## **Supplementary Information**

# Regio-, chemo- and stereoselective deuterium labeling method of sugars based on ruthenium-catalyzed C-H bond activation

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## General

All the catalysts used in this study were obtained from the N.E. Chemcat Corporation.  $D_2O$  (>99.9% D atom) was purchased from Spectra Gases, Inc. All other reagents were purchased from commercial sources and were used without purification. The <sup>1</sup>H and <sup>2</sup>H spectra were recorded by a JEOL AL-400, EX-400 (<sup>1</sup>H: 400 MHz, <sup>2</sup>H: 61 MHz) or ECA-500 spectrometer (<sup>1</sup>H: 500 MHz, <sup>2</sup>H: 61 MHz). Chemical shifts ( $\delta$ ) are expressed in ppm and are internally referenced to trimethylsilane or residual solvents (<sup>1</sup>H NMR: 0.00 ppm for TMS for CDCl<sub>3</sub>; <sup>2</sup>H NMR: 7.26 ppm for CHCl<sub>3</sub>). The FAB mass spectra were taken by a JEOL JMS-SX102A instrument at the Mass Spectrometry Laboratory of the Gifu Pharmaceutical University. Optical rotations were taken by a DIP-360 instrument. The heating reactions were carried out using a personal organic synthesizer, Chemist Plaza (Shibata Science Technology, Ltd.).

## Typical Procedure for the Ru/C-Catalyzed H-D Exchange of Sugars (Table 2).

A suspension of the sugar (Entries 1-8: 0.5 mmol; Entry 9: 0.25 mmol) and 10% Ru/C (Entries 1–7: 5 mol% of the substrate; Entries 8 and 9: 10 mol% of the sugar) in 2 cm<sup>3</sup> of D<sub>2</sub>O was stirred at 80 °C in a test tube under a hydrogen atmosphere (balloon). After the appropriate time, the mixture was cooled to room temperature and filtered using a membrane filter (Millipore, Millex<sup>®</sup>-LH, 0.45 µm) to remove the catalyst. The filtrate was then concentrated in vacuo. To the residue were added 2.5 mmol of acetic anhydride and 5.0 mmol of pyridine, and the mixture was stirred at room temperature for 24 h and concentrated in vacuo to give the deuterated *O*-acetylated glycoside.

#### **Evaluation of Deuterium Content**

The deuterium content was determined by the <sup>1</sup>H NMR on the basis of the integration of the methyl protons of the methoxy group. For Entry 9, the integration of the proton at the 1 (anomeric) positions was employed instead of the methyl protons. The deuterium incorporation was also assigned by the <sup>2</sup>H NMR and FAB Mass spectra.

## **Compound Data**

Methyl 2,3,4,6-tetraacetyl-α-D-glucopyranoside-d<sub>5</sub> (Table 2, Entry 1)



 $[\alpha]_{D}^{20} = 107.4 (c = 0.1 \text{ in CHCl}_3); {}^{1}\text{H NMR (CDCl}_3): \delta 4.95 (s, 1\text{H}), 3.98 (s, 1\text{H}), 3.42 (s, 3\text{H}), 2.10 (s, 3\text{H}), 2.08 (s, 3\text{H}), 2.03 (s, 3\text{H}), 2.01 (s, 3\text{H}); {}^{2}\text{H NMR (CHCl}_3): \delta 5.36 (br s), 4.95 (br s), 4.8 (br s), 4.14 (br s), 3.99 (br s).$ 

Methyl 2,3,4,6-tetraacetyl-β-D-glucopyranoside-d<sub>5</sub> (Table 2, Entry 2)



 $[\alpha]_{D}^{21} = -16.5$  (c = 0.1 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  4.43 (s, 1H), 3.69 (s, 1H), 3.51 (s, 3H), 2.10 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 2.01 (s, 3H); <sup>2</sup>H NMR (CHCl<sub>3</sub>):  $\delta$  5.16 (br s), 5.04 (br s), 4.93 (br s), 4.21 (br s), 4.01 (br s).

**Methyl 2,3,4,6-tetraacetyl-α-D-mannopyranoside-***d*<sub>5</sub> (Table 2, Entry 3)



 $[\alpha]_{D}^{22} = 44.4 (c = 0.1 \text{ in CHCl}_{3}); {}^{1}\text{H NMR (CDCl}_{3}): \delta 5.29-5.27 (m, 0.17\text{H}), 4.71 (s, 1\text{H}), 3.96 (s, 1\text{H}), 3.41 (s, 3\text{H}), 2.16 (s, 3\text{H}), 2.11 (s, 3\text{H}), 2.04 (s, 3\text{H}), 1.99 (s, 3\text{H}); {}^{2}\text{H NMR (CHCl}_{3}): \delta 5.16 (\text{br s}), 4.16 (\text{br s}), 4.01 (\text{br s}).$ 

Methyl 2,3,4-triacetyl-β-D-xylopyranoside-d<sub>3</sub> (Table 2, Entry 4)



 $[\alpha]_{D}^{20} = -65.2$  (c = 0.1 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  4.40 (s, 1H), 4.14–4.11 (d, J = 11.7 Hz, 1H), 3.47 (s, 3H), 3.39–3.36 (d, J = 11.7 Hz, 1H), 2.06 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H); <sup>2</sup>H NMR (CHCl<sub>3</sub>):  $\delta$  5.06 (br s), 4.82 (br s).

Methyl 2,3,4-triacetyl-α-L-fucopyranoside-d<sub>3</sub> (Table 2, Entry 5)



 $[\alpha]_{D}^{21} = -127.0$  (c = 0.1 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  4.94 (s, 1H), 4.15–4.10 (q, J = 6.59 Hz, 1H), 3.40 (s, 3H), 2.17 (s, 3H), 2.09 (s, 3H), 1.98 (s, 3H), 1.16–1.15 (d, J = 6.59 Hz, 3H); <sup>2</sup>H NMR (CHCl<sub>3</sub>):  $\delta$  5.22 (br s), 5.04 (br s).

1-Deoxy-2,3,4,6-tetraacetyl-D-glucopyranoside-d<sub>5</sub> (Table 2, Entry 6)



 $[\alpha]_{D}^{21} = 29.6 (c = 0.1 \text{ in CHCl}_{3}); {}^{1}\text{H NMR (CDCl}_{3}): \delta 4.40 (s, 1\text{H}), 4.19-4.12 (m, 1.25\text{H}), 3.59 (s, 1\text{H}), 3.32-3.30 (d,$ *J* $= 8.7 Hz, 1\text{H}), 2.10 (s, 3\text{H}), 2.04-2.03 (s, 9\text{H}); {}^{2}\text{H NMR (CHCl}_{3}): 5.19 (br s), 5.01 (br s), 4.18 (br s), 4.11 (br s).$ 

Methyl 2,3,4,6-tetraacetyl-β-D-galactopyranoside-d<sub>5</sub> (Table 2, Entry 8)



 $[\alpha]_D^{21} = 103.5$  (c = 0.1 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  5.19 (m, 0.12H), 4.40 (s, 1H), 3.90 (s, 1H), 3.52 (s, 3H), 2.15 (s, 3H), 2.07 (s, 3H), 2.05 (s, 3H), 1.98 (s, 3H); <sup>2</sup>H NMR (CHCl<sub>3</sub>):  $\delta$  5.30 (br s), 5.10 (br s), 4.95 (br s), 4.08 (br s).

2,2',3,3',4,4',6,6'-octaacetyl-treharose-d<sub>10</sub> (Table 2, Entry 9)



 $[\alpha]_{D}^{20} = 148.0 \text{ (c} = 0.1 \text{ in CHCl}_3); {}^{1}\text{H NMR (CDCl}_3): \delta 5.28 \text{ (s}, 1\text{H}), 4.04 \text{ (s}, 1\text{H}), 2.09 \text{ (s}, 3\text{H}), 2.08 \text{ (s}, 3\text{H}), 2.06 \text{ (s}, 3\text{H}), 2.04 \text{ (s}, 3\text{H}); {}^{2}\text{H NMR (CHCl}_3): \delta 5.44 \text{ (br s)}, 4.99 \text{ (br s)}, 4.16 \text{ (br s)}, 3.94 \text{ (br s)}.$ 





#### MASS

File: FAB09B882 Instrument: SX102A Inlet: Direct

Date Run: 10-2-2009 (Time Run: 15:32:51) Ionization mode: FAB+





Supplementary Material (ESI) for Chemical Communications



#### MASS



Date Run: 11-4-2009 (Time Run: 13:24:56) Ionization mode: FAB+



#Ions: 260

600





File: FAB09C200 Instrument: SX102A Inlet: Direct Date Run: 11-4-2009 (Time Run: 14:00:21) Ionization mode: FAB+

#Ions: 167









File: FAB09C205 Instrument: SX102A Inlet: Direct

Date Run: 11-4-2009 (Time Run: 14:23:43) Ionization mode: FAB+



#Ions: 393



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File: FAB09C208 Instrument: SX102A Inlet: Direct Date Run: 11-4-2009 (Time Run: 14:51:22) Ionization mode: FAB+

#Ions: 111

Scan: 3 R.T.: .3 Base: m/z 185; 86%FS TIC: 42671649 HI-1-158/Gly HO











File: FAB09C635 Instrument: SX102A Inlet: Direct

Date Run: 12-7-2009 (Time Run: 18:11:38) Ionization mode: FAB+





