

Supporting Information
for
Silver(I)-Mediated Highly Enantioselective Synthesis
of Axially Chiral Allenes under Thermal and
Microwave-Assisted Conditions

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General Experimental Section: Reagents were obtained commercially and used without further purification unless indicated otherwise. Solvent was removed under reduced pressure and the residue obtained was chromatographed on a silica gel column (230-400 mesh) using a gradient solvent system (EtOAc / *n*-hexane as eluant unless specified otherwise). ¹H and ¹³C NMR spectra were measured on either Bruker DPX-500, DPX-400 or DPX-300 spectrometer. Chemical shifts (δ ppm) were determined with tetramethylsilane (TMS) as internal reference. Mass spectra were determined on a Finnigan MAT 95 mass spectrometer. IR spectra were recorded on a Bio-RAD PTS-165 spectrometer. Chiral HPLC spectra were measured on an Agilent 1100 series using Chiralcel-OD column.

General Procedure for Synthesis of Propargylamines

A mixture of Au(Salen)PF₆ (0.01 mmol), aldehyde (1.0 mmol), amine (1.1 mmol) and alkyne (1.5 mmol) in water (1 mL) was stirred at 40 °C for 24 h in the absence of light under N₂ atmosphere. The reaction mixture was extracted with diethyl ether. The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The product was purified by flash column chromatography on silica gel using ethyl acetate-hexane as eluent. [V. K.-Y. Lo; Y. Liu; M.-K. Wong and C.-M. Che, *Org. Lett.*, 2006, **8**, 1529.]

The preparation of propargylamine **1n** and **1s** was the same as the others, despite the reactions were performed under reflux condition.

General Procedure for Silver-Mediated Synthesis of Axially Chiral Allenes from Optically Active Propargylamines under Thermal Condition

A mixture of propargylamine (0.1 mmol) and AgNO₃ (0.05 mmol) in CH₃CN (2 mL) was stirred at 40 °C for 24 h in the absence of light. Solvent was removed under reduced pressure. The

product was purified by flash column chromatography on silica gel using ethyl acetate-hexane as eluent. The % ee value was determined by HPLC using Chiralcel OD column with IPA-hexane as eluent.

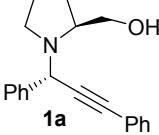
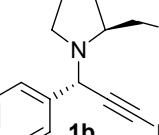
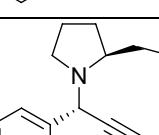
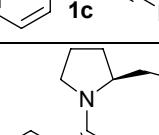
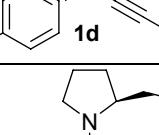
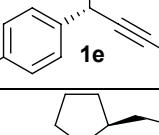
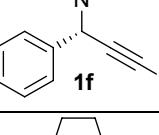
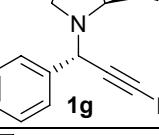
General Procedure for Silver-Mediated Synthesis of Axially Chiral Allenes from Optically Active Propargylamines under Microwave-Assisted Condition

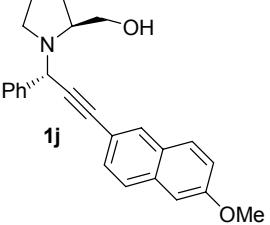
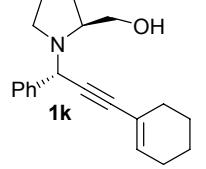
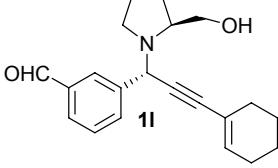
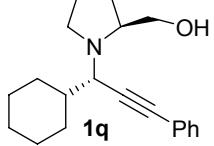
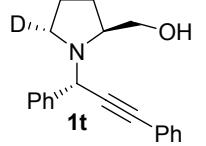
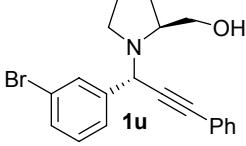
A mixture of propargylamine (0.1 mmol) and AgNO₃ (0.05 mmol) in CH₃CN (1 mL) was stirred for 20 min at 70 °C under microwave irradiation (60 W). After cooling to room temperature, solvent was removed under reduced pressure. The product was purified by flash column chromatography on alumina using ethyl acetate-hexane as eluent. The % ee value was determined by HPLC using Chiralcel OD column with IPA-hexane as eluent, or by GC using Cyclodex-β column.

General Procedure for Preparation of Racemic Allenes

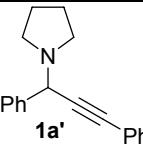
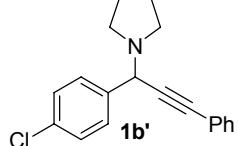
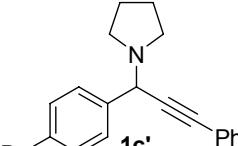
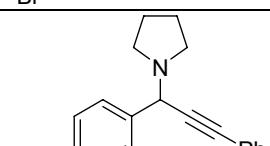
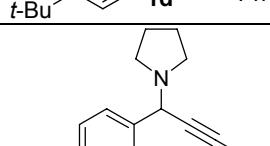
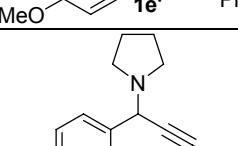
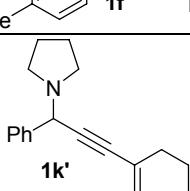
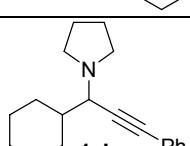
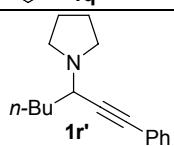
Racemic propargylamines **1a'-1h'**, **1j'-1s'** were synthesized as described above, despite using pyrrolidine as the amine component. The racemic propargylamines then were transformed into racemic allenes using procedures described above.

Literature References of Chiral Propargylamines

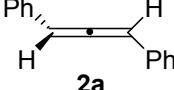
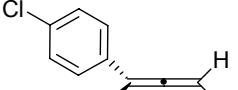
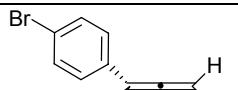
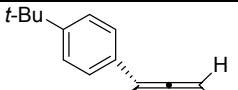
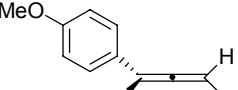
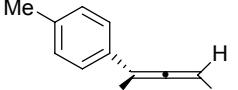
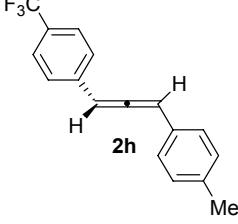
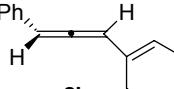
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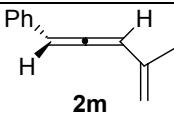
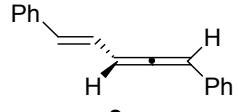
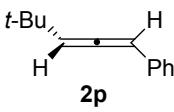
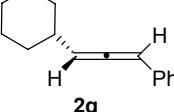
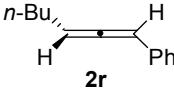
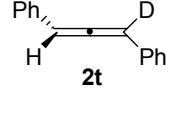
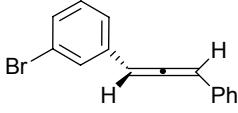
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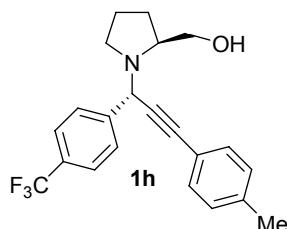
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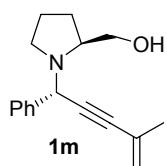
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 <p>2p</p>	<p>1. H. Inoue, H. Tsubouchi, Y. Nagaoka and K. Tomioka, <i>Tetrahedron</i>, 2002, 58, 83.</p> <p>2. C. J. Elserier and P. Vermeer, <i>J. Org. Chem.</i>, 1989, 54, 3726.</p>
 <p>2q</p>	<p>1. M. A. Tius and S. K. Pal, <i>Tetrahedron Lett.</i>, 2001, 42, 2605.</p> <p>2. A. G. Myers and B. Zheng, <i>J. Am. Chem. Soc.</i>, 1996, 118, 4492.</p>
 <p>2r</p>	<p>1. Ramu, E.; Varala, R.; Sreelatha. N.; Adapa, S. R. <i>Tetrahedron Lett.</i> 2007, <i>48</i>, 7184-7190.</p> <p>2. C. J. Elserier and P. Vermeer, <i>J. Org. Chem.</i>, 1989, 54, 3726.</p>
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 <p>2u</p>	V. K.-Y. Lo, M.-K. Wong and C.-M. Che, <i>Org. Lett.</i> , 2008, 10 , 517.

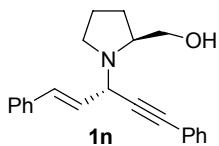
Characterization Data of Optically Active Propargylamines



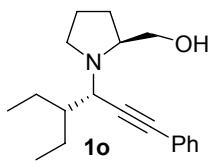
Yellow oil; analytical TLC (silica gel 60) (30 % EtOAc in hexane) $R_f = 0.37$; ^1H NMR (300 MHz, CDCl_3) δ 7.75-7.73 (d, $J = 8.2$ Hz, 2H), 7.62-7.59 (d, $J = 8.2$ Hz, 2H), 7.43-7.40 (d, $J = 8.0$ Hz, 2H), 7.15-7.12 (d, $J = 8.0$ Hz, 2H), 5.17 (s, 1H), 3.80 (dd, $J = 11.0, 3.7$ Hz, 1H), 3.57 (dd, $J = 11.0, 2.7$ Hz, 1H), 3.32-3.25 (m, 1H), 2.83-2.75 (m, 1H), 2.59-2.53 (m, 1H), 2.35 (s, 3H), 2.01-1.61 (m, 4H); ^{13}C NMR (75 MHz, CDCl_3) δ 143.35, 138.60, 131.64, 129.67 (q, $J = 32$ Hz), 129.07, 128.38, 125.14 (q, $J = 4$ Hz), 124.11 (q, $J = 270$ Hz), 119.46, 88.44, 83.60, 62.31, 61.88, 56.11, 47.78, 27.82, 23.45, 21.32; IR (NaCl, neat, cm^{-1}) 3400, 2361, 2216, 1619; EIMS m/z 342 ($\text{M}^+ \text{-CH}_2\text{OH}$); HRMS (EI) for $\text{C}_{21}\text{H}_{19}\text{F}_3\text{N}$, calcd 342.1469, found 342.1463.



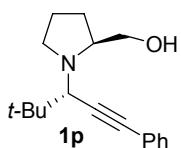
Yellow oil; analytical TLC (silica gel 60) (30% EtOAc in hexane) $R_f = 0.30$; ^1H NMR (400 MHz, CDCl_3) δ 7.54-7.52 (m, 2H), 7.36-7.33 (m, 2H), 7.29-7.25 (m, 1H), 5.36 (s, 1H), 5.27 (t, $J = 1.6$ Hz, 1H), 5.00 (s, 1H), 3.78 (dd, $J = 10.9, 3.6$ Hz, 1H), 3.51 (dd, $J = 10.9, 2.3$ Hz, 1H), 3.21-3.16 (m, 1H), 3.18 (td, $J = 5.9, 3.0$ Hz, 1H), 2.74-2.68 (m, 1H), 1.96 (s, 3H), 1.95-1.90 (m, 1H), 1.86-1.83 (m, 1H), 1.72-1.58 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.15, 128.24, 128.01, 127.51, 126.50, 121.80, 89.00, 84.30, 61.81, 61.62, 56.06, 47.74, 27.98, 23.82, 23.53; IR (NaCl, neat, cm^{-1}) 3400, 2361, 1616; EIMS m/z 224 ($\text{M}^+ \text{-CH}_2\text{OH}$); HRMS (EI) for $\text{C}_{16}\text{H}_{18}\text{N}$, calcd 224.1434, found 224.1430.



Yellow oil; analytical TLC (silica gel 60) (50% EtOAc in hexane) $R_f = 0.26$; ^1H NMR (400 MHz, CDCl_3) δ 7.61-7.60 (m, 2H), 7.52-7.50 (m, 2H), 7.39-7.28 (m, 8H), 5.13 (s, 1H), 3.83 (dd, $J = 11.0, 3.5$ Hz, 1H), 3.54 (dd, $J = 11.0, 2.5$ Hz, 1H), 3.32-3.27 (m, 1H), 2.82 (td, $J = 9.3, 7.3$ Hz, 1H), 2.66-2.61 (m, 1H), 2.01-1.61 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.06, 131.87, 128.39, 128.36, 128.14, 127.68, 122.97, 87.89, 85.31, 61.86, 56.37, 47.95, 29.72, 28.04, 23.62; IR (NaCl, neat, cm^{-1}) 3429, 2361, 1653; EIMS m/z 317 (M^+); HRMS (EI) for $\text{C}_{22}\text{H}_{23}\text{NO}$, calcd 317.1774, found 317.1767.

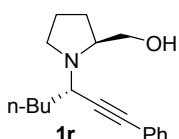


Yellow oil; analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.23$; ^1H NMR (300 MHz, CDCl_3) δ 7.43-7.38 (m, 2H), 7.27-7.22 (m, 3H), 3.51-3.48 (d, $J = 7.4$ Hz, 1H), 2.76-2.61 (m, 4H), 1.79-1.75 (m, 5H), 1.61-1.45 (m, 4H), 0.95-0.88 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3) δ 131.61, 128.09, 127.56, 123.70, 88.03, 85.40, 58.33, 50.20, 43.76, 23.54, 22.33, 21.79, 10.89, 10.75; IR (NaCl, neat, cm^{-1}) 3430, 2362; EIMS m/z 254 ($\text{M}^+ - \text{CH}_2\text{OH}$); HRMS (EI) for $\text{C}_{18}\text{H}_{24}\text{N}$, calcd 254.1909, found 254.1909.

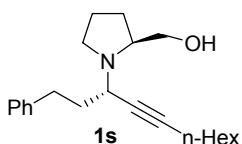


Yellow oil, analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.11$; ^1H NMR (400 MHz, CDCl_3) δ 7.43-7.40 (m, 2H), 7.31-7.24 (m, 3H), 3.63-3.59 (m, 1H), 3.43-3.40 (m, 1H), 3.41 (s, 1H), 3.07-3.02 (m, 2H), 2.97-2.92 (m, 1H), 1.87-1.70 (m, 4H), 1.08 (s, 9H); ^{13}C NMR (100 MHz,

CDCl_3) δ 131.51, 128.14, 127.74, 123.36, 86.55, 86.51, 64.81, 63.86, 61.56, 49.49, 35.75, 27.76, 27.22, 25.00; IR (NaCl, neat, cm^{-1}) 3436, 2360; EIMS m/z 256 ($\text{M}^+ \text{-CH}_3$); HRMS (EI) for $\text{C}_{17}\text{H}_{22}\text{NO}$, calcd 256.1701, found 256.1701.

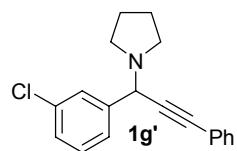


Yellow oil; analytical TLC (silica gel 60) (30 % EtOAc in hexane) $R_f = 0.31$; ^1H NMR (400 MHz, CDCl_3) δ 7.42-7.38 (m, 2H), 7.30-7.25 (m, 3H), 3.75 (t, $J = 7.7$ Hz, 1H), 3.63 (dd, $J = 10.8, 4.0$ Hz, 1H), 3.44-3.41 (m, 2H), 3.10-3.06 (m, 1H), 2.96-2.91 (m, 1H), 2.85-2.78 (m, 1H), 1.91-1.86 (m, 1H), 1.80-1.69 (m, 4H), 1.50-1.34 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 131.49, 128.01, 127.69, 123.09, 87.83, 85.01, 61.79, 61.74, 52.55, 47.13, 34.69, 28.74, 27.62, 23.49, 22.22, 13.83; IR (NaCl, neat, cm^{-1}) 3435, 2361, 2335; EIMS m/z 240 ($\text{M}^+ \text{-CH}_2\text{OH}$); HRMS (EI) for $\text{C}_{17}\text{H}_{22}\text{N}$, calcd 240.1752, found 240.1754.

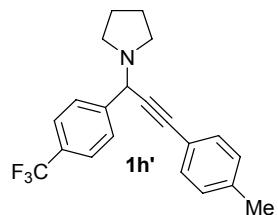


Yellow oil; analytical TLC (silica gel 60) (50% EtOAc in hexane) $R_f = 0.16$; ^1H NMR (400 MHz, CDCl_3) δ 7.28-7.23 (m, 2H), 7.19-7.14 (m, 3H), 3.54-3.49 (m, 2H), 3.34 (dd, $J = 10.8, 2.3$ Hz, 1H), 2.99-2.95 (m, 1H), 2.90-2.85 (m, 1H), 2.77-2.68 (m, 3H), 2.22-2.18 (m, 2H), 1.92 (q, $J = 7.8$ Hz, 2H), 1.87-1.71 (m, 4H), 1.53-1.46 (m, 2H), 1.44-1.38 (m, 2H), 1.34-1.28 (m, 5H), 0.90-0.87 (m, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 141.61, 129.26, 128.75, 83.48, 77.63, 61.65, 61.58, 51.50, 47.03, 37.03, 32.83, 31.22, 28.95, 28.39, 27.72, 23.66, 22.50, 18.52, 13.95; IR (NaCl, neat, cm^{-1}) 3429, 2364; EIMS m/z 296 ($\text{M}^+ \text{-CH}_2\text{OH}$); HRMS (EI) for $\text{C}_{21}\text{H}_{30}\text{N}$, calcd 296.2373, found 296.2374.

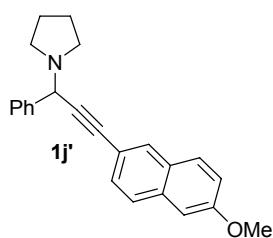
Characterization Data of Racemic Propargylamines



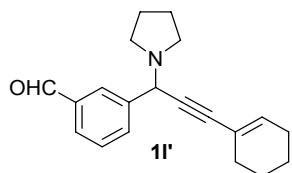
Pale yellow oil; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.25$; ^1H NMR (300 MHz, CDCl_3) δ 7.62 (s, 1H), 7.50-7.47 (m, 3H), 7.32-7.30 (m, 3H), 7.26-7.25 (m, 2H), 4.87 (s, 1H), 2.67-2.65 (m, 4H), 1.79-1.77 (m, 4H); ^{13}C NMR (75 MHz, CDCl_3) δ 141.70, 134.13, 131.79, 129.45, 128.28, 128.23, 127.69, 126.35, 122.93, 87.40, 85.73, 58.46, 50.09, 23.50; IR (KBr, neat, cm^{-1}) 2362; EIMS m/z 295 (M^+); HRMS (EI) for $\text{C}_{19}\text{H}_{18}\text{NCl}$, calcd 295.1127, found 295.1122.



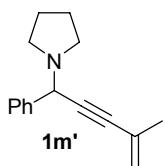
Yellow oil; analytical TLC (silica gel 60) (30 % EtOAc in hexane) $R_f = 0.58$; ^1H NMR (400 MHz, CDCl_3) δ 7.74 (d, $J = 8.2$ Hz, 2H), 7.59 (d, $J = 8.2$ Hz, 2H), 7.38 (d, $J = 8.1$ Hz, 2H), 7.10 (d, $J = 8.0$ Hz, 2H), 4.92 (s, 1H), 2.68-2.64 (m, 4H), 2.32 (s, 3H), 1.79-1.76 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 143.78, 138.38, 131.65, 129.66 (q, $J = 32$ Hz), 129.03, 128.49, 125.11 (q, $J = 4$ Hz), 124.22 (q, $J = 270$ Hz), 119.78, 87.74, 84.73, 58.54, 50.04, 23.48, 21.32; IR (NaCl, neat, cm^{-1}) 2361, 2224, 1619; EIMS m/z 343 (M^+); HRMS (EI) for $\text{C}_{21}\text{H}_{30}\text{NF}_3$, calcd 343.1548, found 343.1540.



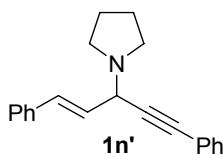
Pale yellow solid; analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.18$; ^1H NMR (300 MHz, CDCl_3) δ 7.91 (s, 1H), 7.65-7.61 (m, 4H), 7.49 (dd, $J = 8.5, 1.6$ Hz, 1H), 7.38-7.33 (m, 2H), 7.30-7.24 (m, 1H), 7.12 (dd, $J = 9.0, 2.5$ Hz, 1H), 7.05 (d, $J = 2.4$ Hz, 1H), 4.91 (s, 1H), 3.84 (s, 3H), 2.74-2.69 (m, 4H), 1.81-1.79 (m, 4H); ^{13}C NMR (75 MHz, CDCl_3) δ 158.16, 139.62, 133.99, 131.23, 129.23, 129.12, 128.43, 128.24, 128.20, 127.49, 126.67, 119.30, 118.09, 105.72, 87.35, 86.22, 59.18, 55.15, 50.25, 23.47, 14.07; IR (NaCl, neat, cm^{-1}) 2352; EIMS m/z 341 (M^+); HRMS (EI) for $\text{C}_{24}\text{H}_{23}\text{NO}$, calcd 341.1779, found 341.1774.



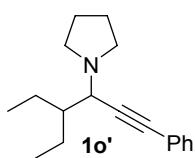
Yellow oil; analytical TLC (silica gel 60) (30% EtOAc in hexane) $R_f = 0.27$; ^1H NMR (300 MHz, CDCl_3) δ 10.03 (s, 1H), 8.06 (s, 1H), 7.85-7.79 (m, 2H), 7.52-7.47 (m, 1H), 6.18-6.15 (m, 1H), 4.87 (s, 1H), 2.65-2.57 (m, 4H), 2.20-2.11 (m, 4H), 1.82-1.76 (m, 4H), 1.70-1.59 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.21, 141.23, 126.37, 134.76, 134.19, 129.72, 128.77, 128.38, 120.25, 89.49, 82.45, 58.32, 49.89, 29.45, 25.51, 23.41, 22.21, 21.40; IR (NaCl, neat, cm^{-1}) 2361, 1699, 1653; EIMS m/z 293 (M^+); HRMS (EI) for $\text{C}_{20}\text{H}_{23}\text{NO}$, calcd 293.1774, found 293.1776.



Yellow oil; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.31$; ^1H NMR (400 MHz, CDCl_3) δ 7.54-7.52 (m, 2H), 7.35-7.31 (m, 2H), 7.28-7.23 (m, 1H), 5.32 (s, 1H), 5.22-5.21 (t, $J = 1.7$ Hz, 1H), 4.75 (s, 1H), 2.62-2.59 (m, 4H), 1.93 (s, 3H), 1.80-1.73 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.54, 128.10, 127.44, 126.68, 121.43, 88.03, 85.68, 58.96, 50.17, 23.76, 23.41; IR (NaCl, neat, cm^{-1}) 2361, 1615; EIMS m/z 225 (M^+); HRMS for $\text{C}_{16}\text{H}_{19}\text{N}$, calcd 224.1434, found 224.1437.

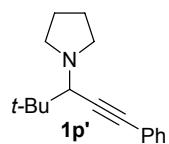


Yellow oil; analytical TLC (silica gel 60) (30% EtOAc in hexane) $R_f = 0.33$; ^1H NMR (400 MHz, CDCl_3) δ 7.61-7.59 (m, 2H), 7.49-7.47 (m, 2H), 7.36-7.33 (m, 3H), 7.30-7.27 (m, 5H), 4.87 (s, 1H), 2.70-2.67 (m, 4H), 1.82-1.74 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.47, 131.75, 128.45, 128.22, 128.04, 127.54, 123.20, 86.88, 86.68, 59.11, 50.25, 23.45; IR (NaCl, neat, cm^{-1}) 2362; EIMS m/z 232 ($\text{M}^+ - \text{C}_4\text{H}_7$); HRMS (EI) for $\text{C}_{17}\text{H}_{14}\text{N}$, calcd 232.1121, found 232.1120.

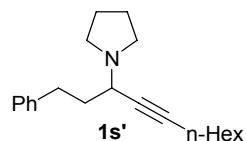


Yellow oil; analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.37$; ^1H NMR (300 MHz, CDCl_3) δ 7.43-7.39 (m, 2H), 7.29-7.24 (m, 3H), 3.49 (d, $J = 7.4$ Hz, 1H), 2.75-2.61 (m, 4H), 1.79-1.75 (m, 5H), 1.61-1.42 (m, 4H), 0.95-0.88 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3) δ 131.61, 128.08, 127.56, 123.71, 88.03, 85.40, 58.33, 50.19, 43.77, 23.55, 22.34, 21.80, 10.89, 10.75; IR

(NaCl, neat, cm^{-1}) 2362; EIMS m/z 255 (M^+); HRMS (EI) for $\text{C}_{18}\text{H}_{25}\text{N}$, calcd 255.1987, found 255.1994.

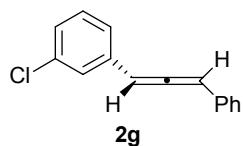


Yellow oil; analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.39$; ^1H NMR (400 MHz, CDCl_3) δ 7.44-7.40 (m, 2H), 7.28-7.23 (m, 3H), 3.46 (s, 1H), 2.82-2.73 (m, 4H), 1.77-1.68 (m, 4H), 1.06 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 131.65, 128.13, 128.58, 123.84, 86.95, 65.14, 51.69, 36.11, 27.72, 24.19; IR (NaCl, neat, cm^{-1}) 2361; EIMS m/z 241 (M^+); HRMS (EI) for $\text{C}_{17}\text{H}_{23}\text{N}$, calcd 241.1831, found 241.1847.

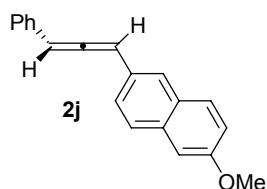


Yellow oil; analytical TLC (silica gel 60) (50% EtOAc in hexane) $R_f = 0.28$; ^1H NMR (300 MHz, CDCl_3) δ 7.28-7.15 (m, 5H), 3.44-3.38 (m, 1H), 2.75-2.57 (m, 6H), 2.25-2.20 (m, 2H), 2.00-1.80 (m, 2H), 1.76-1.74 (m, 4H), 1.54-1.39 (m, 4H), 1.32-1.26 (m, 4H), 0.89 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 141.96, 128.45, 128.19, 128.65, 85.61, 77.87, 53.91, 49.46, 36.95, 32.81, 31.28, 29.02, 28.46, 23.41, 22.54, 18.61, 13.98; IR (NaCl, neat, cm^{-1}) 2364; EIMS m/z 297 (M^+); HRMS (EI) for $\text{C}_{21}\text{H}_{31}\text{N}$, calcd 297.2457, found 297.2446.

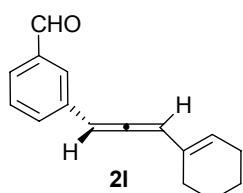
Characterization Data of Allenes



Light yellow solid; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.66$; ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.30 (m, 5H), 7.26-7.18 (m, 4H), 6.62 (d, *J* = 6.5 Hz, 1H), 6.53 (d, *J* = 6.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 208.05, 135.70, 134.71, 133.09, 129.92, 128.83, 127.59, 127.35, 126.83, 125.16, 98.91, 97.55; IR (NaCl, neat, cm⁻¹) 1937; EIMS *m/z* 226 (M⁺); HRMS (EI) for C₁₅H₁₁Cl, calcd 226.0549, found 226.0546.

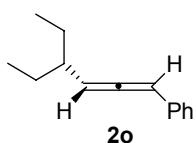


Light yellow solid; analytical TLC (silica gel 60) (10 % EtOAc in hexane) $R_f = 0.47$; ¹H NMR (300 MHz, CDCl₃) δ 7.70-7.64 (m, 3H), 7.48 (dd, *J* = 8.6, 1.1 Hz, 1H), 7.43-7.29 (m, 4H), 7.25-7.20 (m, 1H), 7.10 (s, 2H), 6.74 (d, *J* = 6.5 Hz, 1H), 6.64 (d, *J* = 6.5 Hz, 1H), 3.91 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 208.08, 157.75, 134.03, 133.84, 129.29, 129.20, 128.78, 127.33, 127.29, 127.07, 125.81, 125.37, 119.01, 106.02, 98.78, 98.61, 78.16, 55.33; IR (NaCl, neat, cm⁻¹) 2359, 1633; EIMS *m/z* 272 (M⁺); HRMS (EI) for C₂₀H₁₆O, calcd 272.1201, found 272.1200.

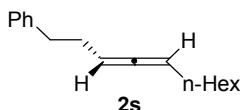


Yellow solid; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.66$; ¹H NMR (400 MHz, CDCl₃) δ 10.01 (s, 1H), 7.79 (m, 1H), 7.72-7.70 (m, 1H), 7.57-7.55 (m, 1H), 7.49-7.45 (m,

1H), 6.46-6.45 (m, 1H), 6.32 (d, $J = 6.3$ Hz, 1H), 5.82-5.79 (m, 1H), 2.17-1.97 (m, 4H), 1.68-1.58 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 206.99, 192.34, 136.87, 136.14, 132.50, 131.30, 129.27, 128.16, 127.97, 127.74, 102.25, 96.69, 25.97, 25.84, 22.43, 22.31; IR (NaCl, neat, cm^{-1}) 1931, 1699, 1601; EIMS m/z 224 (M^+); HRMS (EI) for $\text{C}_{16}\text{H}_{16}\text{O}$, calcd 224.1196, found 224.1196.



Yellow oil; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.74$; ^1H NMR (300 MHz, CDCl_3) δ 7.30-7.26 (m, 4H), 7.20-7.14 (m, 1H), 6.14 (dd, $J = 6.4, 2.0$ Hz, 1H), 5.41-5.37 (m, 1H), 2.01-1.98 (m, 1H), 1.60-1.33 (m, 4H), 0.99-0.91 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3) δ 204.83, 135.27, 128.54, 126.55, 98.91, 94.55, 43.11, 29.72, 27.71, 27.47, 11.87, 11.67; IR (NaCl, neat, cm^{-1}) 1963; EIMS m/z 186 (M^+); HRMS (EI) for $\text{C}_{14}\text{H}_{18}$, calcd 186.1403, found 186.1401.



Yellow oil; analytical TLC (silica gel 60) (10% EtOAc in hexane) $R_f = 0.71$; ^1H NMR (400 MHz, CDCl_3) δ 7.29-7.26 (m, 2H), 7.20-7.16 (m, 3H), 5.13-5.07 (m, 2H), 2.72 (t, $J = 7.8$ Hz, 2H), 2.33-2.26 (m, 2H), 1.96-1.91 (m, 2H), 1.37-1.25 (m, 8H), 0.90-0.87 (m, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 204.01, 141.98, 128.51, 128.53, 125.78, 91.56, 90.22, 35.52, 31.72, 30.73, 29.72, 29.17, 28.93, 28.80, 22.67, 14.11; IR (NaCl, neat, cm^{-1}) 1948; EIMS m/z 228 (M^+); HRMS (EI) for $\text{C}_{17}\text{H}_{24}$, calcd 228.1873, found 228.1871.

ESI-MS Spectrum of **1a**-Ag Adduct

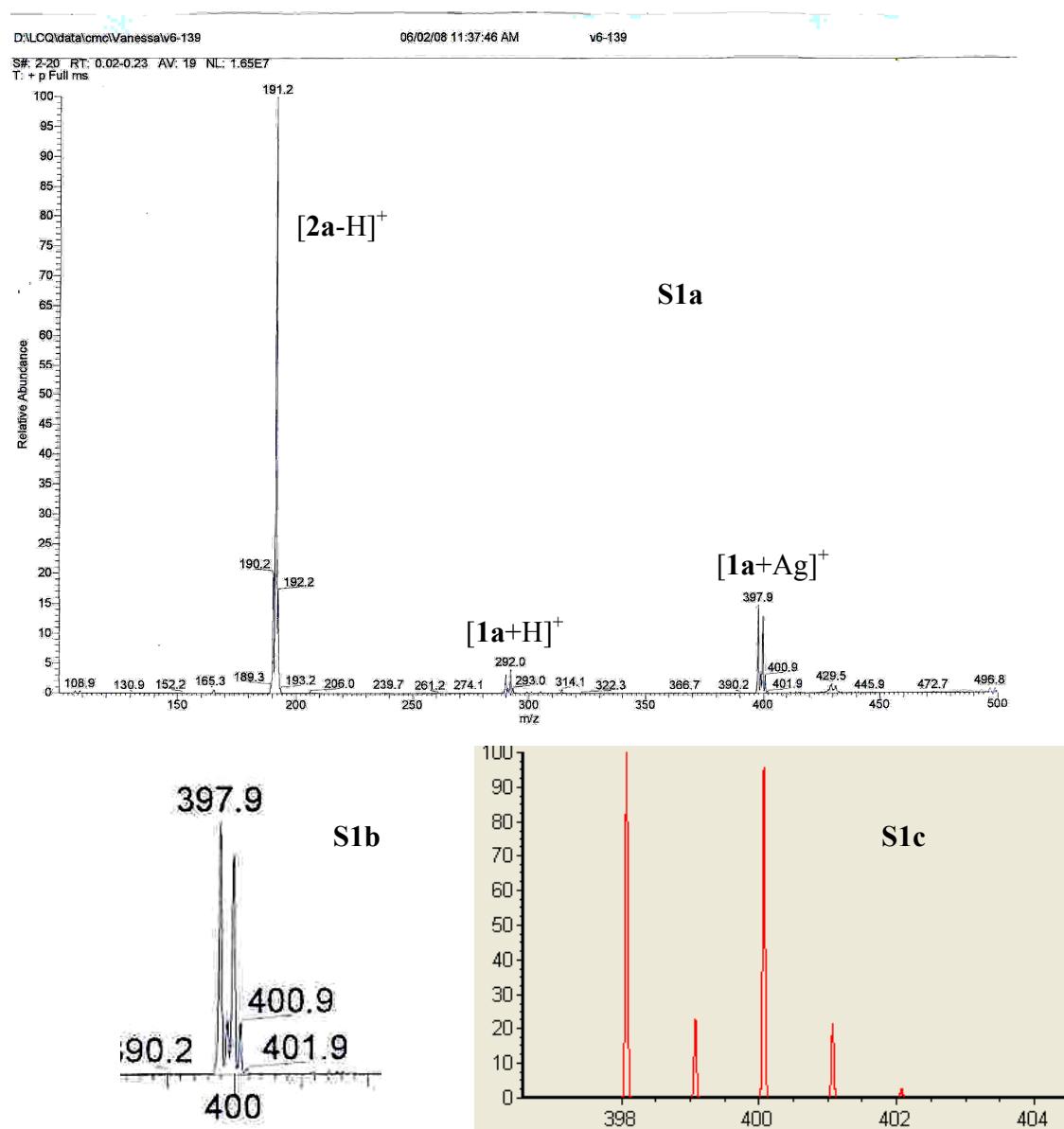


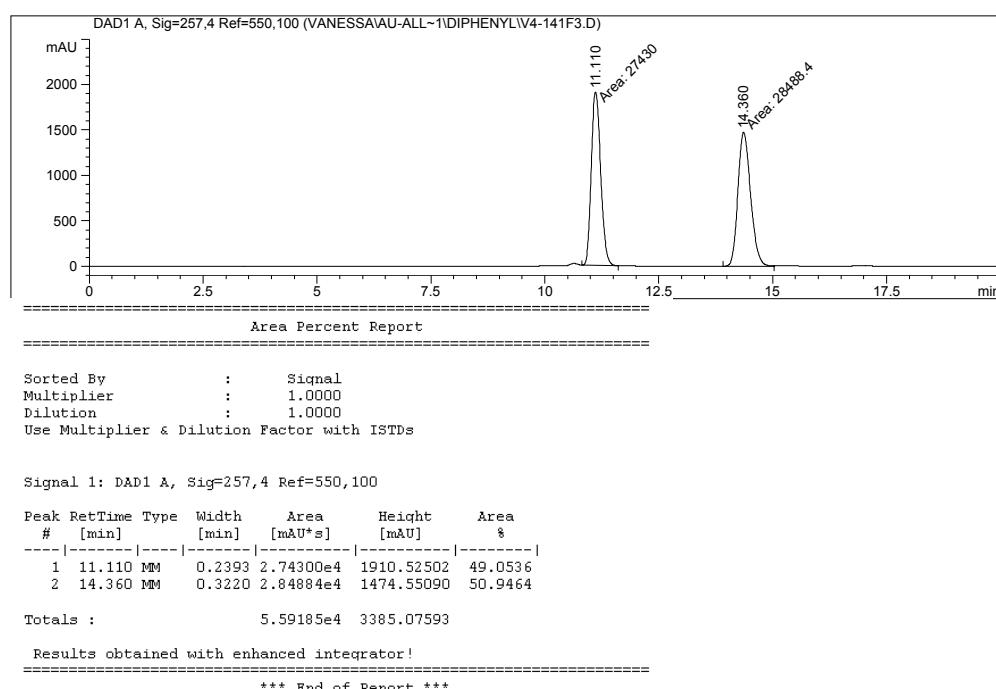
Figure S1a. ESI-MS spectrum of adduct of **1a** and silver(I) (peak at m/z = 398).

Figure S1b. Peak at m/z 398. Figure S1c. Simulated isotopic pattern of $\text{C}_{20}\text{H}_{21}\text{NOAg}$ (**1a** + Ag^+).

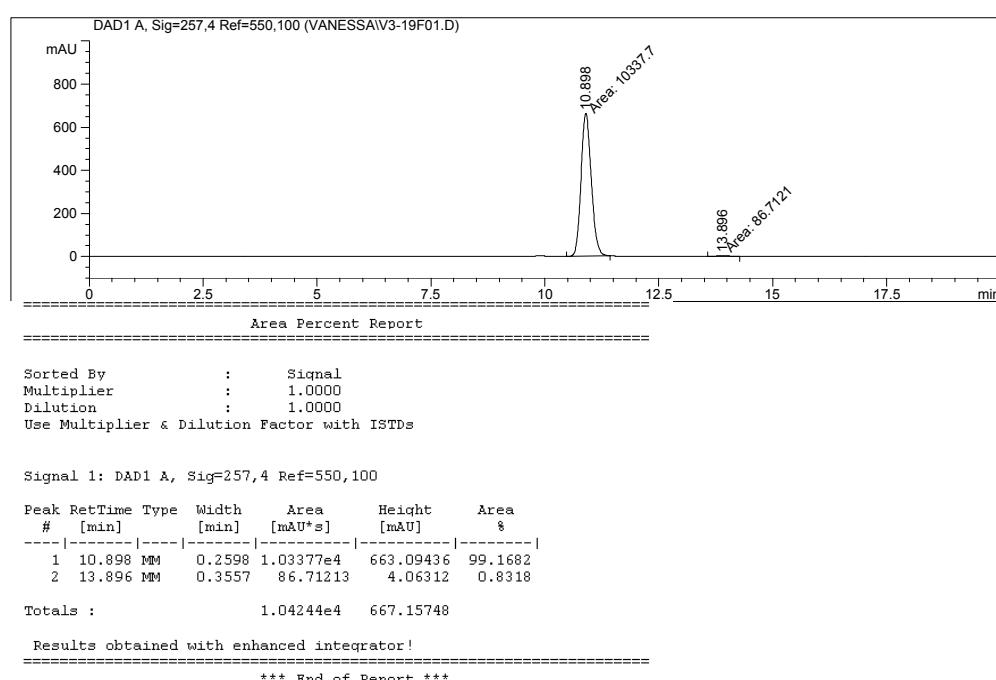
HPLC Spectra of 2a-o, 2q-s

2a (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

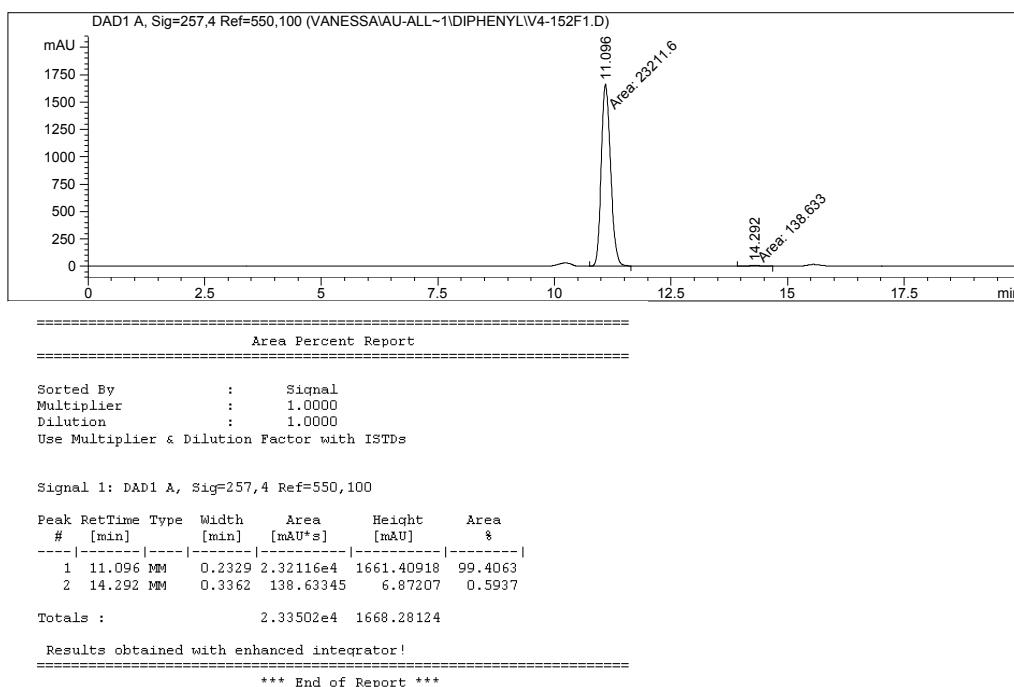
Racemic



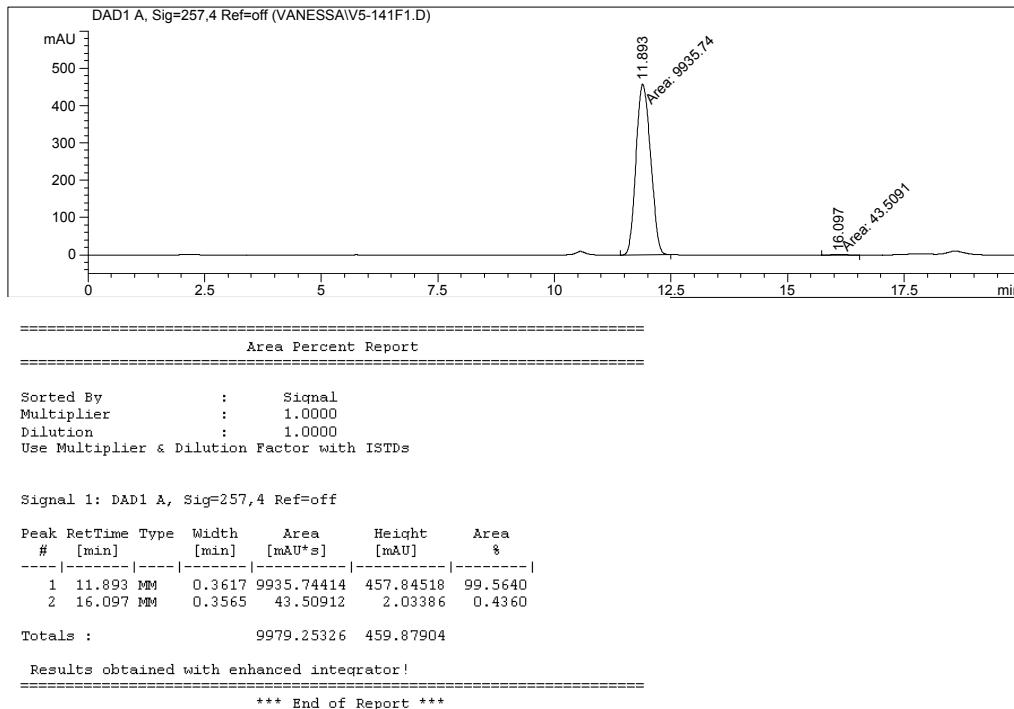
(R)-2a, for Table 1, entry 1



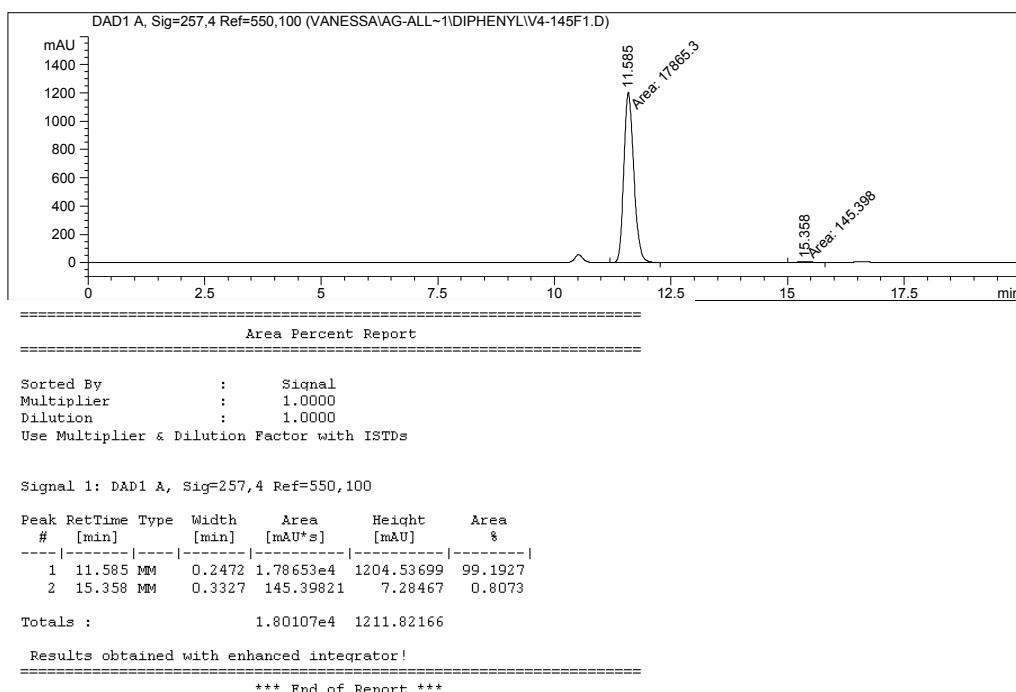
(R)-2a, for Table 1, entry 2



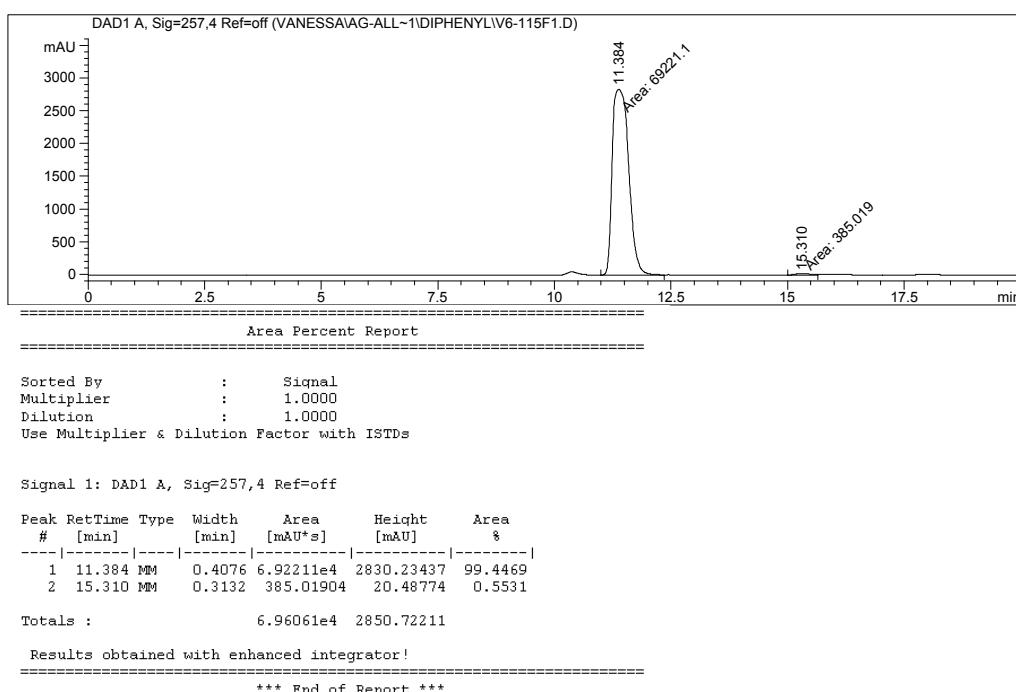
(R)-2a, for Table 1, entry 3



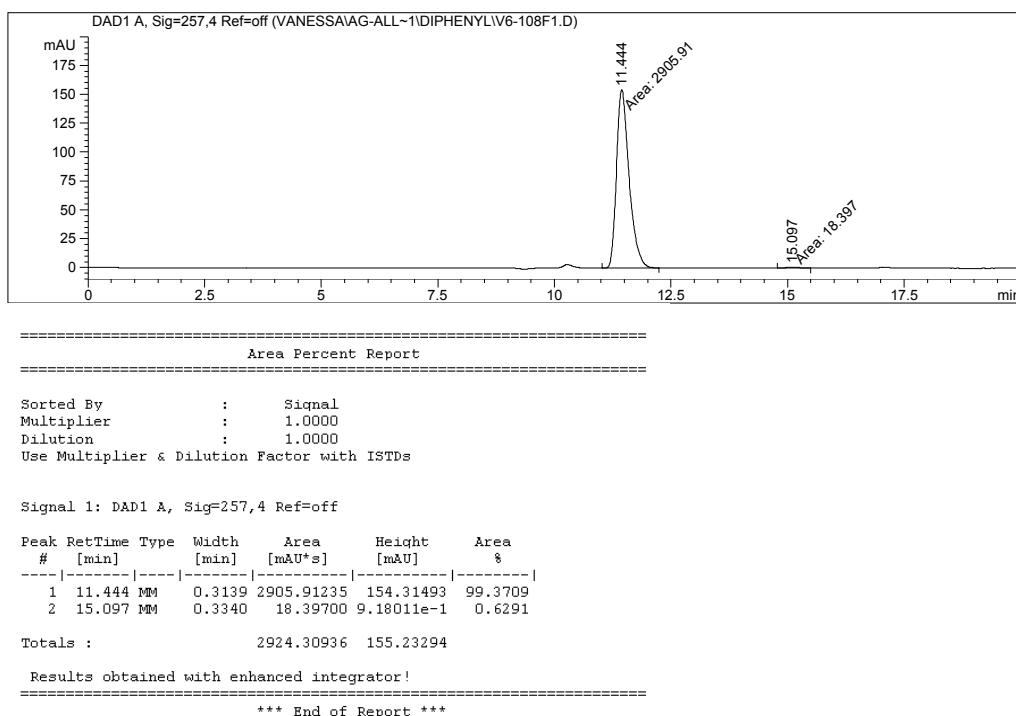
(R)-2a, for Table 1, entry 4



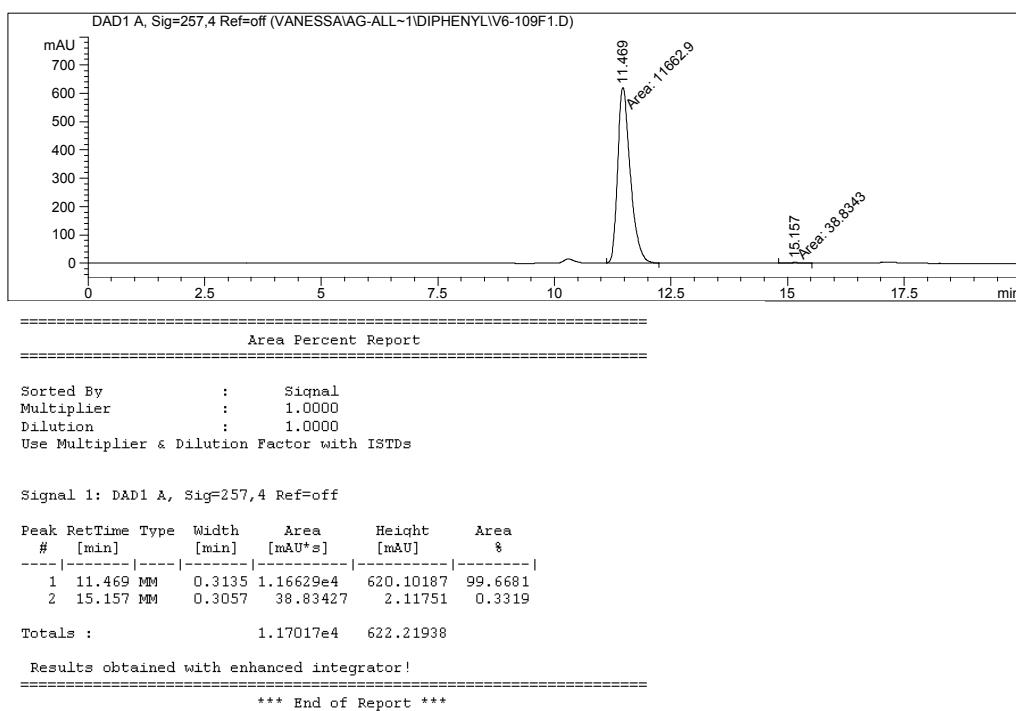
(R)-2a, for Table 1, entry 5



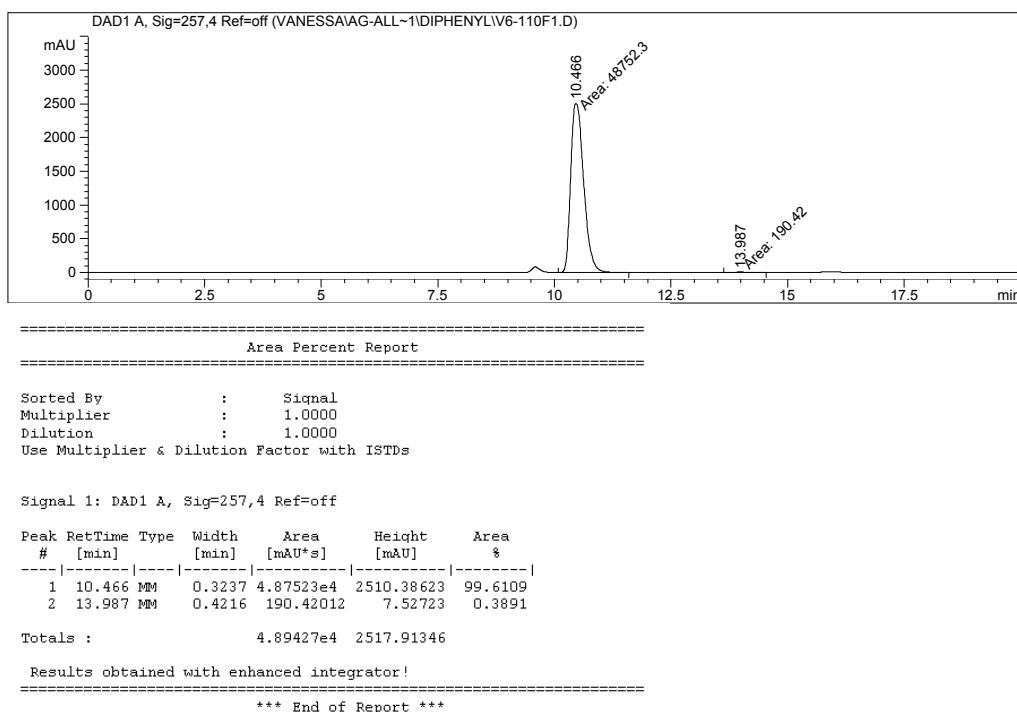
(R)-2a, for Table 1, entry 6



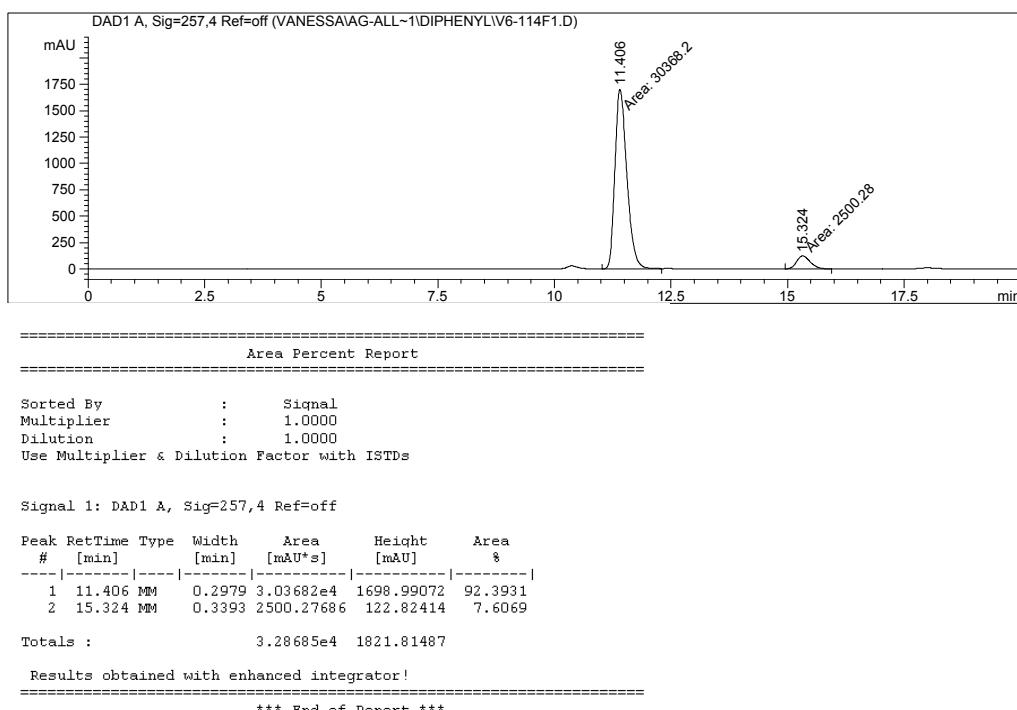
(R)-2a, for Table 1, entry 7



(R)-2a, for Table 1, entry 8

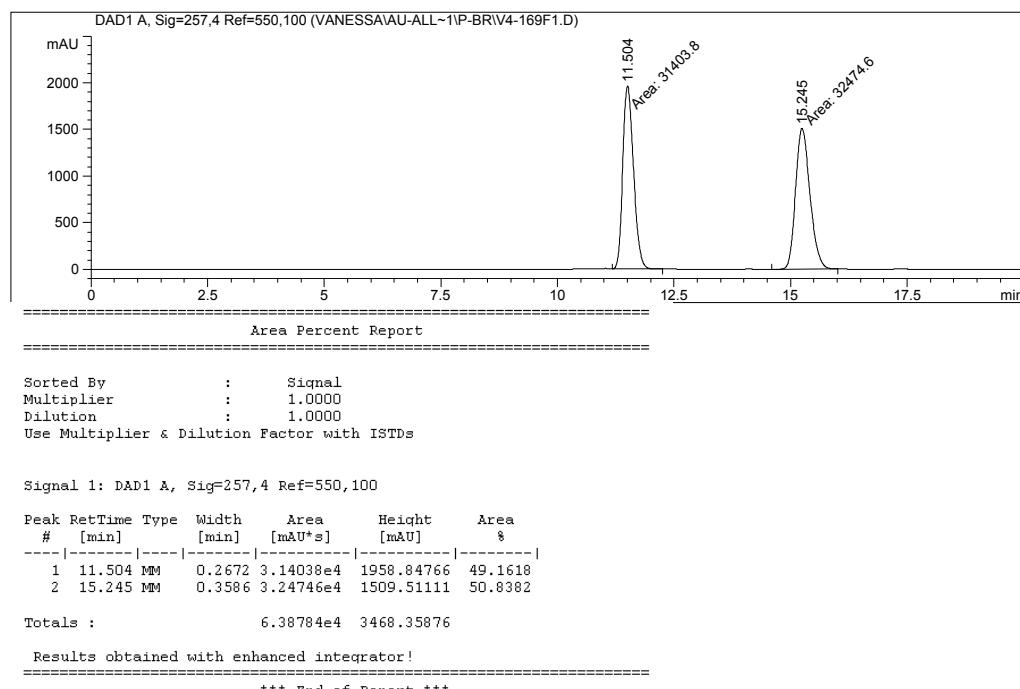


(R)-2a, for Table 1, entry 9

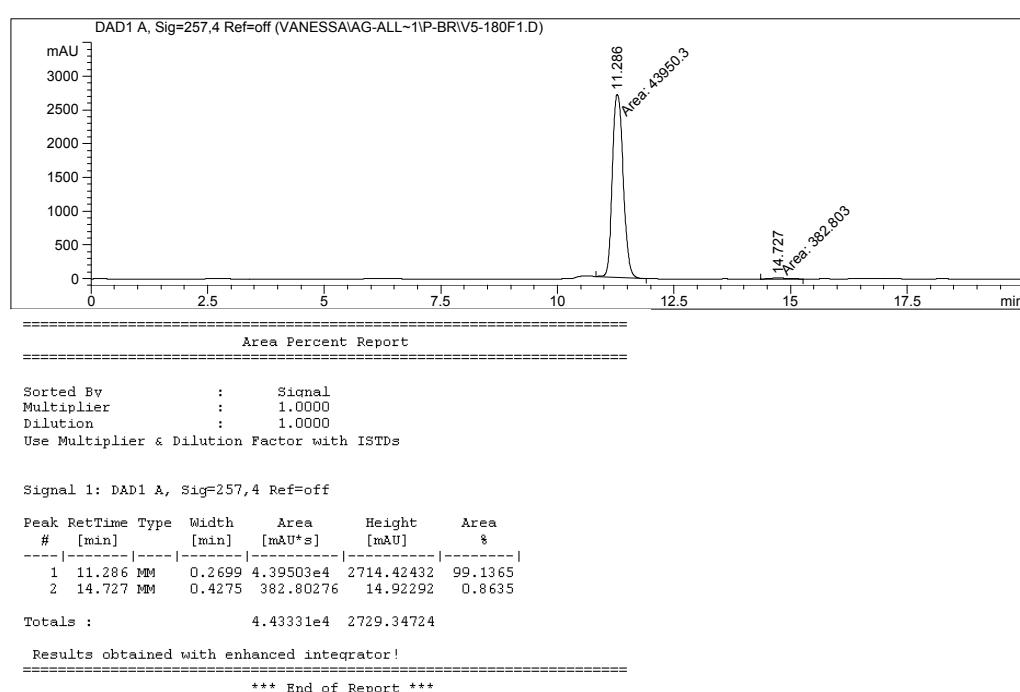


2c (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

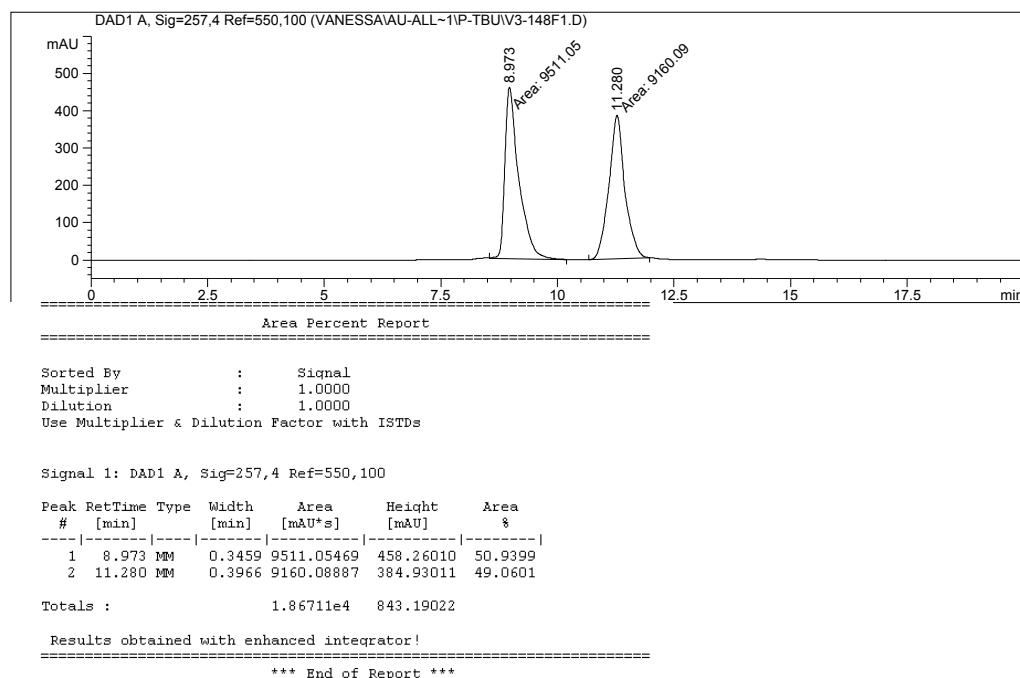


(R)-**2c**, for Table 2, entry 2

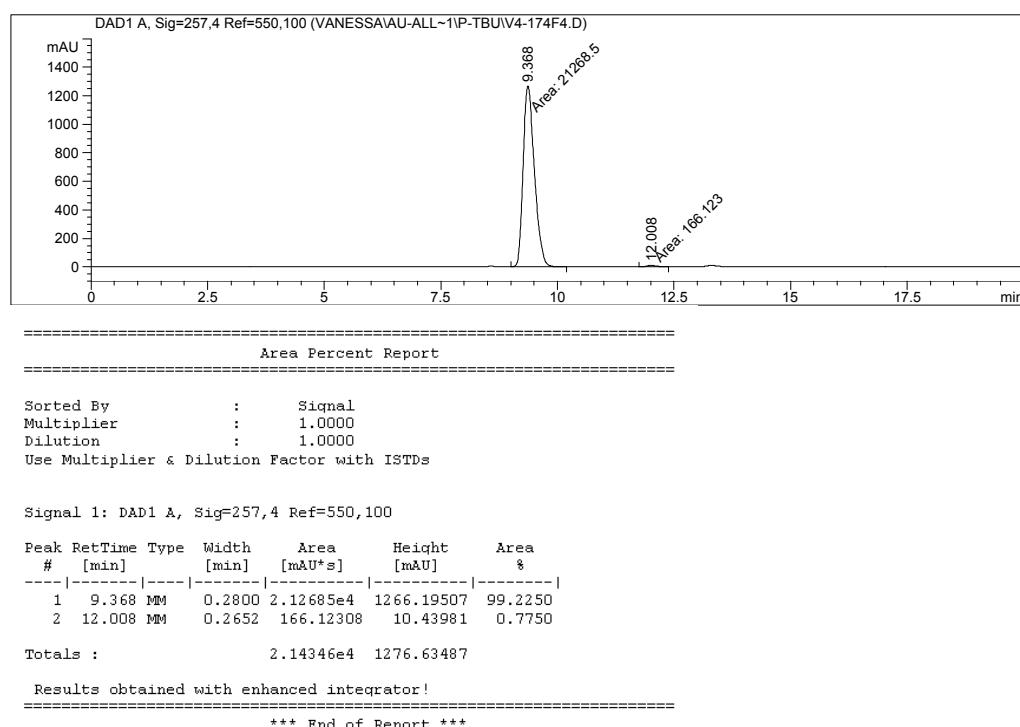


2d (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

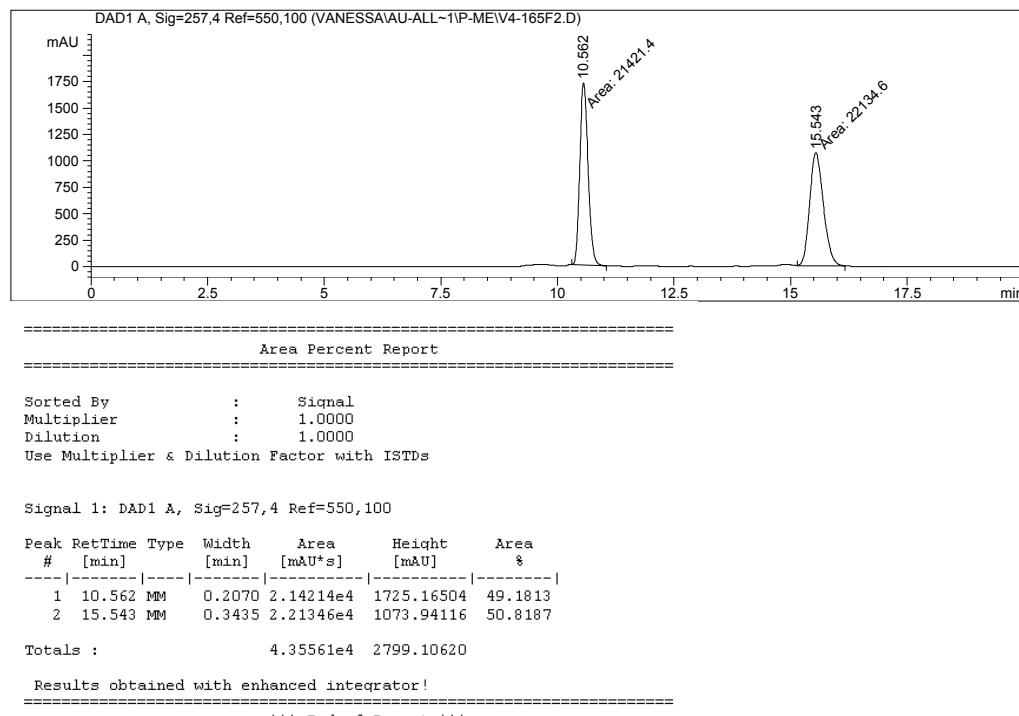


(R)-**2d**, for Table 2, entry 3

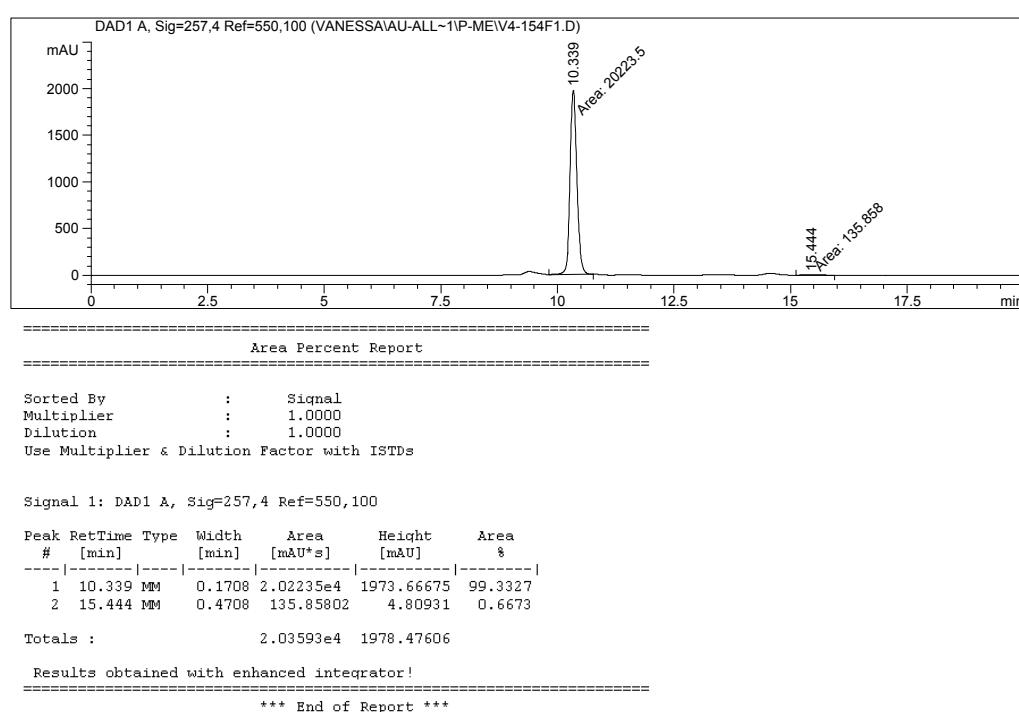


2f (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

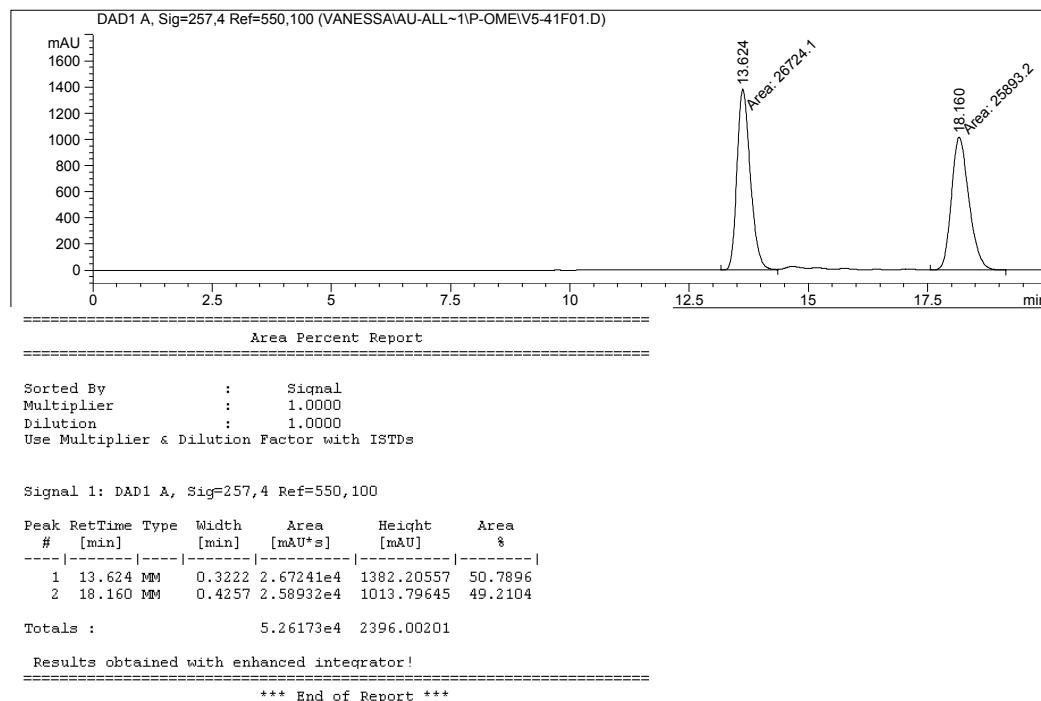


(R)-2f, for Table 2, entry 4

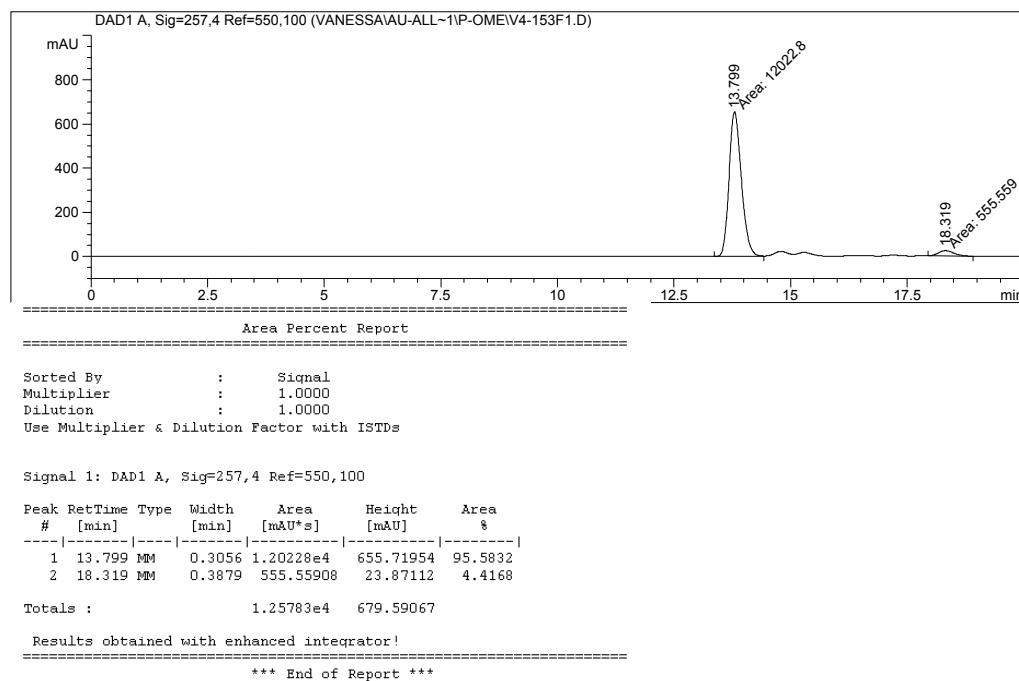


2e (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

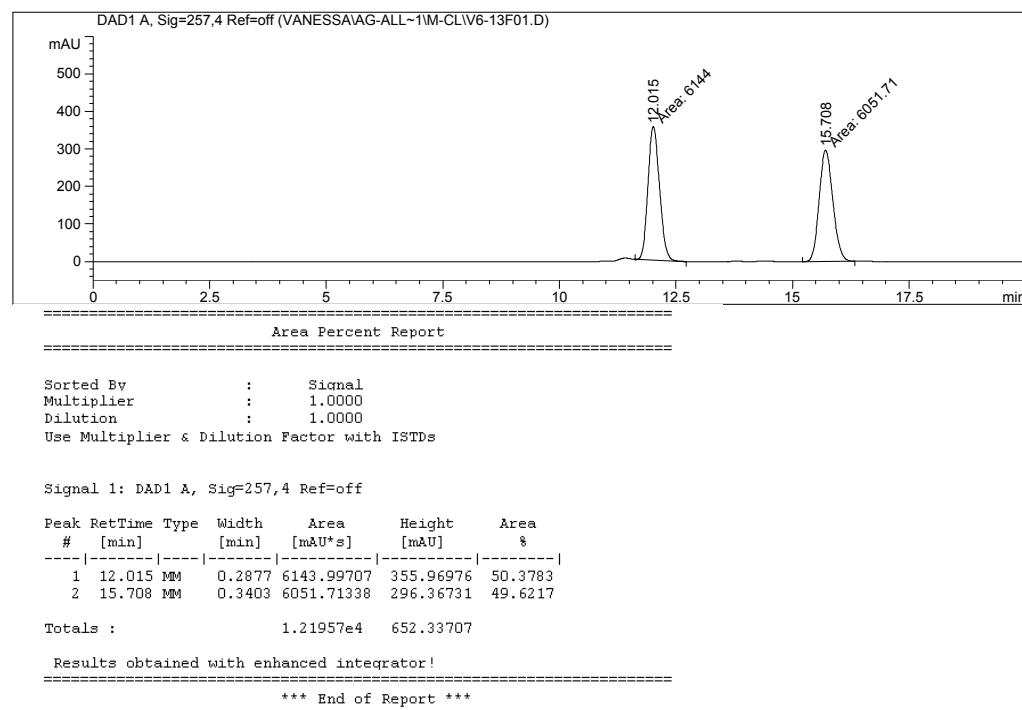


(R)-**2e**, for Table 2, entry 5

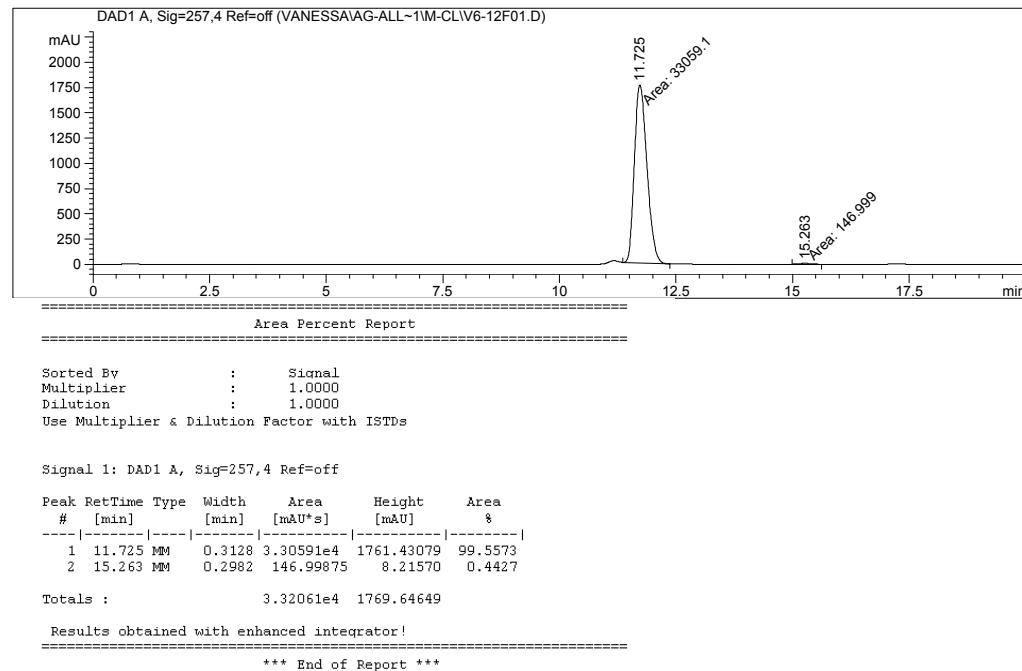


2g (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

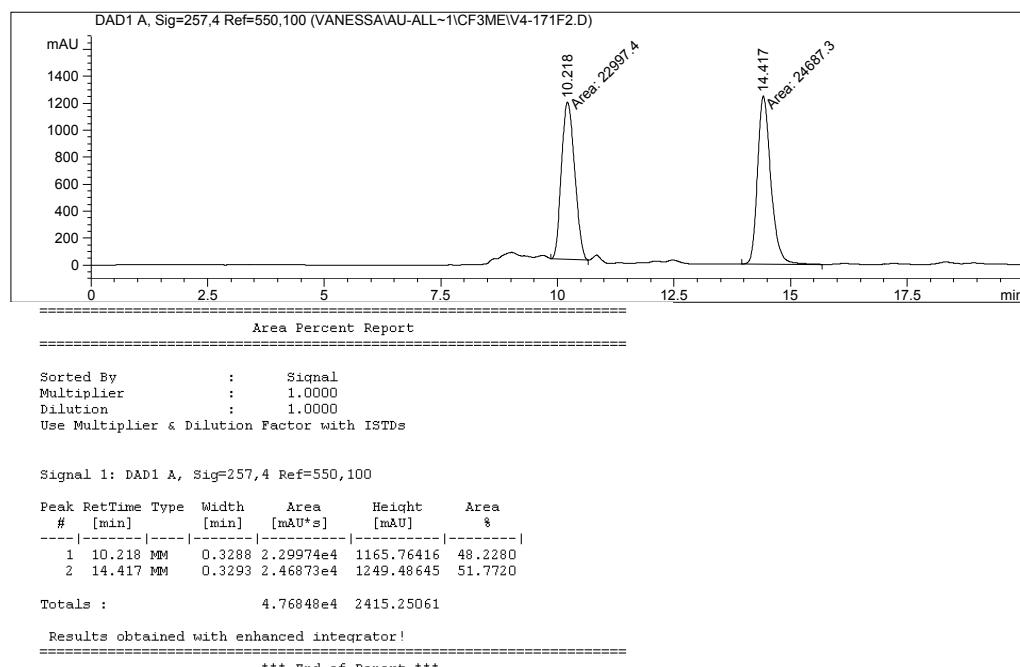


(R)-2g, for Table 2, entry 6

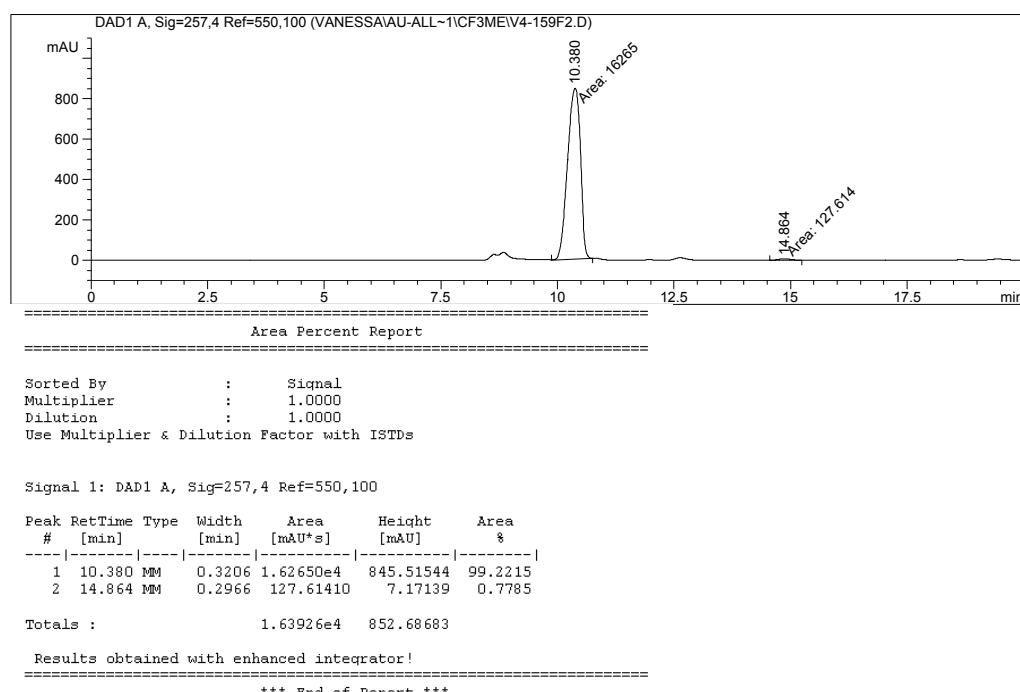


2h (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

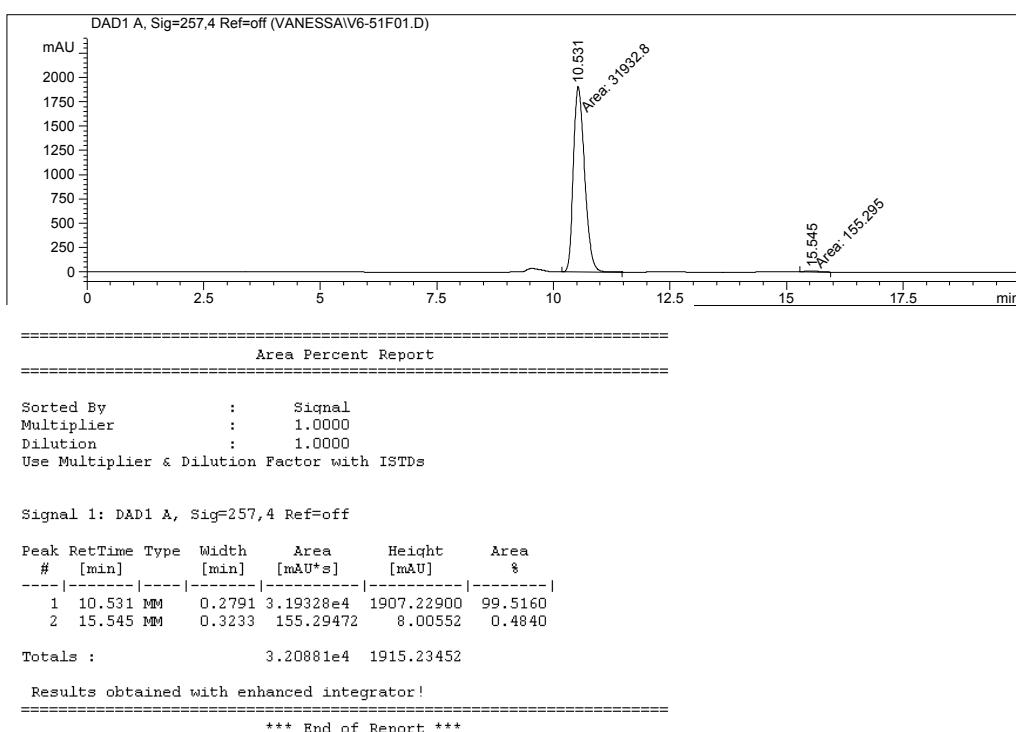
Racemic



(R)-2h, for Table 2, entry 7

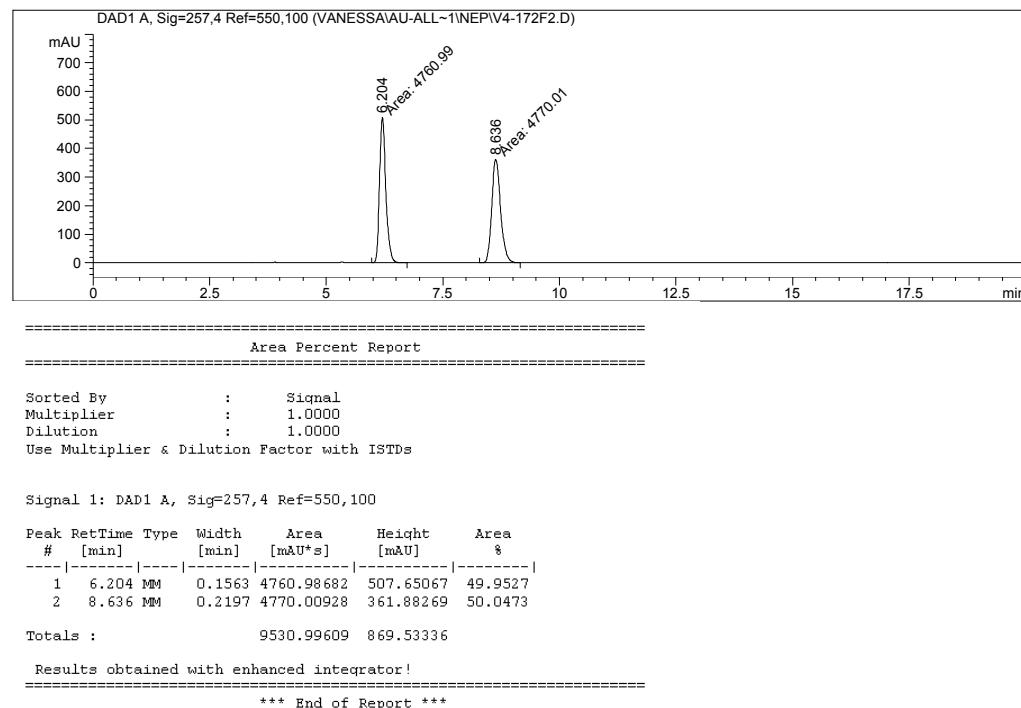


(R)-2f, for Table 2, entry 8

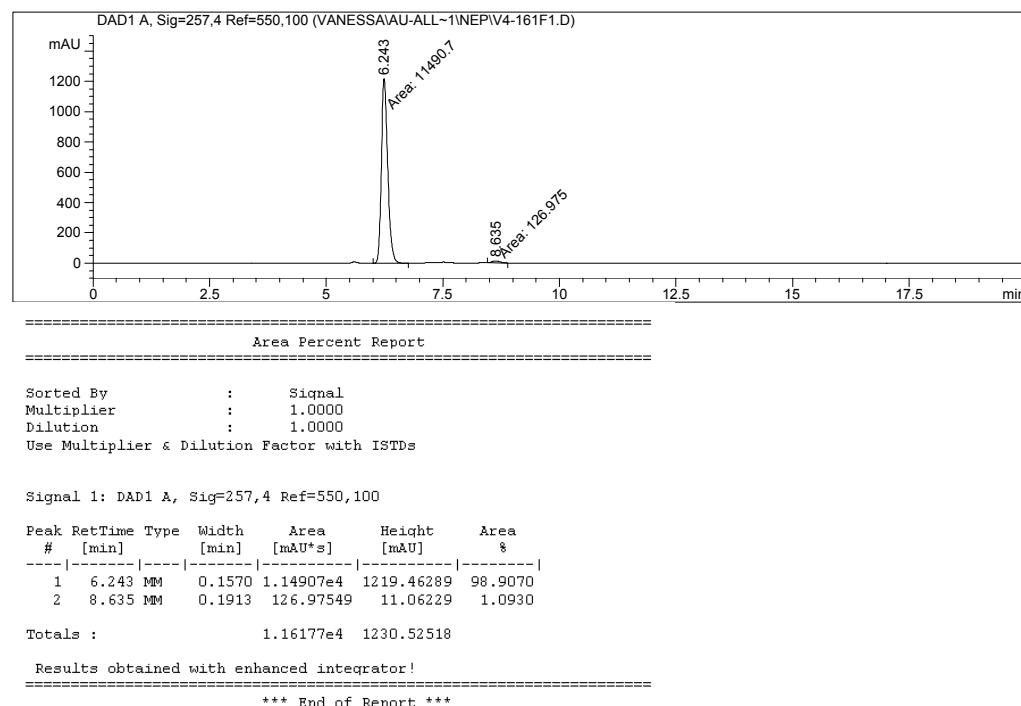


2j (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 1 mL / min)

Racemic

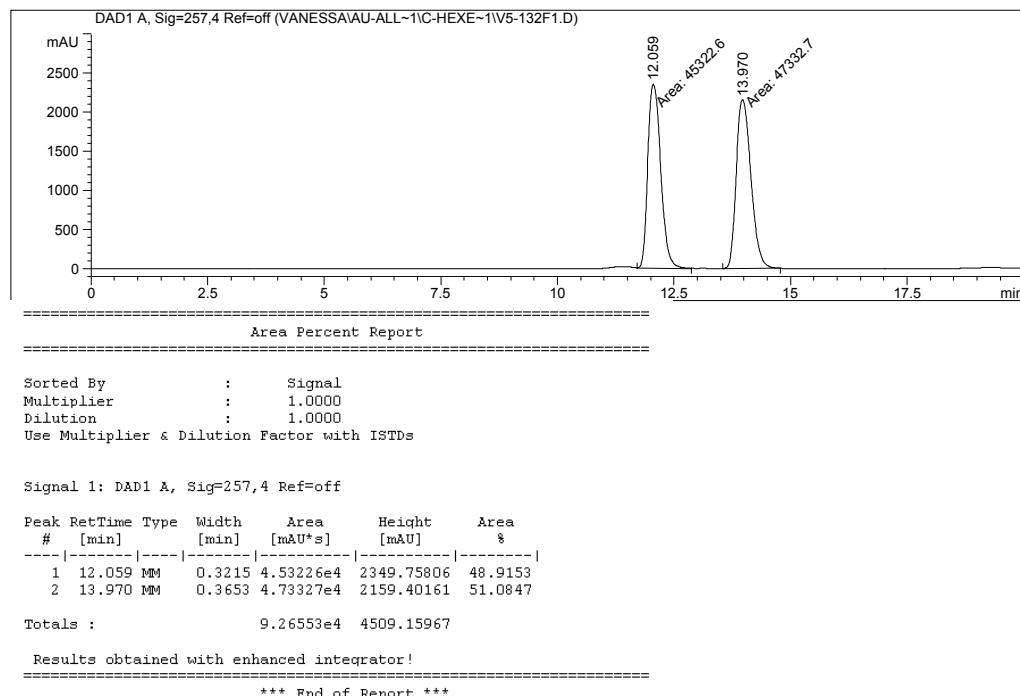


(R)-**2j**, for Table 2, entry 9

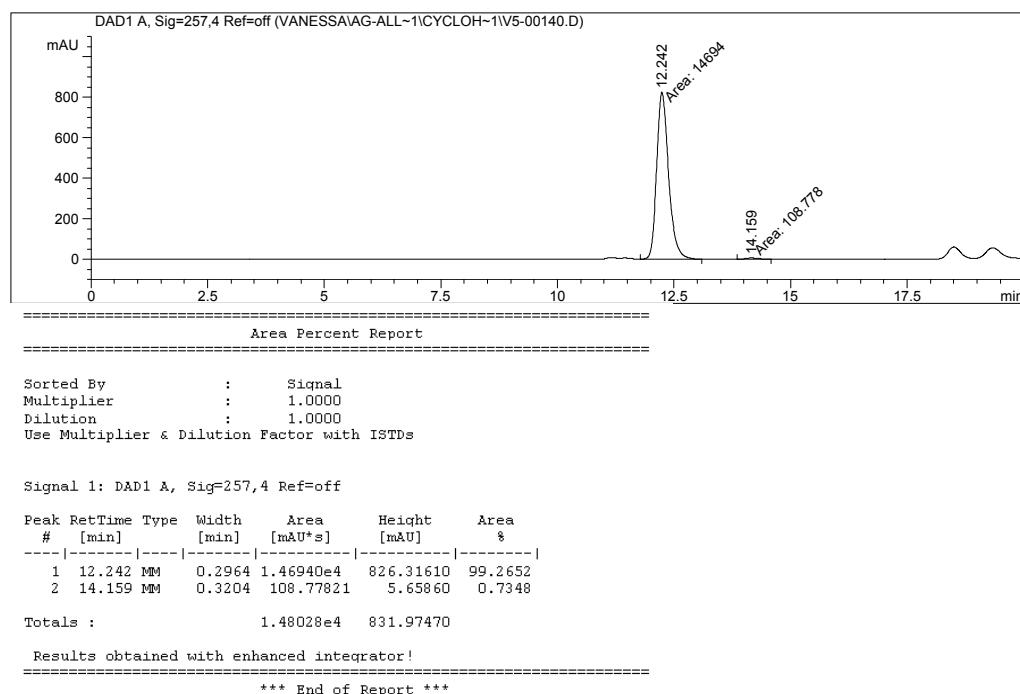


2k (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

Racemic

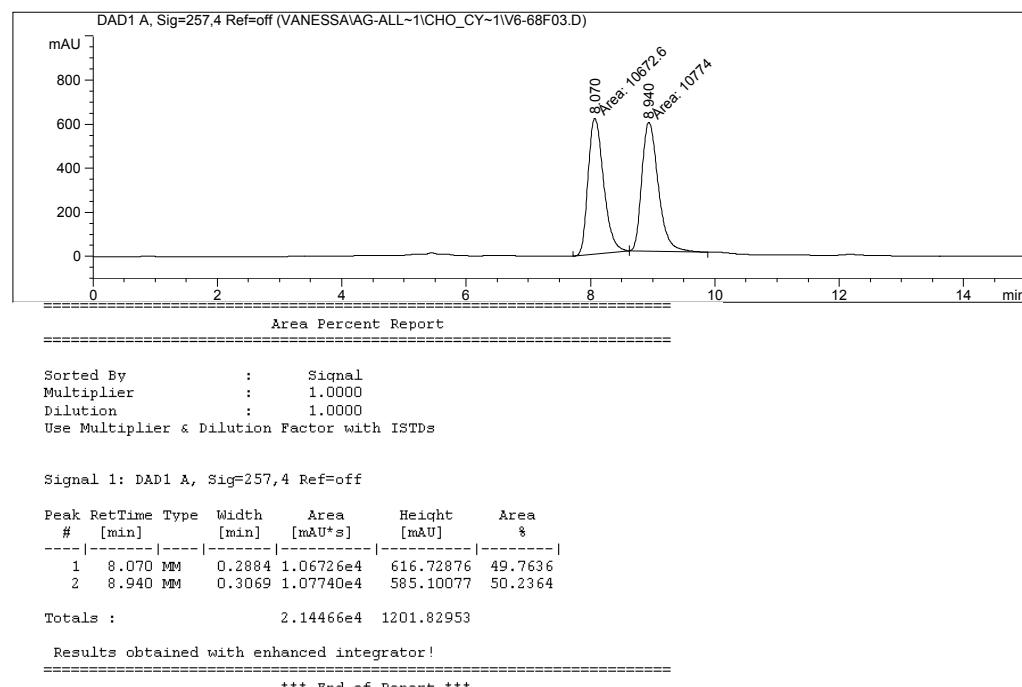


(R)-2k, for Table 2, entry 10

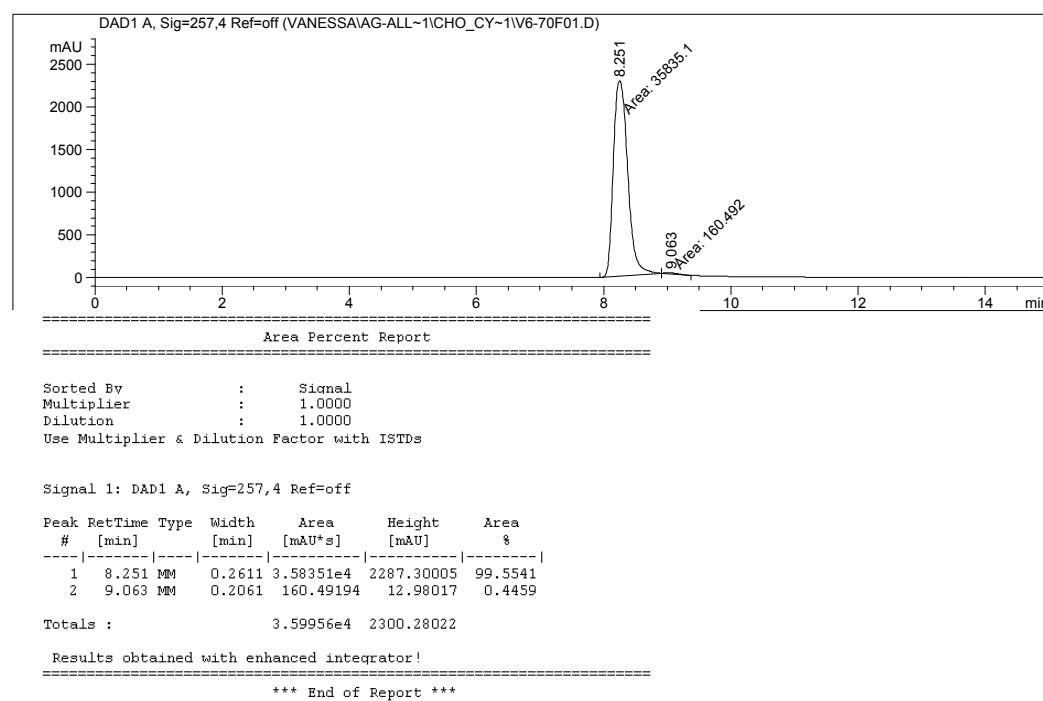


2l (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 1 mL / min)

Racemic

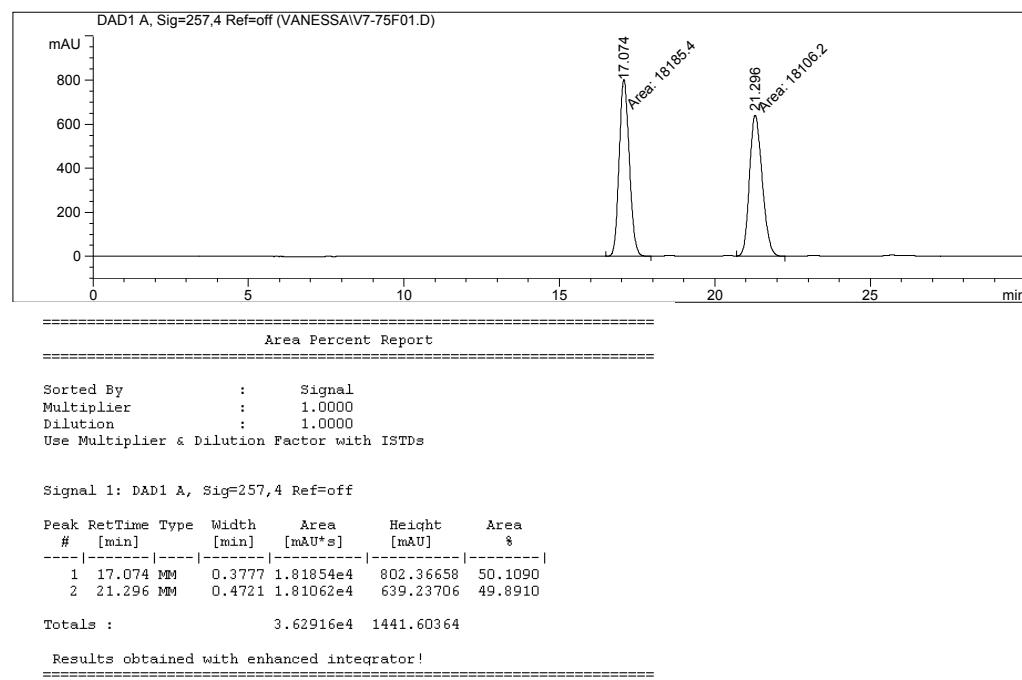


(R)-**2l**, for Table 2, entry 11

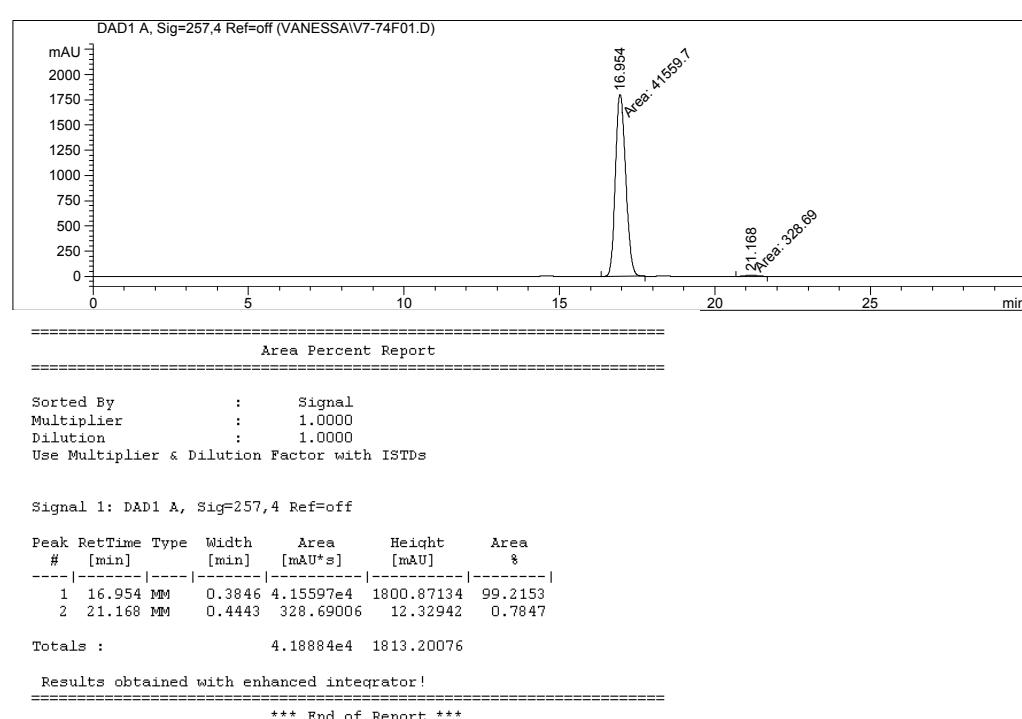


2m (Chiralcel-OD column 0.46 cm x 25 cm, 100% hexane, 0.4 mL / min)

Racemic

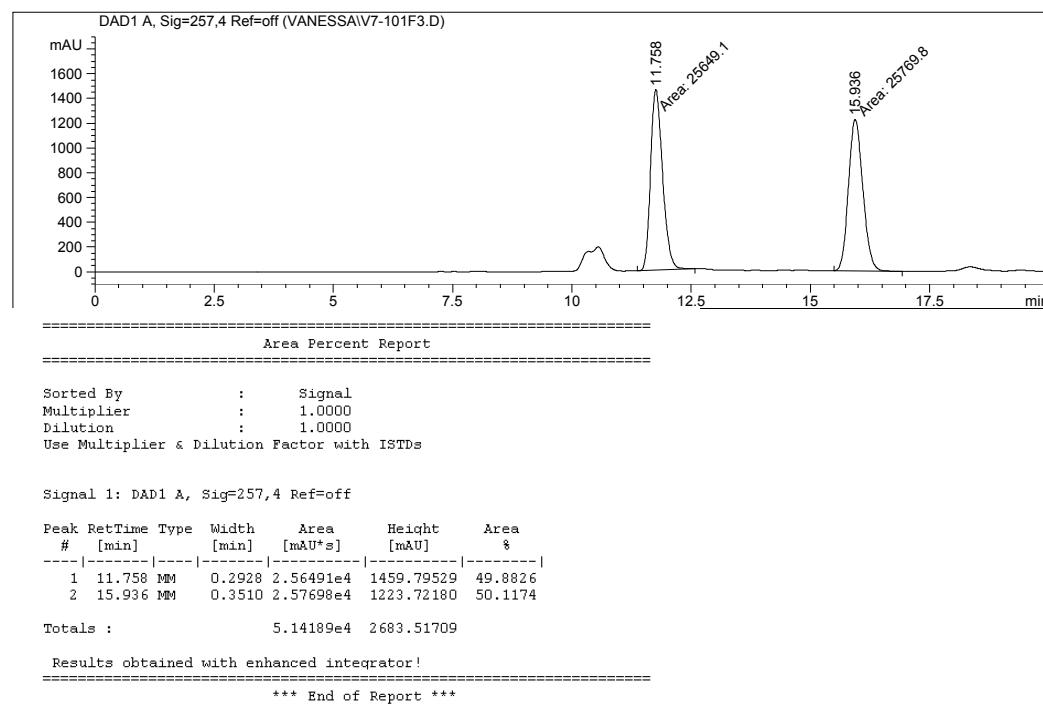


(R)-**2m**, for Table 2, entry 12

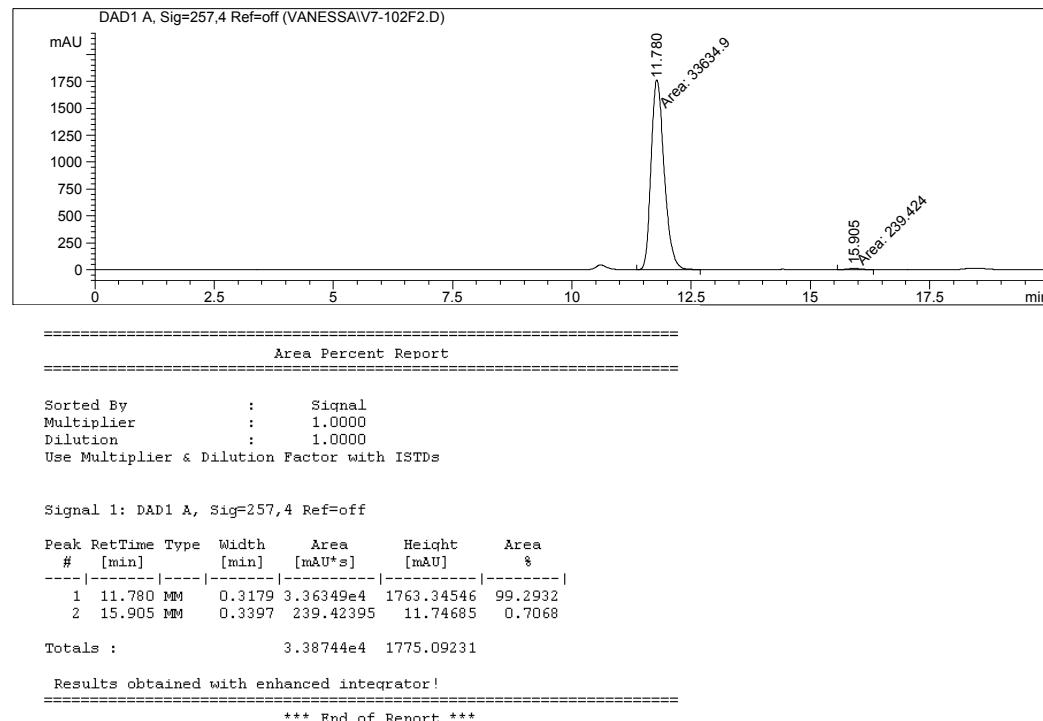


2n (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.4 mL / min)

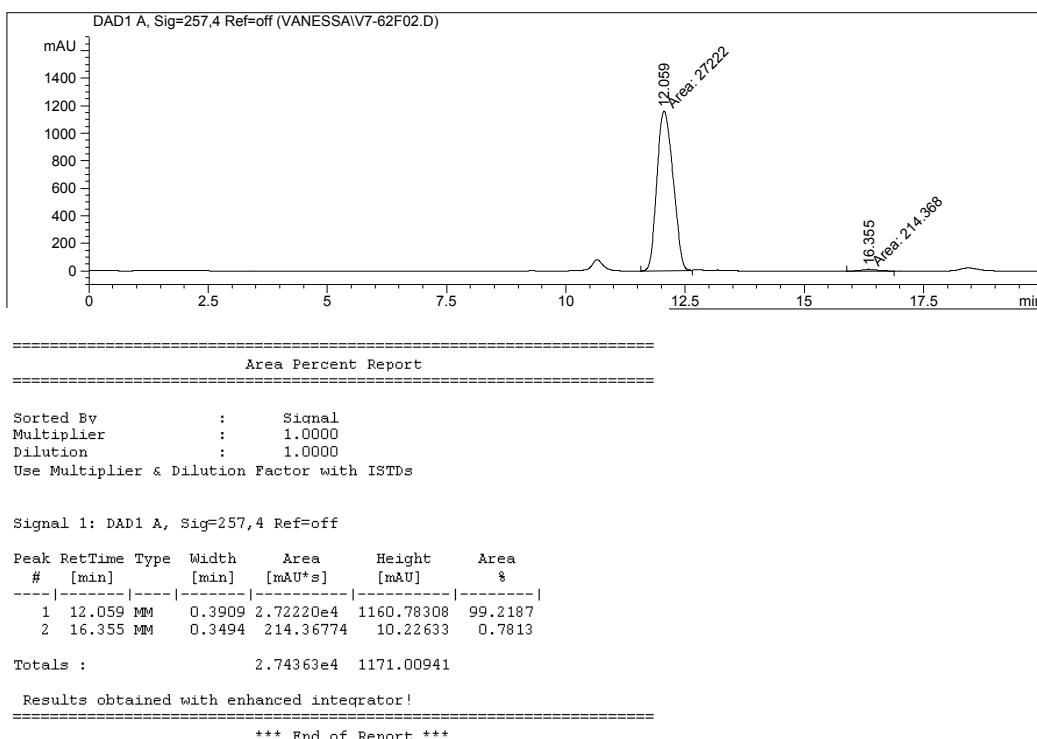
Racemic



(R)-**2n**, for Table 2, entry 13

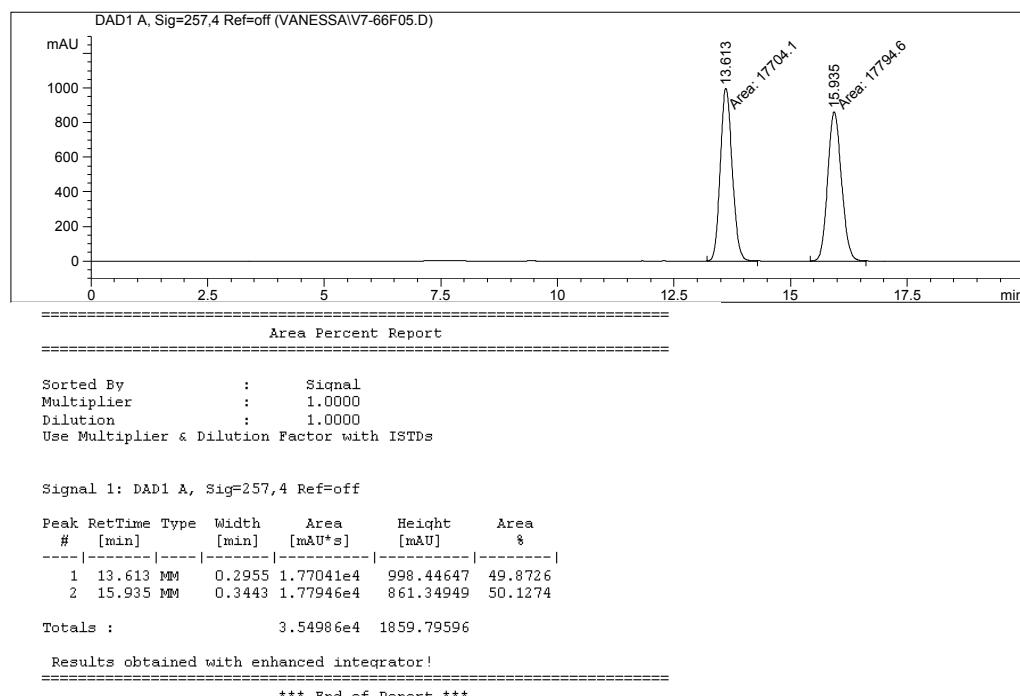


(R)-2a, for Table 3, entry 1

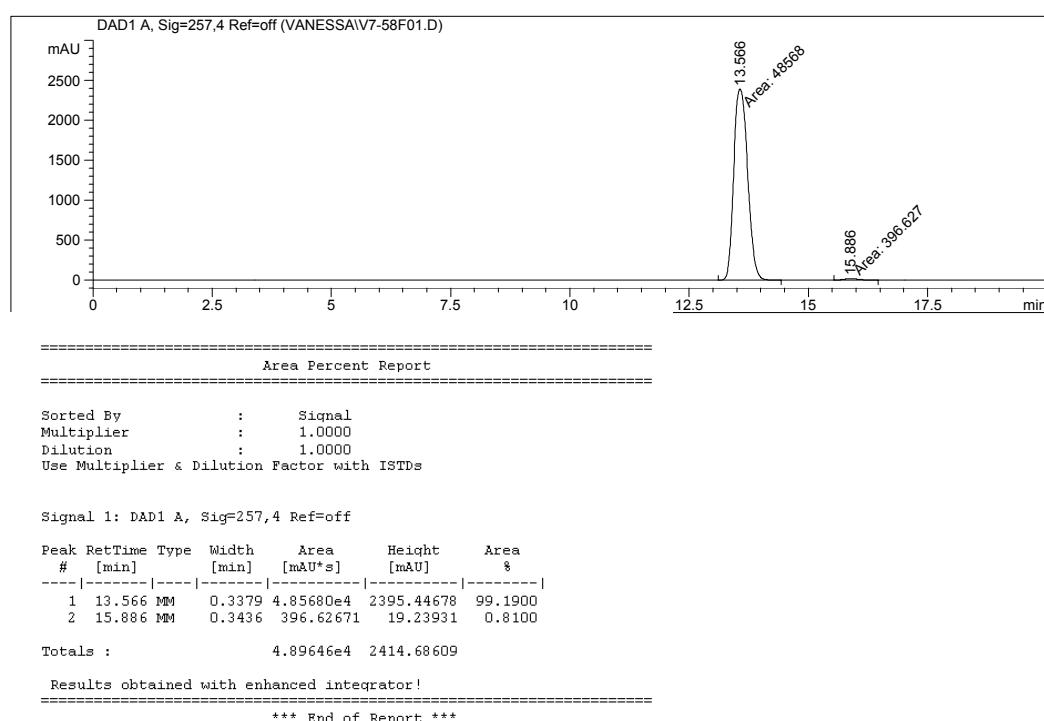


2o (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.7 mL / min)

Racemic

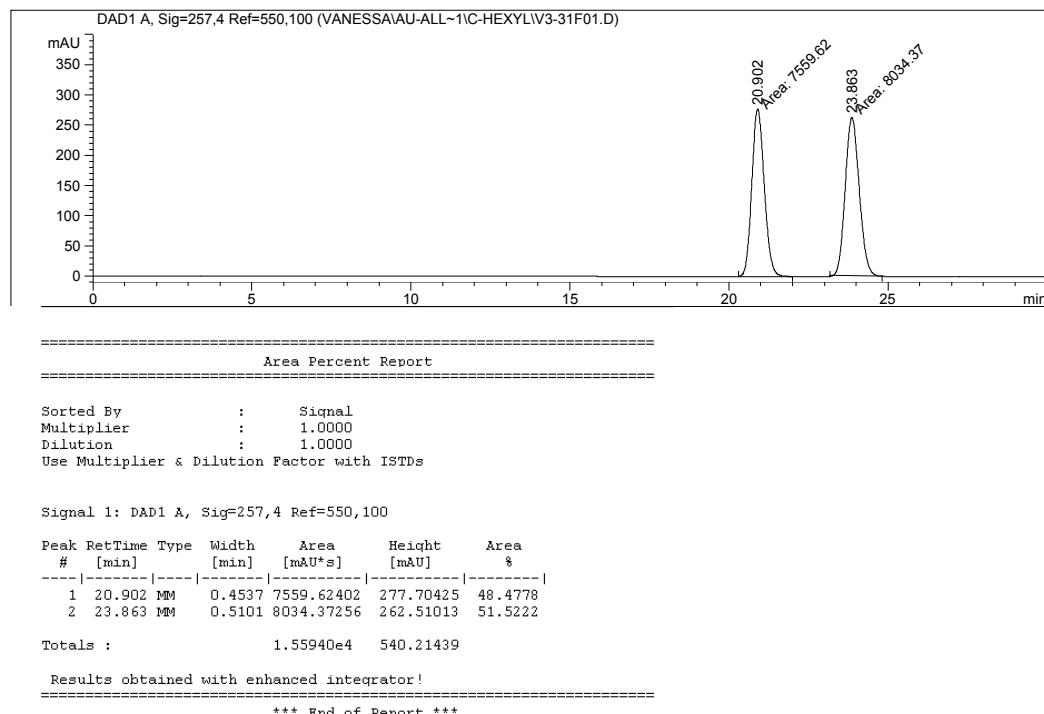


(R)-**2o**, for Table 3, entry 2

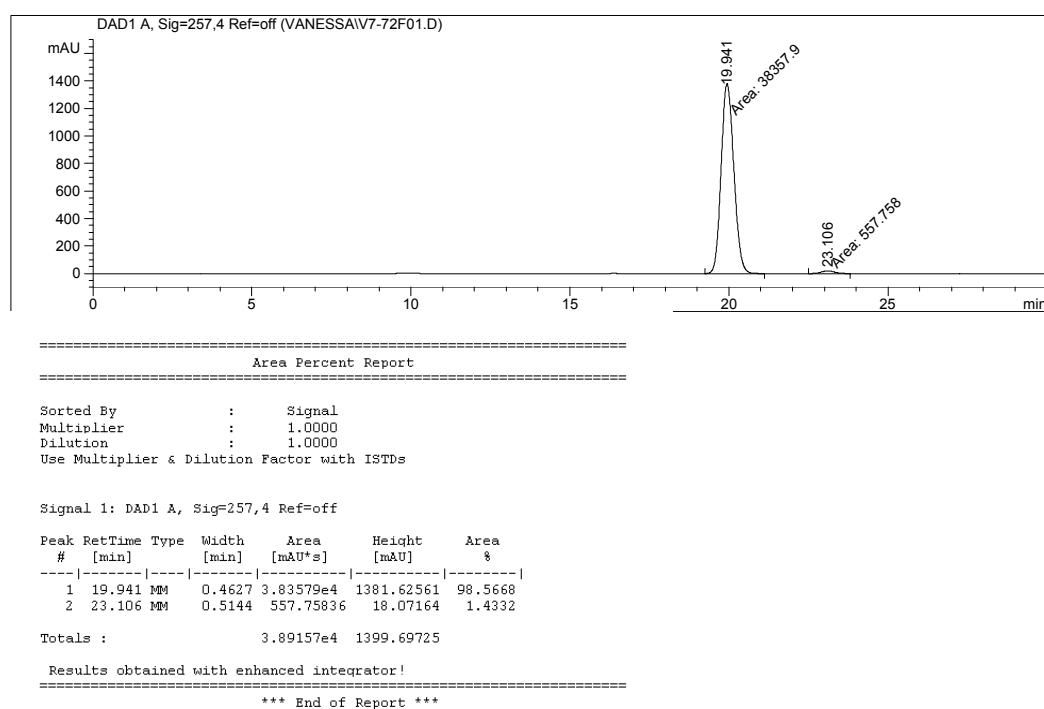


2q (Chiralcel-OD column 0.46 cm x 25 cm, 1% IPA in hexane, 0.3 mL / min)

Racemic

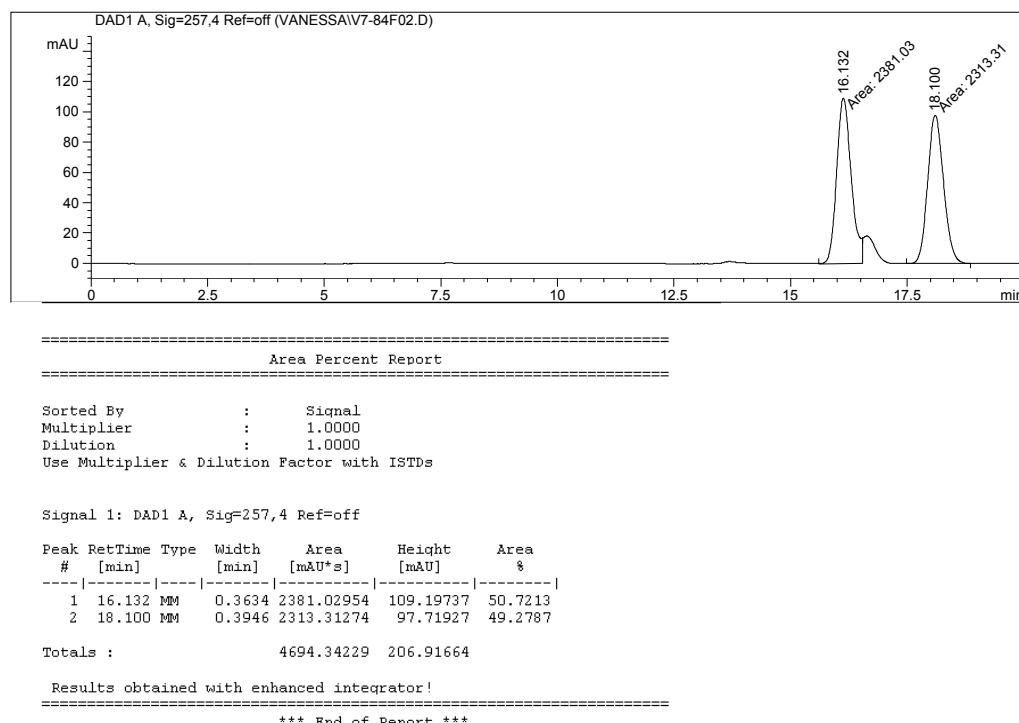


(R)-2q, for Table 3, entry 4

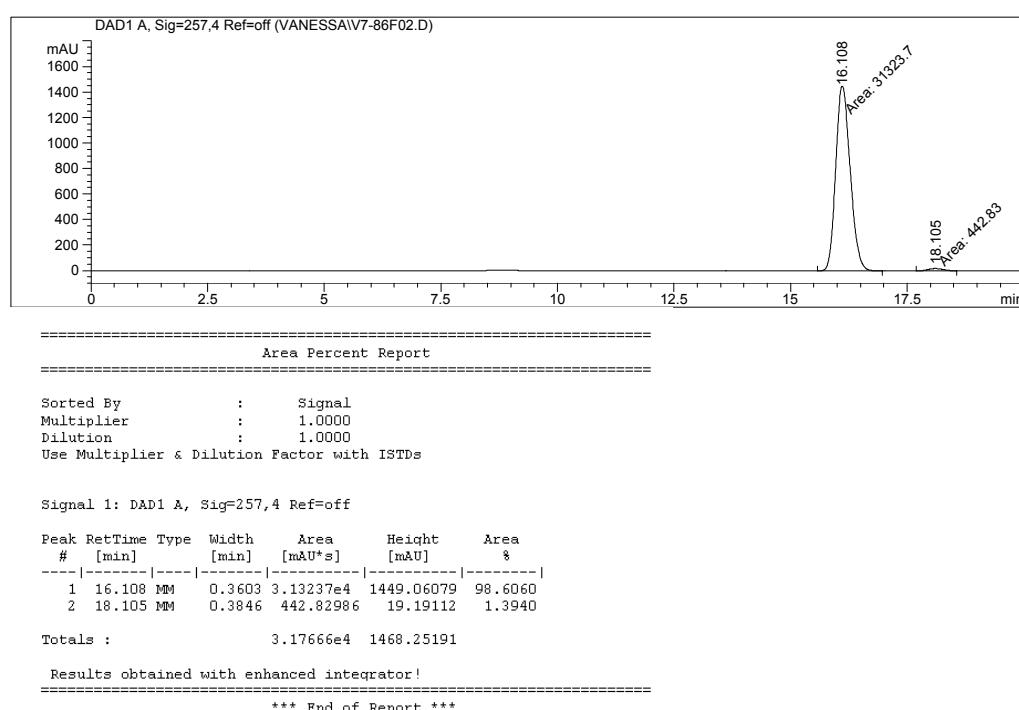


2r (Chiralcel-OD column 0.46 cm x 25 cm, 0.1% IPA in hexane, 0.4 mL / min)

Racemic

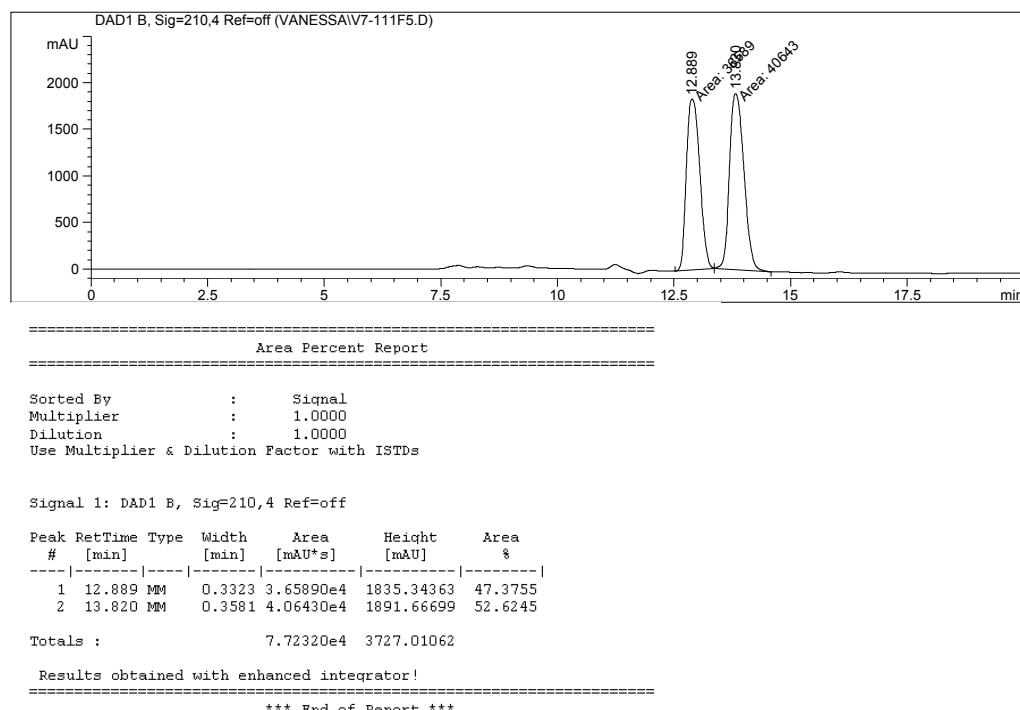


(R)-**2r**, for Table 3, entry 5

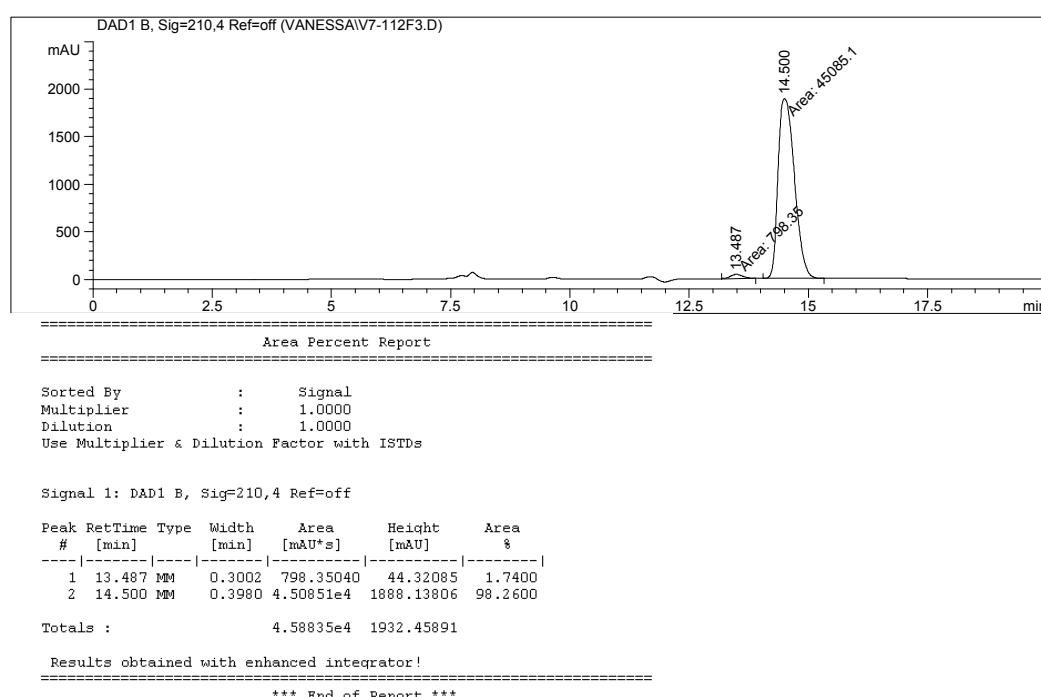


2s (Chiralcel-OD column 0.46 cm x 25 cm, 100% hexane, 0.4 mL / min)

Racemic

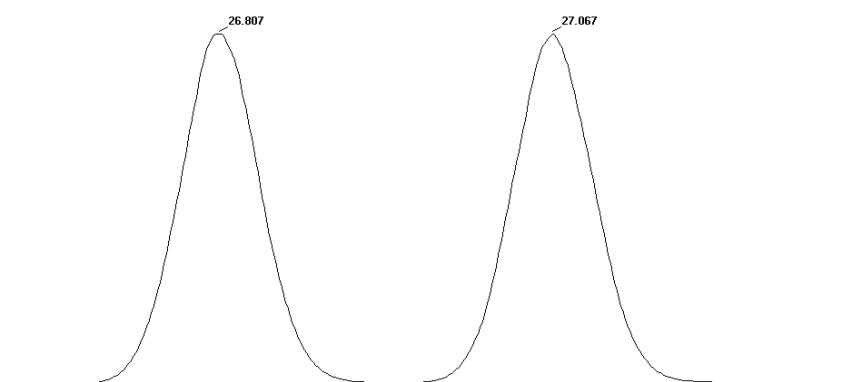


(R)-2s, for Table 3, entry 6



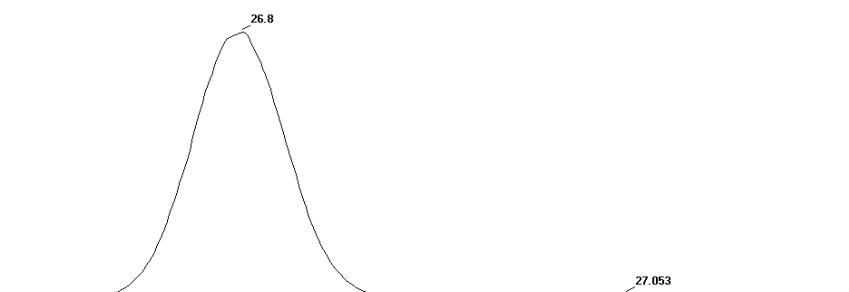
GC Spectra of 2p

Racemic (Cyclodex- β column, He as carrier gas, 1 mL/min)



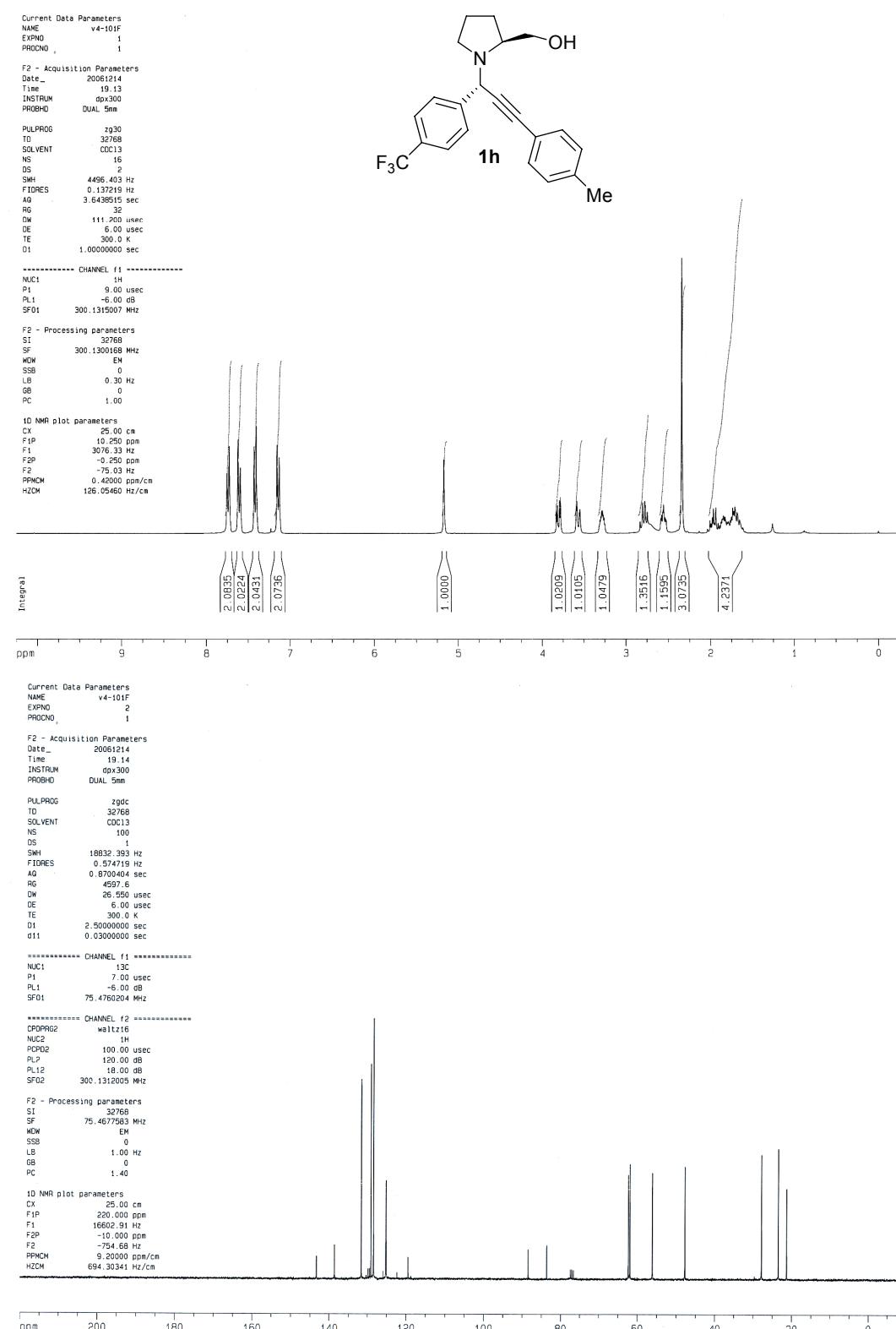
Peak #: 31	Peak area : 935763	Width at 50% : 0.0700
Peak points #: 162	Name :	Width at 10% : 0.1300
Peak ret.time : 26.8067	Concentr. : .4385625	Theor. plates : 812455.1
Peak start : 26.6767	Asymm. at 10% : 1.0526	
Peak end : 26.9467	Asymm. at 5% : 1.0698	
Peak height : 20813	USP Tailing factor : 1.0349	
Peak #: 32	Peak area : 933463	Width at 50% : 0.0717
Peak points #: 154	Name :	Width at 10% : 0.1300
Peak ret.time : 27.0667	Concentr. : .4374846	Theor. plates : 790214.3
Peak start : 26.9550	Asymm. at 10% : 1.0526	
Peak end : 27.2117	Asymm. at 5% : 1.0698	
Peak height : 20739	USP Tailing factor : 1.0349	

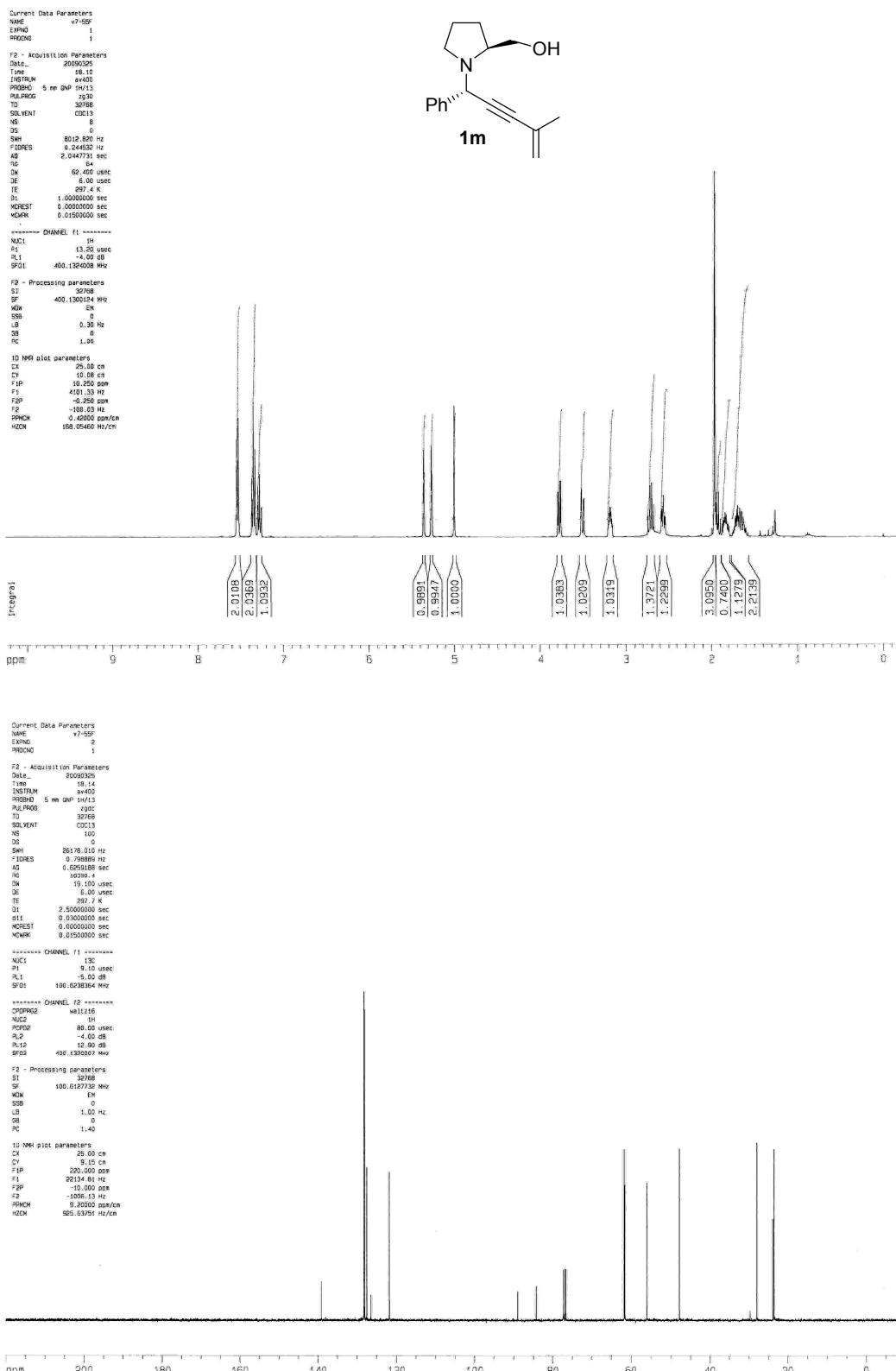
(R)-2p, for Table 3, entry 3

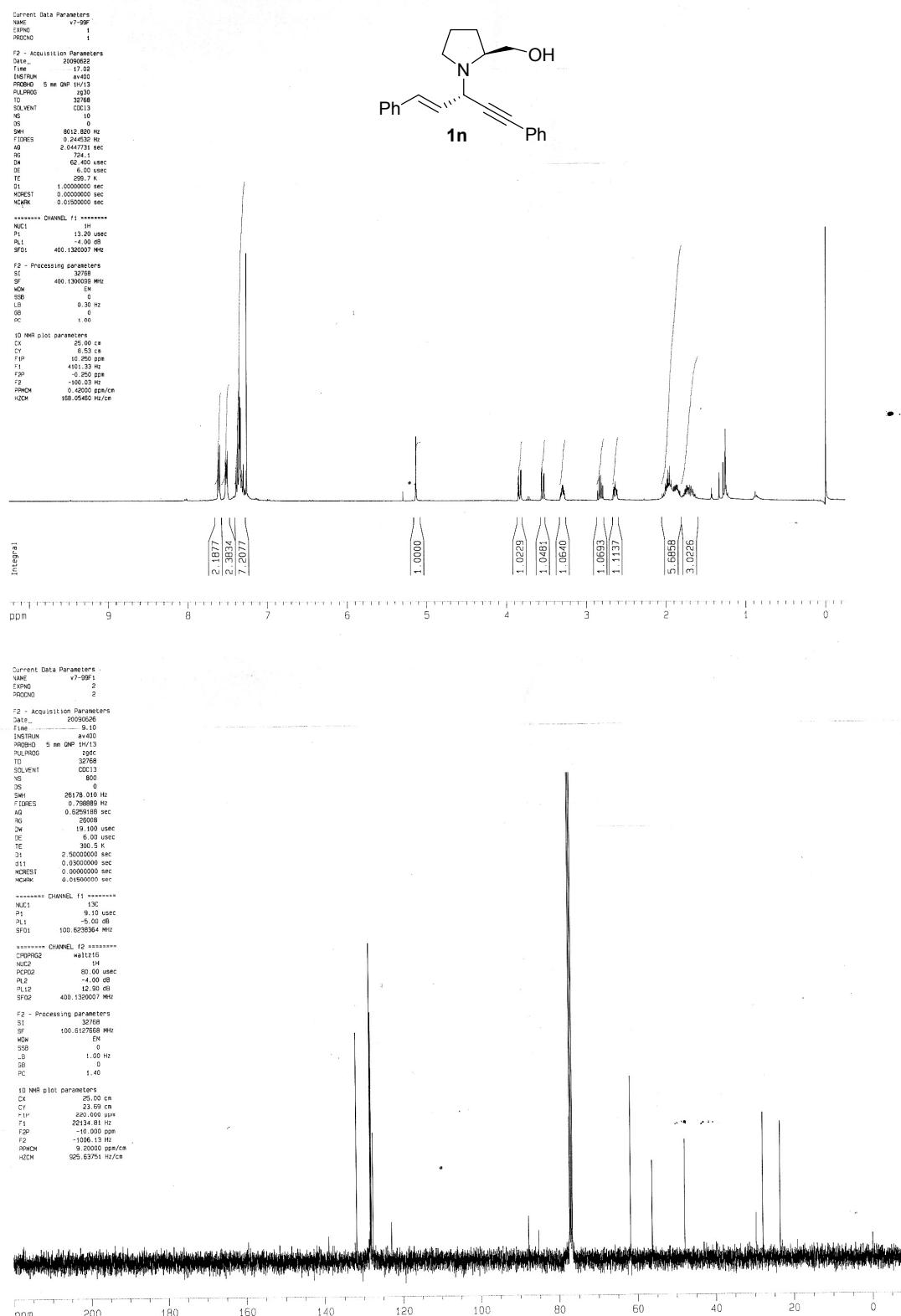


Peak #: 38	Peak area : 509280	Width at 50% : 0.0700
Peak points #: 156	Name :	Width at 10% : 0.1300
Peak ret.time : 26.8000	Concentr. : .2472914	Theor. plates : 812051
Peak start : 26.6800	Asymm. at 10% : 0.95	
Peak end : 26.9400	Asymm. at 5% : 0.9778	
Peak height : 11150	USP Tailing factor : 0.9889	
Peak #: 39	Peak area : 9883	Width at 50% : 0.0700
Peak points #: 121	Name :	Width at 10% : 0.1250
Peak ret.time : 27.0533	Concentr. : 0	Theor. plates : 827475.8
Peak start : 26.9567	Asymm. at 10% : 1.3438	
Peak end : 27.1583	Asymm. at 5% : 1.4118	
Peak height : 225	USP Tailing factor : 1.2059	

¹H and ¹³C NMR Spectra







Current Data Parameters
NAME v3-185F
EXPNO 1
PROCNO 1

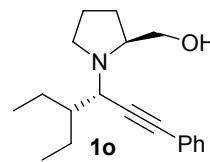
F2 - Acquisition Parameters
Date_ 20081027
Time 18.02
INSTRUM dpx300
PROBHD DUAL 5mm
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 12
DS 2
SWH 4498.403 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 18
DW 111.0 usec
DE 6.00 usec
TE 300.0 K
D1 1.0000000 sec

***** CHANNEL f1 *****

NUC1 1H
P1 9.0 usec
PL1 -5.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters
SI 32768
SF 300.1300221 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 25.00 cm
F1P 10.250 ppm
F1 3076.33 Hz
F2P -10.00 ppm
F2 -75.03 Hz
PPMCH 0.42000 ppm/cm
HZCM 126.05462 Hz/cm



Integral]

ppm 1 9 8 7 6 5 4 3 2 1 0

2.0378
2.9497

1.0000
4.1594

5.0762
4.3531

6.4161

Current Data Parameters
NAME v3-185F
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081027
Time 18.04
INSTRUM dpx300
PROBHD DUAL 5mm
PULPROG zg30c
TD 32768
SOLVENT CDCl3
NS 8
DS 1
SWH 18832.493 Hz
FIDRES 0.574719 Hz
AQ 0.870448 sec
RG 450
DW 26.550 usec
DE 6.00 usec
TE 300.0 K
D1 2.8000000 sec
d1t 0.0300000 sec

***** CHANNEL f1 *****

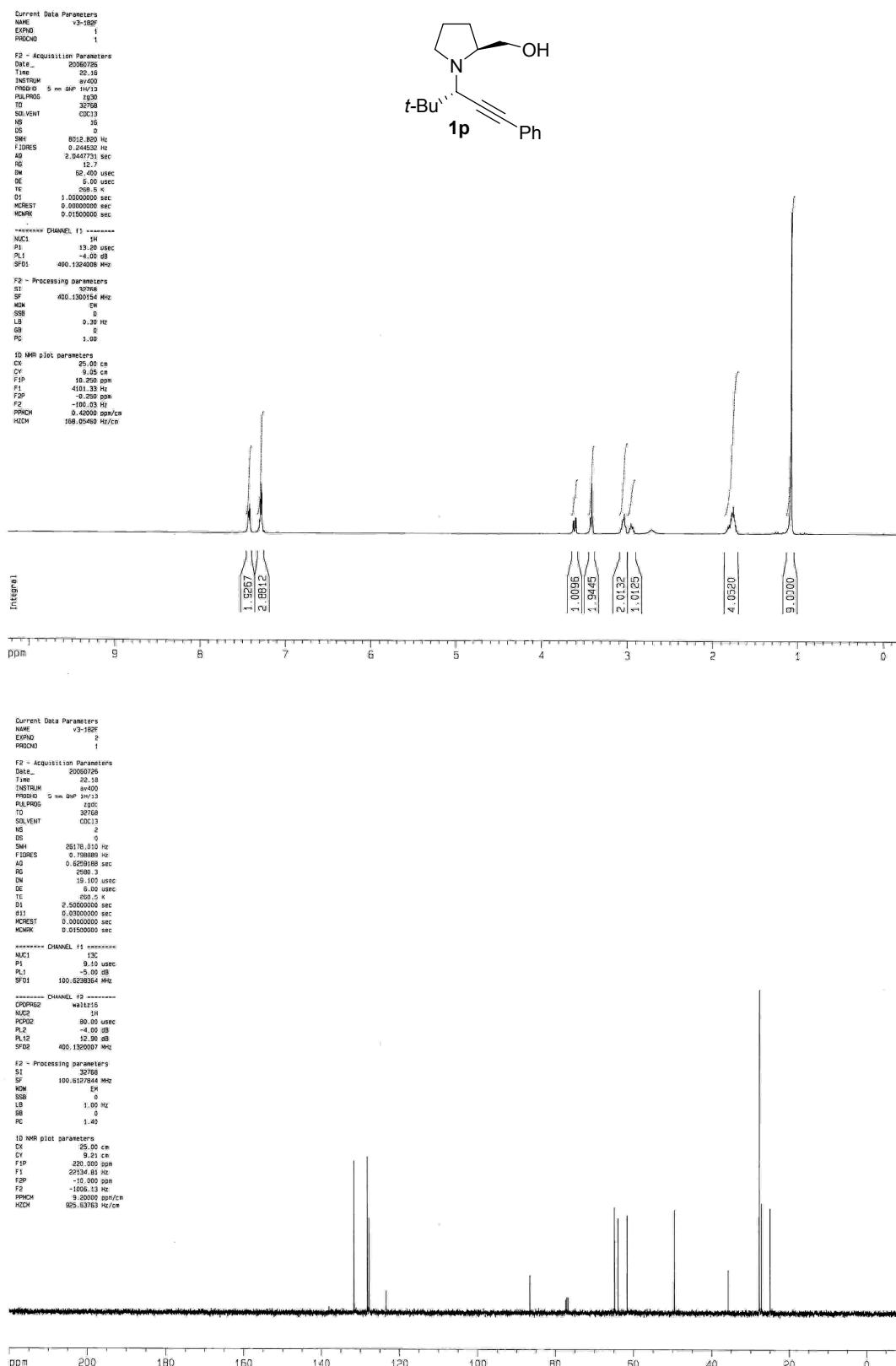
NUC1 13C
P1 7.00 usec
PL1 -6.00 dB
SF01 75.4762004 MHz

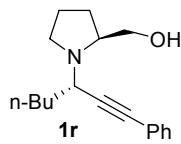
***** CHANNEL f2 *****
CPDPFG2 w811216
CP1C2 1H
CPDPFG1 100.00 usec
PL2 120.00 dB
PL12 18.00 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4675777 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 25.00 cm
F1P 26502.81 ppm
F1 16502.81 Hz
F2P -10.00 ppm
F2 -754.68 Hz
PPMCH 9.20000 ppm/cm
HZCM 694.30341 Hz/cm

ppm 200 180 160 140 120 100 80 60 40 20 0





```

Current Data Parameters
NAME          v3-156F
EXPNO         1
PRDNO        1

P2 - Acquisition Parameters
Data_1       20060711
Time         18.46
INSTRNAME    BRUKER
PROBODIM    5 mm gav
PULPROG     3D_sst_1H3D
TD           32768
SOLVENT      C6C13
NS           16
SWH          8812.000 Hz
FIDRES      0.244682 Hz
TDZ          2.047473 sec
RG           12.7
DW           62.000 usec
DW1          6.000 usec
TC           18.000 K
TEC          1.000000 sec
MCOUNT      0.00000000 sec
MCHEST      0.00000000 sec
NMHCNT     0.01500000 sec

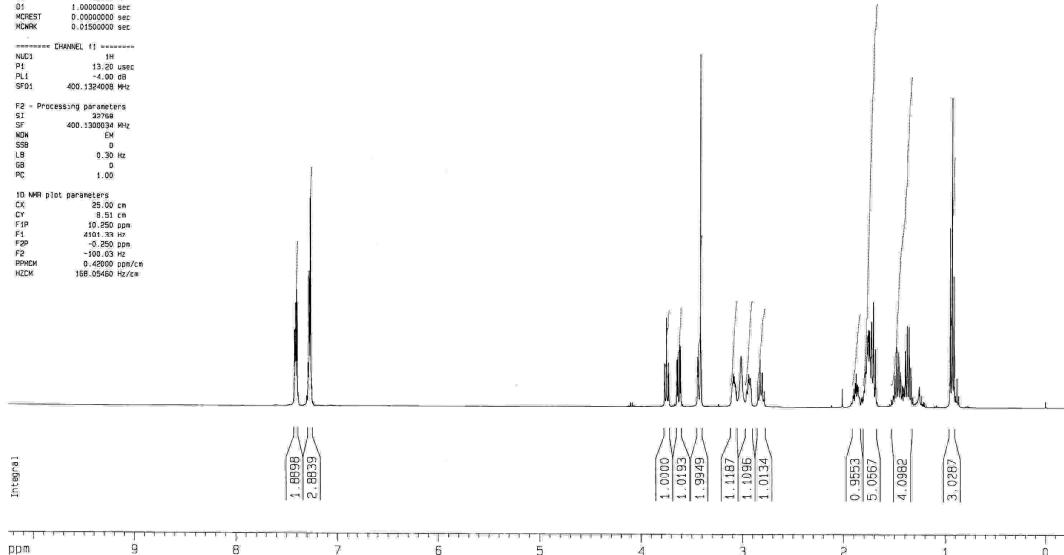
```

```
***** CHANNEL 11 *****

NUC1          1H
PI           13.20 usec
PLI          -4.00 dB
SPD1        400.132000 MHz

F2 - Processing parameters
SI            27984
SF        400.130000 MHz
NDN             EM
SSB              0
LB            0.30 Hz
GB              0
PC            1.00

1D MRI plot parameters
CS           25.00 cm
CY           9.51 cm
FIP          10.25 ppm
FI           400.00 Hz
FPP          -20.00 ppm
RPM          0.42000 ppm/c
KCMZ        156.054624
```



```

Current Data Parameters
NAME          v3-16SF
EXPNO         2
PROCNO        1

Data - Acquisition Parameters
TS             2000.014
Time           18.48
INSTRUM       av400
PROBHD        D MM BNP 1H13
PULPROG      zgpc
TD             32768
SOLVENT        MeOH
NS             19
DS             0
SWH           26178.01 Hz
FIDRES       0.759889 Hz
AQ             0.625000 sec
RG             724.1
DW             10.00 usec
DE             6.00 usec
TC             268.4 K
DT            0.000000 sec
D1           0.0300000 sec
M1           0.0000000 sec
MCRT          0.0150000 sec

```

```

***** CHANNEL 1 *****

MC1      1.13dB
P1       9.10  usec
PL1      -5.00  dB
SF01    100.628264 MHz

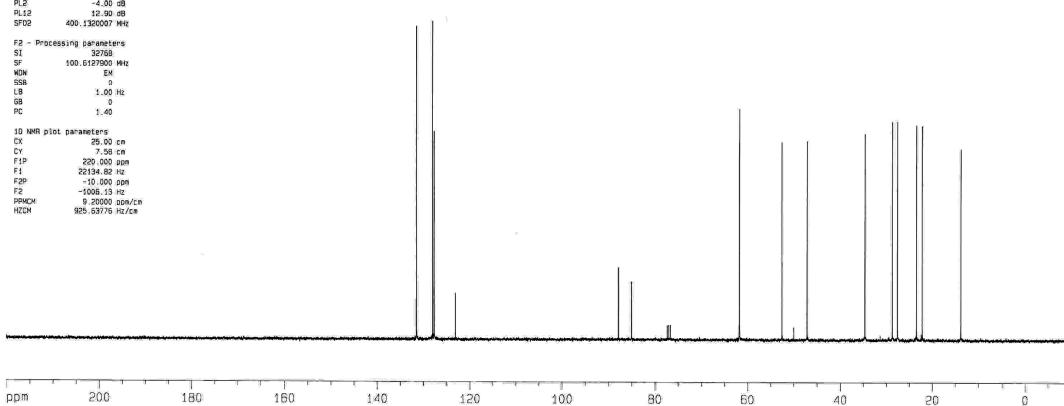
***** CHANNEL 2 *****

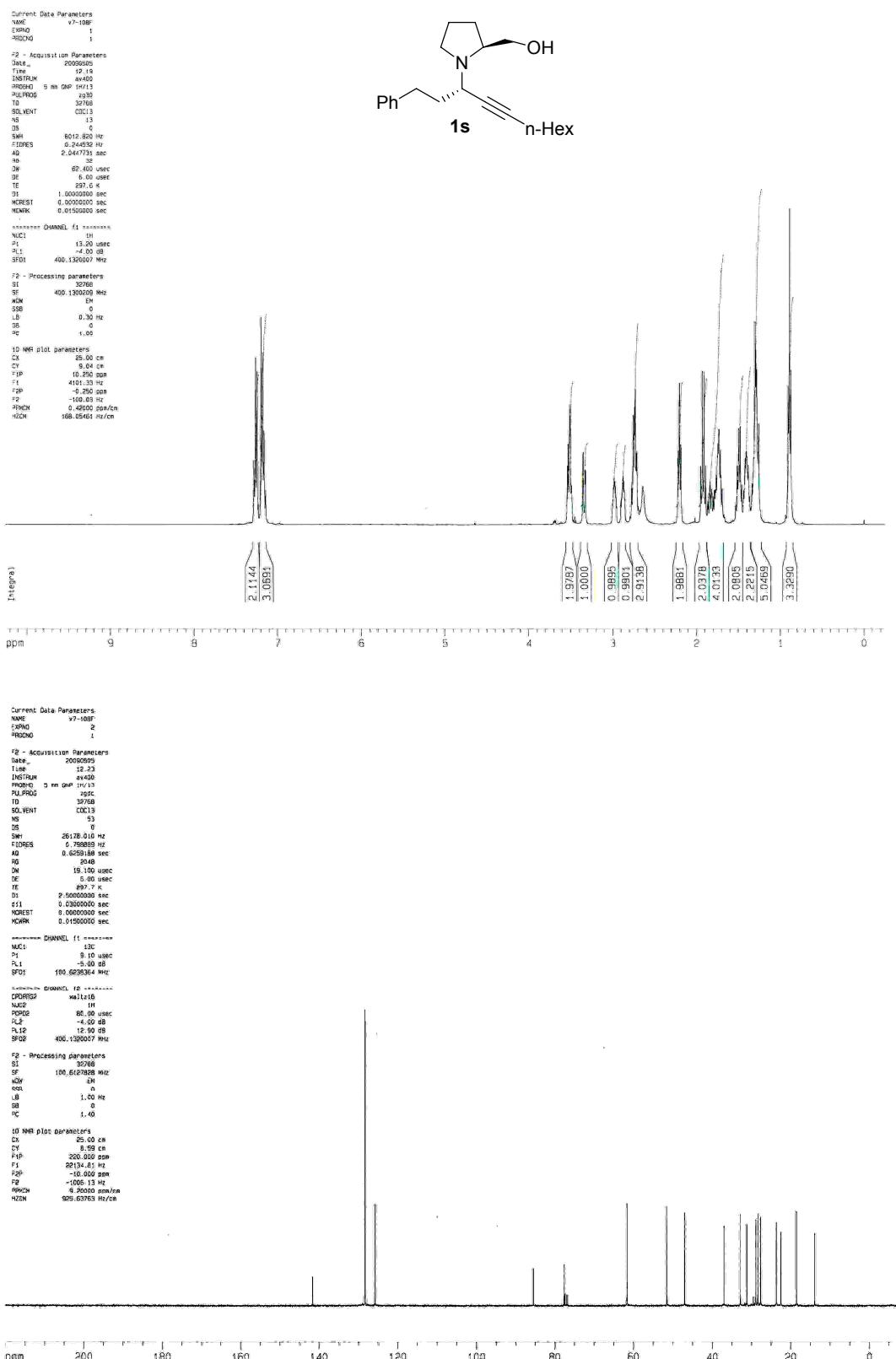
PRG2      9.11dB
PCP02     80.00  usec
PL2       -4.00  dB
PL12     12.00  dB
SF02    400.130000 MHz

#4 - Processing parameters
SI          32768
NOI      100.627990 MHz
MWN        9
SSB        9
LB       1.00  Hz
GB        1.40

#5 - WVR plot parameters
CX        25.00 cm
CY        7.56 cm
PP1      200.00 ppm
F1       2034.13 Hz
F2P      -10.000 ppm
F2C      -1008.15 Hz
PROMCH  0.000000
HZCH  529.537573 Hz

```





Current Data Parameters
NAME v5-180F
EXPNO 1
PROCNO 1

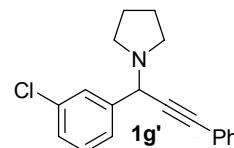
F2 - Acquisition Parameters
Date 20080109
Time 20:13
INSTRUM dpX300
PROBHD DUAL 5mm
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 4
DS 2
SWH 4496.403 Hz
ETDRES 0.137219 Hz
AQ 3.6438515 sec
RG 80.6
DW 111.0 usec
DE 6.00 usec
TE 300.0 K
DI 1.0000000 sec

***** CHANNEL f1 *****

N1C1 1H
P1 9.00 usec
PL1 -6.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters
SI 32768
SF 300.1300159 MHz
WM EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 25.00 ppm
F1P 10.250 ppm
F1 3076.33 Hz
F2P -0.250 ppm
F2 -75.03 Hz
PPMCM 0.42000 ppm/cm
HZCM 125.05460 Hz/cm



Integral

ppm 9 8 7 6 5 4 3 2 1 0

Current Data Parameters
NAME v5-180F
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date 20080109
Time 20:14
INSTRUM dpX300
PROBHD DUAL 5mm
PULPROG zg30c
TD 32768
SOLVENT CDCl3
NS 42
DS 1
SWH 18832.393 Hz
ETDRES 0.574719 Hz
AQ 0.8700000 sec
RG 3251
DW 26.550 usec
DE 6.00 usec
TE 300.0 K
DI 2.5000000 sec
d11 0.0300000 sec

***** CHANNEL f1 *****

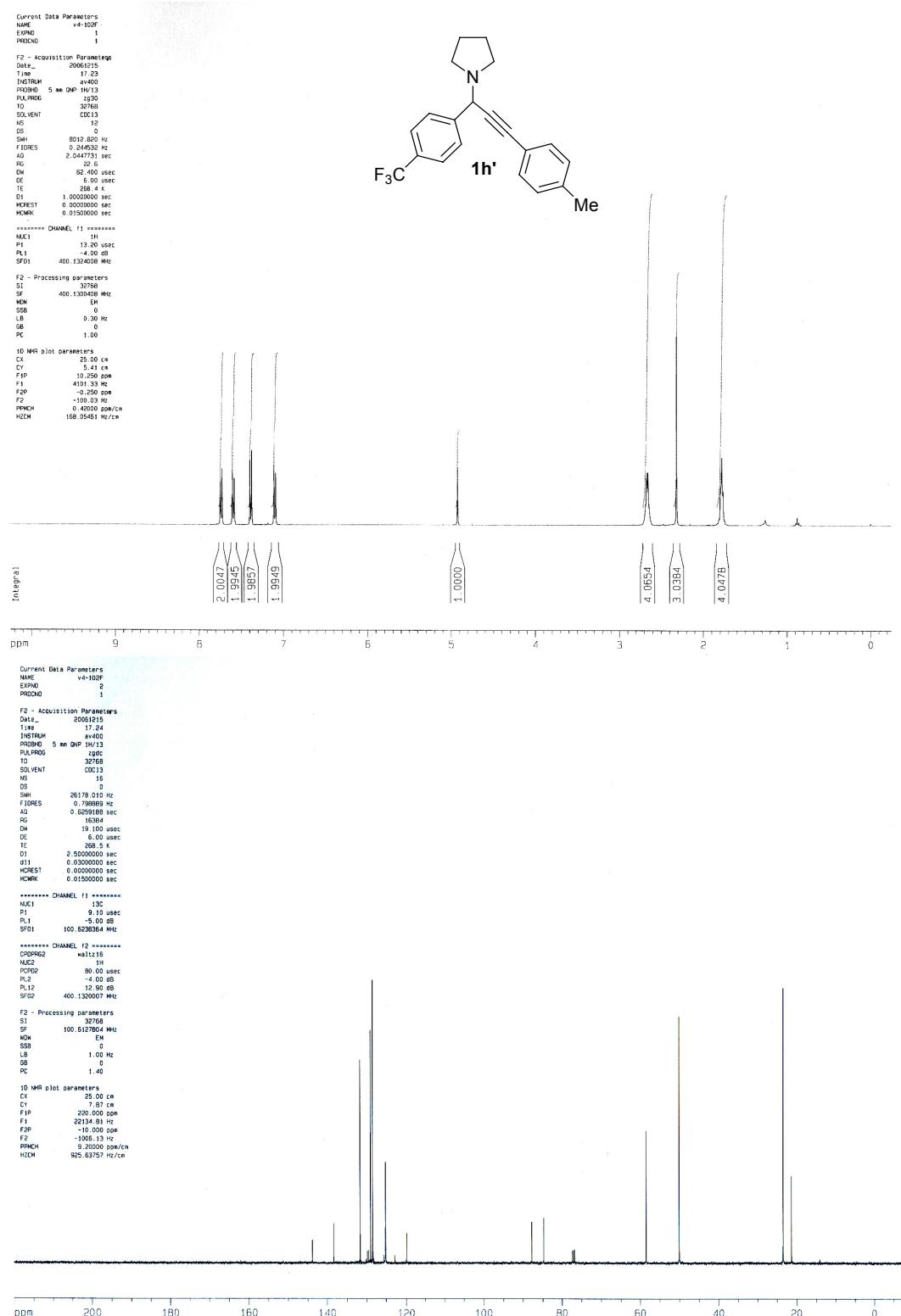
N1C1 13C
P1 7.00 usec
PL1 -6.00 dB
SF01 75.476024 MHz

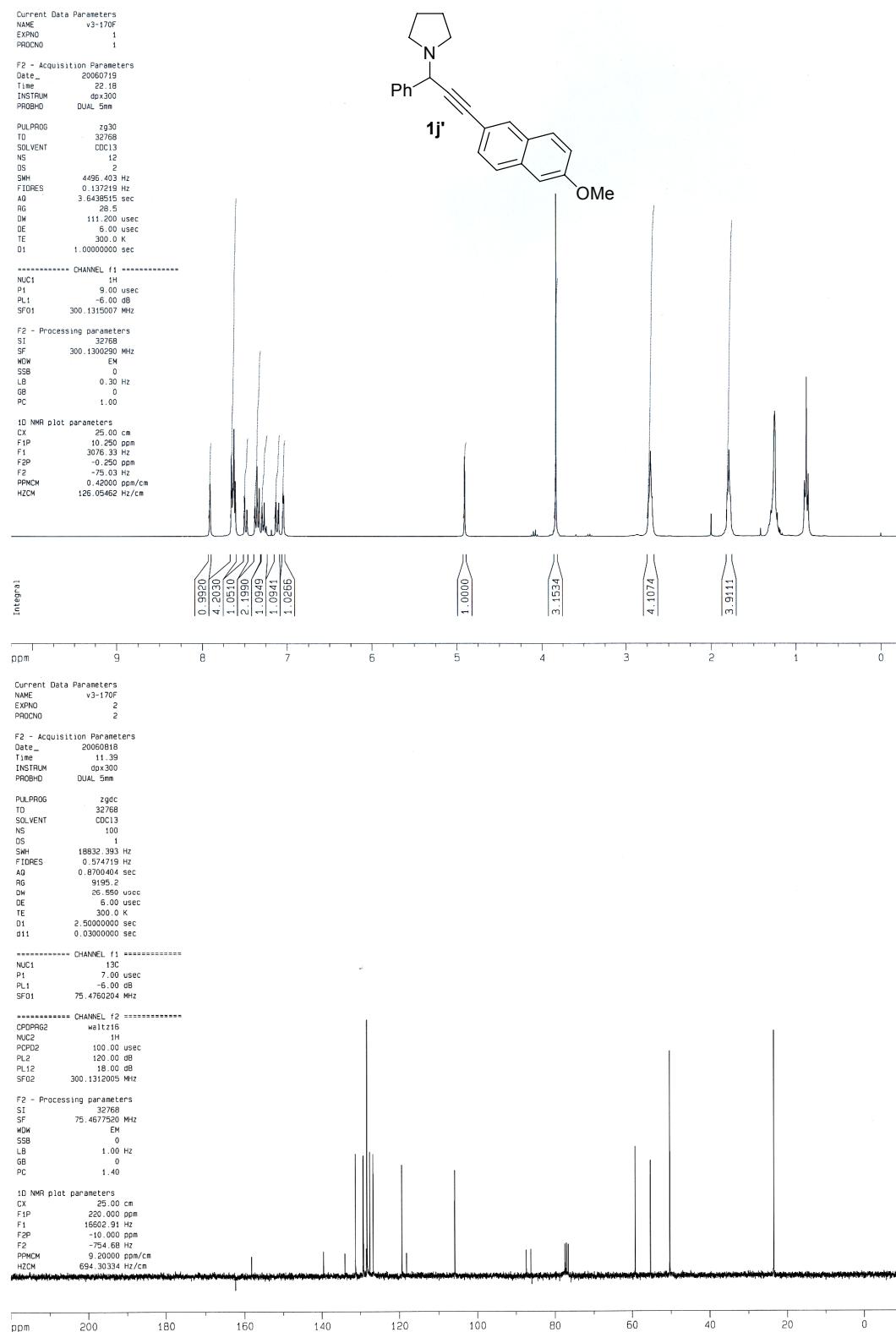
***** CHANNEL f2 *****
CPDPRG2 waltz16
N2C2 1H
PCPDPQ 100.00 usec
PL2 120.00 dB
PL12 18.00 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.467755 MHz
WM EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 25.00 cm
F1P 220.000 ppm
F1 16602.91 Hz
F2P -10.000 ppm
F2 -754.68 Hz
PPMCM 9.20000 ppm/cm
HZCM 694.30334 Hz/cm

ppm 200 180 160 140 120 100 80 60 40 20 0





Current Data Parameters
NAME v6.53F
EXPNO 1
PROCNO 2

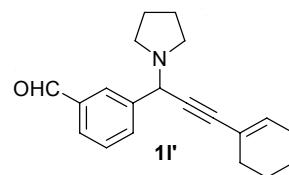
F2 - Acquisition Parameters
Date 20090525
Time 17:04
INSTRUM spect
DUAL 5mm
PROBHD

PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 12
DS 5
SWH 4496.403 Hz
ETDRES 0.137210 Hz
AQ 3.643615 sec
RG 287.4
DW 111.200 usec
DE 6.00 usec
TE 300.0 K
DI 1.0000000 sec

***** CHANNEL F1 *****
NUC1 1H
PI 9.00 usec
PL1 -6.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters
SI 32768
SF 300.13000500 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 25.00 cm
F1P 11.000 ppm
F1 9301.43 Hz
F2P -0.250 ppm
F2 -75.03 Hz
PPMCM 0 45000 ppm/cm
HCDM 135.05850 Hz/cm



Integral
1.0001
1.0288
2.1384
1.1361
1.0062
1.0000
4.4683
4.4254
4.4501
4.4583
2.4285
4.8252



Current Data Parameters
NAME v7.157F
EXPNO 2
PROCNO 1
F2 - Acquisition Parameters
Date 20090706
Time 17:02
INSTRUM av400
PROBHD 5 QNP SH/13
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 12
DS 0
SWH 26178.010 Hz
ETDRES 0.769889 Hz
AQ 0.6299999 sec
RG 26008
DW 19.100 usec
DE 6.00 usec
TE 299.0 K
DI 2.5000000 sec
DT 0.0300000 sec
MCBRT 0.0500000 sec
MCRST 0.0100000 sec

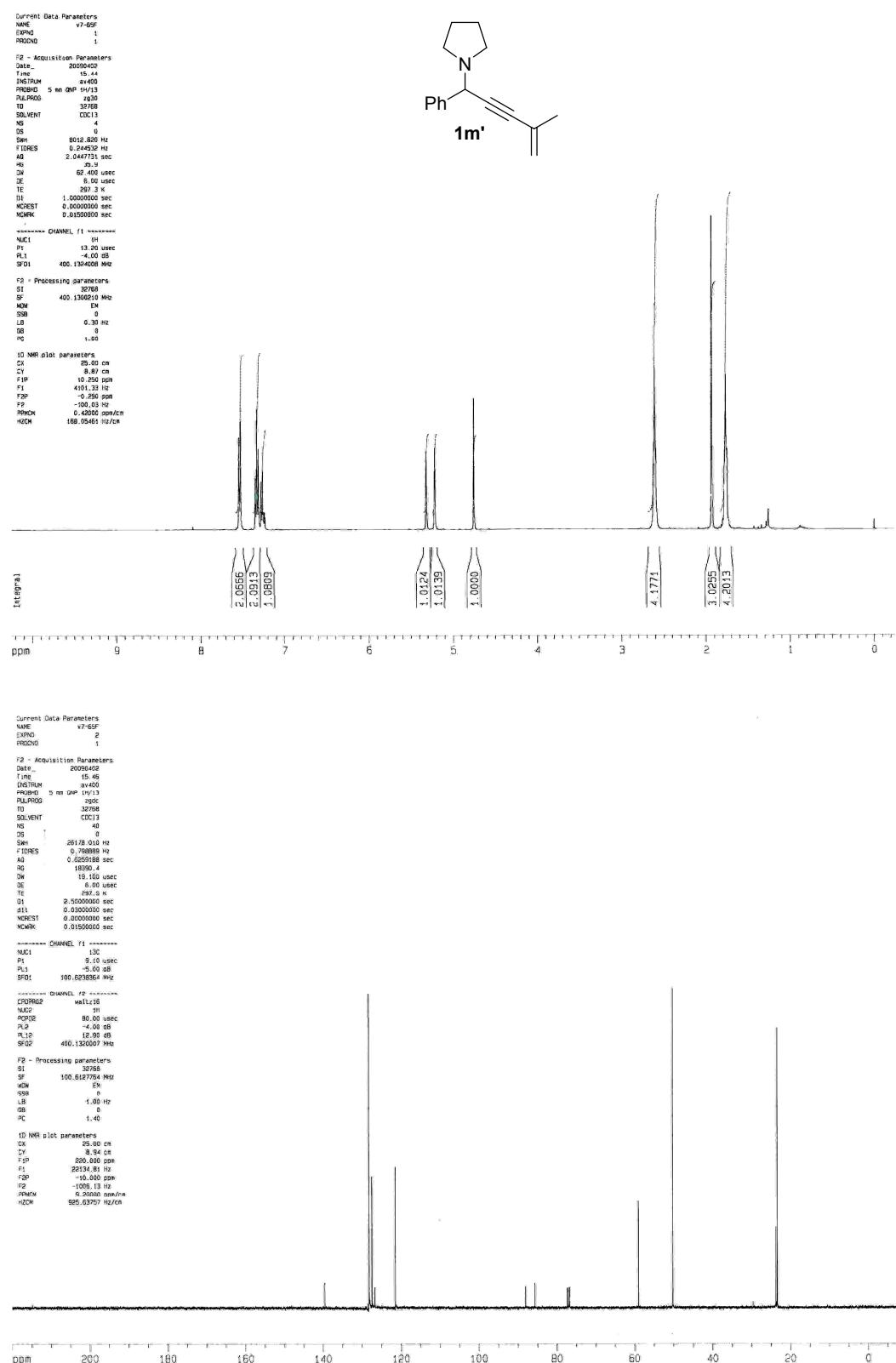
***** CHANNEL F1 *****
NUC1 1H
PI 9.10 usec
PL1 -5.00 dB
SF01 100.6239364 MHz

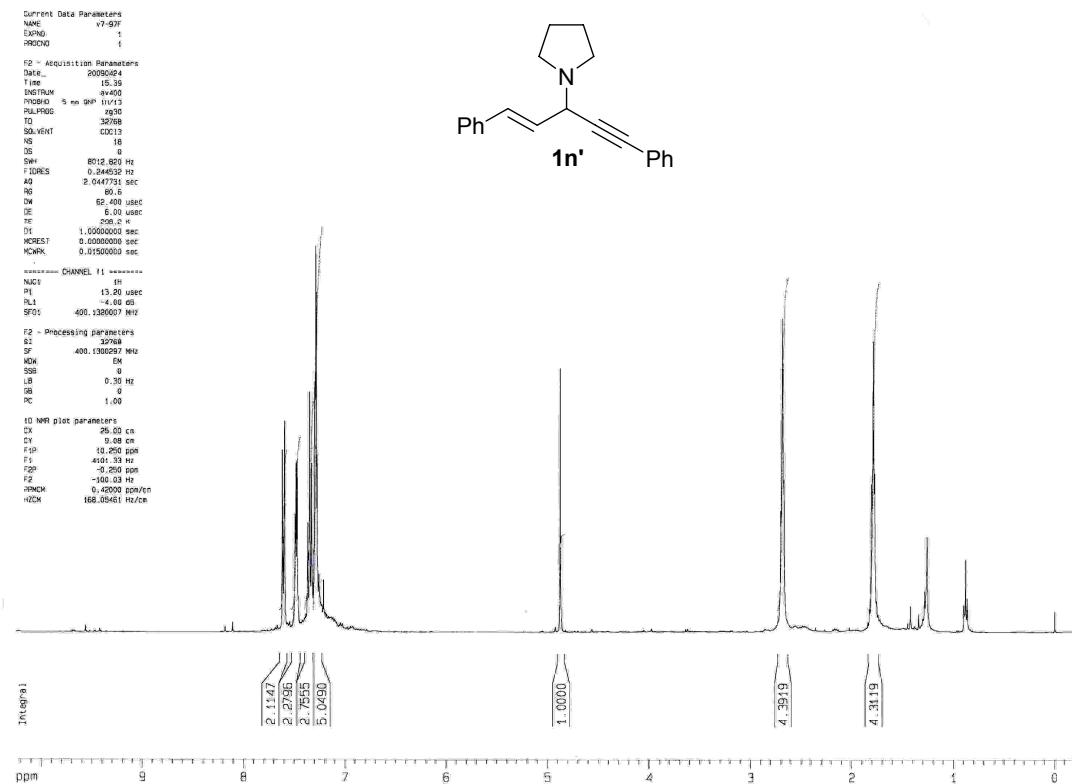
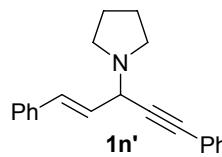
***** CHANNEL F2 *****
CPDPRG2 waltz16
NUC2 13C
PCPDPG 80.00 usec
PL2 -4.00 dB
PL12 -12.80 dB
SF2 400.1300007 MHz

F2 - Processing parameters
SI 32768
SF 100.6127780 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 25.00 cm
CY 9.62 cm
F1P 220.00 ppm
F1 22134.81 Hz
F2P -10.000 ppm
F2 -10.000 ppm
PPMCM 9.20000 ppm/cm
HCDM 925.63379 Hz/cm







Current Data Parameters
NAME v7-97F
EXPNO 2
PROCNID 1

P2 - Acquisition Parameters

INSTRUM av400
 PROBHD 5 mm QNP 1H/13C
 PULPROG zgac
 TD 32768
 SOLVENT CDCl₃

NS 36
 OS 0
 SMH 26178.010 Hz
 FIDGES -0.798889 Hz
 AG 0.676169 Hz

AC	0.6259188 sec
RG	20542.5
SM	19.100 usec
SE	5.00 usec
TC	290.3 K

DI	2.50000000 sec
DII	0.03000000 sec
MCREST	0.00000000 sec
MCHRK	0.01500000 sec

PARAMETER CHANNEL S1

NUC1	130
P _S	9.10 usec
P _{L,S}	-5.00 dB

SF01 100.6238364 MHz
----- CHANNEL 63 -----
CPDPRG2 m11216
SMC2 JH

PCPD2	80.00 usec
PL2	-4.00 dB
PL12	12.90 dB
SF02	400.1320007 MHz

F2 - Processing parameters
SI 32768
SF 100.6127780 kHz
NDM EN

SSR	0
LB	1.00 %
GB	0
PC	1.40

1D NMR plot parameters
 CX 25.00 cm
 CY 10.51 cm
 F1P 220,000 ppm

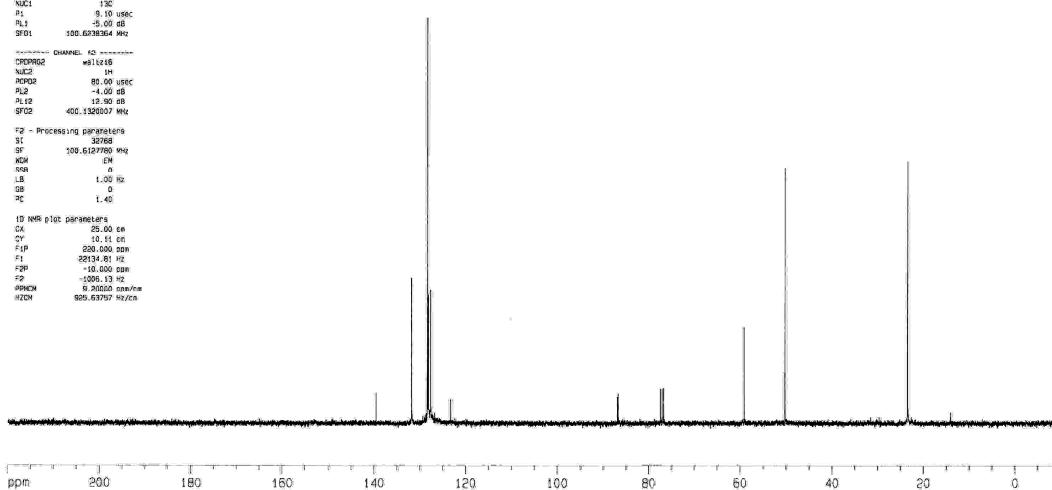
F1	22134.81 Hz
F2P	-10.000 ppm
F2	-1005.13 Hz
SPION	9.20000 ppm
H2CN	924.63252 Hz (C)

WZUN

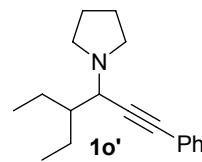
— 1 —

— 4 —

ppm 200



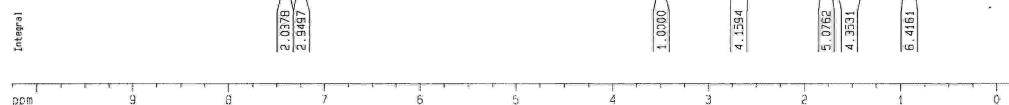
Current Data Parameters:
NAME: v3-185F
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date: 20051027
Time: 18.02
INSTRUM: dpx300
PROBHD: DUAL 5mm
PULPROG: zg30
TD: 32768
SOLVENT: CDCl3
NS: 12
DS: 5
SWH: 4496.403 Hz
FIDRES: 0.137219 Hz
AQ: 3.6436515 sec
RG: 18
DW: 111.200 usec
DE: 6.00 usec
TE: 300.0 K
D1: 1.0000000 sec



===== CHANNEL F1 =====
NUC1: 1H
P1: 9.00 usec
PL1: -6.00 dB
SF01: 300.1315007 MHz

F2 - Processing parameters
SI: 32768
SF: 300.1300221 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

1D NMR plot parameters
CX: 25.00 cm
F1P: 10.250 ppm
F1: 20.32 Hz
F2P: -0.250 ppm
F2: -75.03 Hz
PPMCH: 0.42000 ppm/cm
HDM: 126.05462 Hz/cm



Current Data Parameters:
NAME: v3-185F
EXPNO: 2
PROCNO: 1
F2 - Acquisition Parameters

Date: 20051027
Time: 18.04
INSTRUM: dpx300
PROBHD: DUAL 5mm

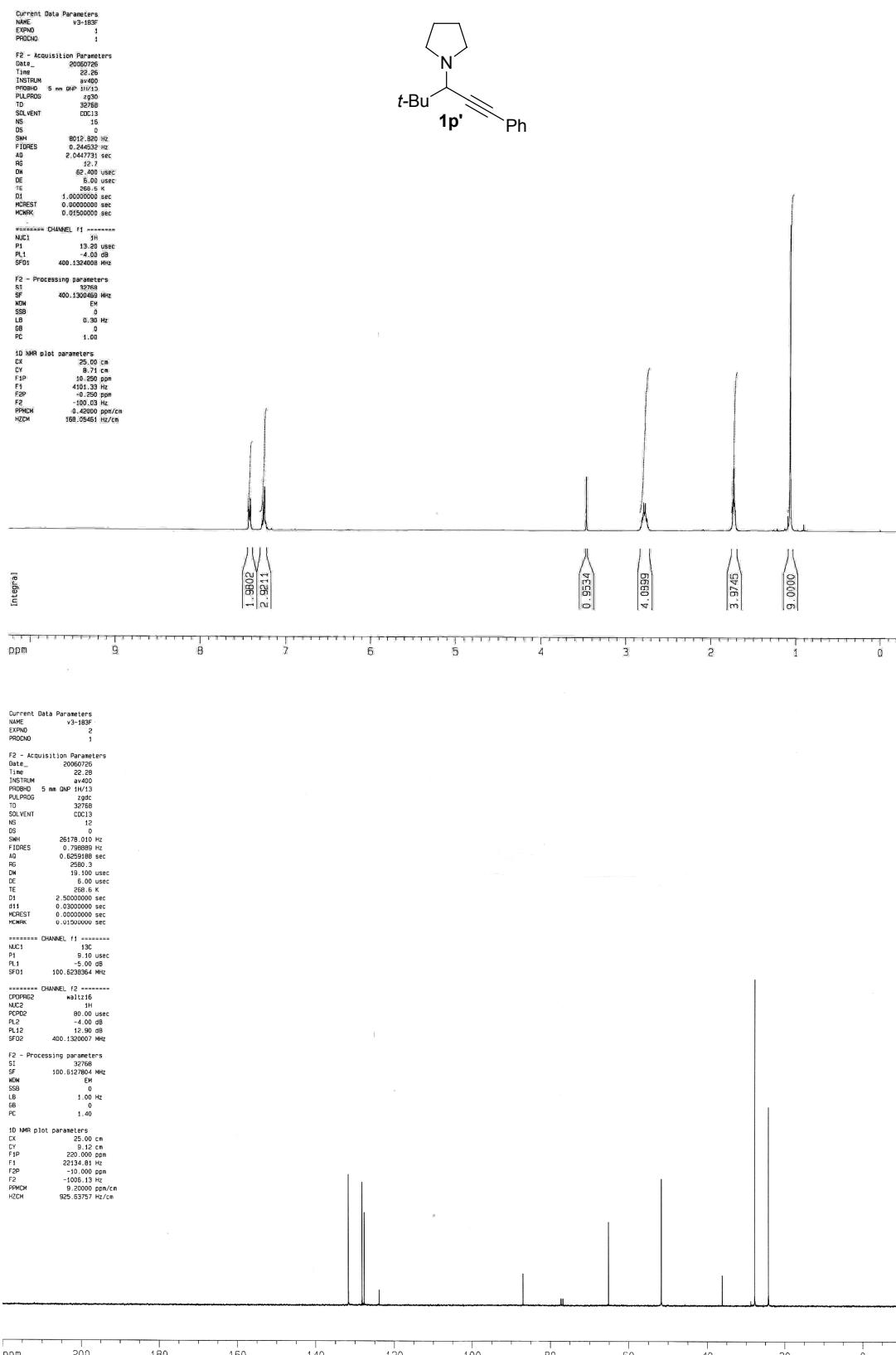
PULPROG: zgdc
TD: 32768
SOLVENT: CDCl3
NS: 50
DS: 1
SWH: 10892.302 Hz
FIDRES: 0.574719 Hz
AQ: 0.8700404 sec
RG: 4997.6
DW: 26.666 usec
DE: 6.00 usec
TE: 300.0 K
D1: 2.5000000 sec
d11: 0.03000000 sec

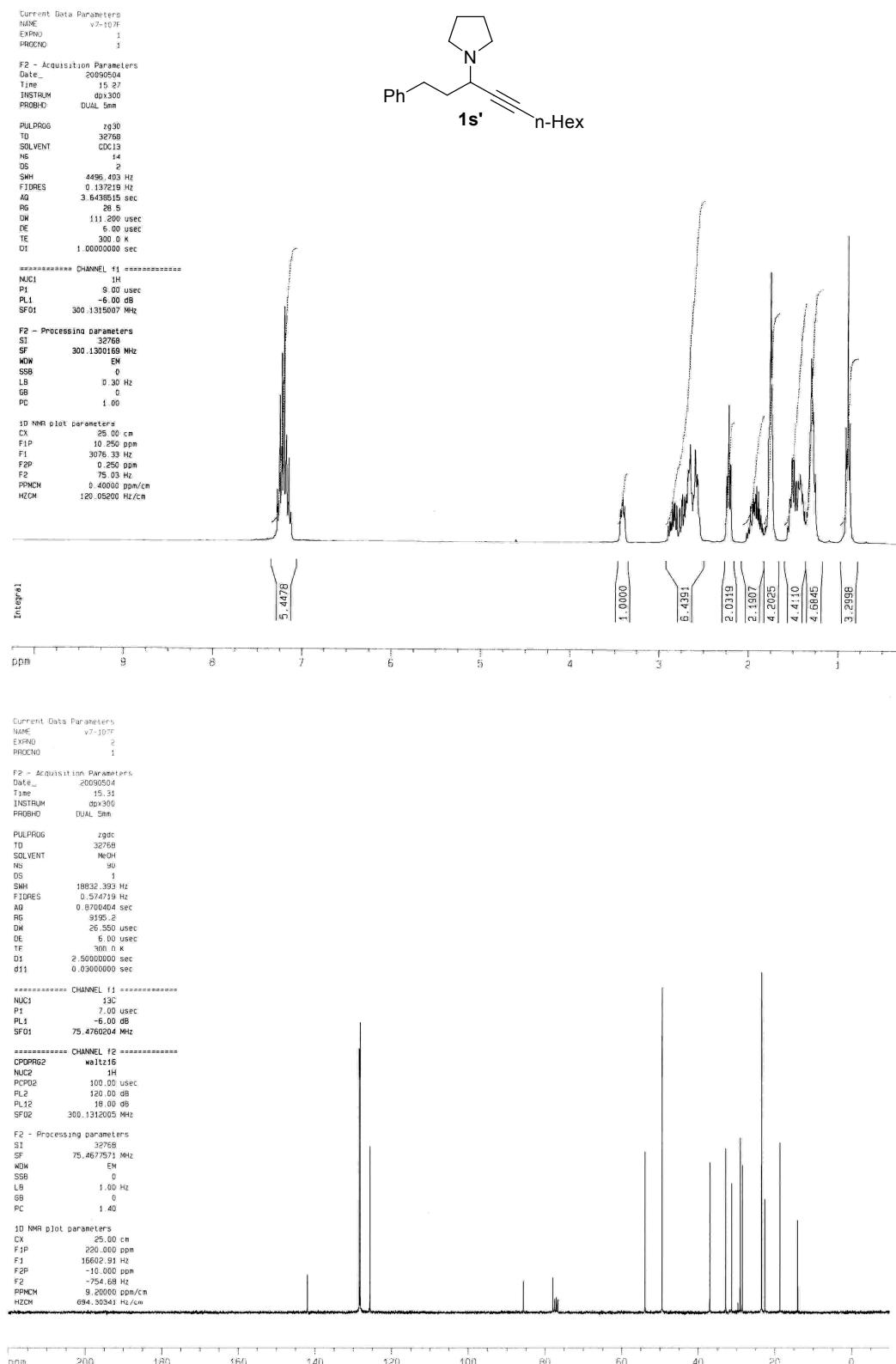
===== CHANNEL 11 =====
NUC1: 13C
P1: 7.00 usec
PL1: -6.00 dB
SF01: 75.4766294 MHz

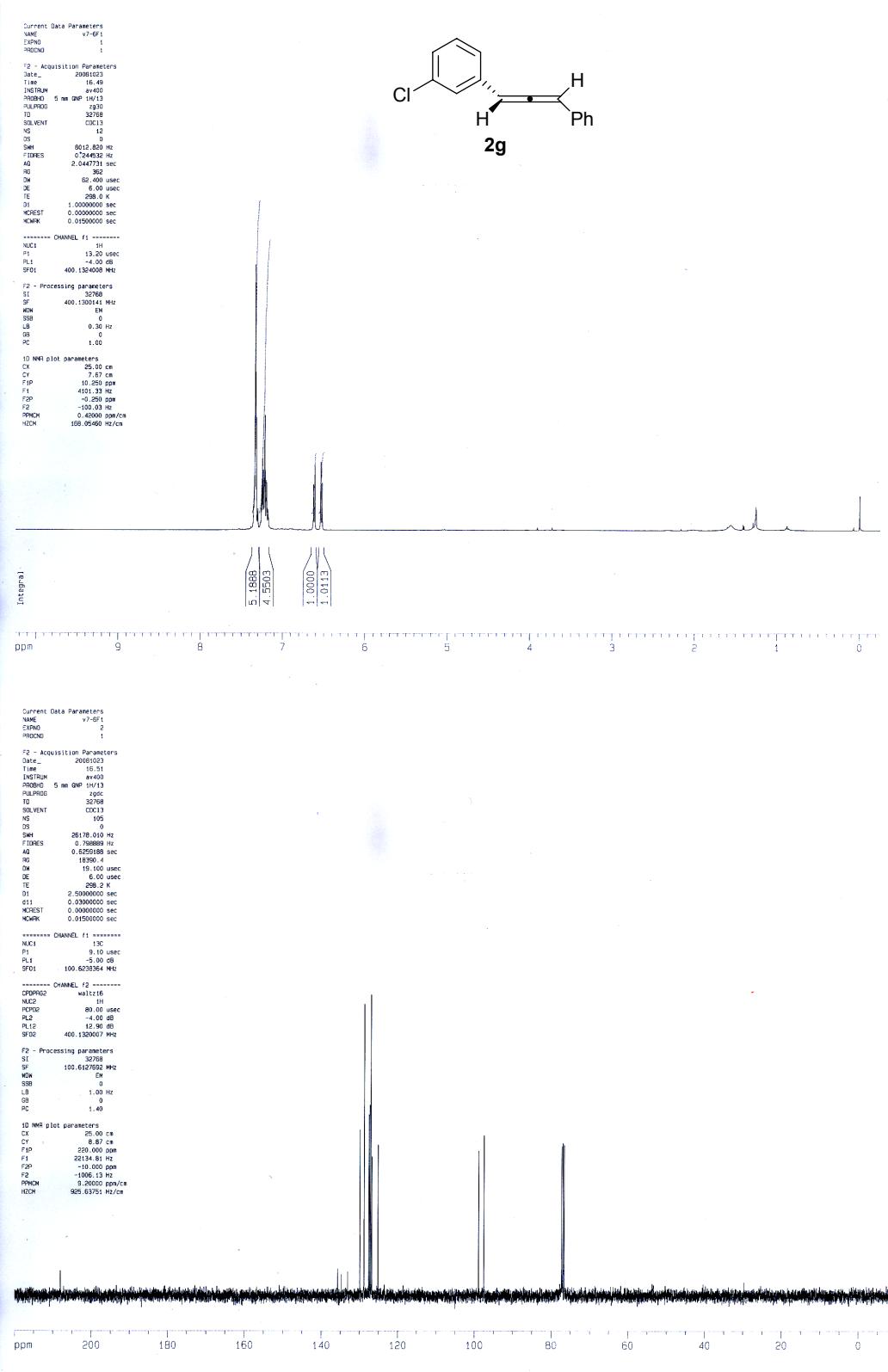
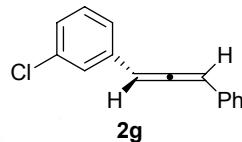
===== CHANNEL 12 =====
CPDPRG2: waltz16
NUC2: 1H
PCP02: 100.00 usec
PL2: 120.00 dB
PL12: 18.00 dB
SF02: 300.1315005 MHz

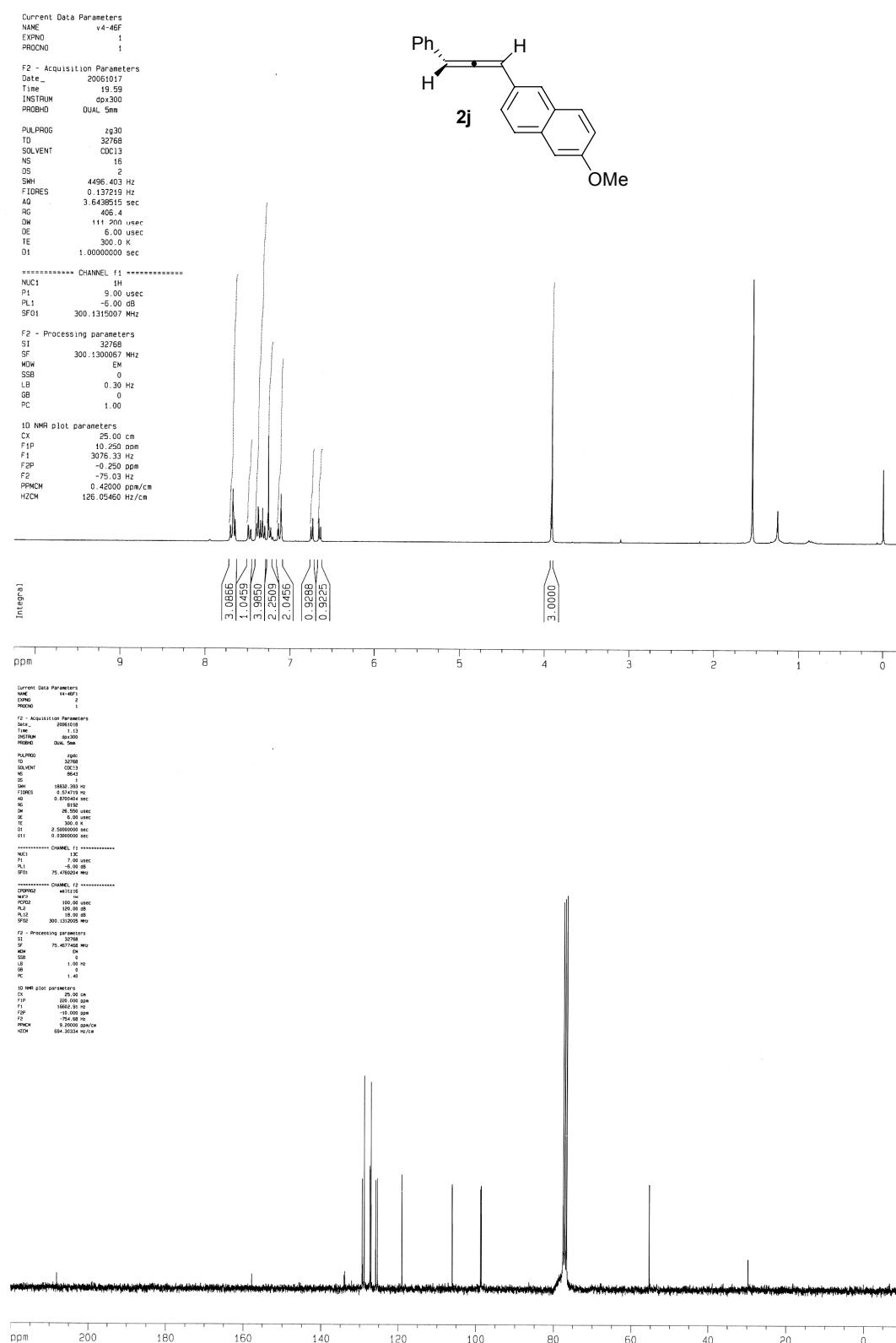
F2 - Processing parameters
SI: 32768
SF: 75.4677577 MHz
WDW: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 1.40
1D NMR plot parameters
CX: 25.00 cm
F1P: 220.000 ppm
F1: 16602.91 Hz
F2P: -10.000 ppm
F2: -754.68 Hz
PPMCH: 9.20000 ppm/cm
HDM: 694.30341 Hz/cm









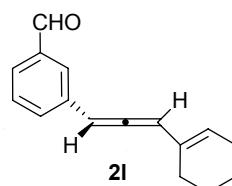


```
Current Data Parameters
NAME: v6-70F
EXPNO: 1
PROCNO: 2
F2 - Acquisition Parameters
Date: 20090317
Time: 19:46
INSTRUM: av400
PROBHD: 5 mm QNP 1H/13
RHOFOID: 300
SL: 32768
TD: 32768
SOLVENT: CDCl3
NS: 30
DS: 0
SWH: 8012.820 Hz
XWPPD: 0.04840 Hz
AQ: 2.0447731 sec
RG: 1448.2
DW: 60.0 usec
DE: 6.00 usec
TE: 297.0 K
D1: 1.0000000 sec
M1: 0.0000000 sec
NCPLST: 0.0000000 sec
NCPLST: 0.01500000 sec

***** CHANNEL F1 *****
NUC1: 1H
P1: 13.20 usec
P1L: -4.00 dB
SF01: 400.1324000 MHz

F2 - Processing parameters
SI: 32768
SF: 400.1300088 MHz
MW: EH
SSB: 0
LB: 0.30 Hz
BB: 0
PC: 1.00

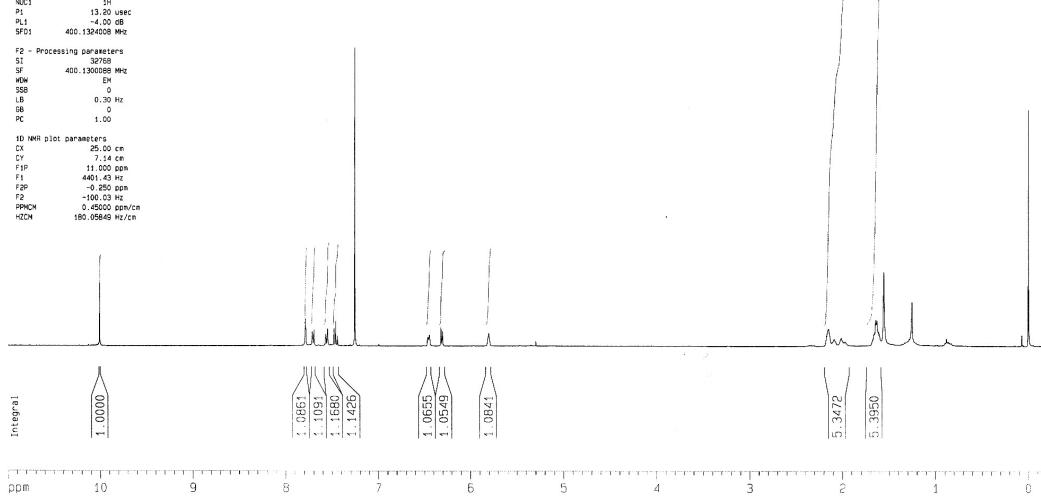
ID NMR plot parameters
CX: 25.00 cm
CY: 7.14 cm
F1P: 11.000 ppm
F1: 400.143 Hz
F2P: -200.00 ppm
F2: -100.03 Hz
PR1: 0.45000 ppm/cm
PR2: 180.05649 Hz/cm
```



```
***** CHANNEL F1 *****
NUC1: 1H
P1: 13.20 usec
P1L: -4.00 dB
SF01: 400.1324000 MHz

F2 - Processing parameters
SI: 32768
SF: 400.1300088 MHz
MW: EH
SSB: 0
LB: 0.30 Hz
BB: 0
PC: 1.00

ID NMR plot parameters
CX: 25.00 cm
CY: 7.14 cm
F1P: 11.000 ppm
F1: 400.143 Hz
F2P: -200.00 ppm
F2: -100.03 Hz
PR1: 0.45000 ppm/cm
PR2: 180.05649 Hz/cm
```

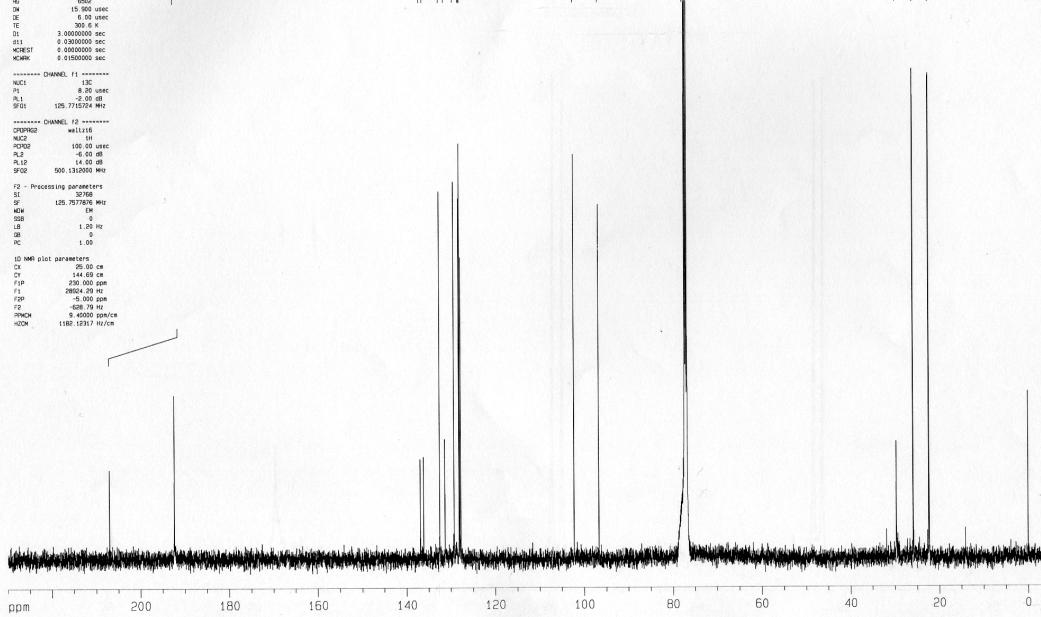


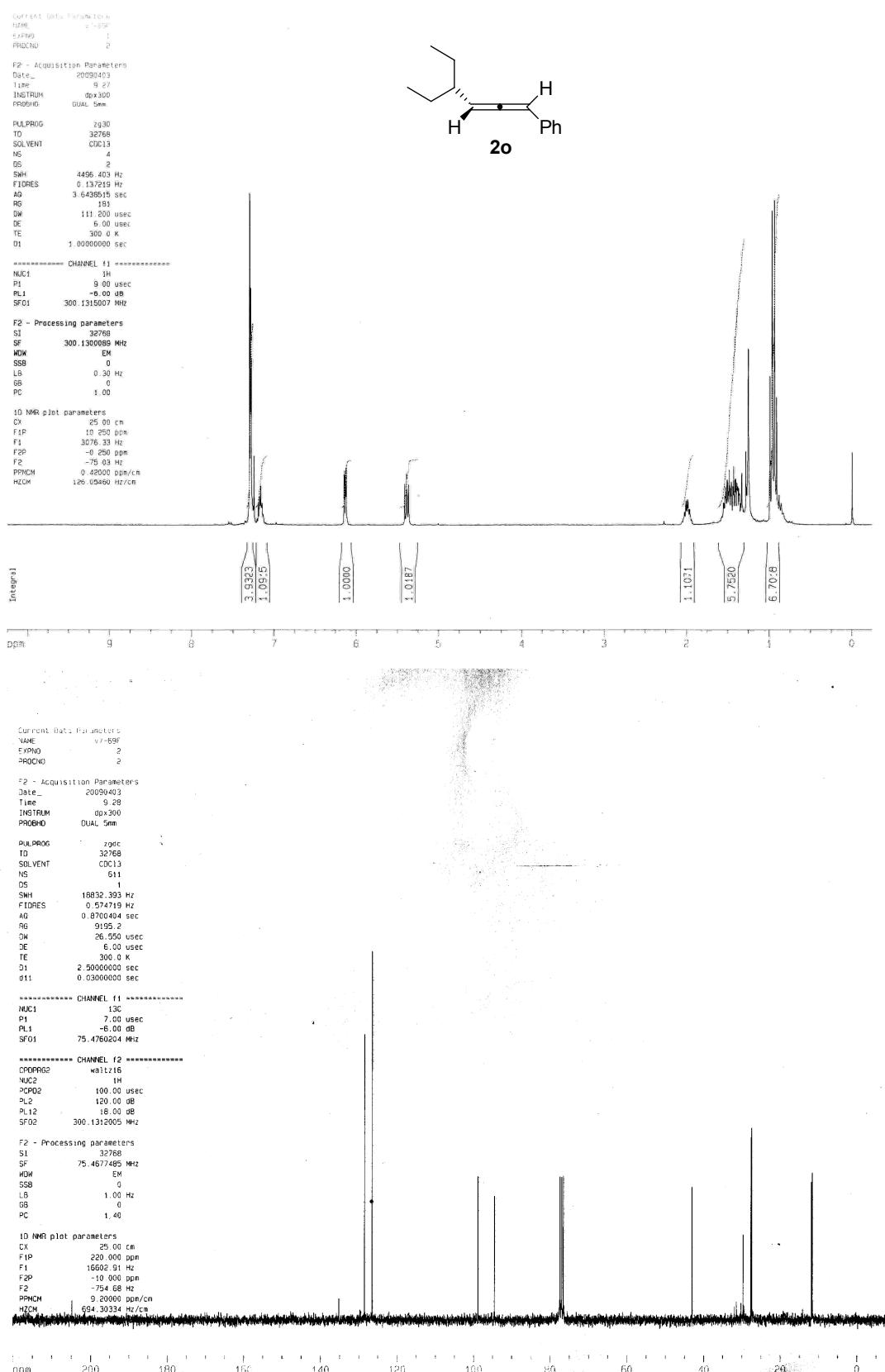
```
Current Data Parameters
NAME: v6-70C
EXPNO: 3
PROCNO: 1
F2 - Acquisition Parameters
Date: 20090317
Time: 16:41
INSTRUM: av400
PROBHD: 8mm QNP 1H/13
RHOFOID: 300
SL: 32768
TD: 32768
SOLVENT: CDCl3
NS: 30
DS: 0
SWH: 8016.140 Hz
XWPPD: 0.04892 Hz
AQ: 6.010612 sec
RG: 15.600 usec
DW: 15.600 usec
DE: 30.5 usec
TE: 305.5 K
D1: 1.0000000 sec
M1: 0.0000000 sec
NCPLST: 0.0000000 sec
NCPLST: 0.01500000 sec

***** CHANNEL F1 *****
NUC1: 13C
P1: 8.20 usec
P1L: -2.00 dB
SF01: 125.771524 MHz

F2 - Processing parameters
SI: 32768
SF: 125.7677875 MHz
MW: EH
SSB: 0
LB: 1.00 Hz
BB: 0
PC: 1.00

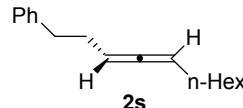
ID NMR plot parameters
CX: 25.00 cm
CY: 144.69 cm
F1P: 2804.29 Hz
F2P: -5.000 ppm
PR1: 0.20000 ppm/cm
PR2: 640000 ppm/cm
PR3: 180.12317 Hz/cm
```





Current Data Parameters
NAME v7-10P
EXPNO 1
PROCNO 1

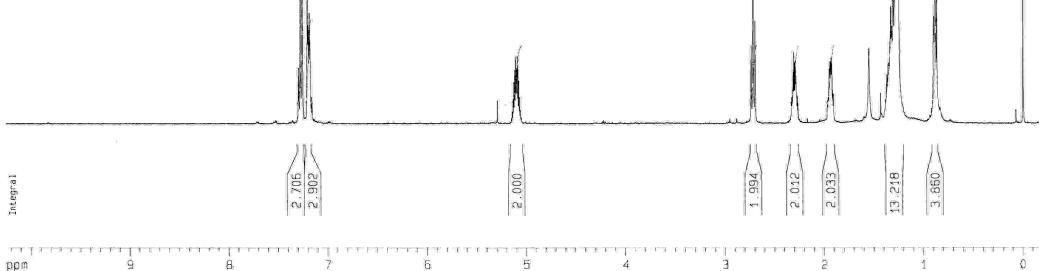
F2 - Acquisition parameters
DATE_ 20090508
TIME 10:23
INSTRUM av400
PROBHD 5 mm GRF TBI/13
PULPROG 3D90
TD 32768
SOLVENT CDCl3
NS 12
DS 0
SWH 8812.820 Hz
FIDRES 0.244532 Hz
AQ 2.044764 sec
RG 128
DW 62.400 usec
DE 6.000 usec
TE 293.6 K
D1 1.0000000 sec
RD1 0.0000000 sec
NCYC 1
DW1 0.0190000 sec



***** CHANNEL f1 *****
NUC1 1H
P1 13.20 usec
PL1 -4.00 dB
SF01 400.130007 MHz

F2 - Processing parameters
SI 32768
SF 400.130007 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 25.00 cm
CP 1.00 cm
TIP 10.250 deg
F1 401.33 Hz
F2 20.000000 Hz
SP 2.000000
TD 40960 points
DWCH 0.40000 ppm/sec
ZFCR 168.05460 Hz/cm



Current Data Parameters
NAME v7-10P
EXPNO 2
PROCNO 1

F2 - Acquisition parameters
DATE_ 20090702
TIME 21:11
INSTRUM dpx300
PROBHD DUAL 5mm
PULPROG 2DP90C
TD 32768
SOLVENT CDCl3
NS 707
DS 0
SWH 16812.363 Hz
FIDRES 0.574710 Hz
AQ 0.8705404 sec
RG 5195.1
DW 26.550 usec
DE 6.00 usec
TE 300.0 K
D1 2.5000000 sec
D11 0.0300000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 7.00 usec
PL1 -6.00 dB
SF01 75.4760204 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
CPDPRG2 100.00 usec
DQ2 120.00 dB
PL12 18.00 dB
SF02 300.1312003 MHz

F2 - Processing parameters
SI 32768
SF 75.4677479 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 25.00 cm
TIP 220.000 ppm
F1 15602.91 Hz
F2P -10.000 ppm
F2 -75.68 Hz
SPNCH 9.20000 ppm/cm
ZFCR 694.30334 Hz/cm

