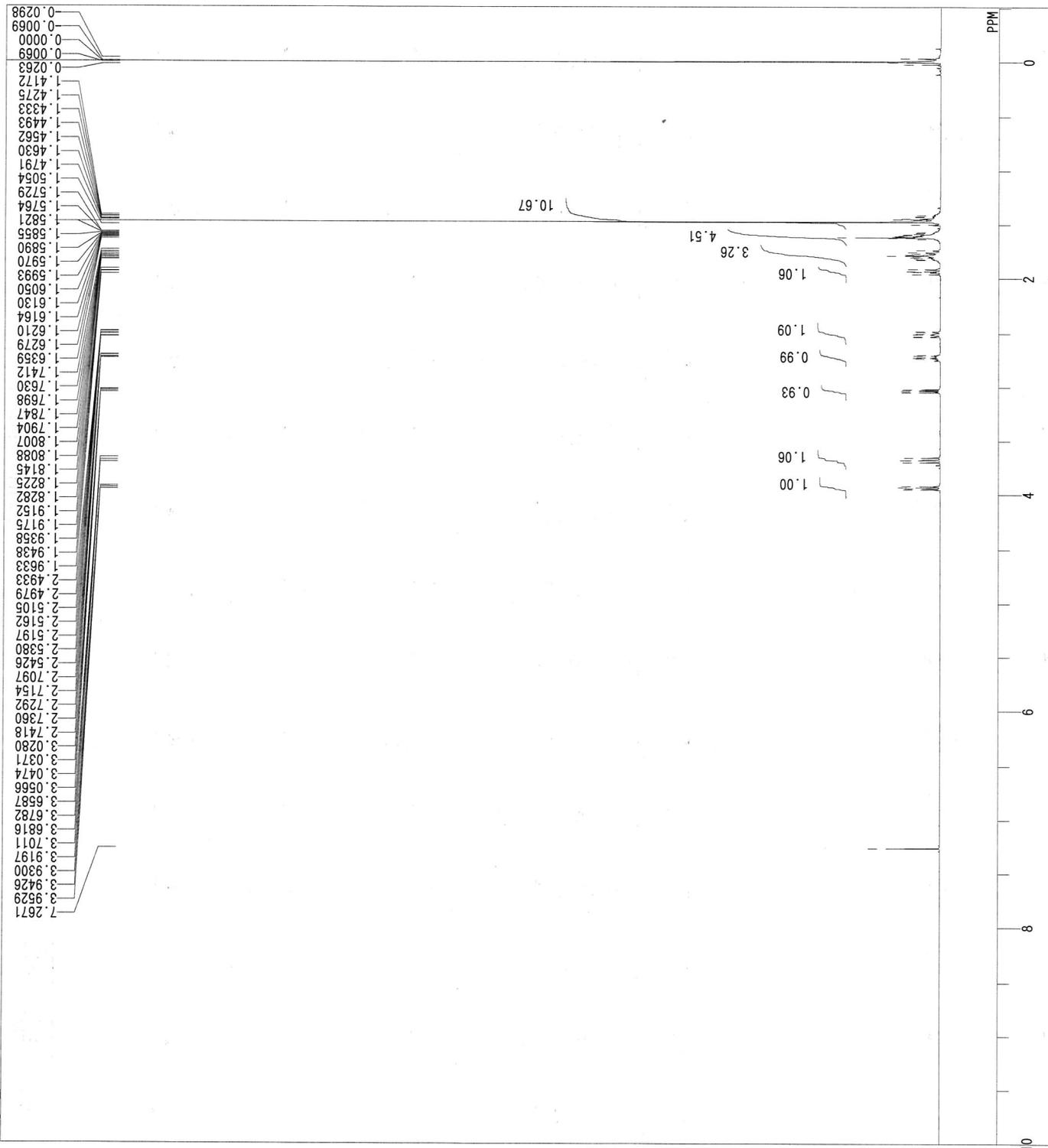
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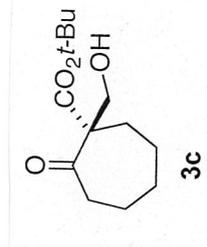
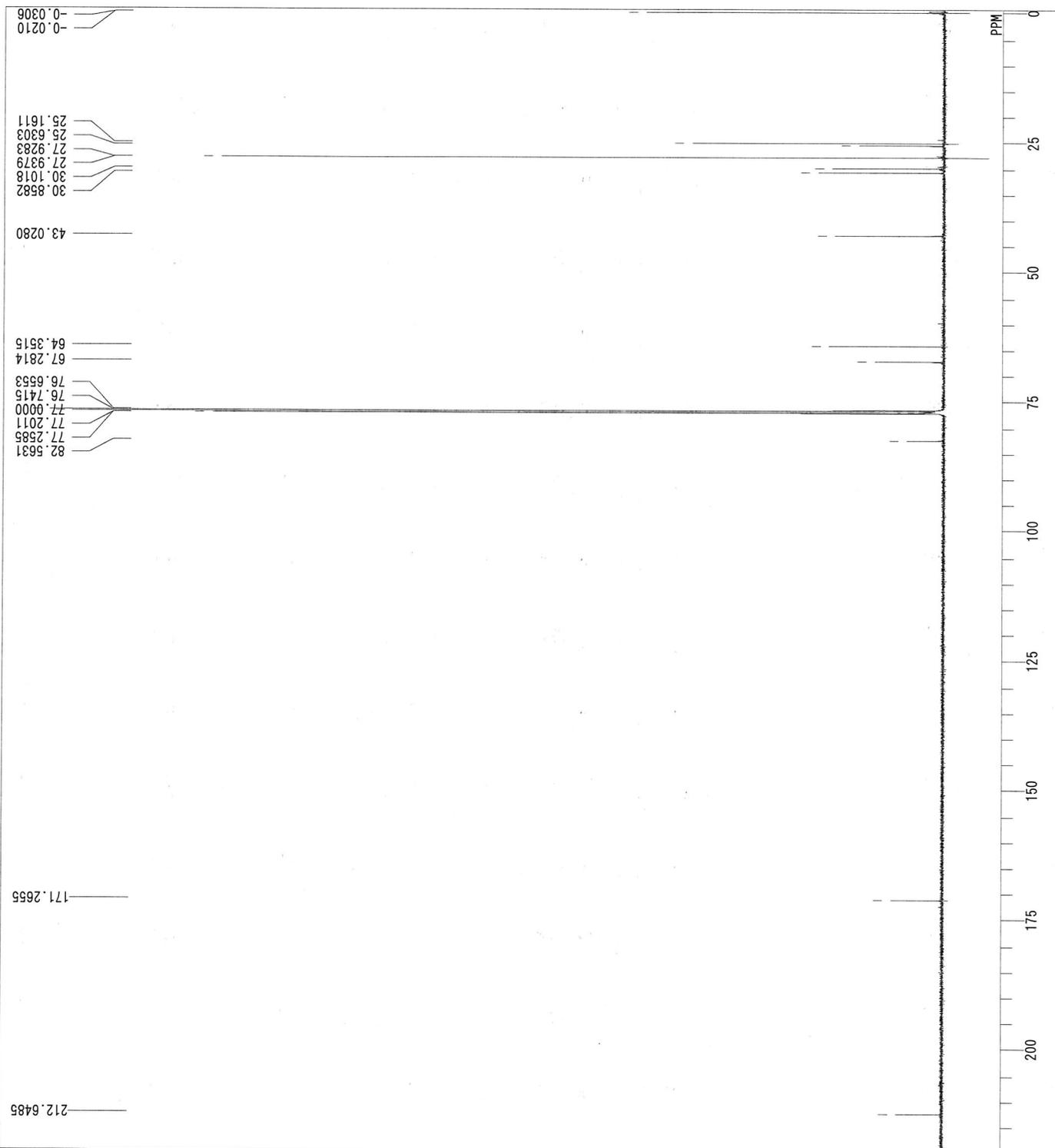
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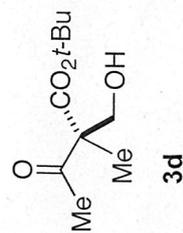
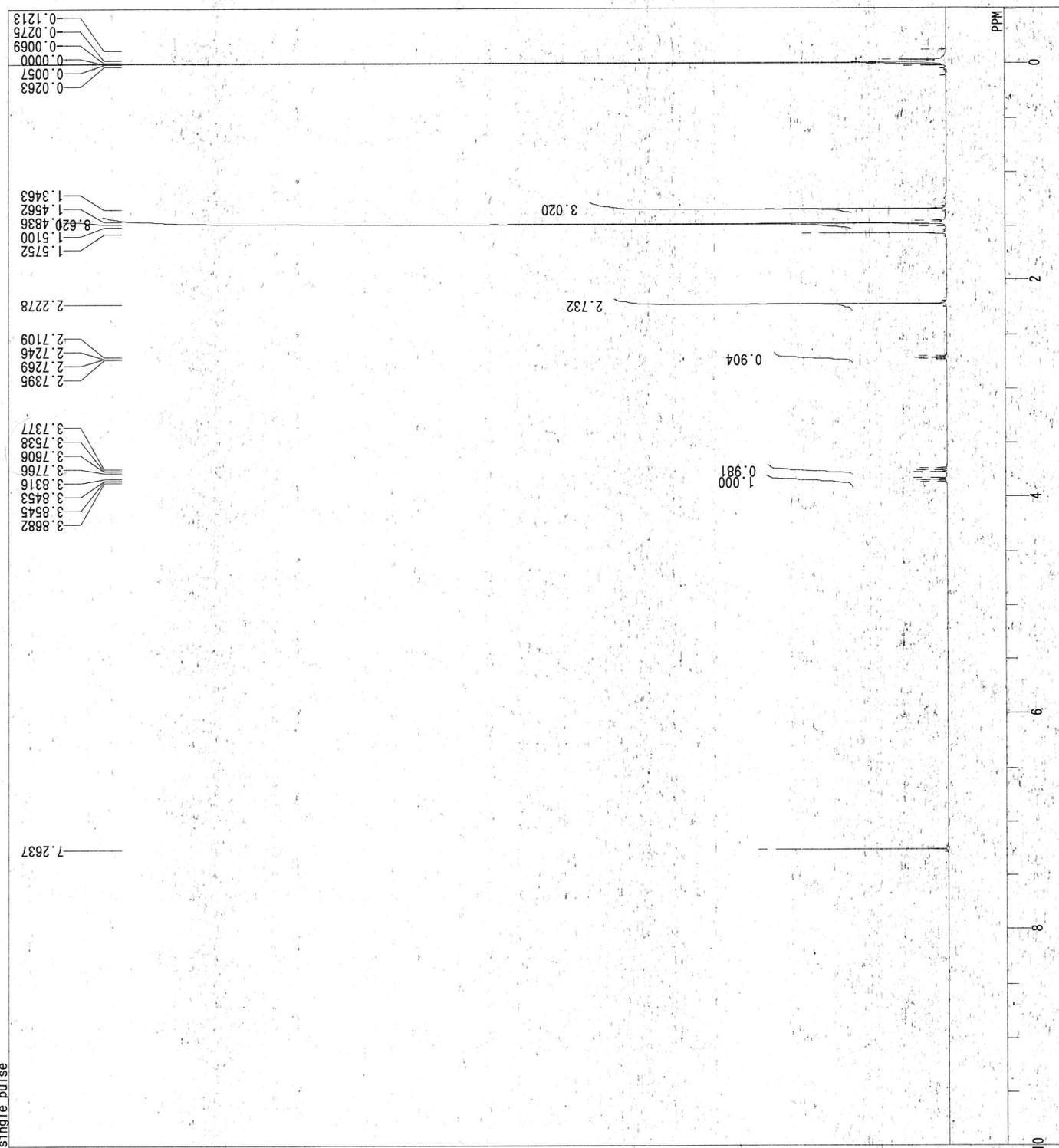
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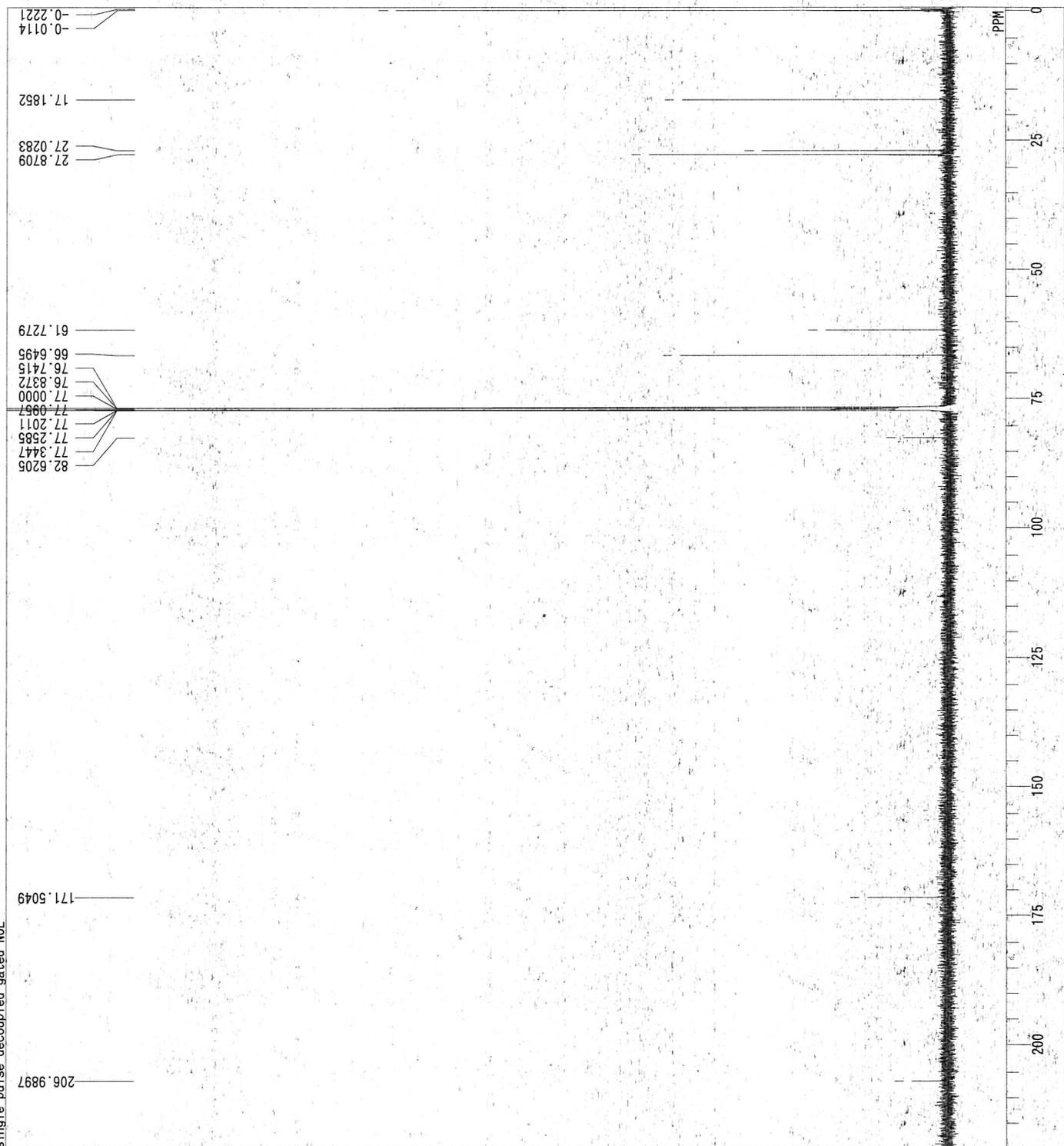


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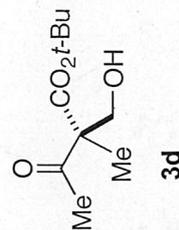


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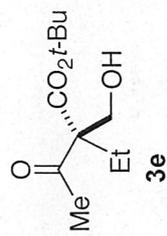
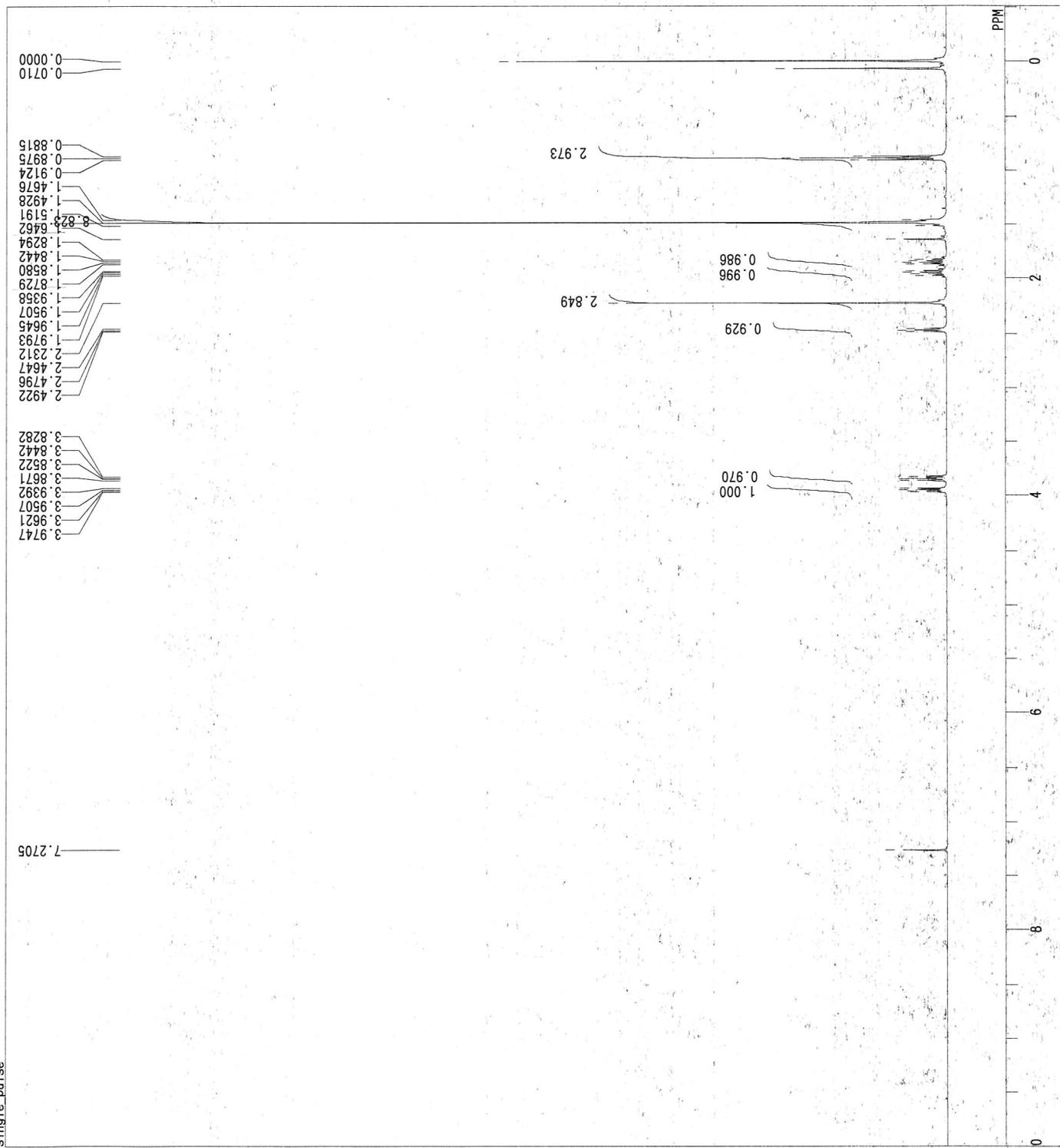
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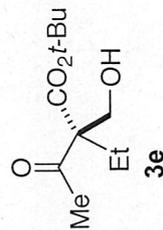
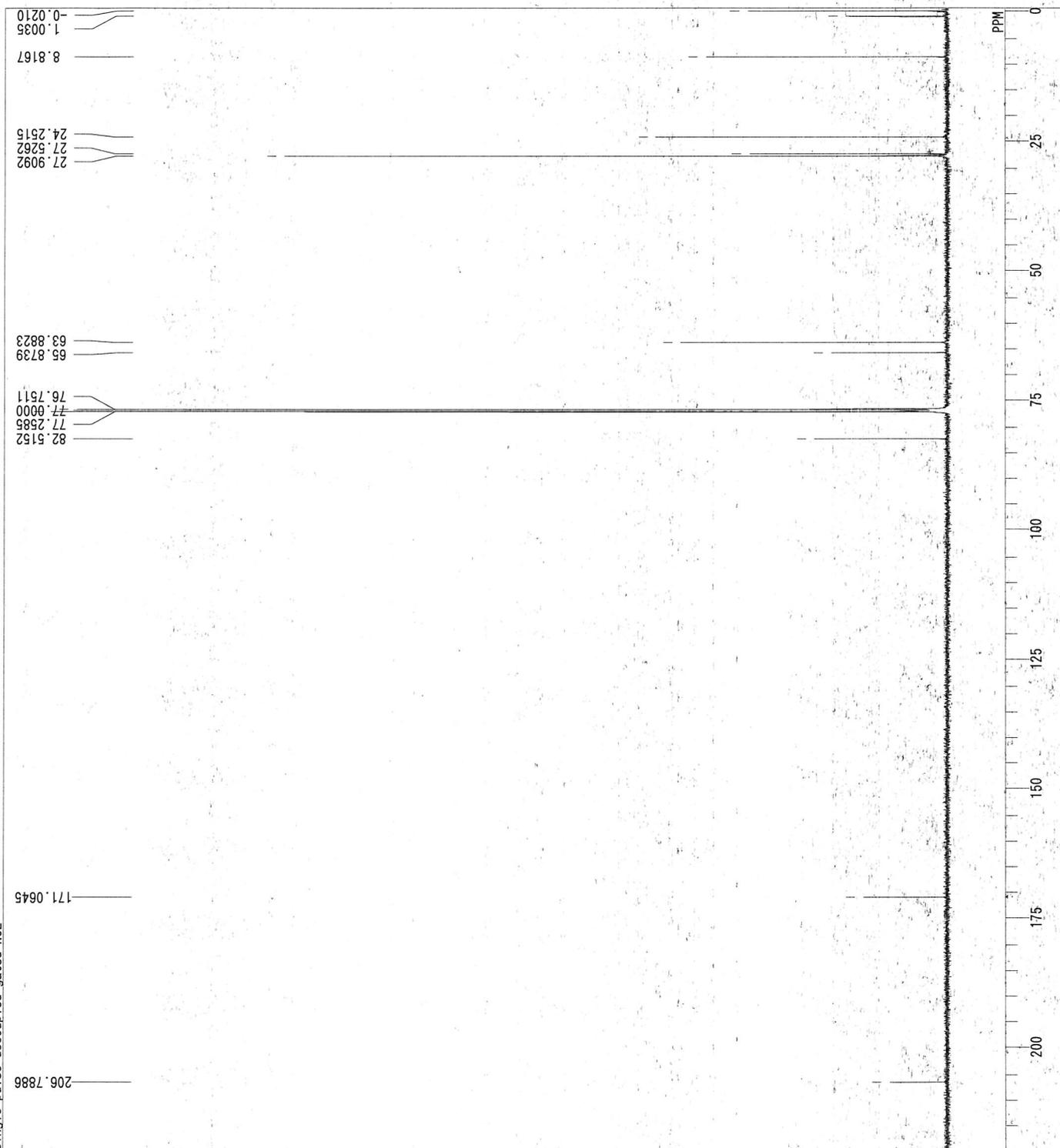
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 05-03-2009 15:13:43
 1H
 single_pulse_ex2
 495.13 MHz
 4.38 KHz
 9.64 Hz
 16384
 9286.78 Hz
 8
 1.7642 sec
 5.0000 sec
 6.20 usec
 1H
 18.6 c
 CDCl3
 0.00 ppm
 1.00 Hz
 44



C:\My Documents\PGousei\SMO\SMO-07-060-13C.1
single pulse decoupled gated NOE

DFILE C:\My Documents\PGousei\SMO\SMO-07-060-13C.1
COMMT single pulse decoupled gated NOE
DATIM 05-03-2009 17:21:48
OBNUC 13C
EXMOD single pulse dec
OBFRQ 124.51 MHz
OBSET 3.45 KHz
OBFIN 6.00 Hz
POINT 32768
FREOU 39062.50 Hz
SCANS 2650
ACQTM 0.8889 sec
PD 2.0000 sec
PW1 3.57 usec
IRNUC 1H
CTEMP 19.7 c
SLVNT CDCl3
EXREF 77.00 ppm
BF 1.00 Hz
RGAIN 60



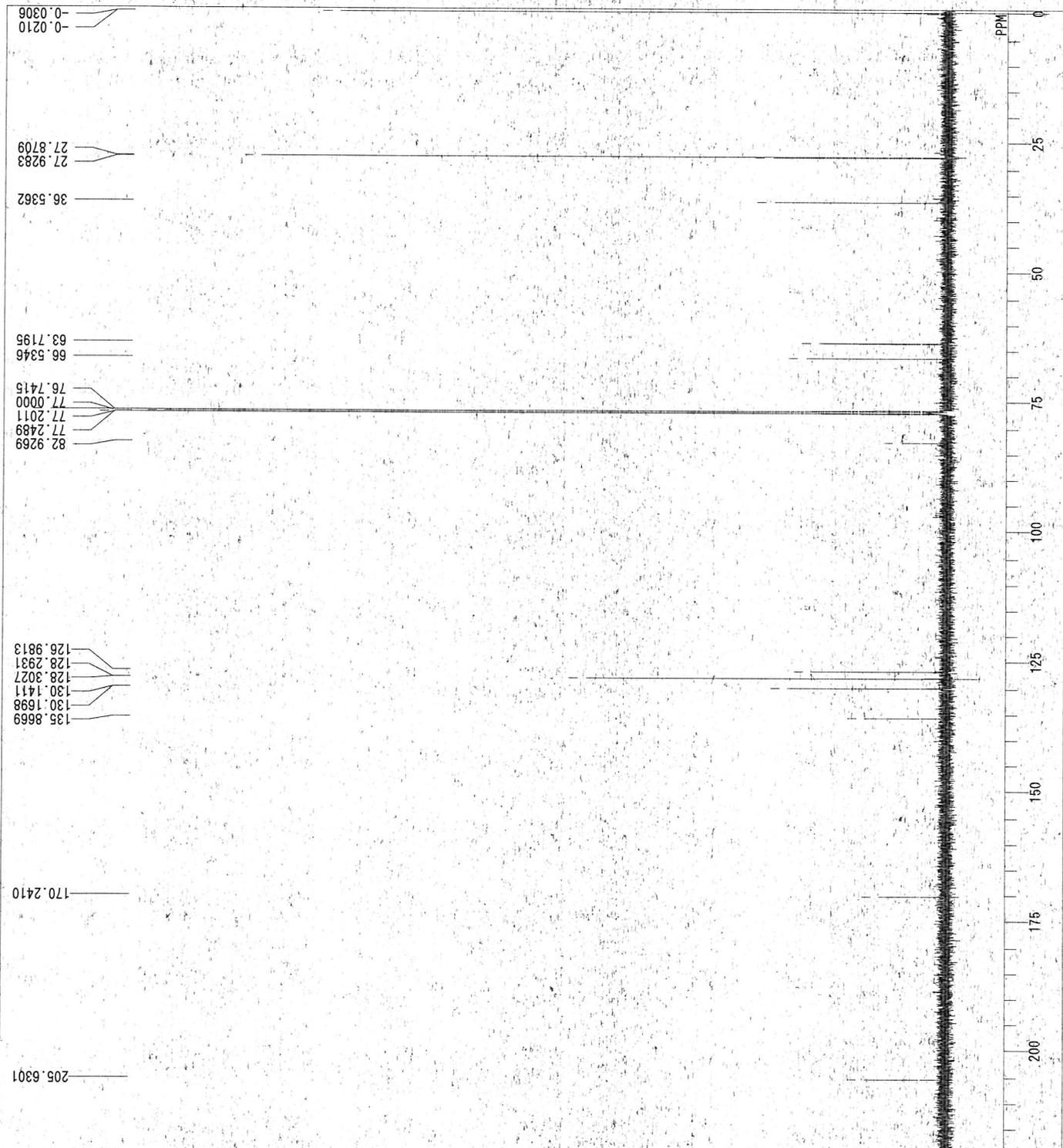
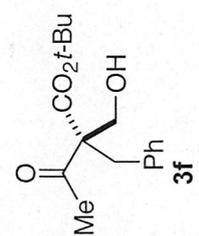
C:\My Documents\FGousei\SMO#07-064\SMO-07-064-13C.1
single_pulse_decoupled_gated_NOE

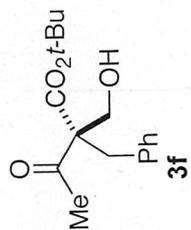
DFILE
COMNT
DATIM
OBNUC
EXMOD
OBFREQ
OBSEI
OBFIN
POINT
FREQU
SCANS
ACQTM
PD
PWI
IRNUC
CTEMP
SLVNT
EXREF
BF
RGAIN

16-02-2009 21:34:10
13C
single_pulse_dec
124.51 MHz
3.45 KHz
6.00 Hz
32768
39062.50 Hz
800
0.8389 sec
2.0000 sec
3.57 usec
1H
19.5 c
CDCl3
77.00 ppm
0.01 Hz
60

C:\My Documents\FGousei\SMO#07-064\SMO-07-064-13C.1
single_pulse_decoupled_gated_NOE

170.2410
205.6301
135.8669
130.1698
130.1411
128.3027
128.2931
126.9813
82.9269
77.2489
77.2011
77.0000
76.7415
66.5346
63.7195
36.5362
27.9283
27.8709
-0.0210
-0.0306





9.096

2.919

1.057
1.012

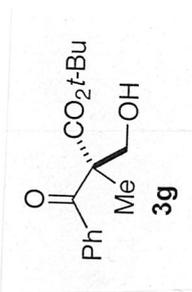
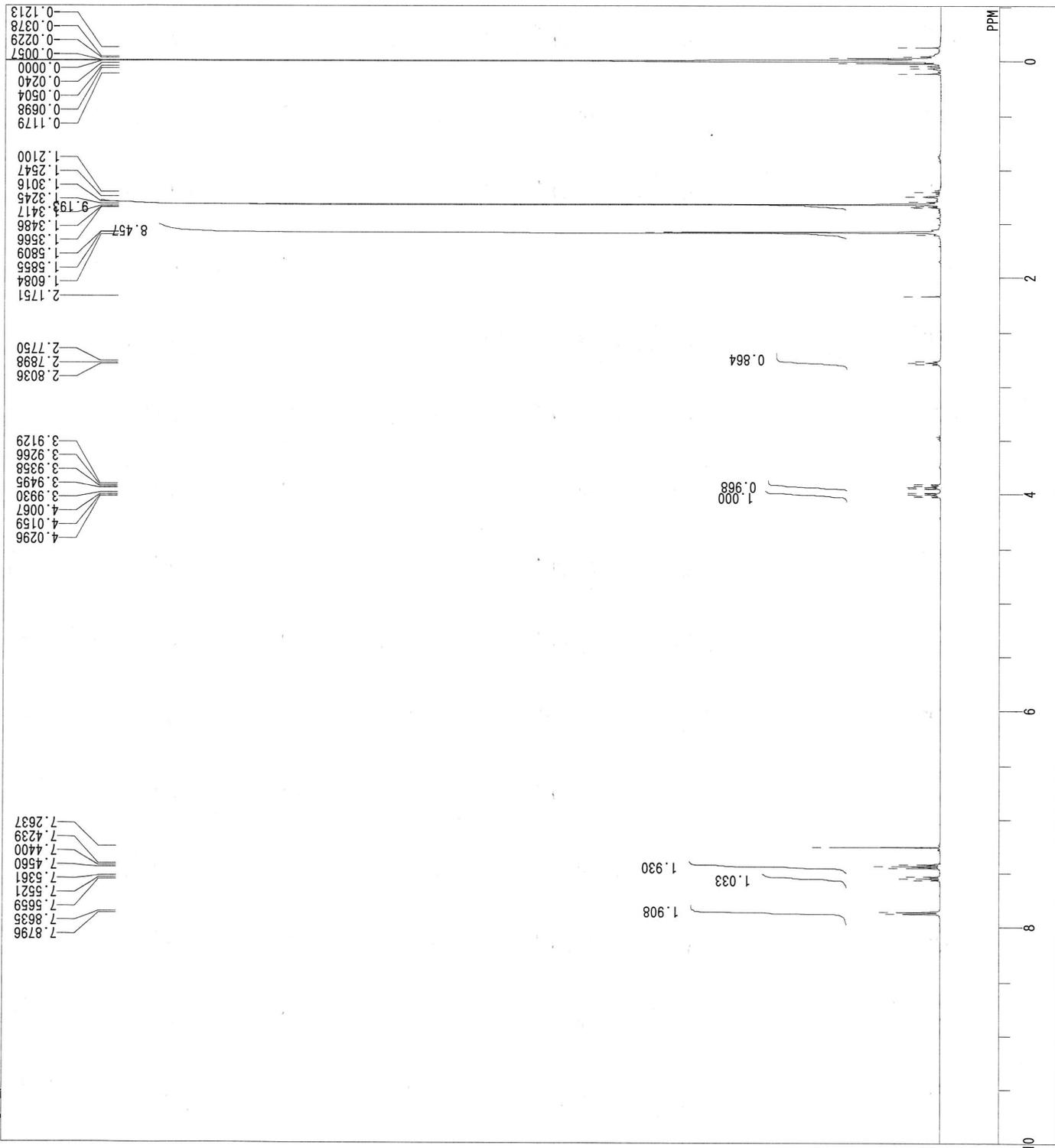
1.018
1.000

5.736



C:\My Documents\PGousei\SMO\SMO-07-180-1H.1
 single_pulse

DFILE C:\My Documents\PGousei\SMO\SMO-07-180-1H.1
 COMNT single_pulse
 DATIM 21-04-2009 22:16:19
 OBNUC 1H
 EXMOD single_pulse.ex2
 OBFREQ 495.13 MHz
 OBSFREQ 4.38 KHz
 OBSFIND 9.64 Hz
 POINT 16384
 FREQU 9286.78 Hz
 SCANS 32
 ACOPTM 1.7642 sec
 PD 5.0000 sec
 PWT 6.20 usec
 IRNUC 1H
 CTEMP 20.2 c
 SLVNT CDCl3
 EXREF 0.00 ppm
 BF 0.00 Hz
 RGAIN 50



C:\My Documents\PGousei\i\SMO\SMO-07-180-13C.1
 single pulse decoupled gated NOE

DETAIL C:\My Documents\PGousei\i\SMO\SMO-07-180-13C.1
 COMNT single pulse decoupled gated NOE
 DATIM 22-04-2009 08:33:53
 OBNUC 13C
 EXMOD single
 EXMOD pulse dec
 OBFRQ 124.51 MHz
 OBSET 3.45 KHz
 OBFTN 6.00 Hz
 POINT 32768
 FREQU 39062.50 Hz
 SCANS 13000
 ACQTM 0.8389 sec
 PD 2.0000 sec
 PW1 3.57 usec
 TRNUC 1H
 CTMP 21.4 c
 SLVNT CDCl3
 EXREF 77.00 ppm
 BF 0.50 Hz
 RGAIN 60

