

SUPPORTING INFORMATION
for

Chiral Cobalt-Catalyzed Enantioselective Aerobic Oxidation of α -Hydroxy Esters

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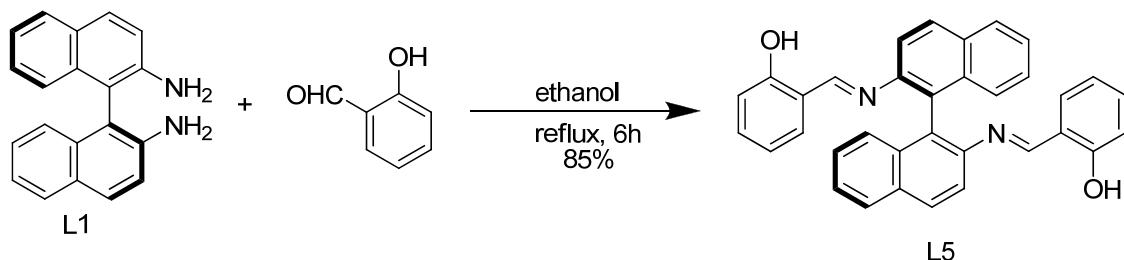
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General considerations

Cobalt(II) acetate tetrahydrate and 2,2,6,6-tetramethyl-piperidin-1-oxyl (TEMPO) were purchased from Sigma-Aldrich chemical company and used as received. (S)-BINAM purchased from Gerchem Labs Pvt. Ltd. Hyderabad, India. Thin-layer chromatography (TLC) was performed using Merck silica gel 60 F₂₅₄ precoated plates (0.25 mm) and visualized by UV fluorescence quenching. Silica gel for column chromatography (particle size 100-200 mesh) purchased from SRL India. Optical rotations were measured with Autopol IV - Rudolph Research Analytical Polarimeter. ¹H and ¹³C NMR spectra were recorded on a Bruker 400 MHz instrument. ¹H NMR spectra were reported relative to Me₄Si (δ 0.0 ppm) or residual CHCl₃ (δ 7.26 ppm). ¹³C NMR were reported relative to CDCl₃ (δ 77.16 ppm). FTIR spectra were recorded on a Nicolet 6700 spectrometer and are reported in frequency of absorption (cm⁻¹). High resolution mass spectra (HRMS) were recorded on Q-Tof Micro mass spectrometer. The enantiomeric excess (%ee) of α -hydroxy esters were determined by JASCO uv-2070 plus HPLC and Shimadzu UFLC systems using Daicel chemical industries, Ltd ChiralPAK AS-H (0.46cm ϕ x 25cm) column. ¹H and ¹³C NMR and HRMS Spectral data have been included for all compounds. HPLC spectra for the %ee determination of all optically active α -hydroxy esters are given in this supporting information.

Synthesis of Ligand L5

Scheme 1



Experimental Procedure

Synthesis of L5¹: In a 50 mL two neck round bottom flask equipped with reflux condenser, a mixture of **L1** (502.7 mg, 1.77 mmol) and salicylaldehyde (0.38 mL, 3.54 mmol) in ethanol (20 mL) was taken and refluxed for 6 hours. The reaction mixture was cooled to room temperature, the solution was filtered off. The solvent was removed under reduced pressure and the residue was recrystallized from benzene : hexane (1:3, V/V) to give **L5** (740 mg, 85%) as a yellow compound. Mp 181 °C (lit.¹ 181-183 °C); R_f 0.41 (in hexanes : ethyl acetate, 80:20 V/V); [α]^D₂₅ = 509.2 (c = 0.5 in chloroform); ¹H NMR (400 MHz, CDCl₃): δ 12.12 (s, 2H), 8.69 (s, 2H), 8.13 (d, J = 9.2 Hz, 2H), 8.00 (d, J = 8 Hz, 2H), 7.67 (d, J = 8.8 Hz, 2H), 7.46-7.50 (m, 2H), 7.27-7.33 (m, 4H), 7.19-7.23 (m, 4H), 6.78-6.82 (m, 2H), 6.73 (d, J = 8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 161.9, 160.8, 143.8, 133.2, 132.8, 132.5, 132.2, 130.0, 129.5, 128.3, 127.0, 126.5, 125.9, 119.3, 118.7, 117.1, 116.9; IR (Neat) 3056, 1608, 1282, 750 cm⁻¹; HRMS (m/z): [M+1]⁺calcd for C₃₄H₂₅N₂O₂, 493.1916; found, 493.1917.

General experimental procedure for oxidative kinetic resolution

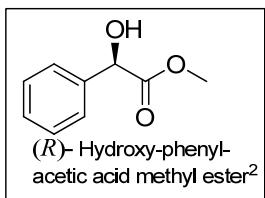
A mixture of **L5** (49.2 mg, 0.1 mmol) and Cobalt(II) acetate tetrahydrate (25 mg, 0.1mmol) in 8 mL of toluene was stirred at room temperature for 10 min, TEMPO (7.82 mg, 0.05 mmol) was then added to the reaction mixture. After stirring for 5 min, (±)-Hydroxy-phenyl-acetic acid methyl ester (166 mg, 1 mmol) was added and then reaction stirred under an O₂ atmosphere (using O₂ balloon) for 11 hours at 90 °C. The reaction mixture was concentrated and the resulting residue was purified by silica gel column chromatography (eluents: hexanes-ethylacetate) to give the Methyl phenylglyoxylate (75.5 mg, yield 46%) and unreacted Hydroxy-phenyl-acetic acid methyl ester (78 mg, yield 47%).

Hydroxy-phenyl-acetic acid methyl ester: Colorless oil; R_f 0.33; (hexanes : ethyl acetate, 80:20 v/v); [α]^D₃₀ = -108.7 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.44-7.41 (m, 2H), 7.40-7.33 (m, 3H), 5.18 (d, J = 5.6 Hz, 1H), 3.77 (s, 3H), 3.14 (d, J = 5.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 174.3, 138.4, 128.8, 128.7, 126.7, 73.0, 53.2; IR

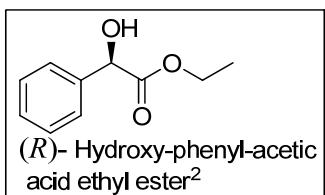
(Neat) 3457, 2954, 1735, 1221, 1071 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₉H₁₀O₃Na₁, 189.0528; found, 189.0526. The enantiomeric excess (%ee) was determined to be 78% by HPLC using ChiralPAK AS-H column (25% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (minor, 5.1 min), t_R (major, 5.6 min).

Methyl phenylglyoxylate: Colorless oil; R_f 0.63; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 8.02 (d, *J* = 7.6 Hz, 2H), 7.67 (t, *J* = 7.2 Hz, 1H), 7.52 (t, *J* = 7.6 Hz, 2H), 3.99 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.1, 164.1, 135.0, 132.4, 130.1, 128.9, 52.8; IR (Neat) 2963, 1741, 1688, 1208, 1003 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₉H₈O₃Na₁, 187.0371; found, 187.0374.

Spectral data for all α-hydroxy esters and α-keto esters

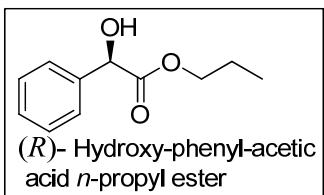


Colorless oil; R_f 0.33; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -108.7 (*c* = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.44-7.41 (m, 2H), 7.40-7.33 (m, 3H), 5.18 (d, *J* = 5.6 Hz, 1H), 3.77 (s, 3H), 3.14 (d, *J* = 5.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 174.3, 138.4, 128.8, 128.7, 126.7, 73.0, 53.2; IR (Neat) 3457, 2954, 1735, 1221, 1071 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₉H₁₀O₃Na₁, 189.0528; found, 189.0526. The enantiomeric excess (%ee) was determined to be 78% by HPLC using ChiralPAK AS-H column (25% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (minor, 5.1 min), t_R (major, 5.6 min).

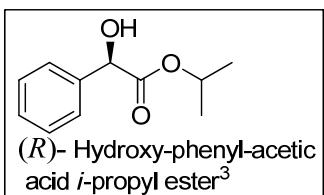


Colorless oil; R_f 0.38; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -127.7 (*c* = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.45 (m, 2H),

7.30-7.39 (m, 3H), 5.16 (d, $J = 5.6$ Hz, 1H), 4.13-4.32 (m, 2H), 3.46 (d, $J = 5.6$ Hz, 1H), 1.23 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.7, 138.5, 128.6, 128.4, 126.6, 73.0, 62.2, 14.0; IR (Neat) 3460, 2982, 1728, 1182, 1065 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{10}\text{H}_{12}\text{O}_3\text{Na}_1$, 203.0684; found, 203.0687. The enantiomeric excess (%ee) was determined to be 97% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_{R} (minor, 8.7 min), t_{R} (major, 9.2 min).

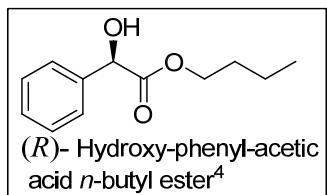


Colorless oil; R_f 0.35; (hexanes : ethyl acetate, 80:20 v/v): $[\alpha]_{30}^D = -67.9$ ($c = 1$ in CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.40-7.45 (m, 2H), 7.29-7.38 (m, 3H), 5.17 (d, $J = 6$ Hz, 1H), 4.06-4.17 (m, 2H), 3.48-3.56 (m, 1H), 1.60 (sextet, $J = 7.2$ Hz, 2H), 0.82 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.9, 138.6, 128.6, 128.4, 126.6, 72.9, 67.7, 21.9, 10.2; IR (Neat) 3453, 2968, 1730, 1180, 1064 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{11}\text{H}_{14}\text{O}_3\text{Na}_1$, 217.0841; found, 217.0843. The enantiomeric excess (%ee) was determined to be 91% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_{R} (minor, 7.4 min), t_{R} (major, 8.5 min).

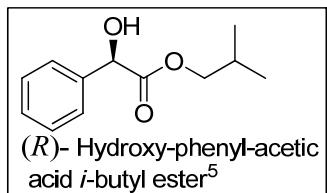


Colorless oil; R_f 0.49; (hexanes : ethyl acetate, 80:20 v/v): $[\alpha]_{30}^D = -173.5$ ($c = 2$ in CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.29-7.34 (m, 2H), 7.15-7.25 (m, 3H), 5.02 (d, $J = 6$ Hz, 1H), 4.94 (septet, $J = 6$ Hz, 1H), 3.77 (d, $J = 4.4$ Hz, 1H), 1.15 (d, $J = 6$ Hz, 3H), 0.98 (d, $J = 6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.3, 138.6, 128.6, 128.4, 126.5, 73.0, 70.2, 21.8, 21.5; IR (Neat) 3458, 2982, 1726, 1181, 1098 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{11}\text{H}_{14}\text{O}_3\text{Na}_1$, 217.0841; found, 217.0845. The enantiomeric excess (%ee) was determined to be 84% by HPLC using ChiralPAK

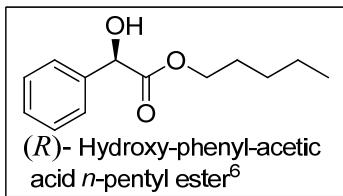
AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (major, 6.7 min), t_R (minor, 7.2 min).



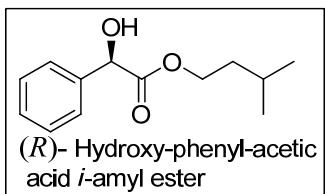
Colorless oil; R_f 0.32; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -62.9 (*c* = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.39-7.44 (m, 2H), 7.28-7.38 (m, 3H), 5.16 (d, *J* = 5.2 Hz, 1H), 4.10-4.22 (m, 2H), 3.53 (d, *J* = 3.6 Hz, 1H), 1.51-1.60 (m, 2H), 1.24 (sextet, *J* = 7.6 Hz, 2H), 0.85 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.8, 138.5, 128.6, 128.4, 126.6, 72.9, 66.0, 30.4, 18.9, 13.6; IR (Neat) 3469, 2959, 1731, 1180, 1066 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₂H₁₆O₃Na₁, 231.0997; found, 231.0996. The enantiomeric excess (%ee) was determined to be 90% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (minor, 6.2 min), t_R (major, 8.3 min).



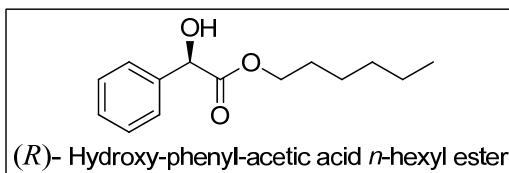
Colorless oil; R_f 0.54; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -118.7 (*c* = 1.3 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.43 (d, *J* = 7.6 Hz, 2H), 7.28-7.39 (m, 3H), 5.18 (d, *J* = 5.2 Hz, 1H), 3.98-4.00 (m, 2H), 3.53 (d, *J* = 5.6 Hz, 1H), 1.87 (septet, *J* = 6.4 Hz, 1H), 0.81 (dd, *J* = 6.8 Hz, 2.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 138.6, 128.6, 128.5, 126.6, 72.9, 72.2, 27.8, 18.9; IR (Neat) 3482, 2962, 1735, 1183, 1099 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₂H₁₆O₃Na₁, 231.0997; found, 231.0995. The enantiomeric excess (%ee) was determined to be 84% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (minor, 5.7 min), t_R (major, 7.2 min).



Colorless oil; R_f 0.42; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -88.2 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.45 (m, 2H), 7.29-7.39 (m, 3H), 5.16 (d, J = 5.6 Hz, 1H), 4.12-4.18 (m, 2H), 3.49 (d, J = 5.6 Hz, 1H), 1.53-1.62 (m, 2H), 1.12-1.28 (m, 4H), 0.83 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 138.6, 128.7, 128.5, 126.7, 73.0, 66.4, 28.2, 27.9, 22.3, 14.0; IR (Neat) 3446, 2957, 2872, 1731, 1183, 1067 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₃H₁₈O₃Na₁, 245.1154; found, 245.1157. The enantiomeric excess (%ee) was determined to be 94% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (minor, 6.5 min), t_R (major, 8.5 min).

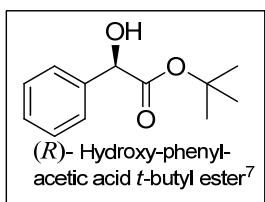


Colorless oil; R_f 0.35; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -80.7 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.39-7.44 (m, 2H), 7.30-7.38 (m, 3H), 5.15 (d, J = 5.6 Hz, 1H), 4.12-4.25 (m, 2H), 3.49 (d, J = 5.6 Hz, 1H), 1.43-1.56 (m, 3H), 0.85 (d, J = 6.4 Hz, 3H), 0.82 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 138.6, 128.7, 128.5, 126.6, 73.0, 65.0, 37.2, 25.0, 22.5, 22.4; IR (Neat) 3445, 2958, 1731, 1183, 1067 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₃H₁₈O₃Na₁, 245.1154; found, 245.1157. The enantiomeric excess (%ee) was determined to be 98% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (minor, 5.9 min), t_R (major, 8.4 min).

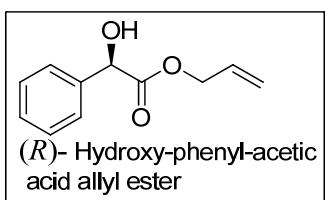


Colorless oil; R_f 0.47; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -70.3 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.42 (d, J =

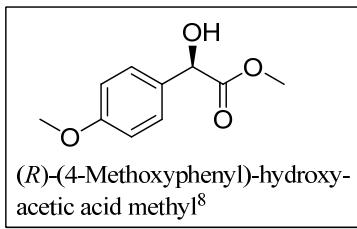
7.2 Hz, 2H), 7.29-7.39 (m, 3H), 5.16 (d, $J = 5.6$ Hz, 1H), 4.15 (t, $J = 6.8$ Hz, 2H), 3.50 (d, $J = 5.6$ Hz, 1H), 1.52-1.60 (m, 2H), 1.15-1.28 (m, 6H), 0.84 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.9, 138.6, 128.7, 128.5, 126.6, 73.0, 66.5, 31.3, 28.5, 25.4, 22.6, 14.0; IR (Neat) 3464, 2956, 2930, 2858, 1731, 1455, 1182, 1067 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{14}\text{H}_{20}\text{O}_3\text{Na}_1$, 259.1310; found, 259.1310. The enantiomeric excess (%*ee*) was determined to be 98% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_{R} (minor, 6.1 min), t_{R} (major, 8.2 min).



White solid; mp = 56 °C (lit. 53-54 °C); R_f 0.48; (hexanes : ethyl acetate, 80:20 v/v): $[\alpha]_{D}^{20} = -86.7$ ($c = 1$ in CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.40 (d, $J = 7.2$ Hz, 2H), 7.28-7.38 (m, 3H), 5.04 (s, 1H), 3.52 (bs, 1H), 1.40 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.0, 139.1, 128.5, 128.2, 126.5, 83.2, 73.1, 27.9; IR (Neat) 3476, 2977, 2930, 1729, 1157, 912 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{12}\text{H}_{16}\text{O}_3\text{Na}_1$, 231.0997; found, 231.0997. The enantiomeric excess (%*ee*) was determined to be 98% by HPLC using ChiralPAK OD-H column (25% *i*-PrOH/ hexanes, 0.5 mL/min, 200 nm): t_{R} (minor, 8.0 min), t_{R} (major, 11.3 min).



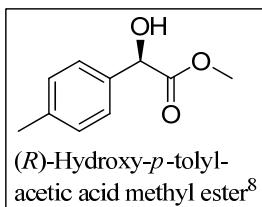
Colorless oil; R_f 0.32; (hexanes : ethyl acetate, 80:20 v/v): $[\alpha]_{D}^{20} = -90.5$ ($c = 0.4$ in CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.41-7.46 (m, 2H), 7.30-7.40 (m, 3H), 5.78-5.89 (m, 1H), 5.21 (s, 2H), 5.15-5.19 (m, 1H), 4.60-4.72 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 173.3, 138.4, 131.2, 128.6, 128.5, 126.6, 118.7, 73.0, 66.4; IR (Neat) 3442, 2932, 1731, 1596, 1454, 1212, 1164 cm^{-1} ; HRMS (m/z): $[\text{MNa}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{O}_3\text{Na}_1$, 215.0684; found, 215.0686. The enantiomeric excess (%*ee*) was determined to be 99.9% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_{R} (major, 5.1 min),



(*R*)-(4-Methoxyphenyl)-hydroxy-

acetic acid methyl ester⁸

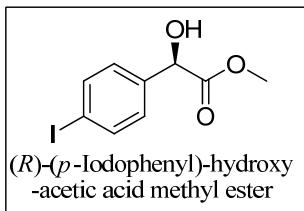
White solid; mp = 62 °C (lit. 63-64 °C,); R_f 0.30; (hexanes : ethyl acetate, 70:30 v/v): [α]^D₃₀ = -129.1 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.33 (dd, J = 2.0 Hz, 6.8 Hz, 2H), 6.90 (dd, J = 2.0 Hz, 6.8 Hz, 2H), 5.13 (d, J = 5.6 Hz, 1H), 3.81 (s, 3H), 3.76 (s, 3H), 3.39 (d, J = 5.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 174.5, 159.9, 130.6, 128.1, 114.2, 72.6, 55.4, 53.1; IR (Neat) 3452, 2948, 1738, 1248, 913 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₀H₁₂O₄Na₁, 219.0997; found, 219.0995. The enantiomeric excess (%ee) was determined to be 90% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (major, 21.5 min), t_R (minor, 24.8 min).



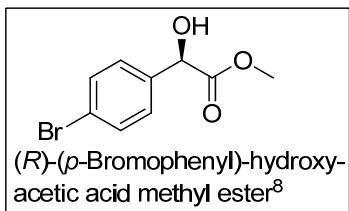
(*R*)-Hydroxy-*p*-tolyl-

acetic acid methyl ester⁸

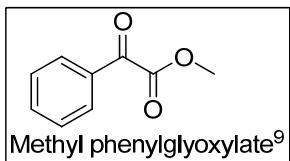
Colorless oil; R_f 0.35; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -82.6 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.30 (d, J = 8 Hz, 2H), 7.18 (d, J = 8 Hz, 2H), 5.15 (d, J = 5.6 Hz, 1H), 3.76 (s, 3H), 3.40 (d, J = 5.6 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 174.4, 138.4, 135.5, 129.4, 126.6, 72.9, 53.0, 21.3; IR (Neat) 3477, 2952, 1738, 1081, 921 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₀H₁₂O₃Na₁, 203.1048; found, 203.1043. The enantiomeric excess (%ee) was determined to be 90% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (major, 10.3 min), t_R (minor, 12.1 min).



(*R*)-(*p*-Iodophenyl)-hydroxy-acetic acid methyl ester
White solid; mp = 64 °C; R_f 0.29; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -55.8 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.70 (d, J = 8 Hz, 2H), 7.18 (d, J = 8 Hz, 2H), 5.12 (s, 1H), 3.76 (s, 3H), 3.47 (bs, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 173.7, 138.0, 137.8, 128.6, 94.4, 72.4, 53.3; IR (Neat) 3443, 2937, 1737, 1224, 1180, 1077 cm⁻¹; HRMS (*m/z*): [MHNa]⁺ calcd for C₉H₁₀O₃I₁Na₁, 315.9570; found, 315.9572. The enantiomeric excess (%ee) was determined to be 99.64% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (major, 11.0 min), t_R (minor, 12.4 min).

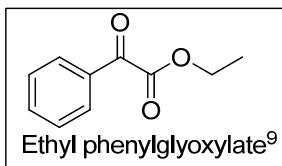


(*R*)-(*p*-Bromophenyl)-hydroxy-acetic acid methyl ester⁸
Colorless oil; R_f 0.25; (hexanes : ethyl acetate, 80:20 v/v): [α]^D₃₀ = -83.0 (c = 1 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.50 (d, J = 8.4 Hz, 2H), 7.31 (d, J = 8 Hz, 2H), 5.14 (d, J = 4.4 Hz, 1H), 3.77 (s, 3H), 3.47 (d, J = 4.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 173.8, 137.3, 131.9, 128.4, 122.7, 72.4, 53.3; IR (Neat) 3401, 2952, 1739, 1252, 913 cm⁻¹; HRMS (*m/z*): [MHNa]⁺ calcd for C₉H₁₀O₃Br₁Na₁, 267.9711; found, 267.9713. The enantiomeric excess (%ee) was determined to be 85% by HPLC using ChiralPAK AS-H column (10% *i*-PrOH/ hexanes, 1 mL/min, 190 nm): t_R (minor, 8.9 min), t_R (major, 10.4 min).

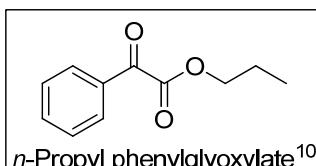


Methyl phenylglyoxylate⁹
Colorless oil; R_f 0.63; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 8.02 (d, J = 7.6 Hz, 2H), 7.67 (t, J = 7.2 Hz, 1H), 7.52 (t, J = 7.6 Hz, 2H), 3.99 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.1, 164.1, 135.0, 132.4,

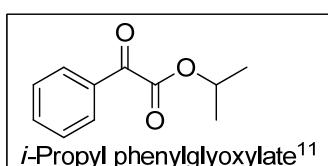
130.1, 128.9, 52.8 ; IR (Neat) 2963, 1741, 1688, 1208, 1003 cm^{-1} ; HRMS (m/z): [MH]⁺ calcd for C₉H₉O₃, 165.0552; found, 165.0547.



Colorless oil; R_f 0.69; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.99-8.04 (m, 2H), 7.66 (t, J = 7.6 Hz, 1H), 7.52 (t, J = 7.6 Hz, 2H), 4.46 (q, J = 7.2 Hz, 2H), 1.43 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.4, 163.8, 134.9, 132.4, 130.0, 128.9, 62.3, 14.1; IR (Neat) 2984, 2920, 1732, 1686, 1196, 1175 cm^{-1} ; HRMS (m/z): [MNa]⁺ calcd for C₁₀H₁₀O₃Na₁, 201.0528; found, 201.0528.

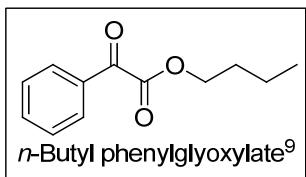


Colorless oil; R_f 0.76; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.98-8.03 (m, 2H), 7.63-7.68 (m, 1H), 7.51 (t, J = 8 Hz, 2H), 4.35 (t, J = 6.8 Hz, 2H), 1.81 (Sextet, J = 7.2 Hz, 2H), 1.01 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.6, 164.1, 135.0, 132.6, 130.1, 129.0, 67.9, 22.0, 10.4; IR (Neat) 2970, 1736, 1690, 1199, 1177, 991 cm^{-1} ; HRMS (m/z): [MNa]⁺ calcd for C₁₁H₁₂O₃Na₁, 215.0684; found, 215.0683.

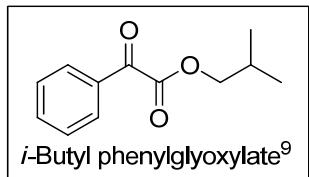


Colorless oil; R_f 0.75; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.98-8.02 (m, 2H), 7.63-7.68 (m, 1H), 7.51 (t, J = 8 Hz, 2H), 5.33 (Septet, J = 6 Hz, 1H), 1.41 (d, J = 6 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 186.8, 163.7, 134.9, 132.5, 130.0, 128.9, 70.7, 21.8; IR (Neat) 2984, 2920, 1728, 1687,

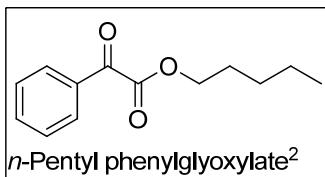
1201, 1176, 1101, 986 cm^{-1} ; HRMS (*m/z*): [MNa]⁺ calcd for C₁₁H₁₂O₃Na₁, 215.0684; found, 215.0681.



Colorless oil; R_f 0.74; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.98-8.03 (m, 2H), 7.63-7.69 (m, 1H), 7.48-7.54 (m, 2H), 4.40 (t, *J* = 6.8 Hz, 2H), 1.77 (Quintet, *J* = 6.8 Hz, 2H), 1.45 (Sextet, *J* = 7.6 Hz, 2H), 0.94-1.0 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.6, 164.1, 135.0, 132.6, 130.1, 129.0, 66.2, 30.6, 19.1, 13.7; IR (Neat) 2961, 2875, 1733, 1687, 1196, 1175 cm^{-1} ; HRMS (*m/z*): [MNa]⁺ calcd for C₁₂H₁₄O₃Na₁, 229.0841; found, 229.0845.

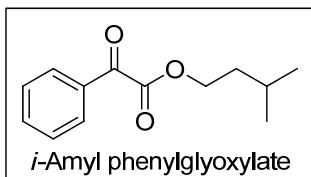


Colorless oil; R_f 0.74; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.98-8.03 (m, 2H), 7.64-7.69 (m, 1H), 7.49-7.55 (m, 2H), 4.18 (d, *J* = 6.8 Hz, 2H), 2.09 (Septet, *J* = 7.2 Hz, 1H), 1.01 (d, *J* = 6.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 186.7, 164.2, 135.0, 132.6, 130.1, 129.0, 72.2, 27.8, 19.1; IR (Neat) 2964, 1733, 1687, 1196, 1174, 995 cm^{-1} ; HRMS (*m/z*): [MNa]⁺ calcd for C₁₂H₁₄O₃Na₁, 229.0841; found, 229.0839.

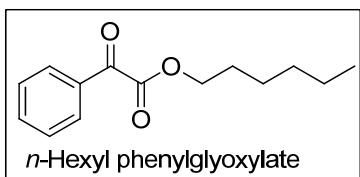


Colorless oil; R_f 0.67; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.98-8.02 (m, 2H), 7.63-7.68 (m, 1H), 7.51 (t, *J* = 8 Hz, 2H), 4.38 (t, *J* = 6.8 Hz, 2H), 1.78 (Quintet, *J* = 7.2 Hz, 2H), 1.30-1.45(m, 4H), 0.914 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.6, 164.1, 135.0, 132.6, 130.1, 129.0,

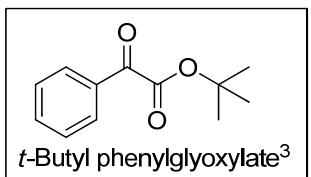
66.5, 28.3, 28.0, 22.4, 14.0; IR (Neat) 2958, 2934, 2872, 1733, 1687, 1196, 1175 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₃H₁₆O₃Na₁, 243.0997; found, 243.0994.



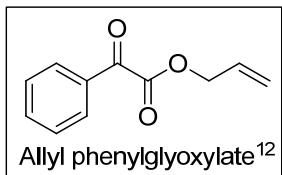
Colorless oil; R_f 0.71; (hexanes : ethyl acetate, 80:20 v/v):
¹H NMR (400 MHz, CDCl₃): δ 7.98-8.02 (m, 2H), 7.66 (t, J = 7.6 Hz, 1H), 7.51 (t, J = 7.6 Hz, 2H), 4.42 (t, J = 7.2 Hz, 2H), 1.75 (q, J = 6.8 Hz, 1H), 1.67 (q, J = 6.8 Hz, 2H), 0.96 (d, J = 6.4 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 186.6, 164.1, 135.0, 132.5, 130.1, 129.0, 65.0, 37.1, 34.2, 25.0, 22.5; IR (Neat) 2959, 1732, 1687, 1196, 1174, cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₃H₁₆O₃Na₁, 243.0997; found, 243.0992.



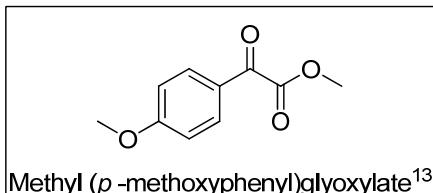
Colorless oil; R_f 0.65; (hexanes : ethyl acetate, 80:20 v/v):
¹H NMR (400 MHz, CDCl₃): δ 7.98-8.02 (m, 2H), 7.62-7.68 (m, 1H), 7.48-7.54 (m, 2H), 4.35-4.41 (m, 2H), 1.77 (Quintet, J = 7.2 Hz, 2H), 1.37-1.44(m, 2H), 1.26-1.34 (m, 4H), 0.85-0.92 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 186.5, 164.1, 134.9, 132.5, 130.0, 128.9, 66.4, 31.3, 28.4, 25.5, 22.5, 14.0; IR (Neat) 2957, 2931, 2860, 1733, 1687, 1194, 1174 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₄H₁₈O₃Na₁, 257.1154; found, 257.1155.



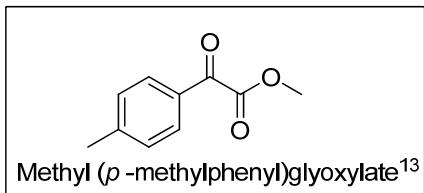
Colorless oil; R_f 0.61; (hexanes : ethyl acetate, 80:20 v/v):
¹H NMR (400 MHz, CDCl₃): δ 7.99-7.95 (m, 2H), 7.64 (t, J = 7.6 Hz, 1H), 7.51 (t, J = 7.6 Hz, 2H), 1.61 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 187.0, 163.9, 134.8, 132.7, 130.0, 129.0, 84.9, 28.2; IR (Neat) 2927, 2861, 1730, 1688, 1211, 1149, 985 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₂H₁₄O₃Na₁, 229.0841; found, 229.0836.



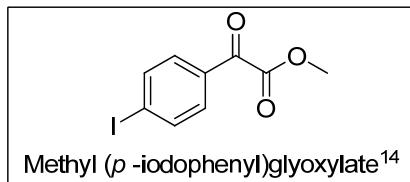
Colorless oil; R_f 0.68; (hexanes : ethyl acetate, 80:20 v/v): ^1H NMR (400 MHz, CDCl_3): δ 7.99-8.04 (m, 2H), 7.64-7.69 (m, 1H), 7.52 (t, J = 8 Hz, 2H), 5.97-6.08 (m, 2H), 5.32-5.49 (m, 1H), 4.88 (dt, J = 6 Hz, 1.2 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 186.1, 163.5, 135.0, 132.5, 130.8, 130.0, 128.9, 120.0, 66.6; IR (Neat) 3067, 1738, 1687, 1599, 1451, 1194, cm^{-1} ; HRMS (m/z): [MNa]⁺ calcd for $\text{C}_{11}\text{H}_{10}\text{O}_3\text{Na}_1$, 213.0528; found, 213.0532.



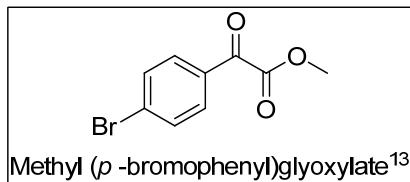
White solid; mp = 52 °C (lit. 51-49 °C,) R_f 0.54; (hexanes : ethyl acetate, 70:30 v/v): ^1H NMR (400 MHz, CDCl_3): δ 8.01 (d, J = 8.8 Hz, 2H), 6.97 (d, J = 8.8 Hz, 2H), 3.96 (s, 3H), 3.89 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 184.6, 165.2, 132.8, 125.6, 114.4, 55.8, 52.8; IR (Neat) 2936, 2859, 1730, 1682, 1213, 1155, 993 cm^{-1} ; HRMS (m/z): [MNa]⁺ calcd for $\text{C}_{10}\text{H}_{11}\text{O}_4$, 195.0657; found, 195.0661.



light yellow oil; R_f 0.70; (hexanes : ethyl acetate, 80:20 v/v): ^1H NMR (400 MHz, CDCl_3): δ 7.96 (d, J = 8.4 Hz, 2H), 7.36 (d, J = 8 Hz, 2H), 4.02 (s, 3H), 2.49 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 184.9, 163.5, 129.5, 129.2, 128.9, 53.1, 21.2; IR (Neat) 1739, 1686, 1210, 1172, 913 cm^{-1} ; HRMS (m/z): [MNa]⁺ calcd for $\text{C}_{10}\text{H}_{11}\text{O}_3\text{Na}_1$, 202.0606; found, 202.0597.



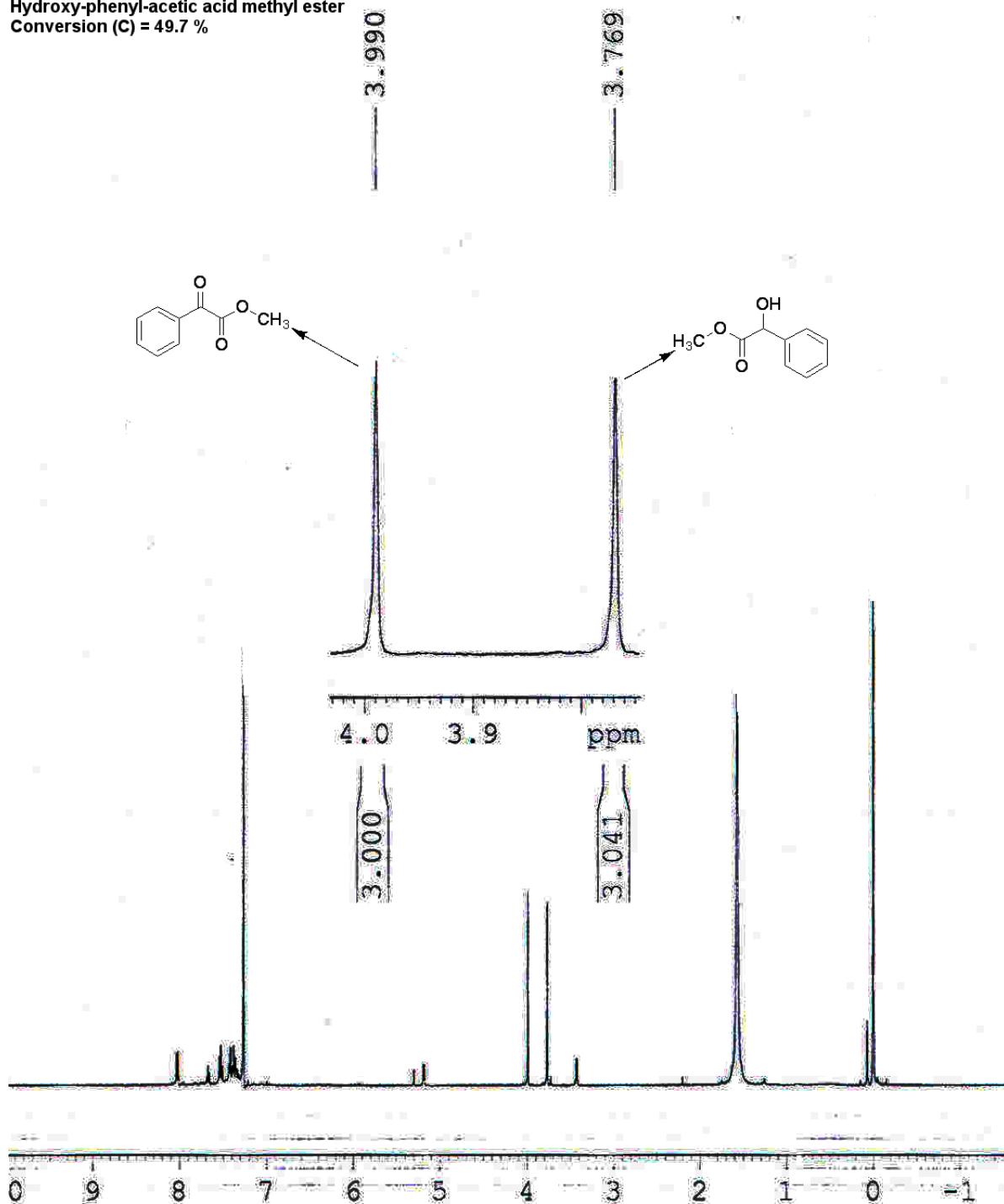
White solid; mp = 58-60 °C; R_f 0.61; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.87-7.91 (m, 2H), 7.74 (dd, J = 1.6 Hz, 8.4 Hz, 2H), 3.97 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 185.2, 163.5, 138.4, 131.8, 131.3, 104.0, 53.1; IR (Neat) 1739, 1689, 1209, 1173, 1002, 912 cm⁻¹; HRMS (m/z): [MHNa]⁺ calcd for C₉H₈O₃I₁Na₁, 313.9416; found, 313.9428.

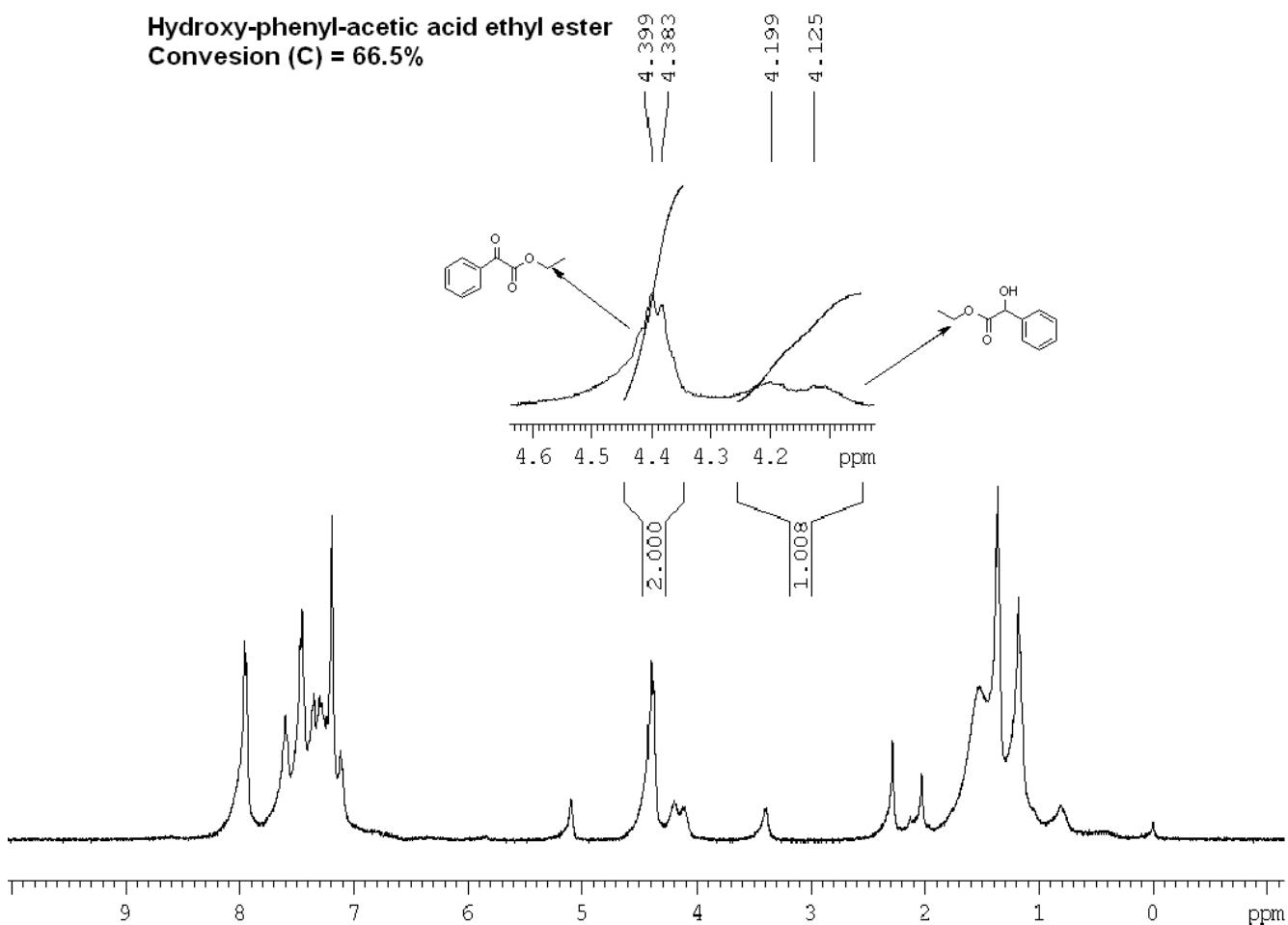


White solid; mp = 62 °C (lit. 63 °C,) R_f 0.53; (hexanes : ethyl acetate, 80:20 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.91 (J = 8.8 Hz, 2H), 7.66 (d, J = 8.4 Hz, 2H), 3.98 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 184.8, 163.5, 132.5, 131.6, 131.4, 130.8, 53.1; IR (Neat) 1737, 1689, 1209, 1171, 1003, 907 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₉H₇O₃Br₁Na₁, 264.9476; found, 264.9473.

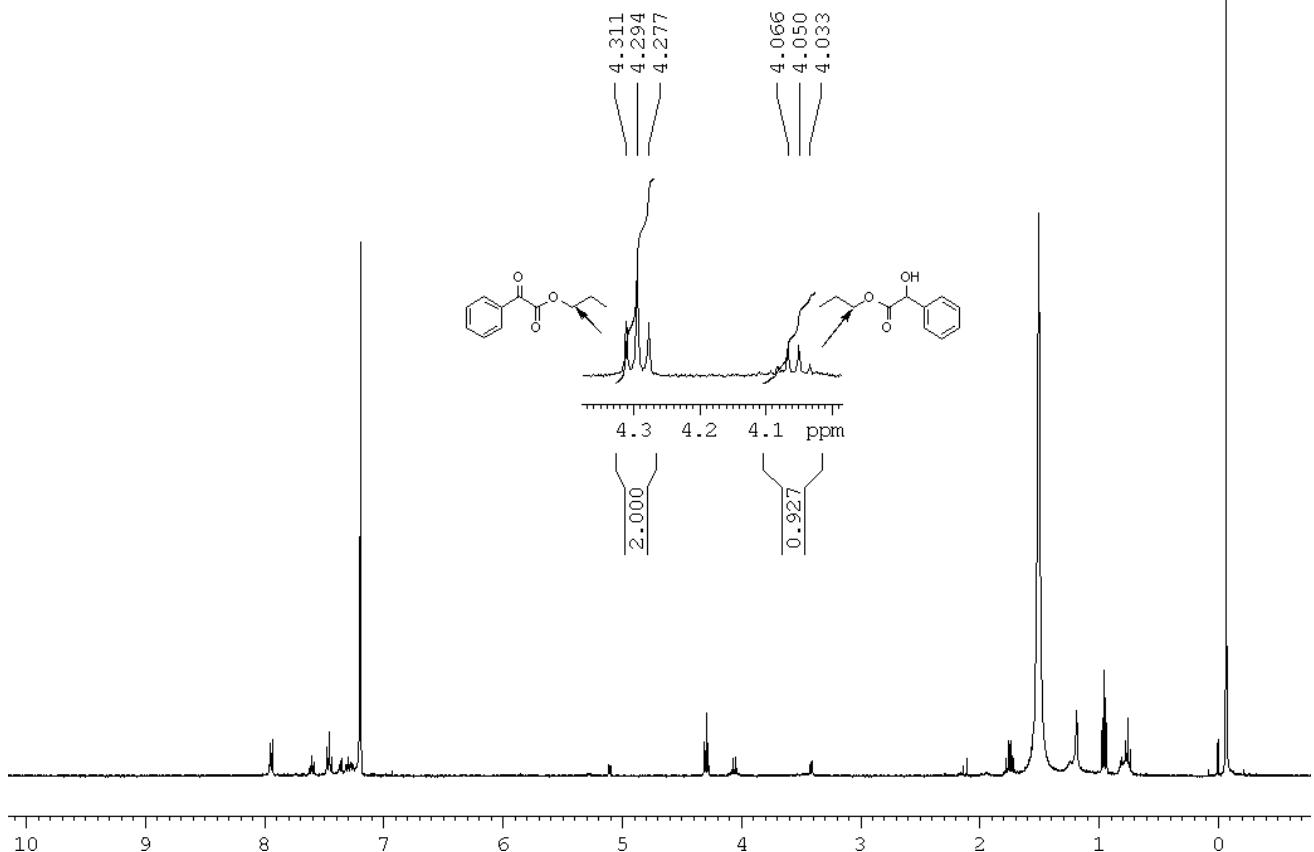
Calculation of conversion C (%) using ^1H NMR

Hydroxy-phenyl-acetic acid methyl ester
Conversion (C) = 49.7 %

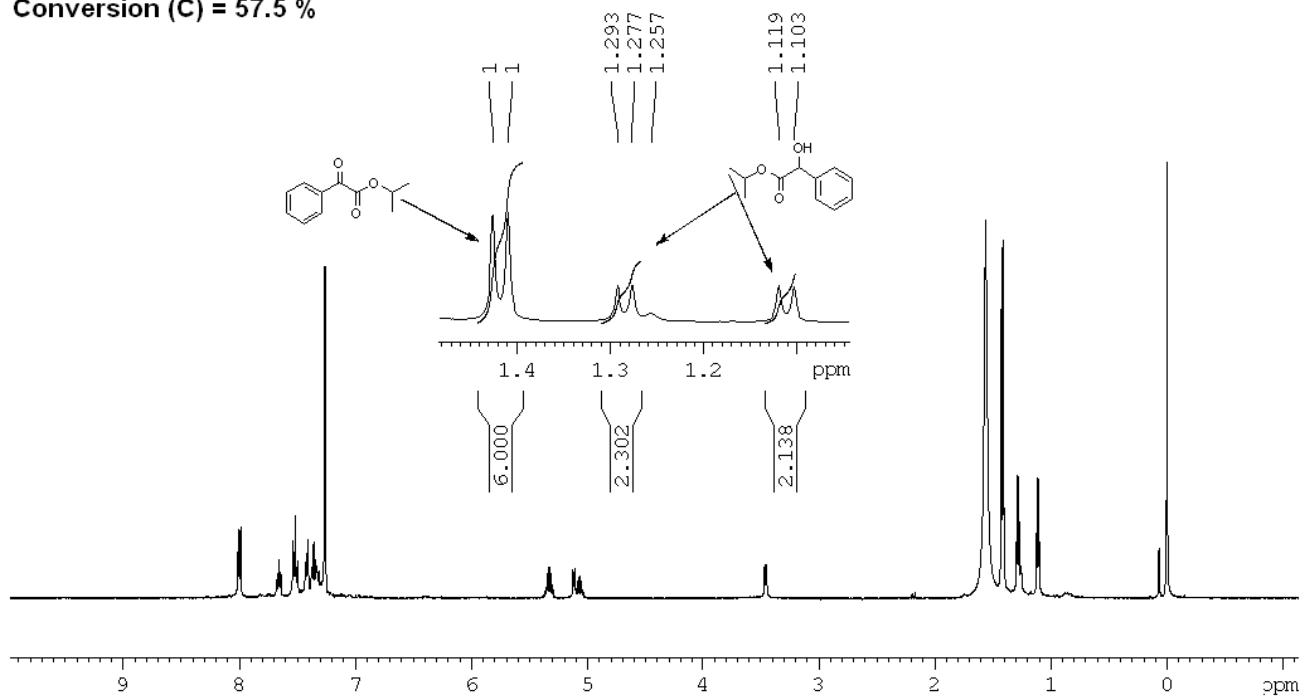




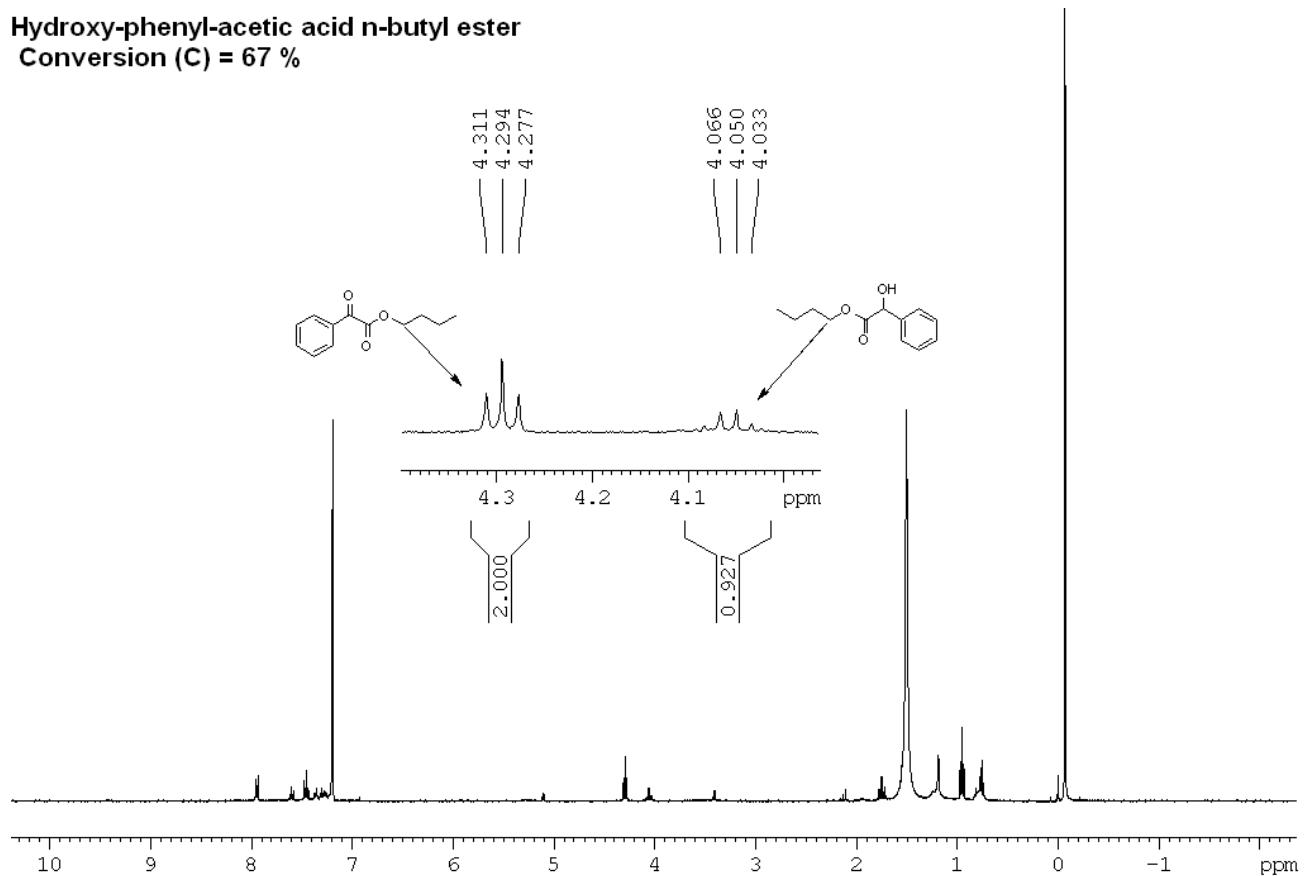
Hydroxy-phenyl-acetic acid n-propyl ester
Conversion (C) = 68 %

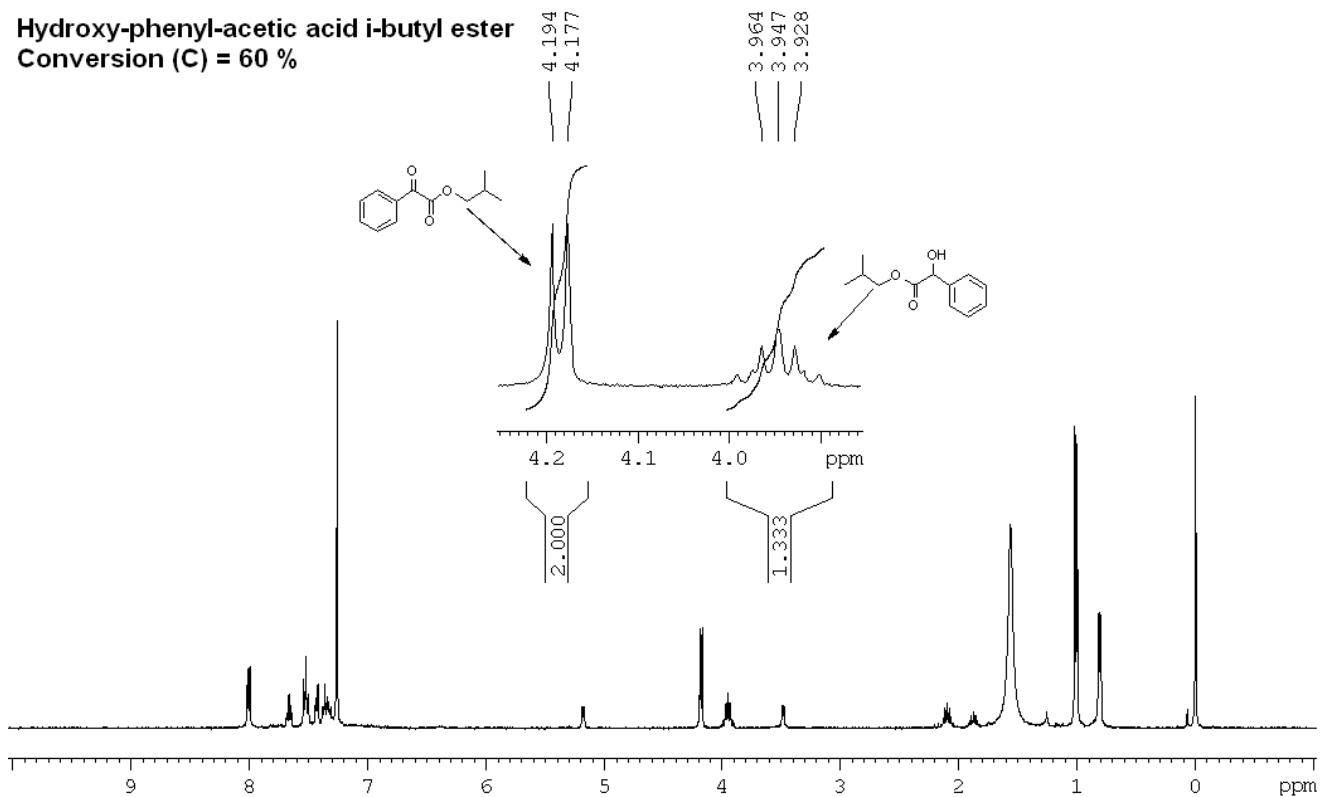


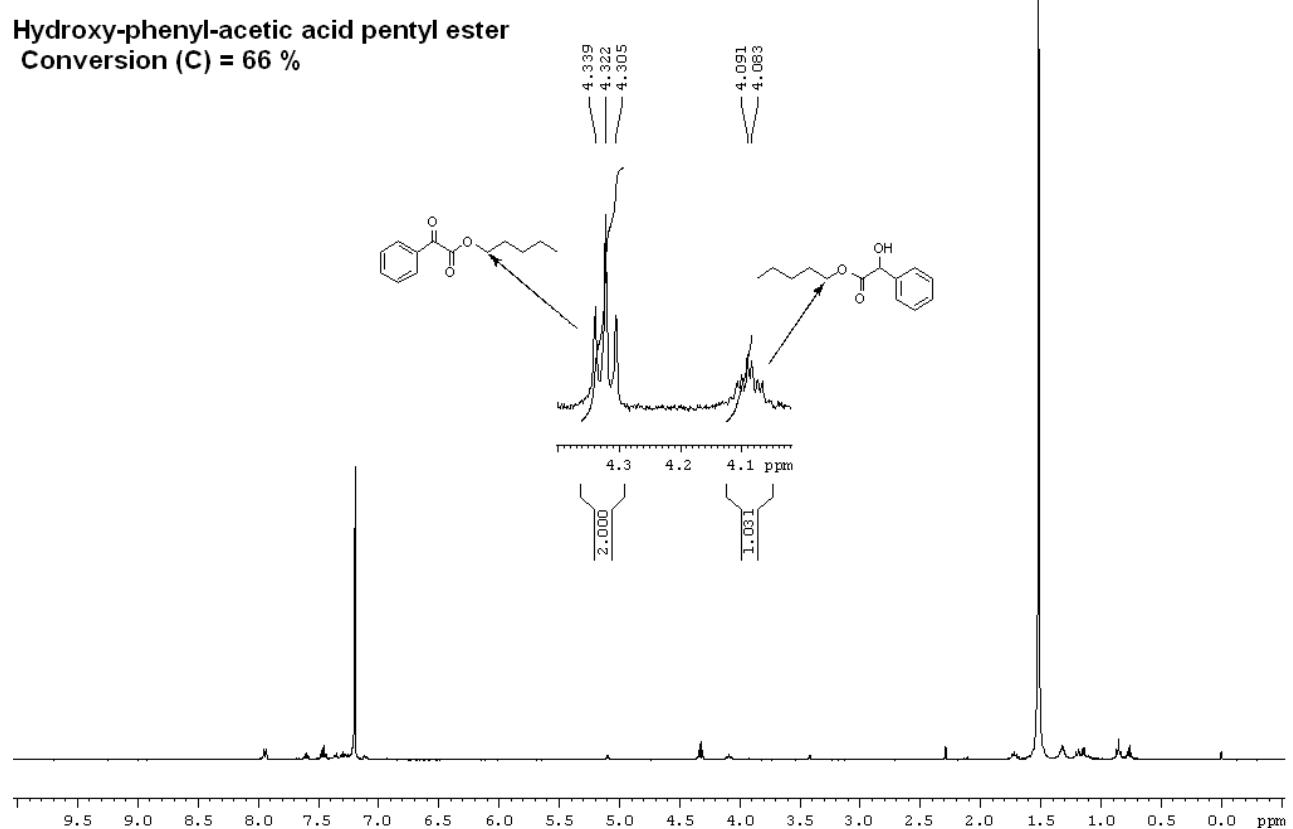
Hydroxy-phenyl-acetic acid i-propyl ester
Conversion (C) = 57.5 %



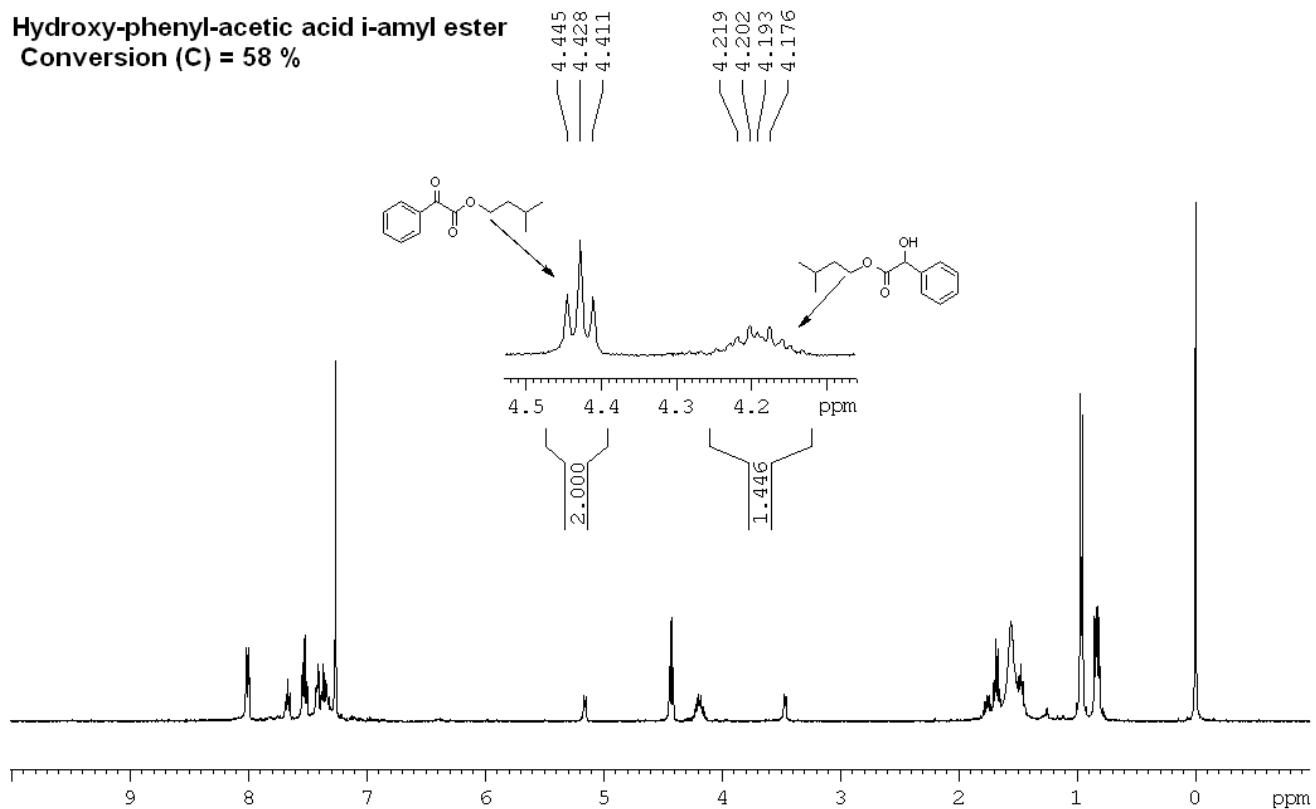
Hydroxy-phenyl-acetic acid n-butyl ester
Conversion (C) = 67 %

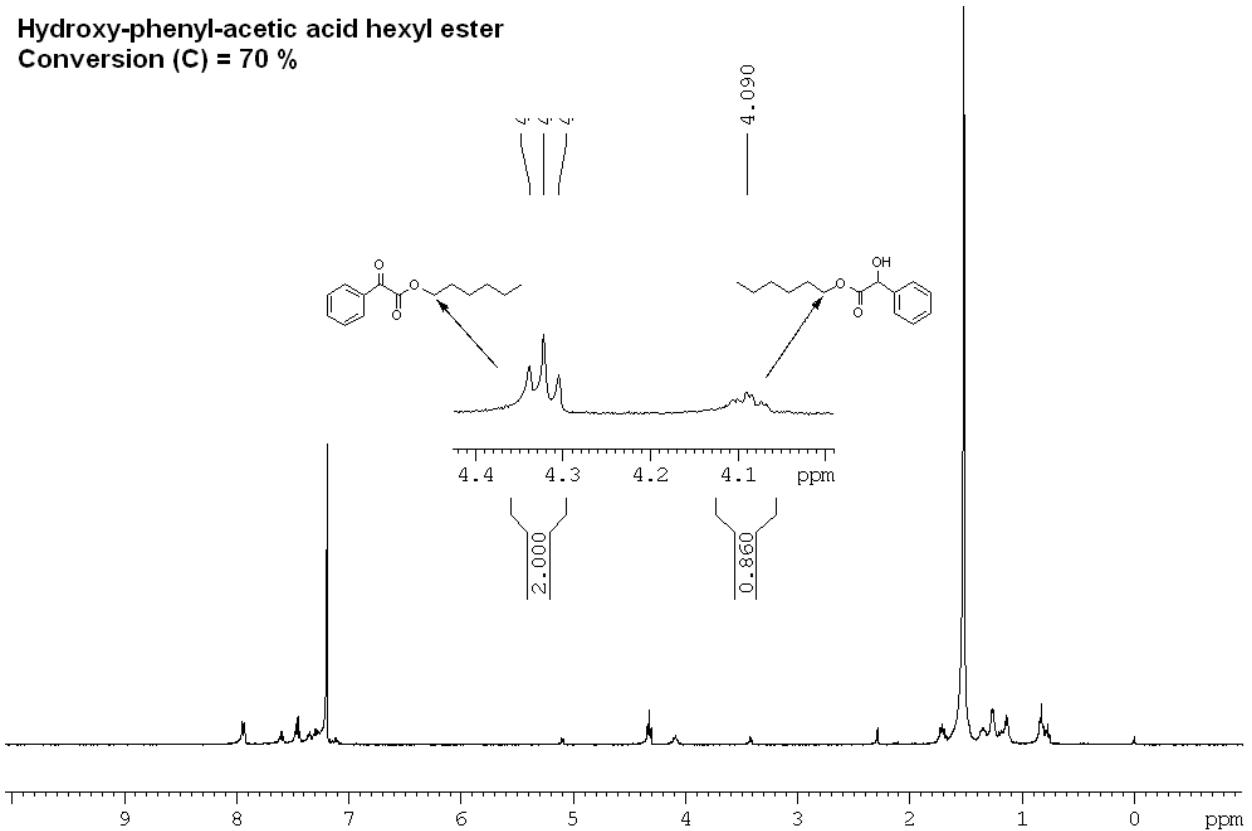


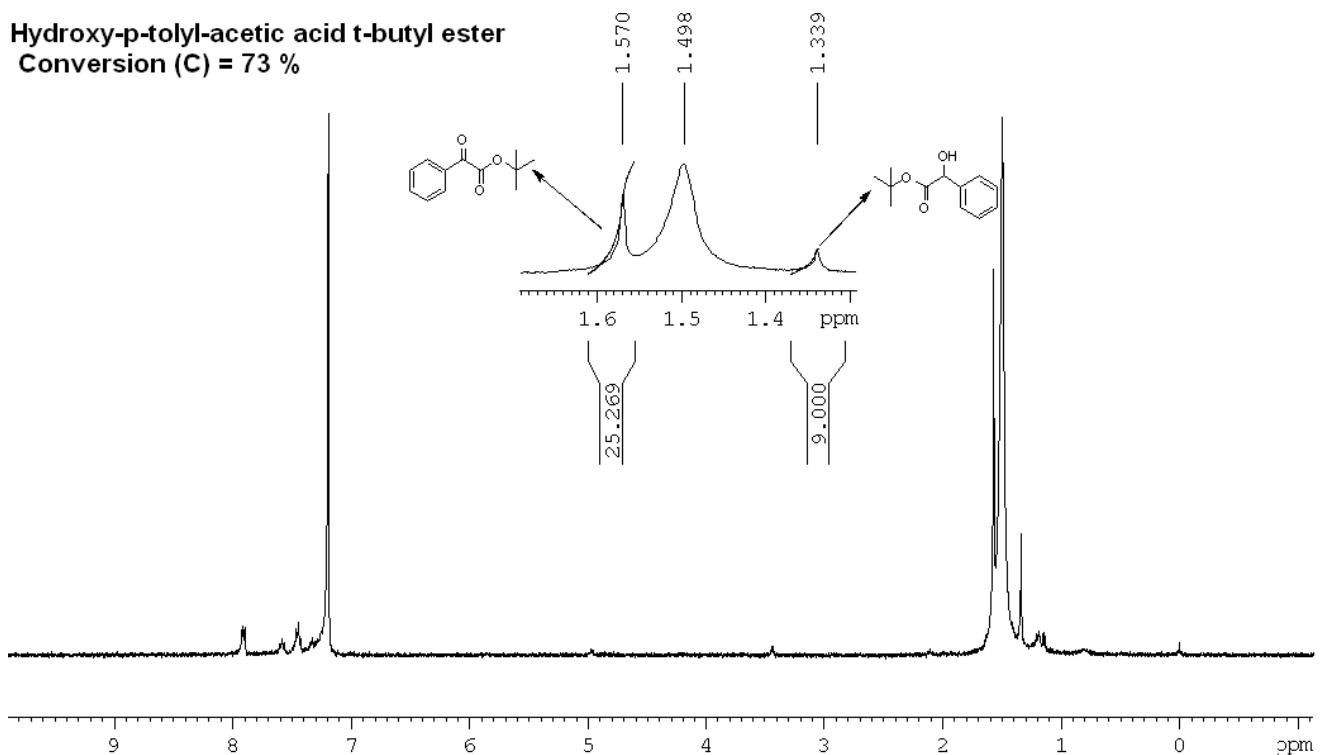




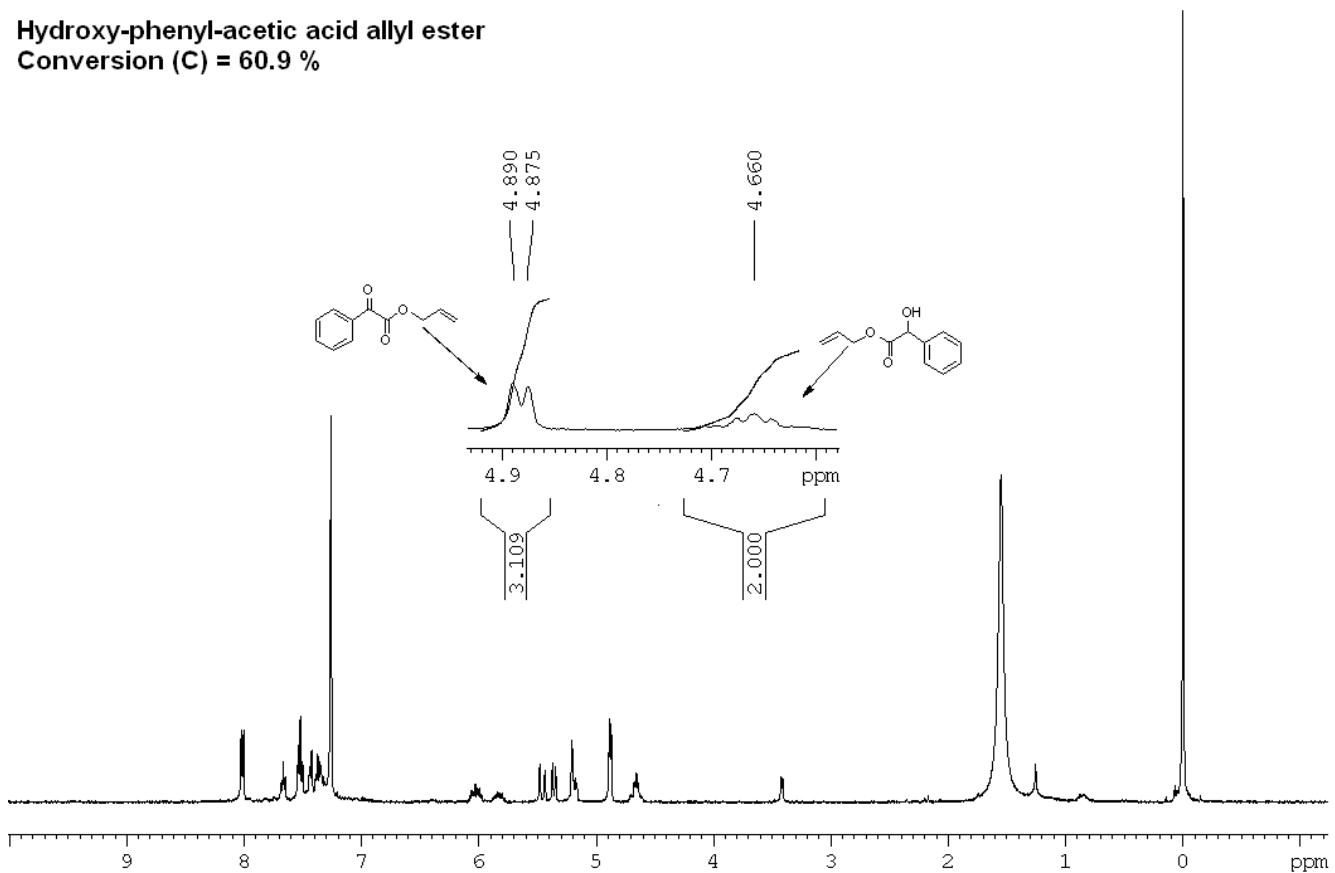
Hydroxy-phenyl-acetic acid i-amyl ester
Conversion (C) = 58 %

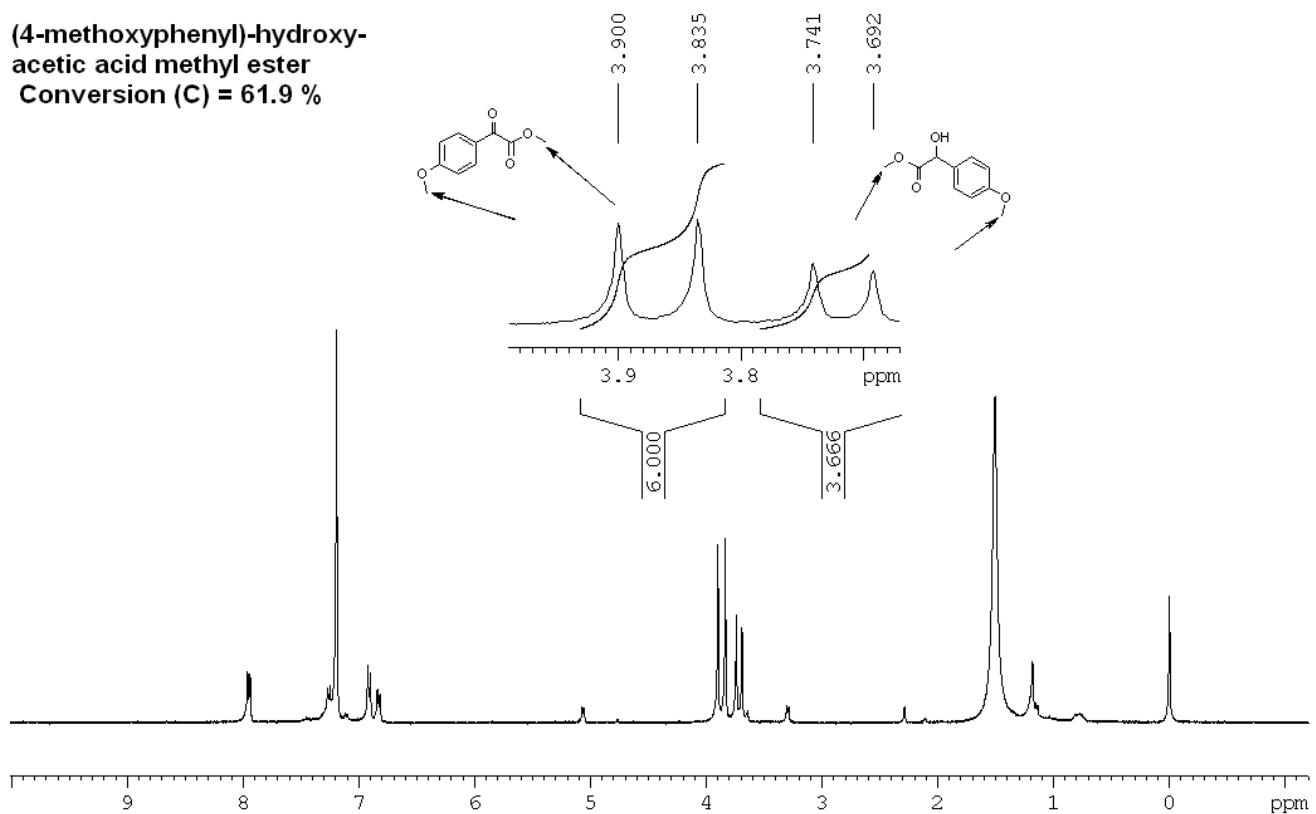




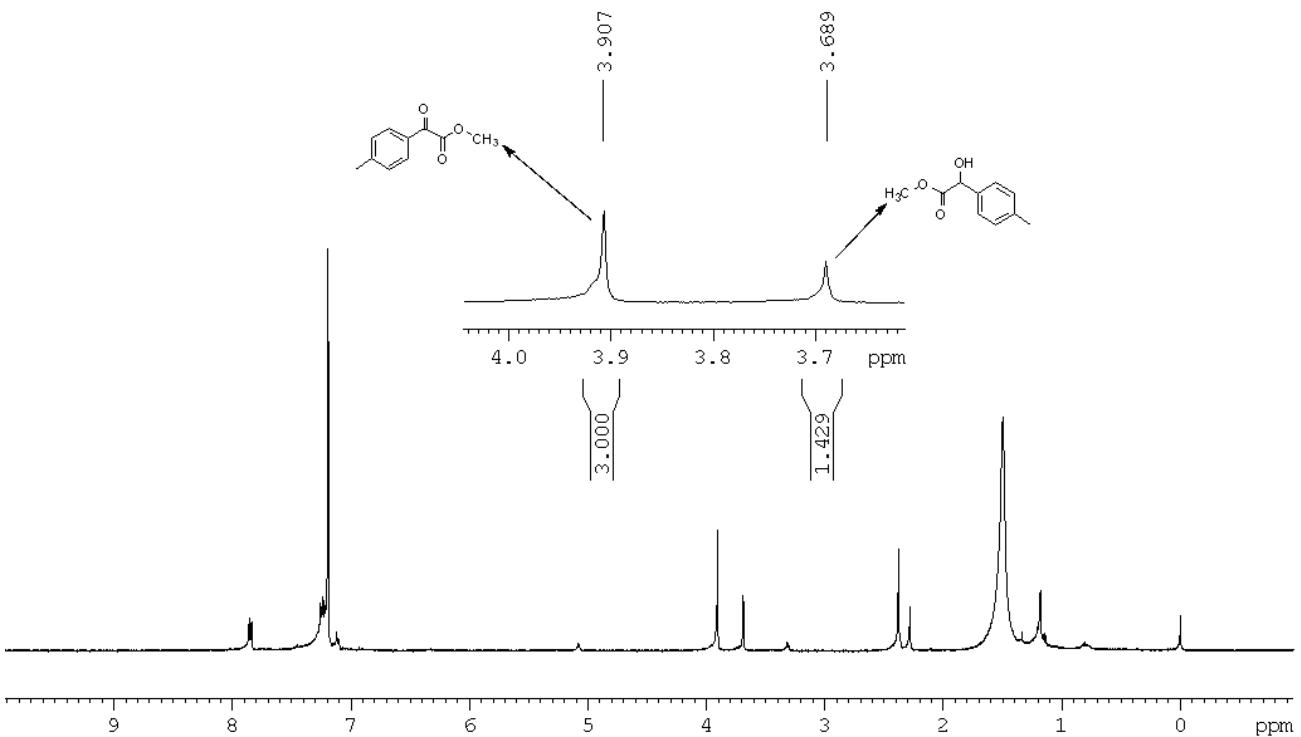


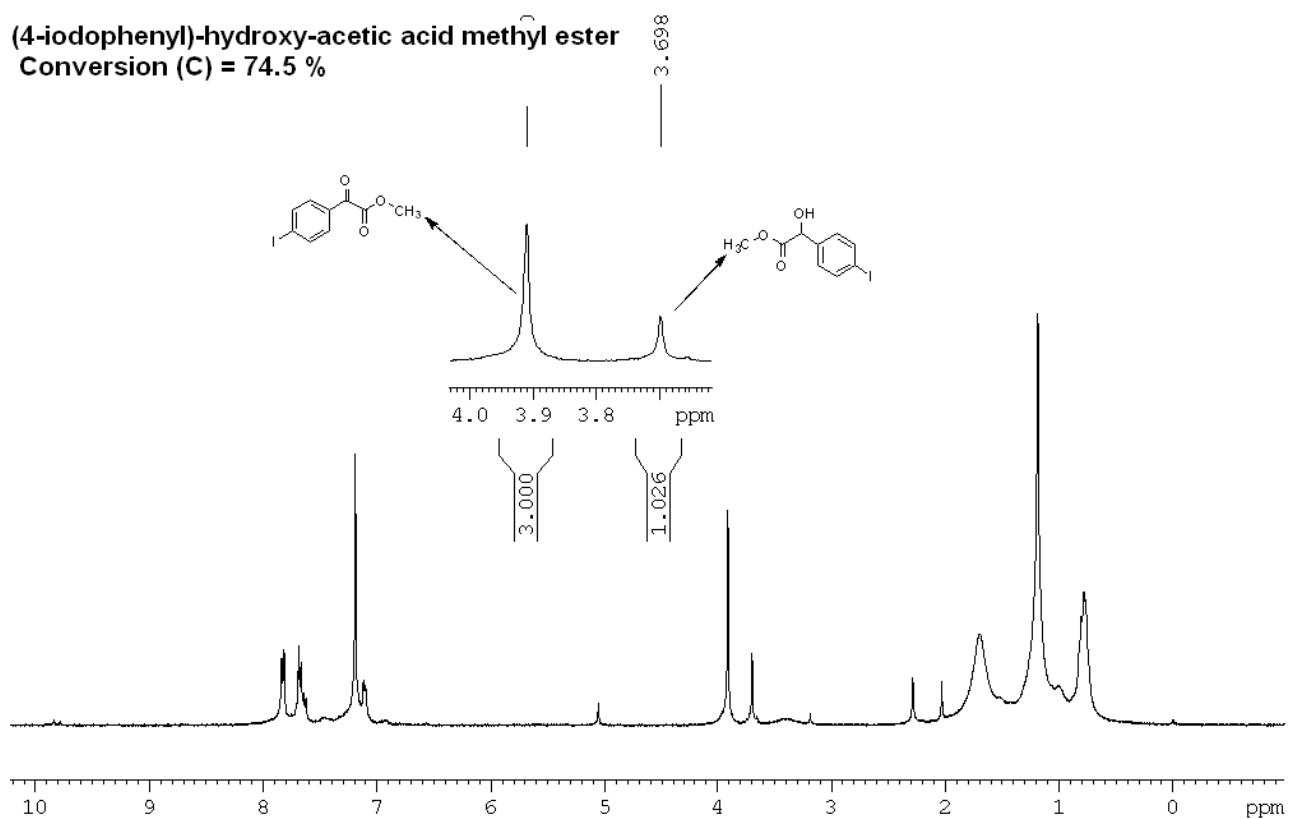
Hydroxy-phenyl-acetic acid allyl ester
Conversion (C) = 60.9 %



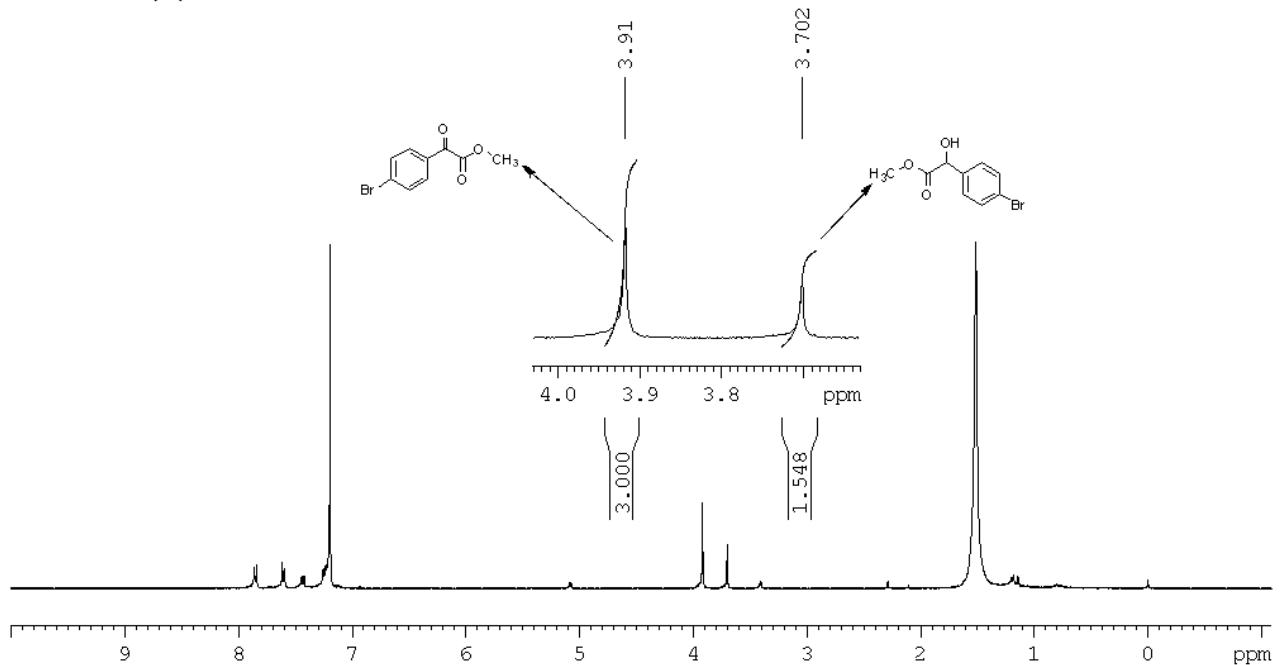


Hydroxy-p-tolyl-acetic acid methyl ester
Conversion (C) = 67.7 %

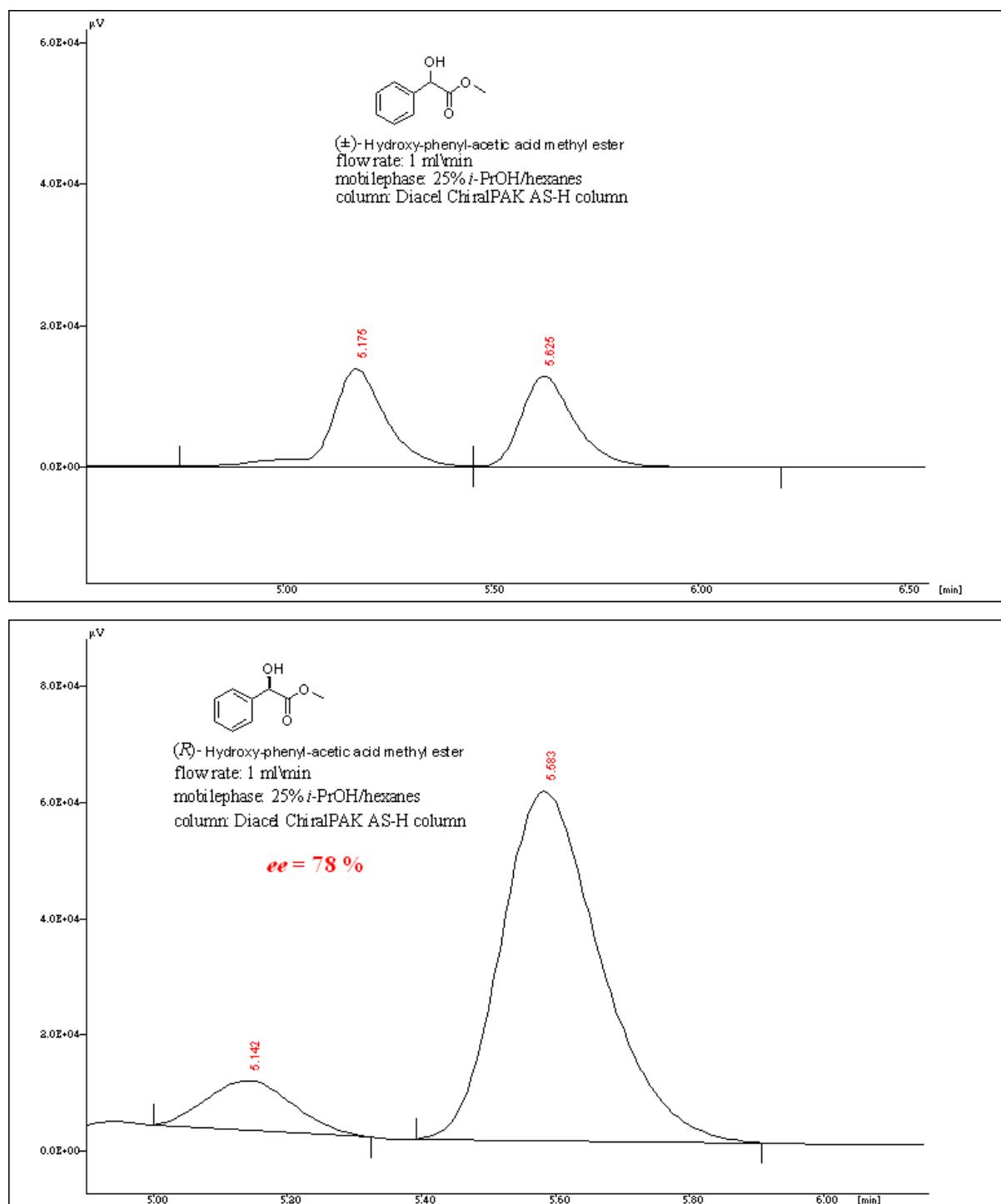




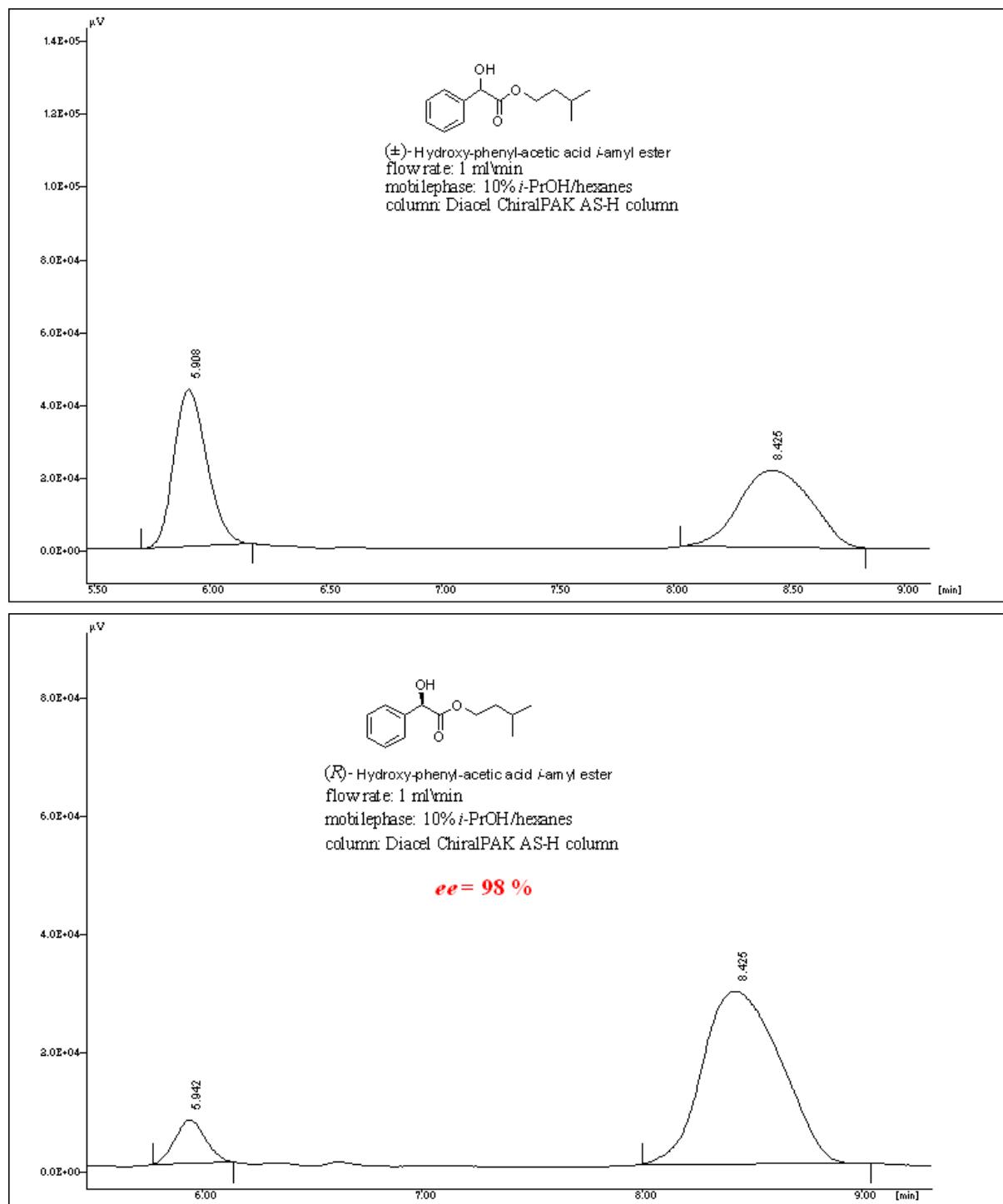
(4-bromophenyl)-hydroxy-acetic acid methyl ester
COnversion (C) = 66 %



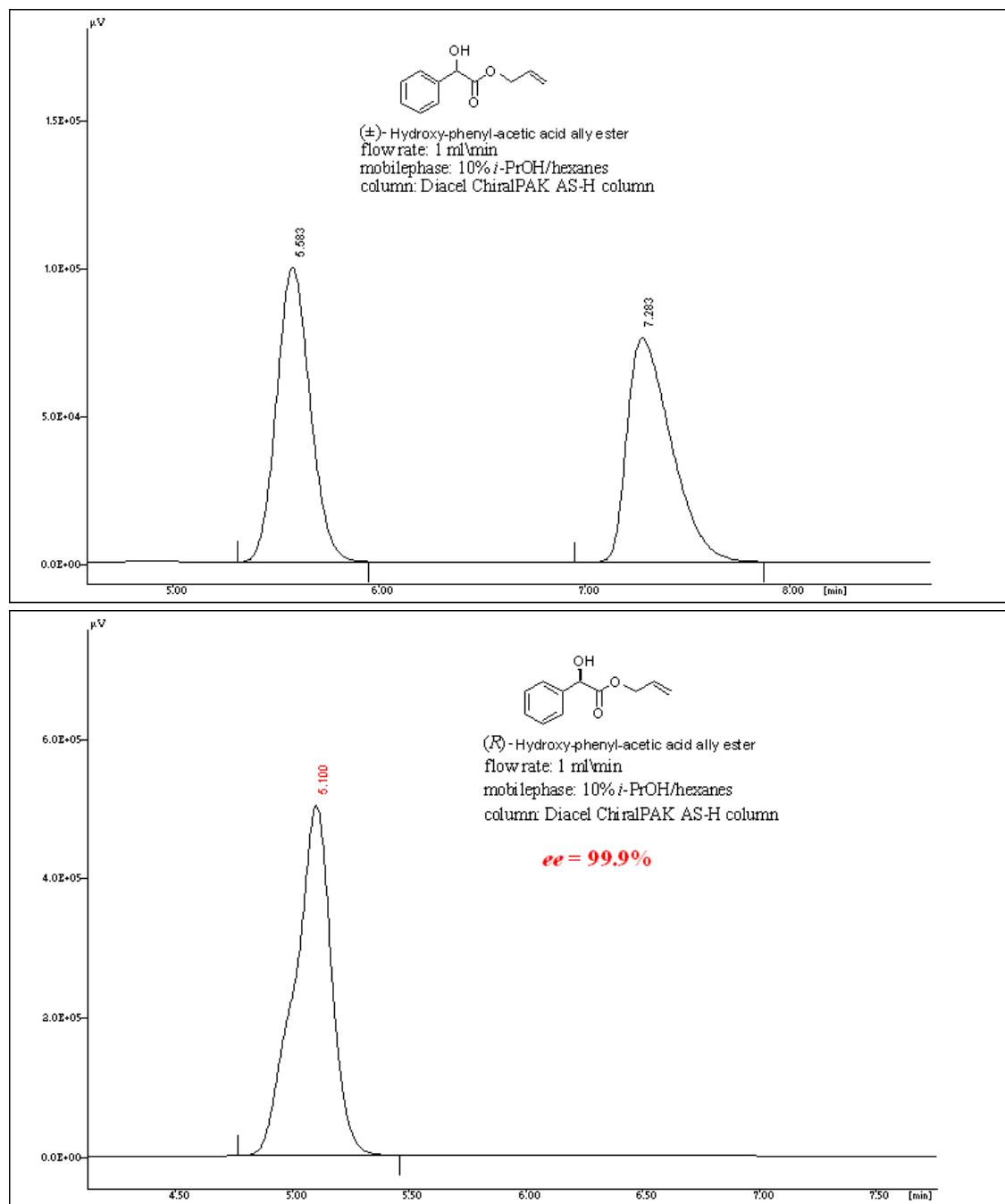
Hydroxy-phenyl-acetic acid methyl ester



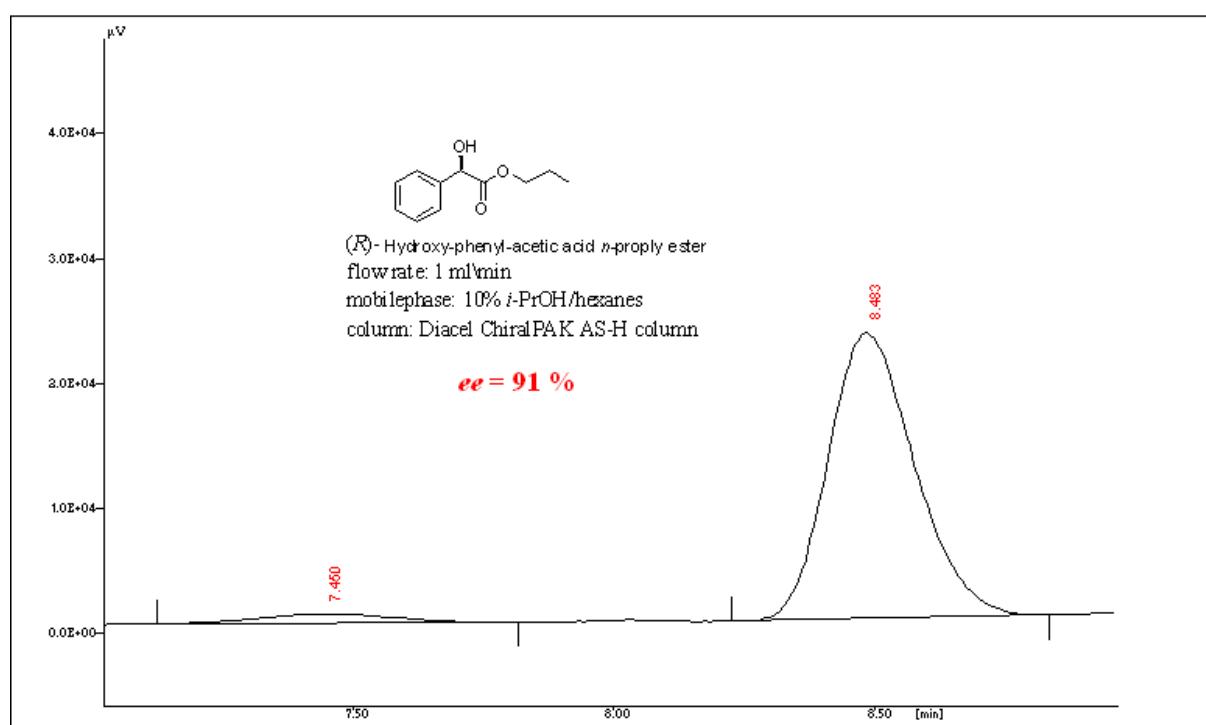
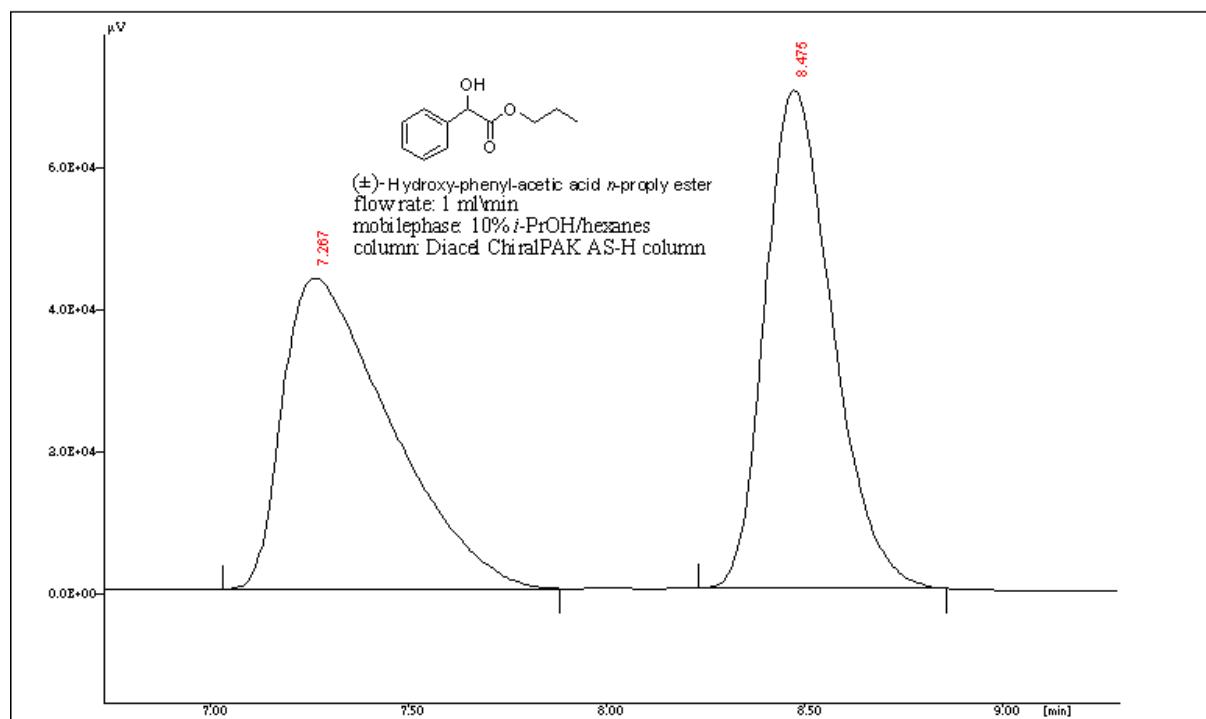
Hydroxy-phenyl-acetic acid *i*-amyl ester



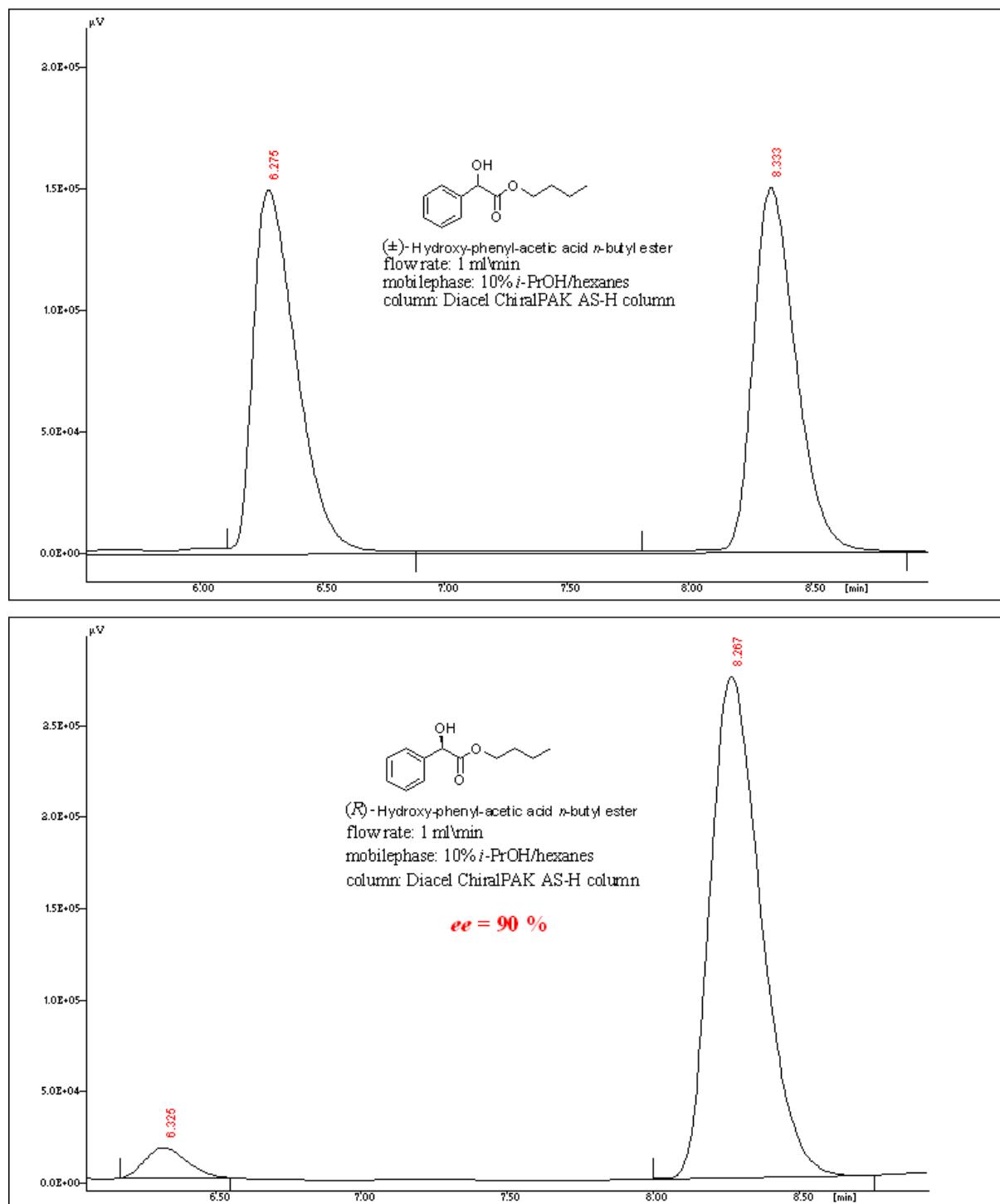
Hydroxy-phenyl-acetic acid allyl ester



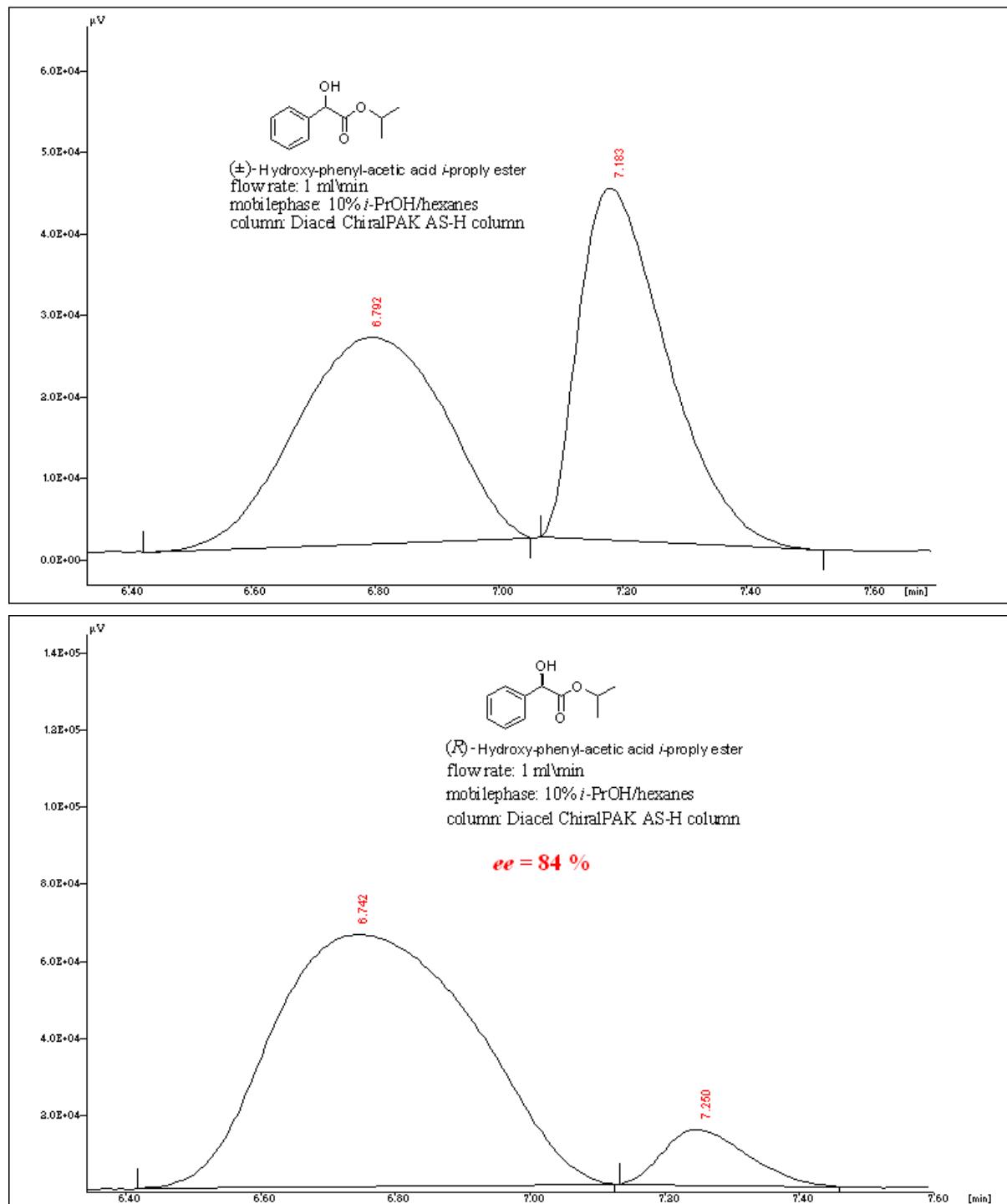
Hydroxy-phenyl-acetic acid *n*-propyl ester



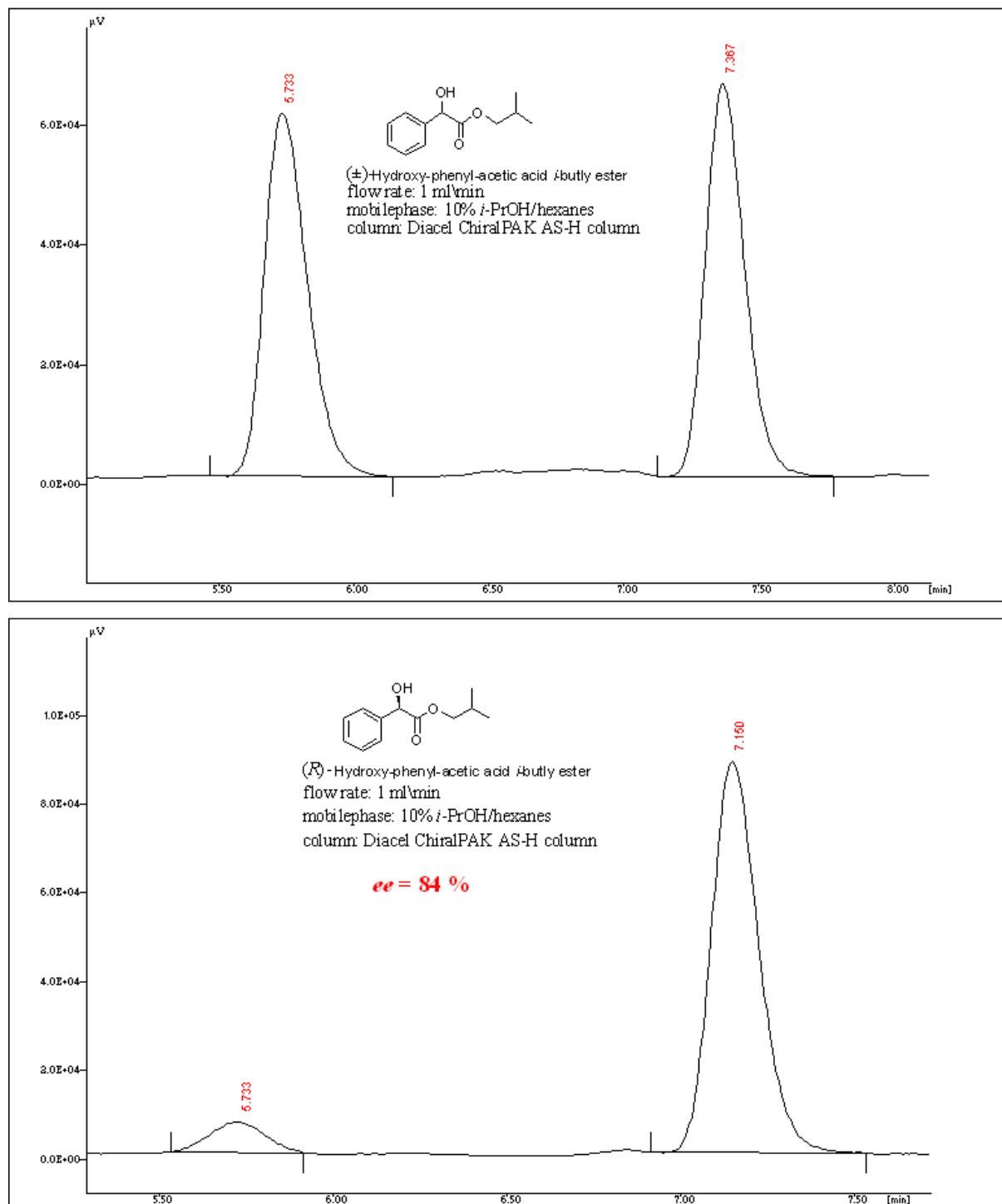
Hydroxy-phenyl-acetic acid *n*-butyl ester



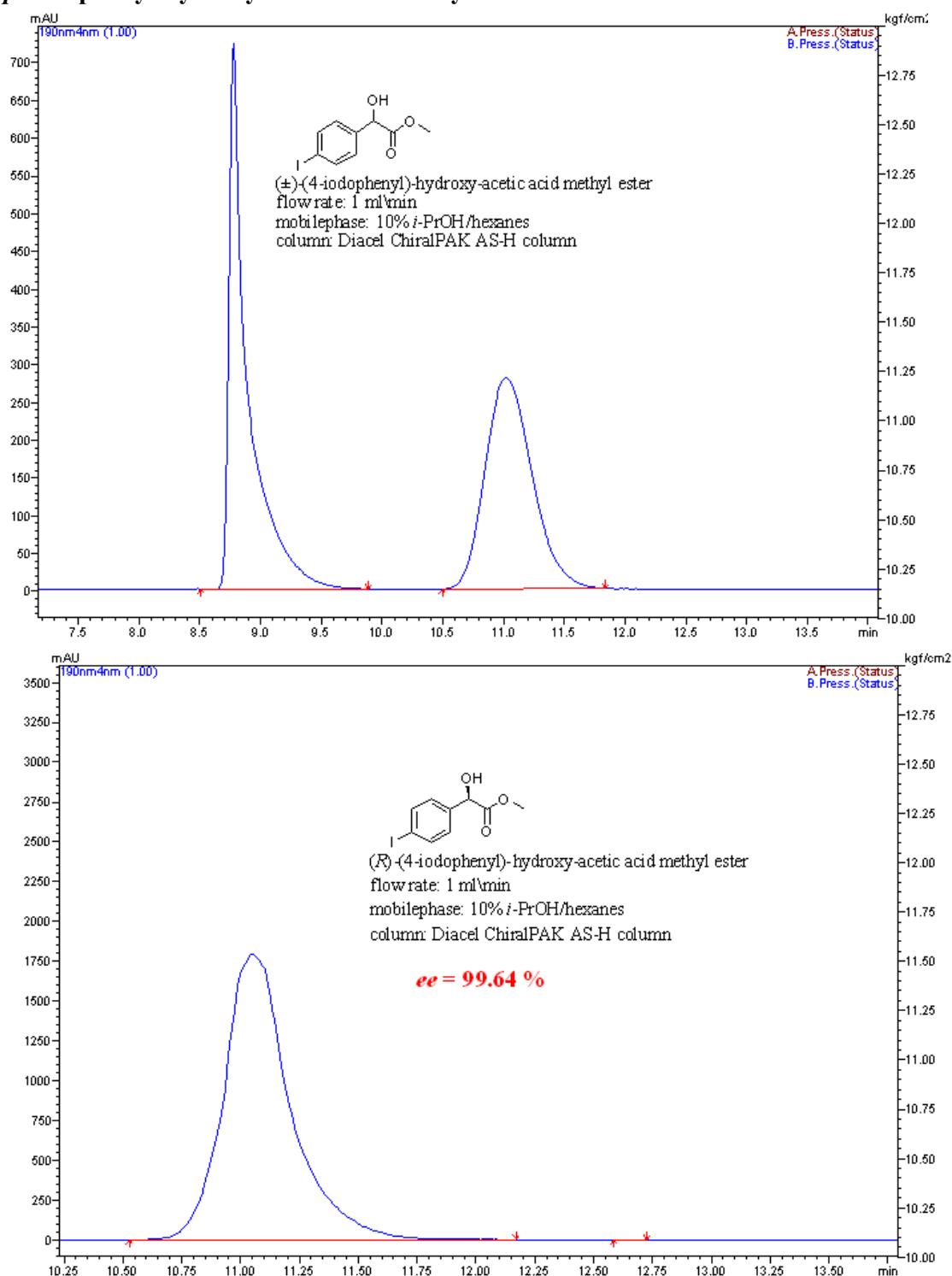
Hydroxy-phenyl-acetic acid *i*-propyl ester



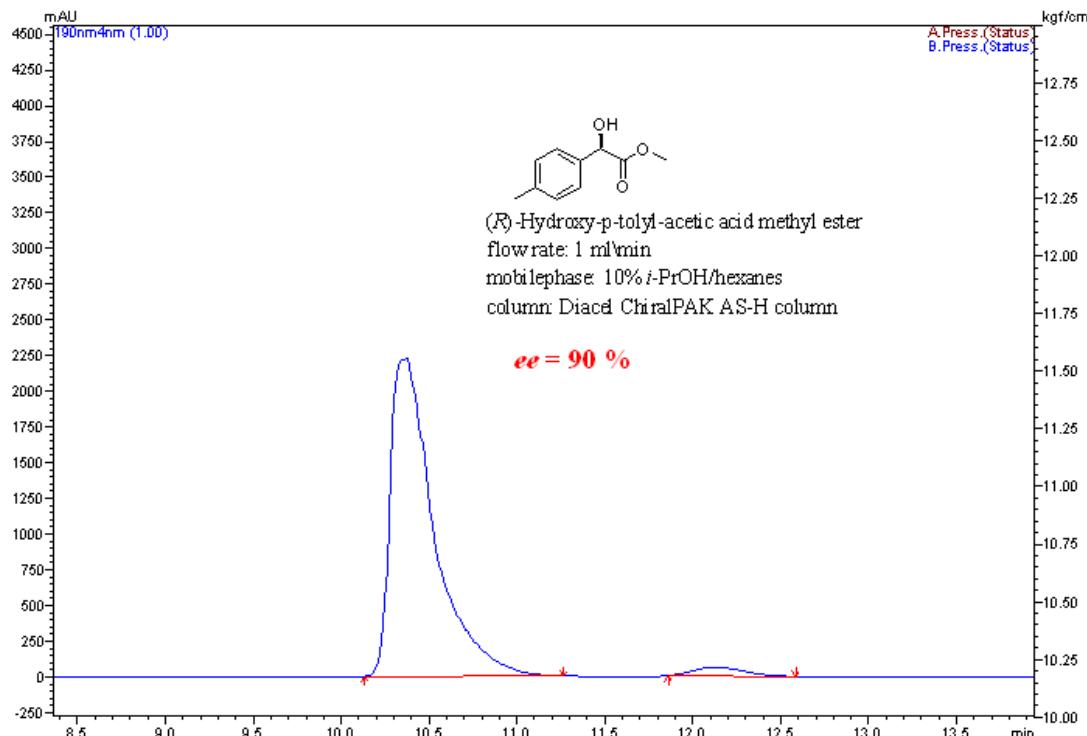
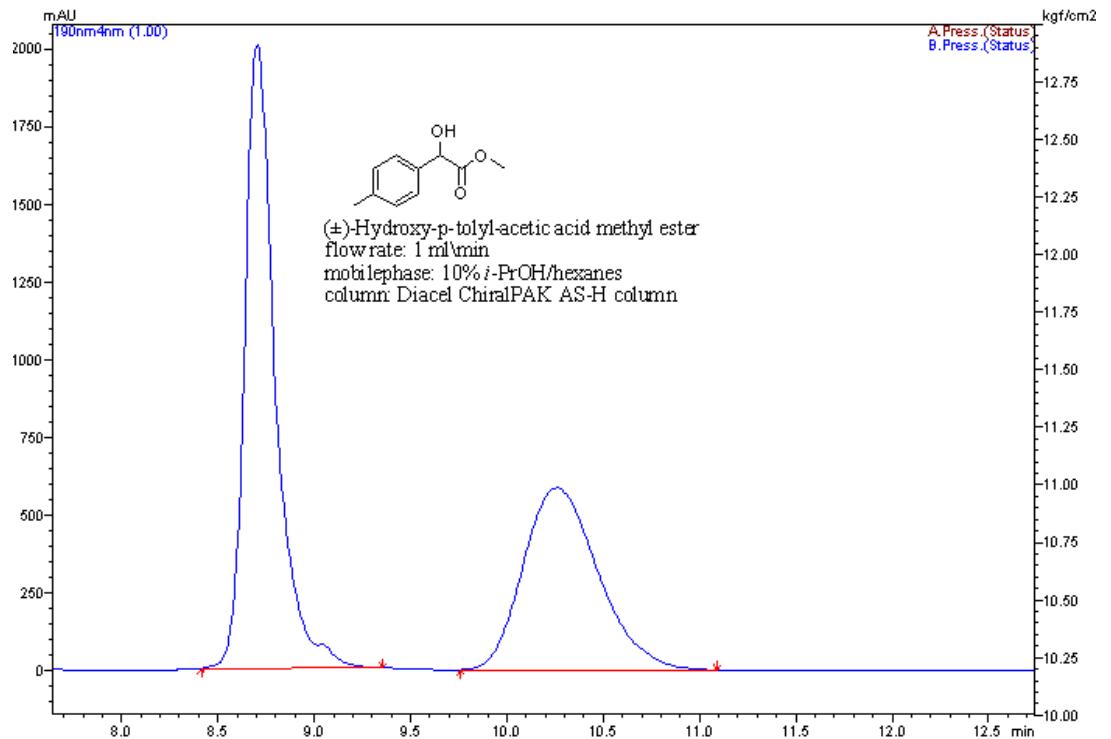
Hydroxy-phenyl-acetic acid *i*-butyl ester



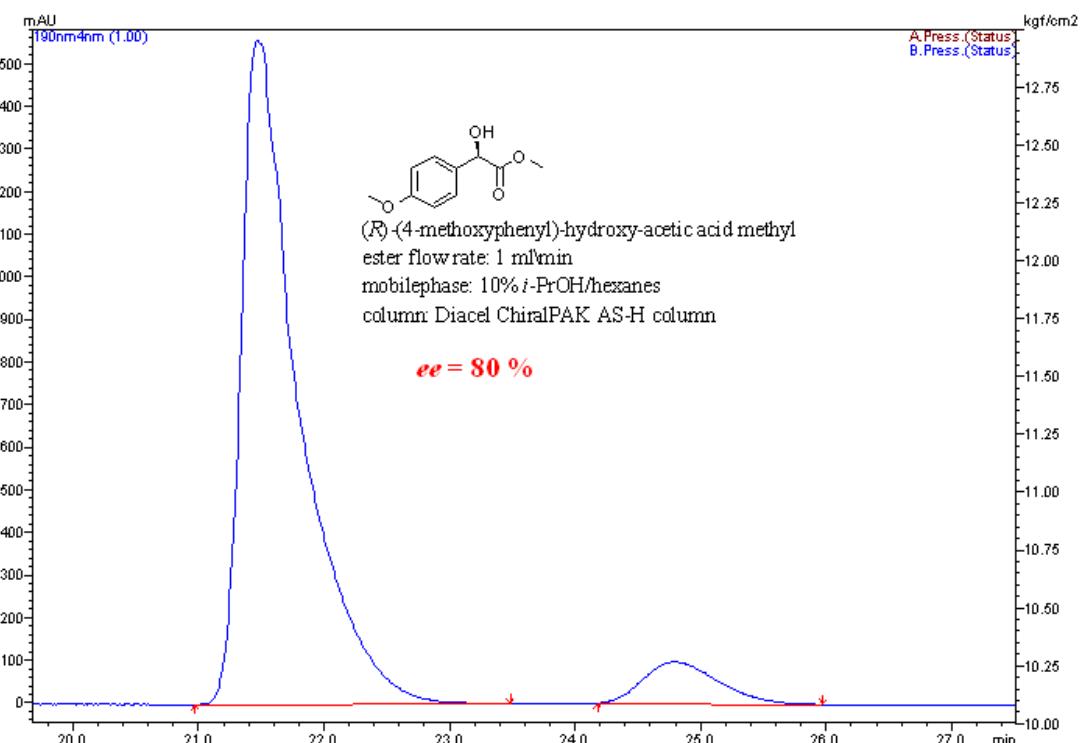
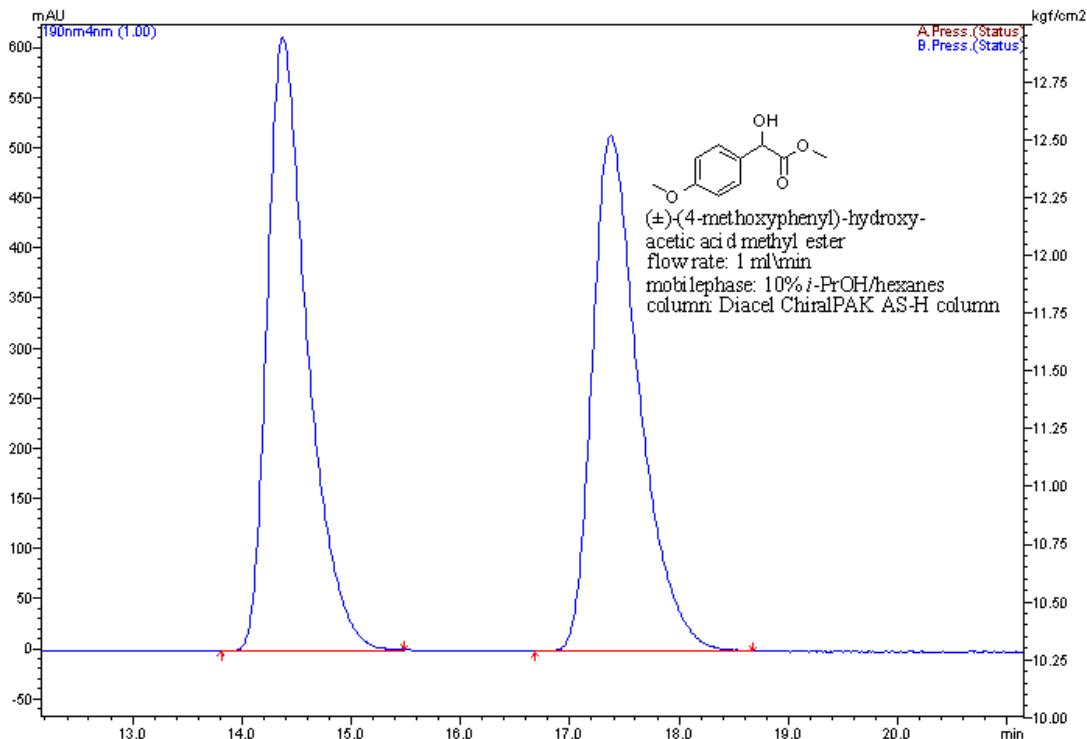
p-Iodophenyl-hydroxy-acetic acid methyl ester



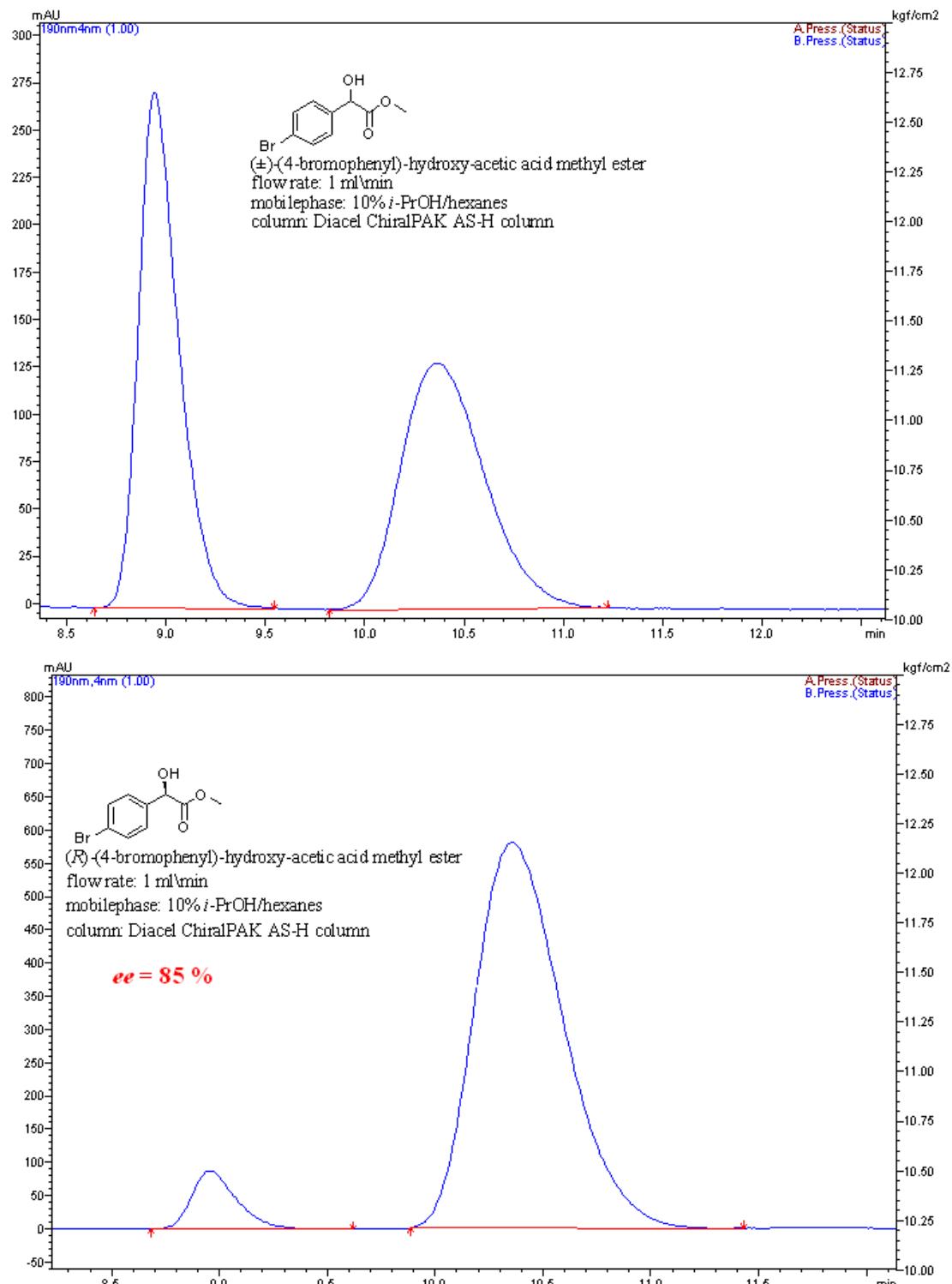
Hydroxy-*p*-tolyl-acetic acid methyl ester



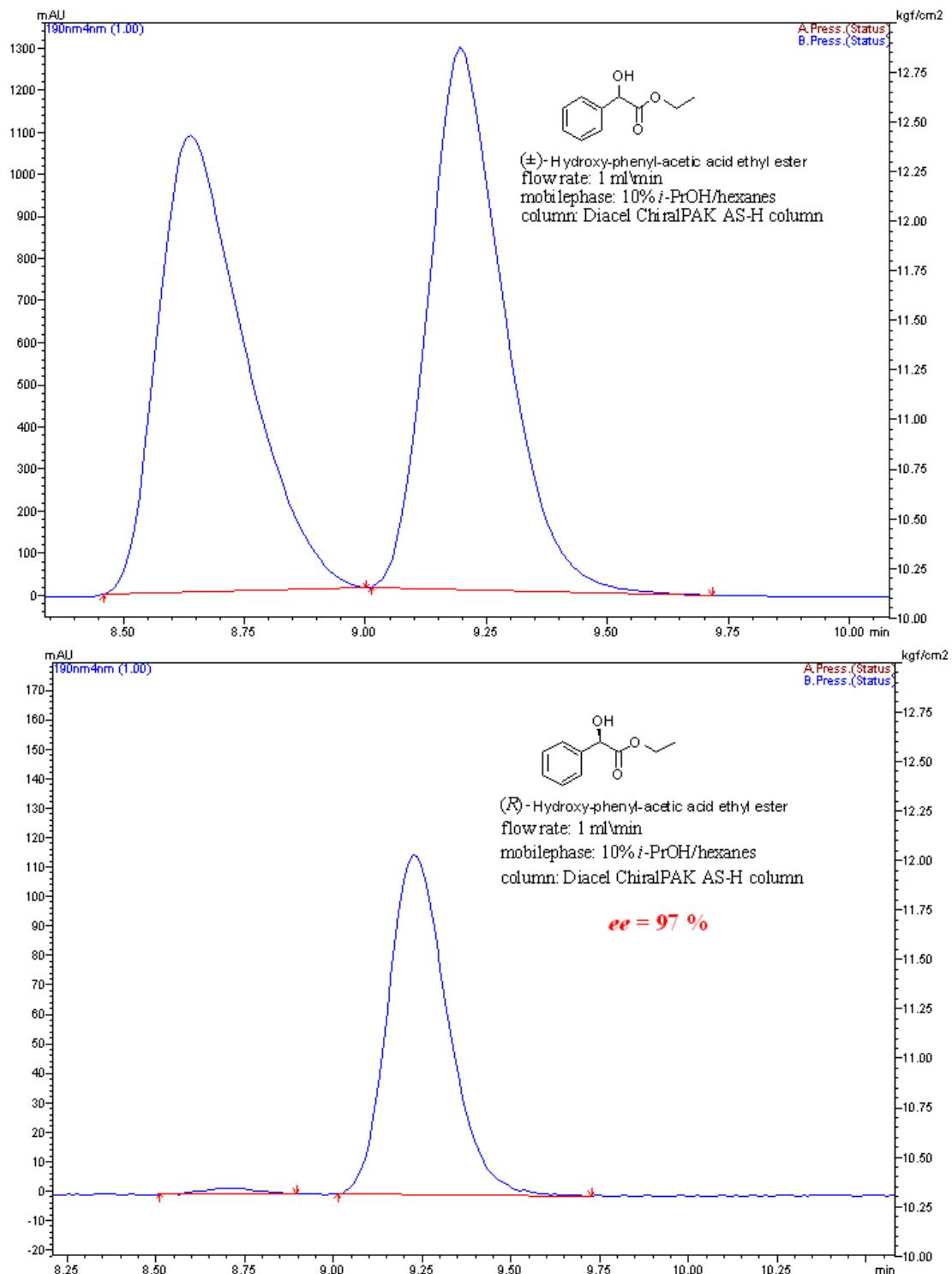
p-Methoxyphenyl-hydroxy-acetic acid methyl



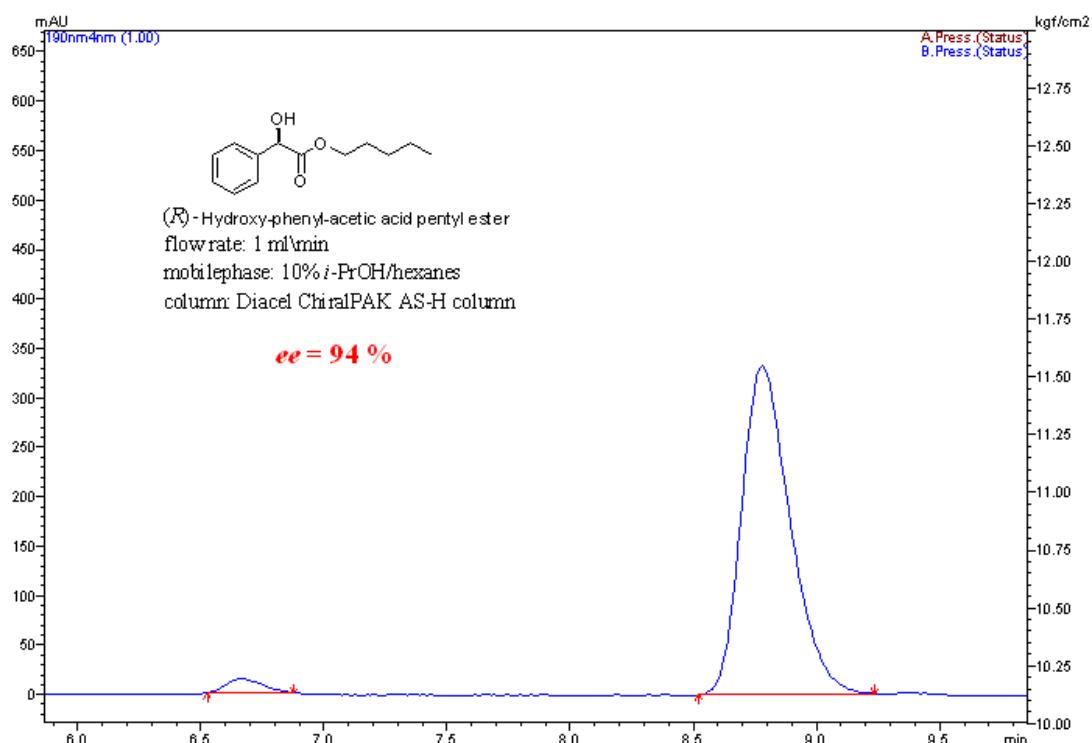
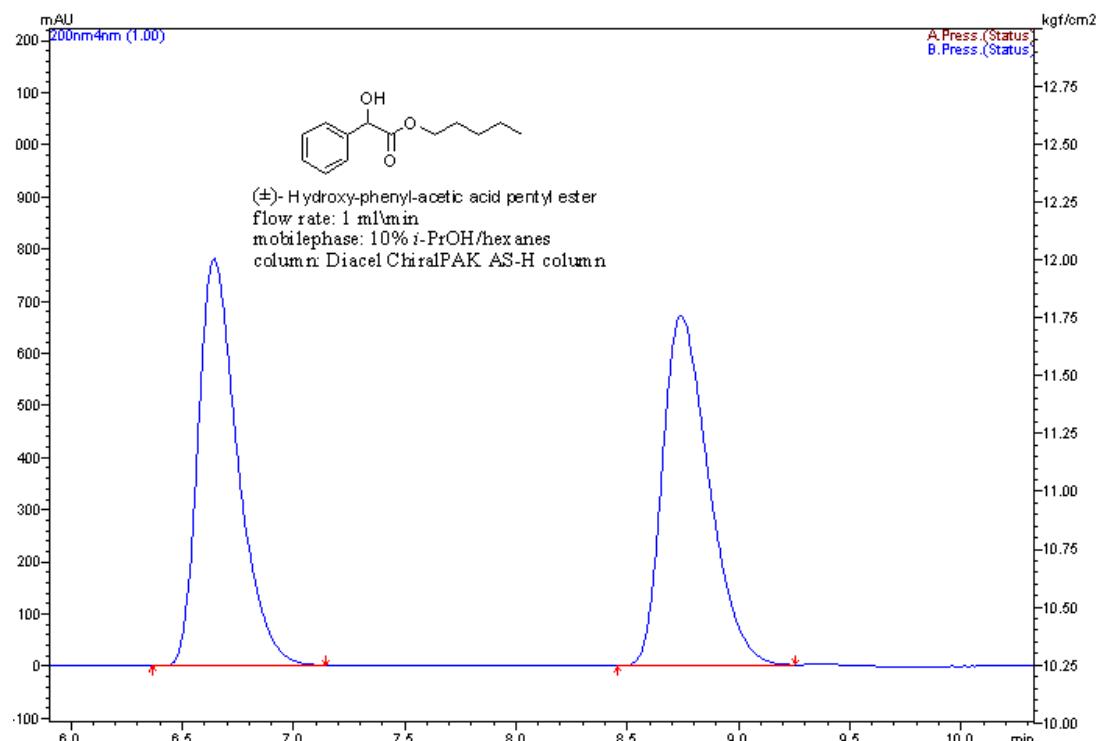
p-Bromophenyl-hydroxy-acetic acid methyl ester



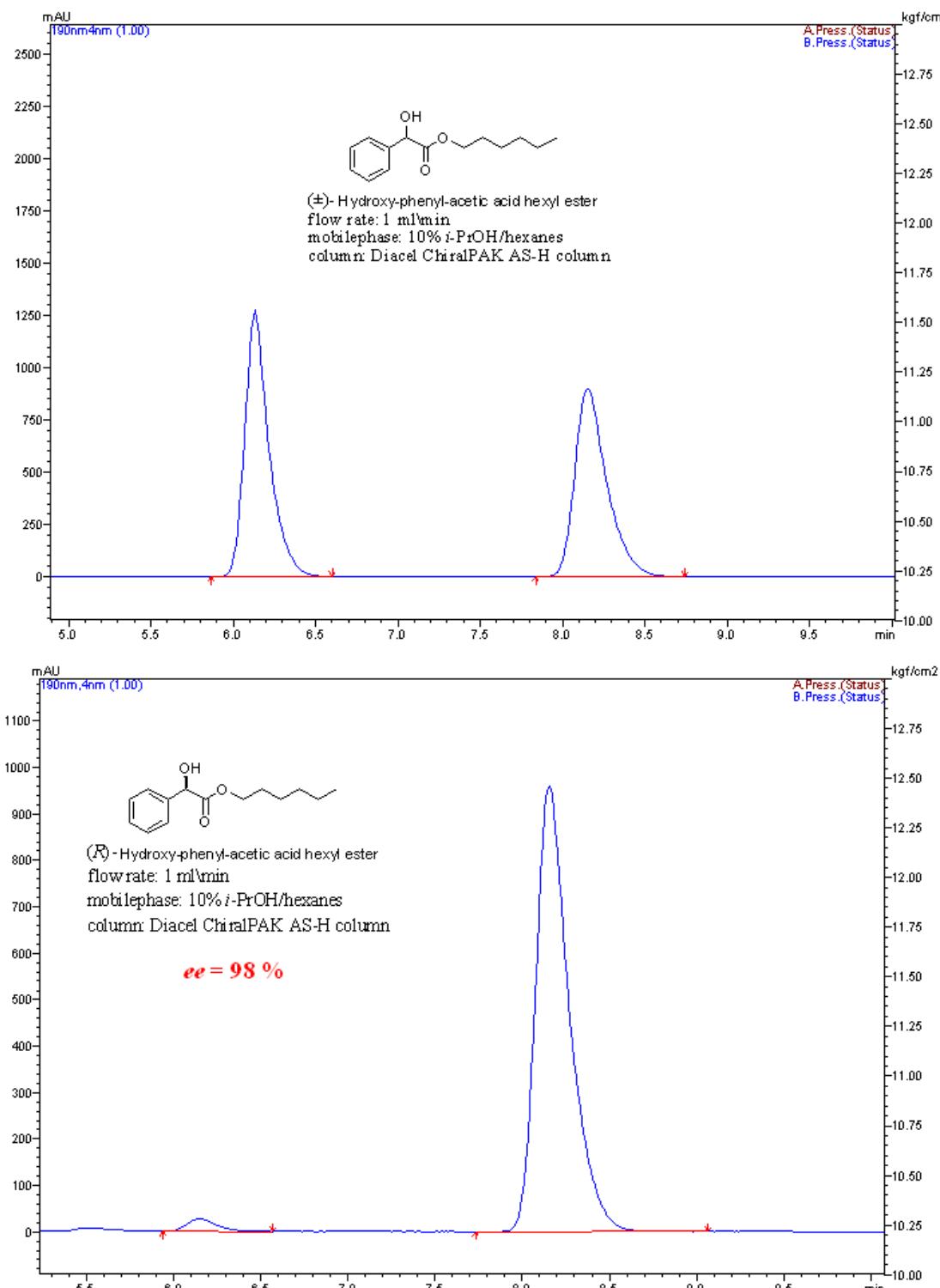
Hydroxy-phenyl-acetic acid ethyl ester



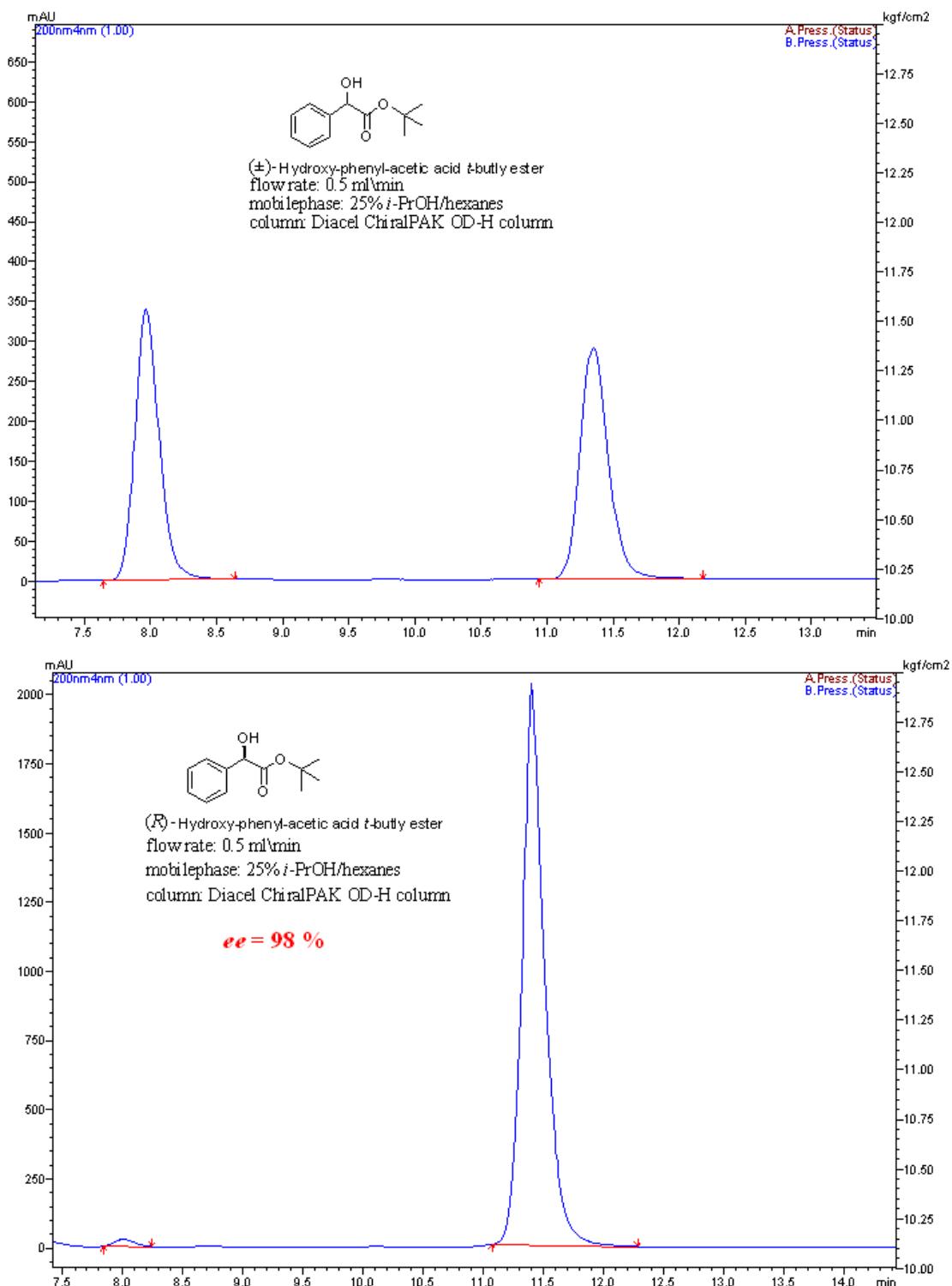
Hydroxy-phenyl-acetic acid *n*-pentyl ester

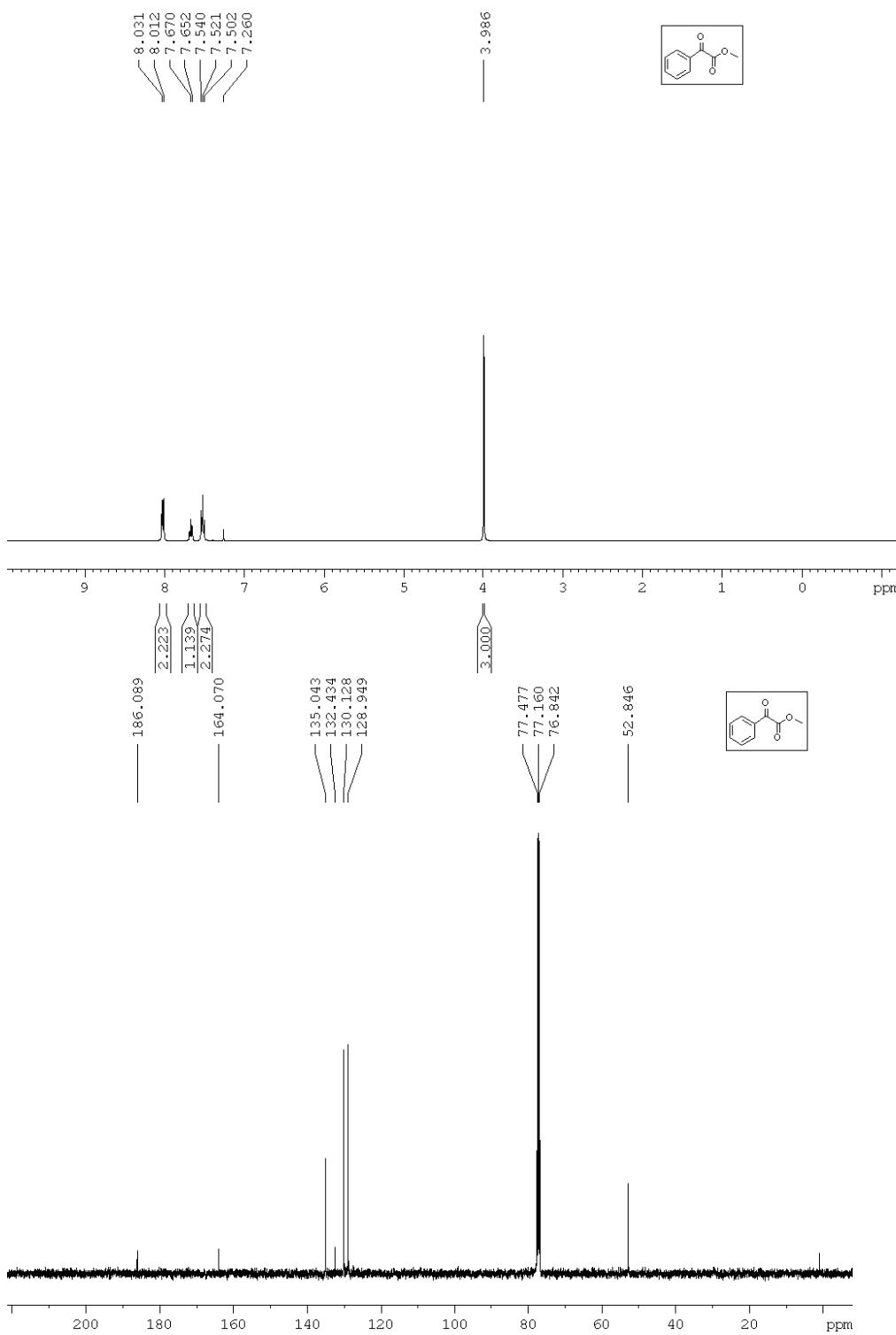


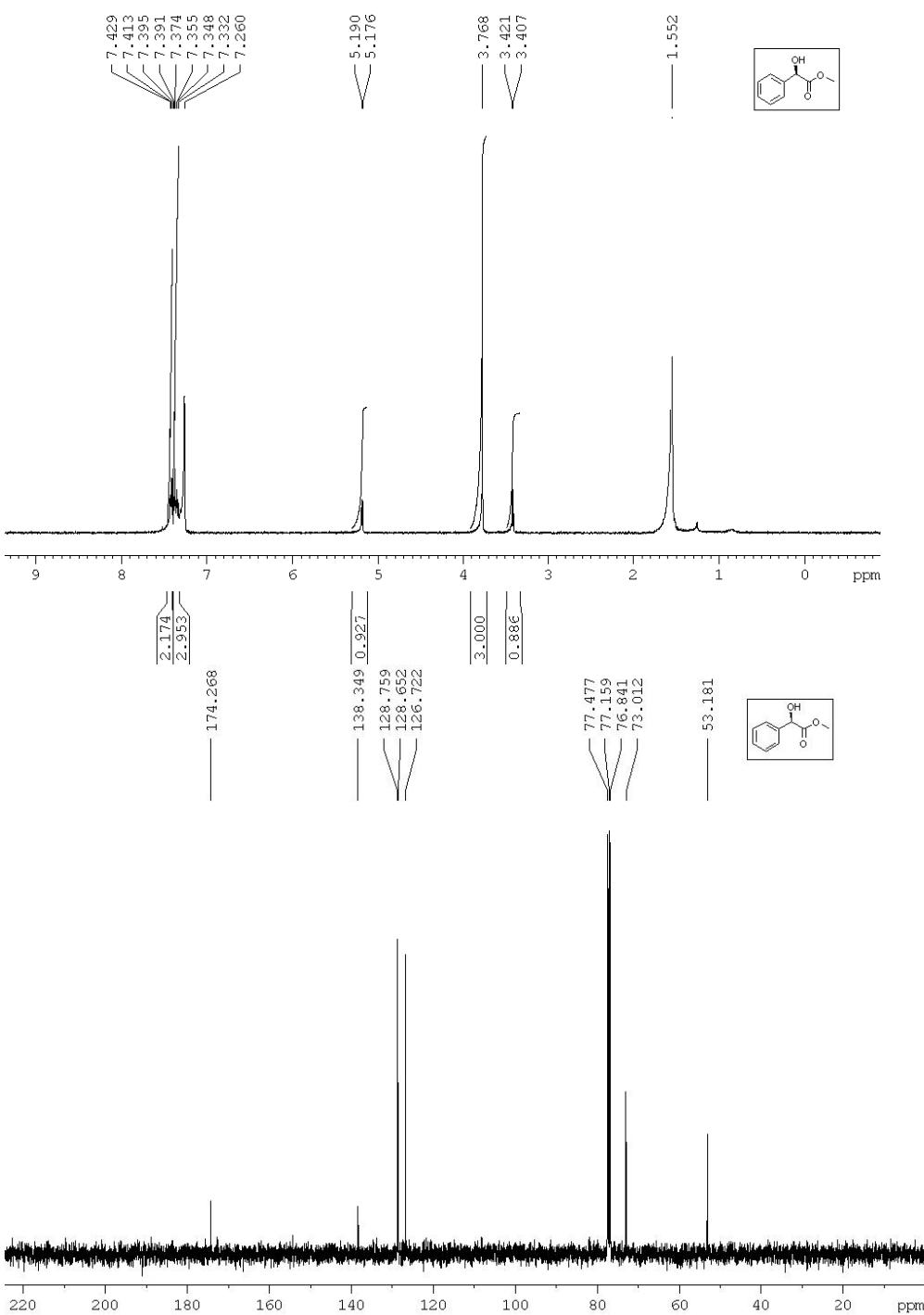
Hydroxy-phenyl-acetic acid *n*-hexyl ester

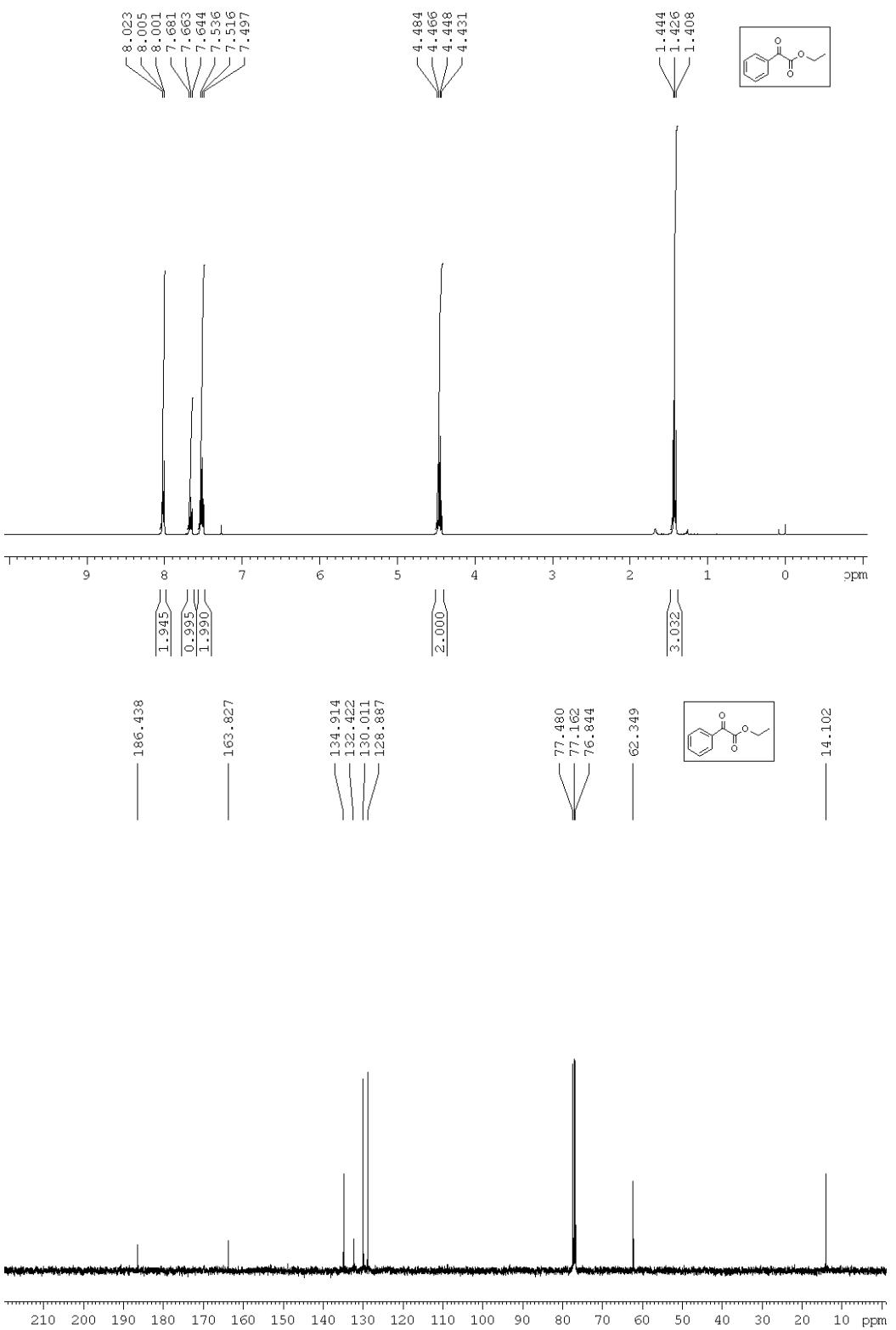


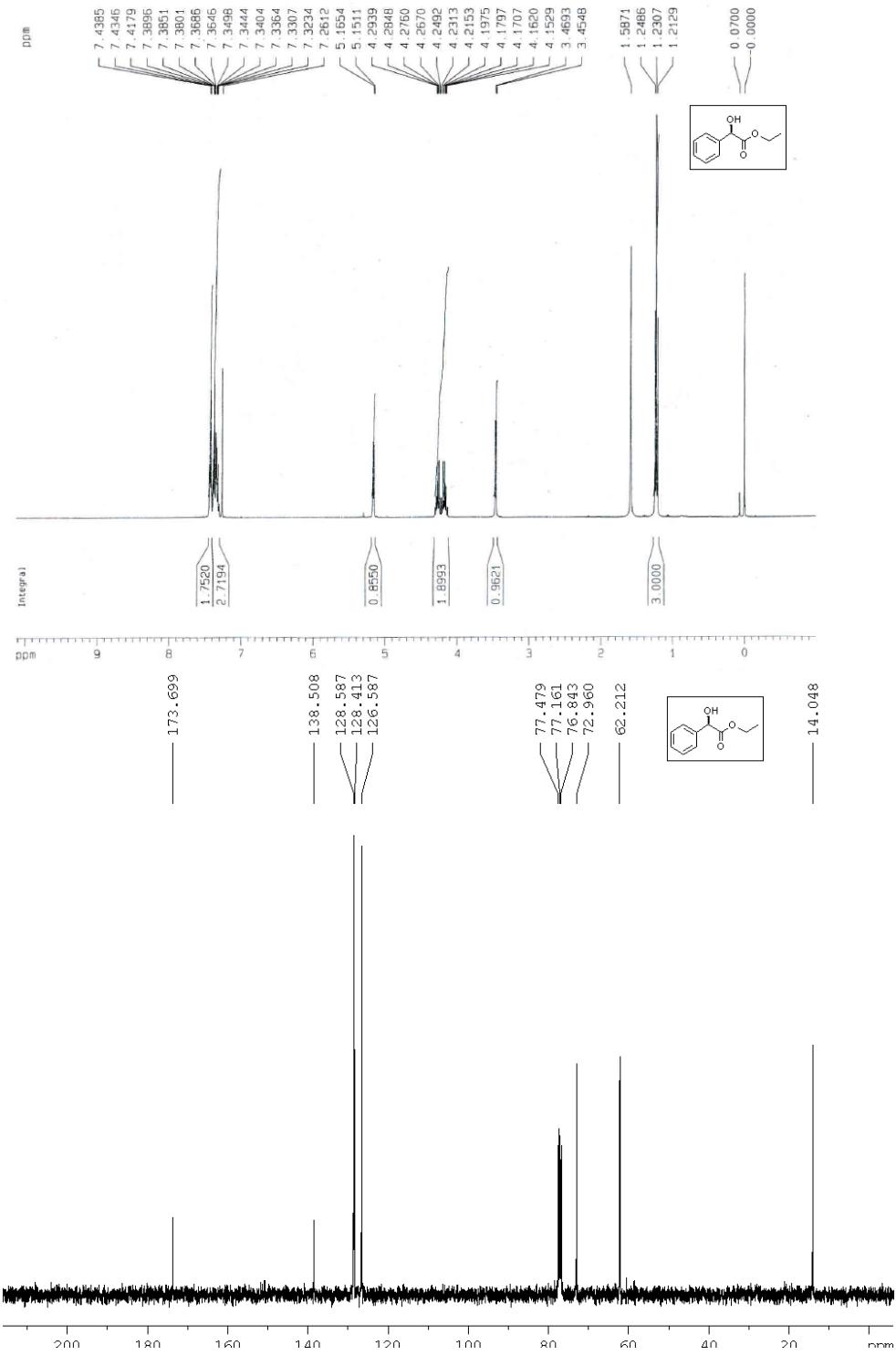
Hydroxy-phenyl-acetic acid *t*-butyl ester

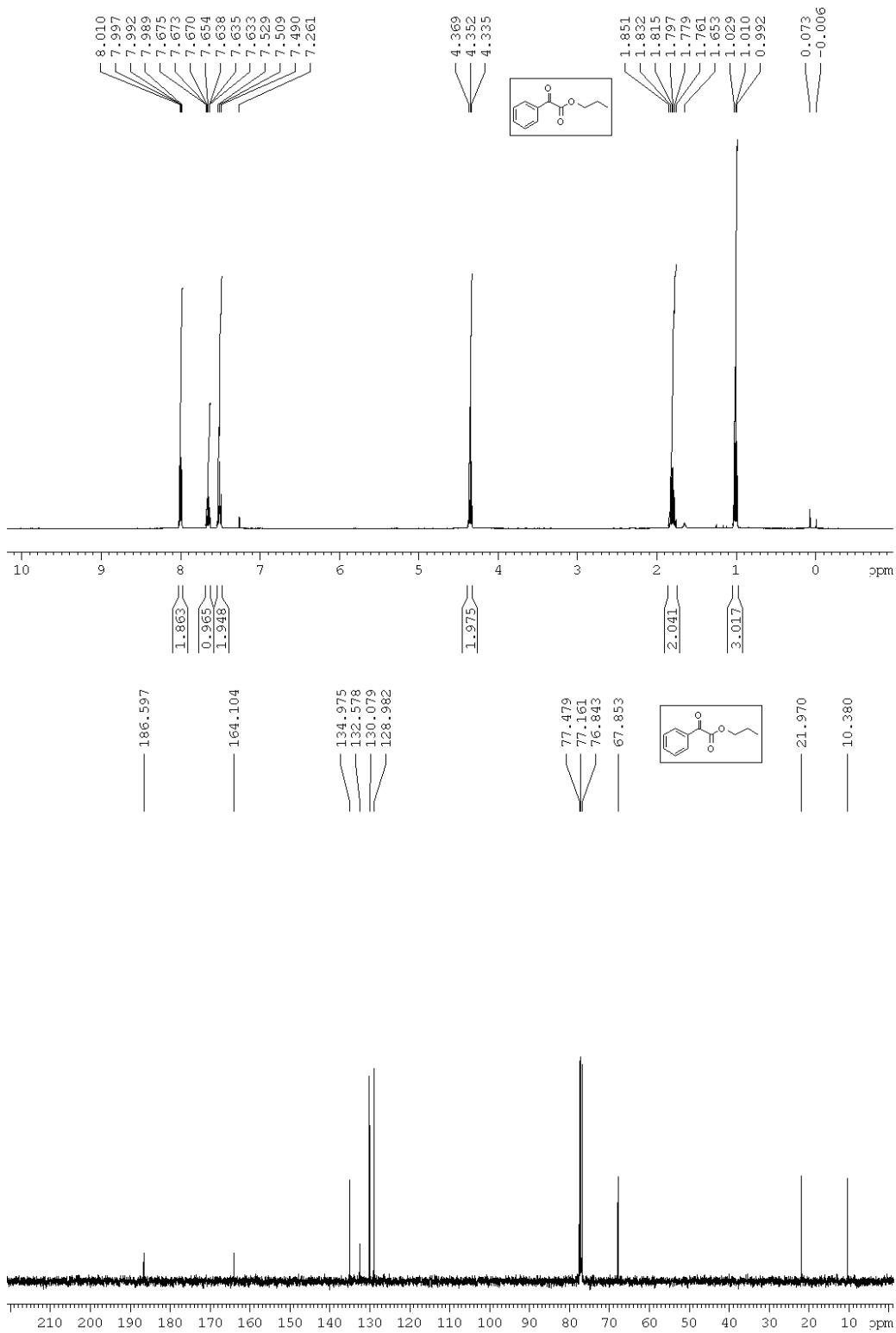


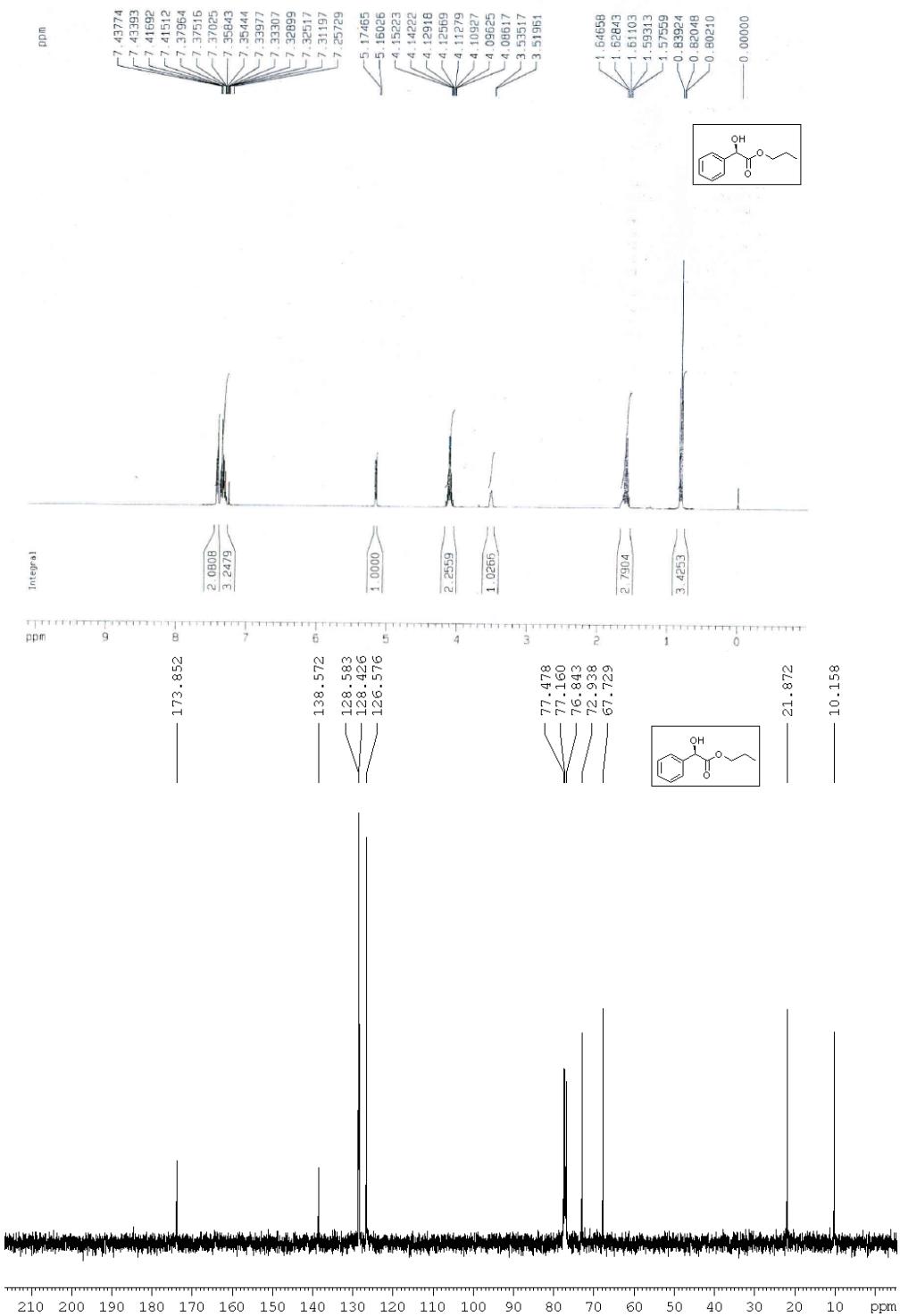


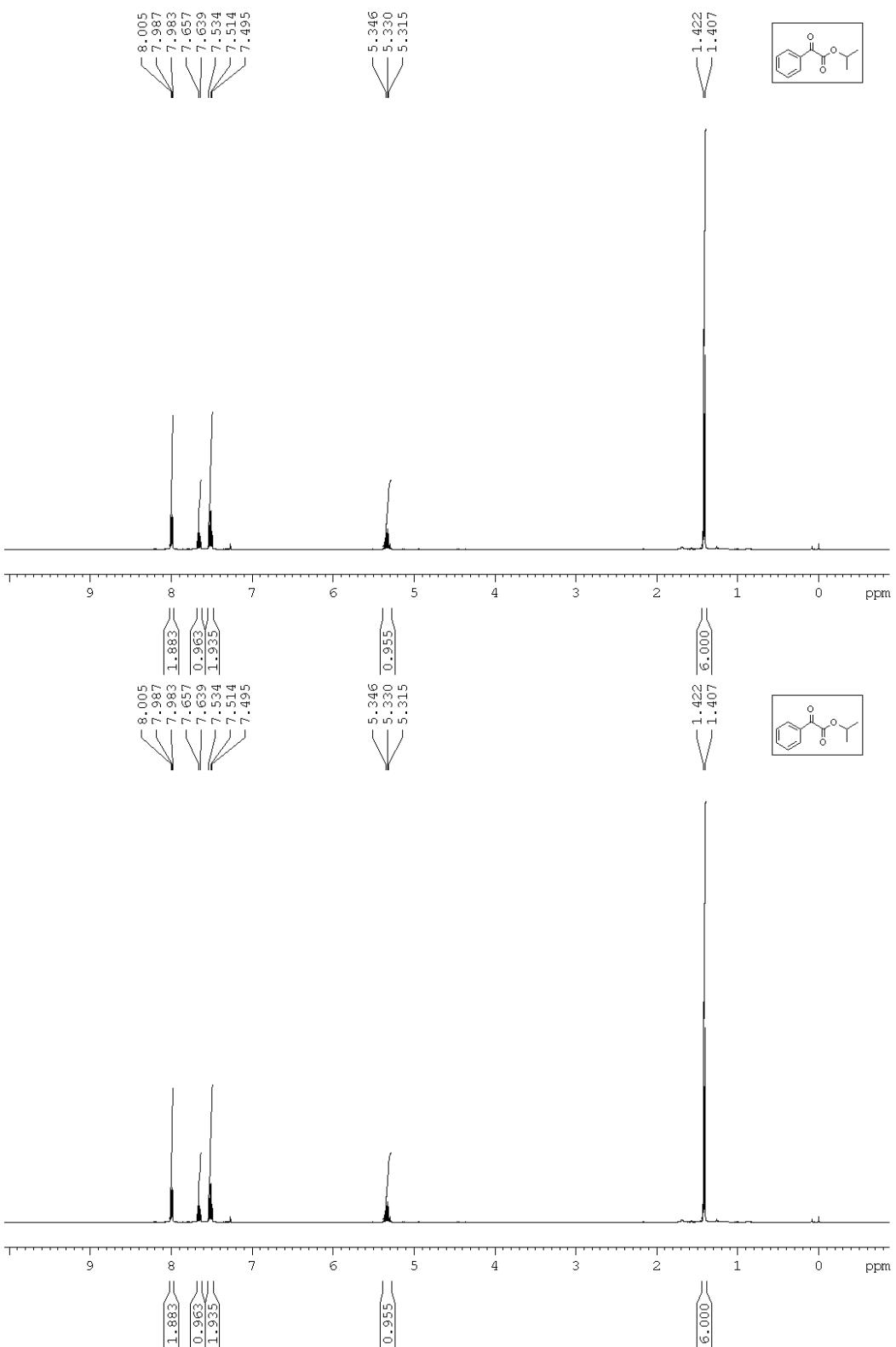


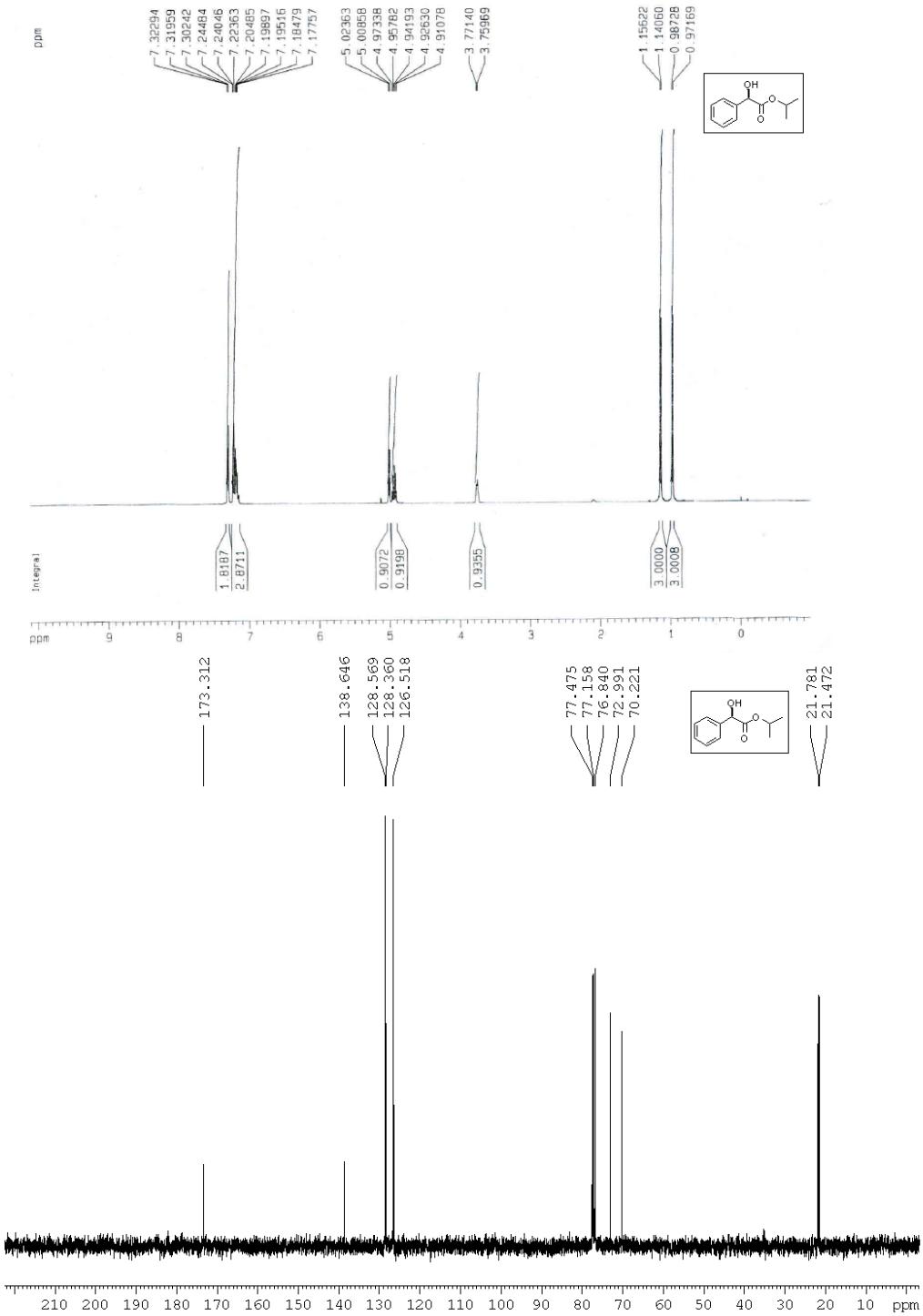


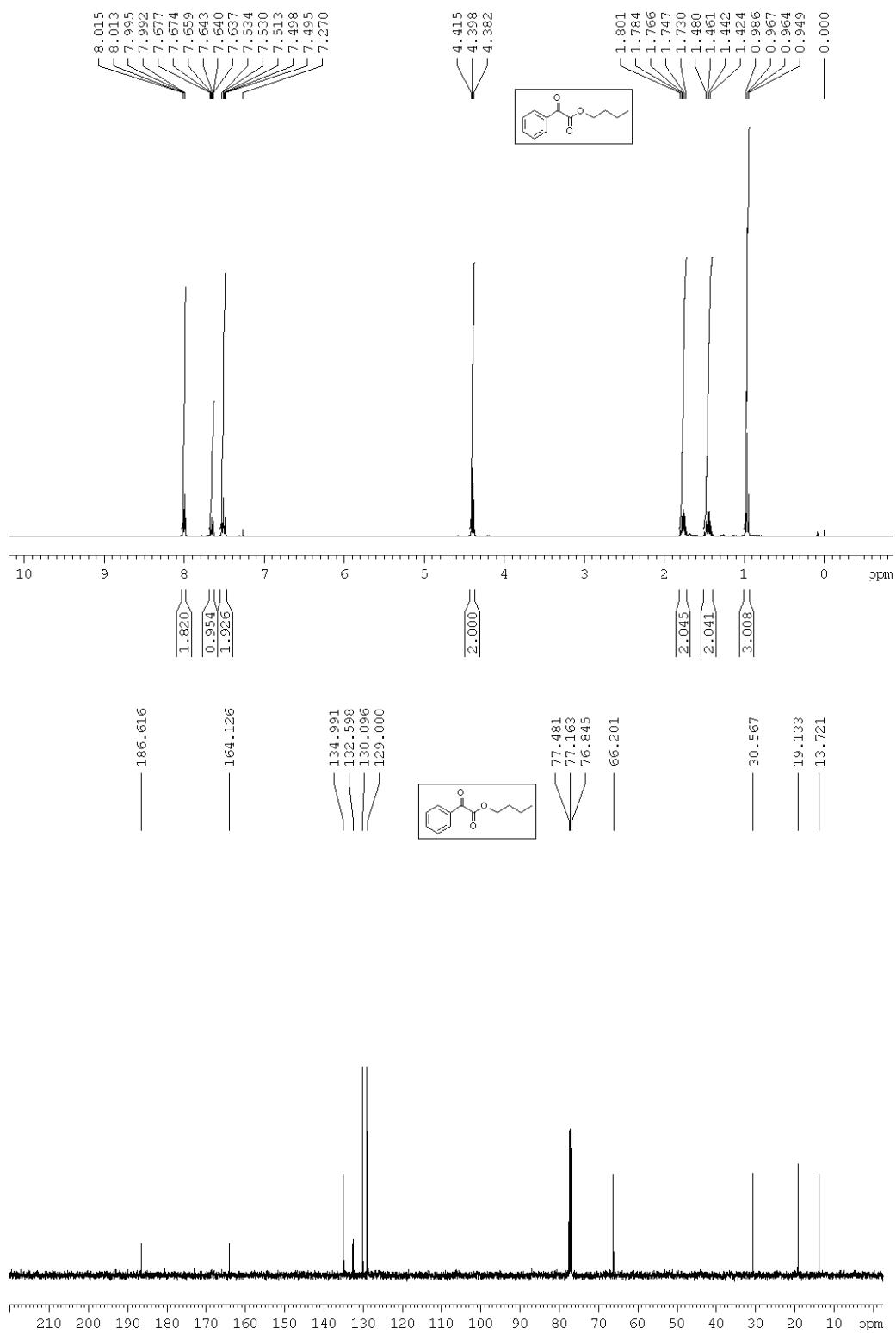


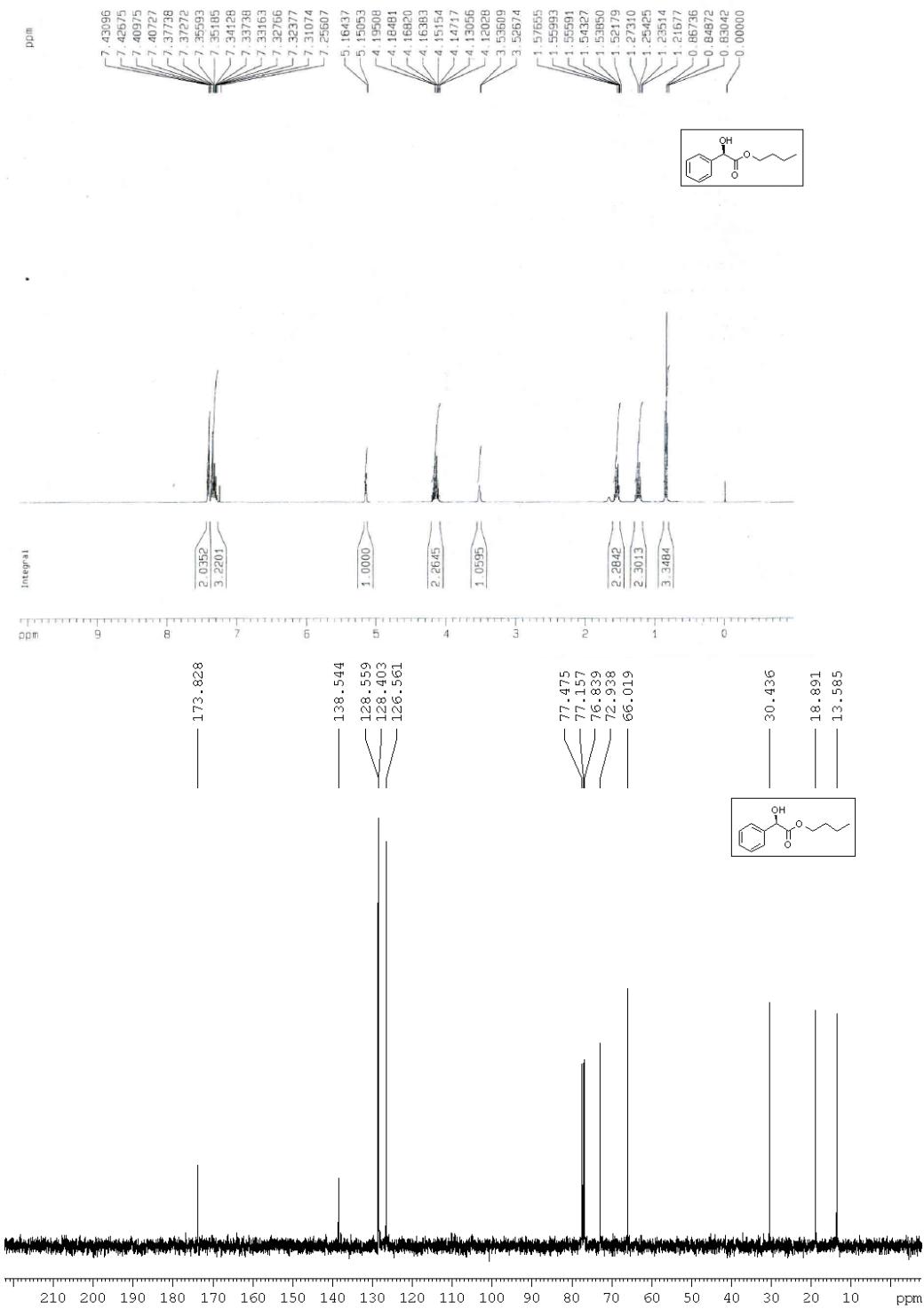


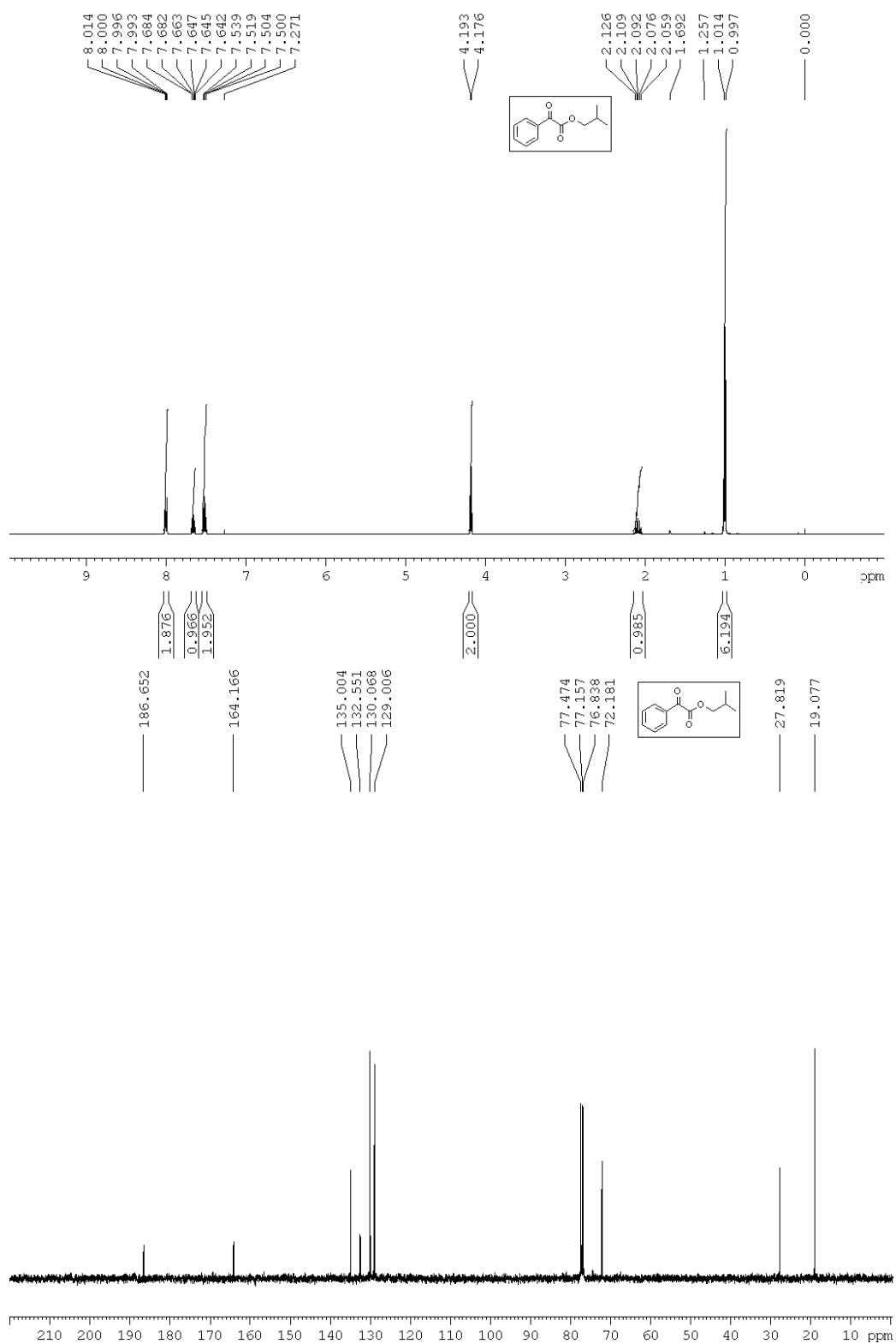


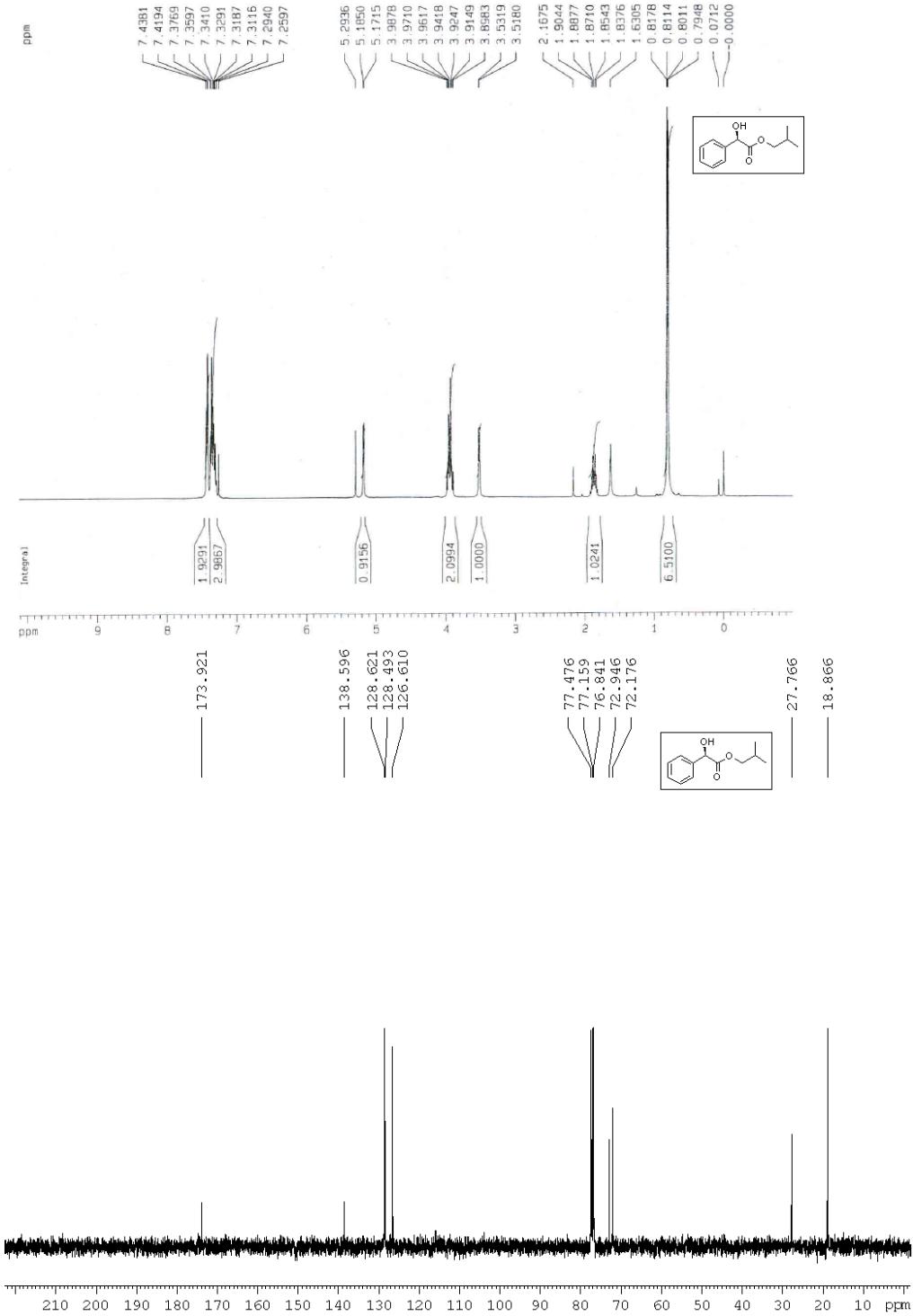


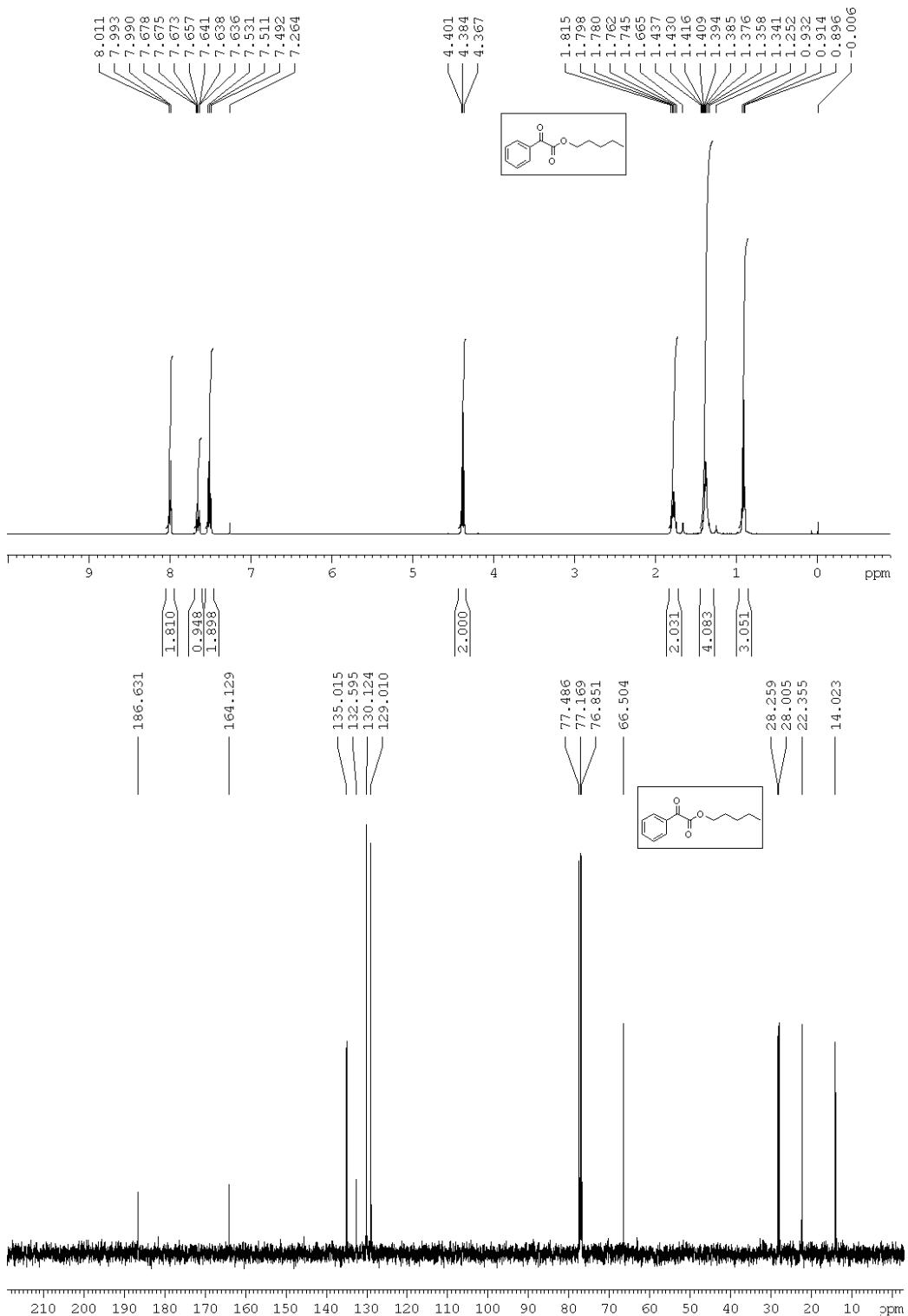


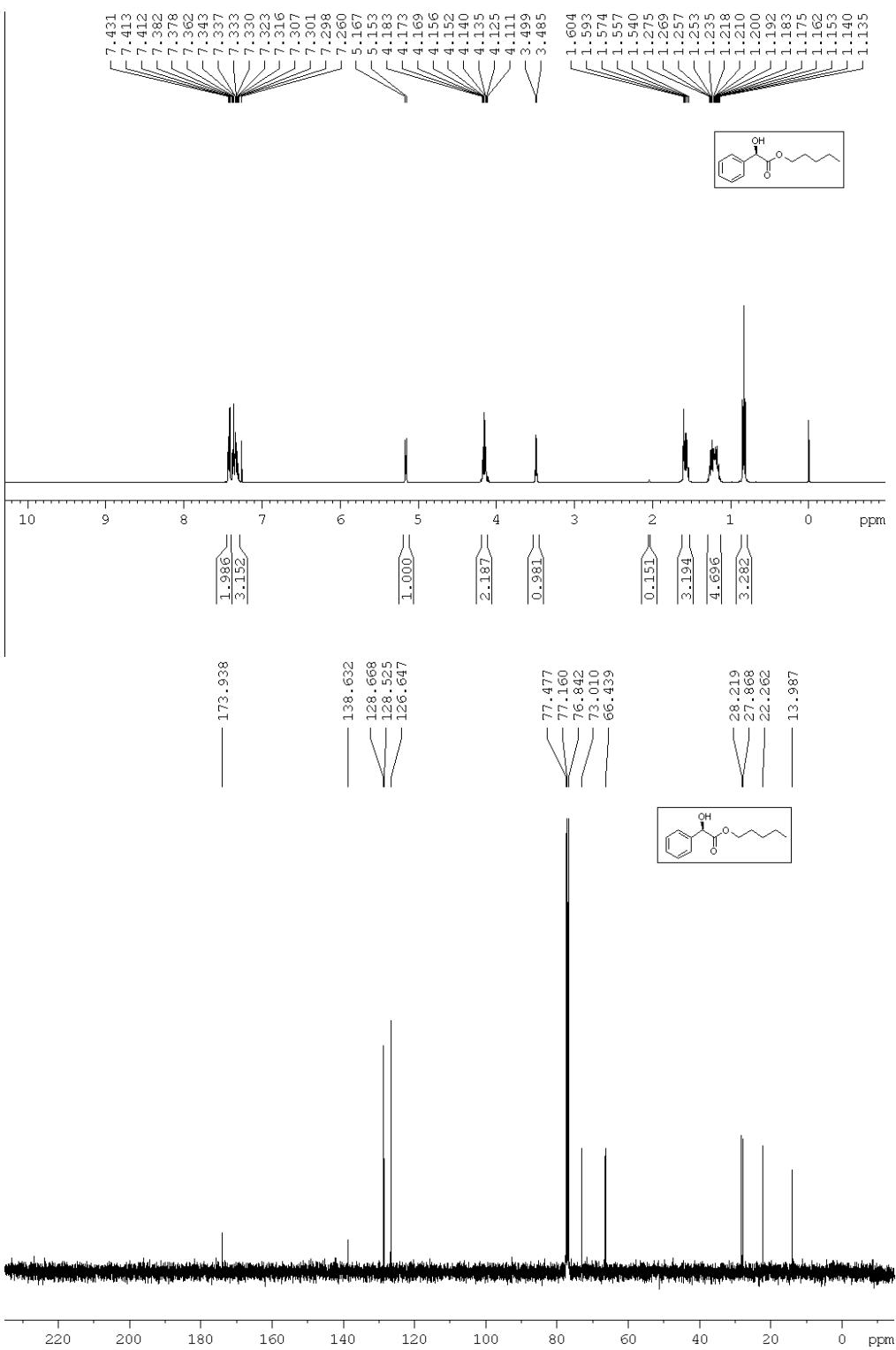


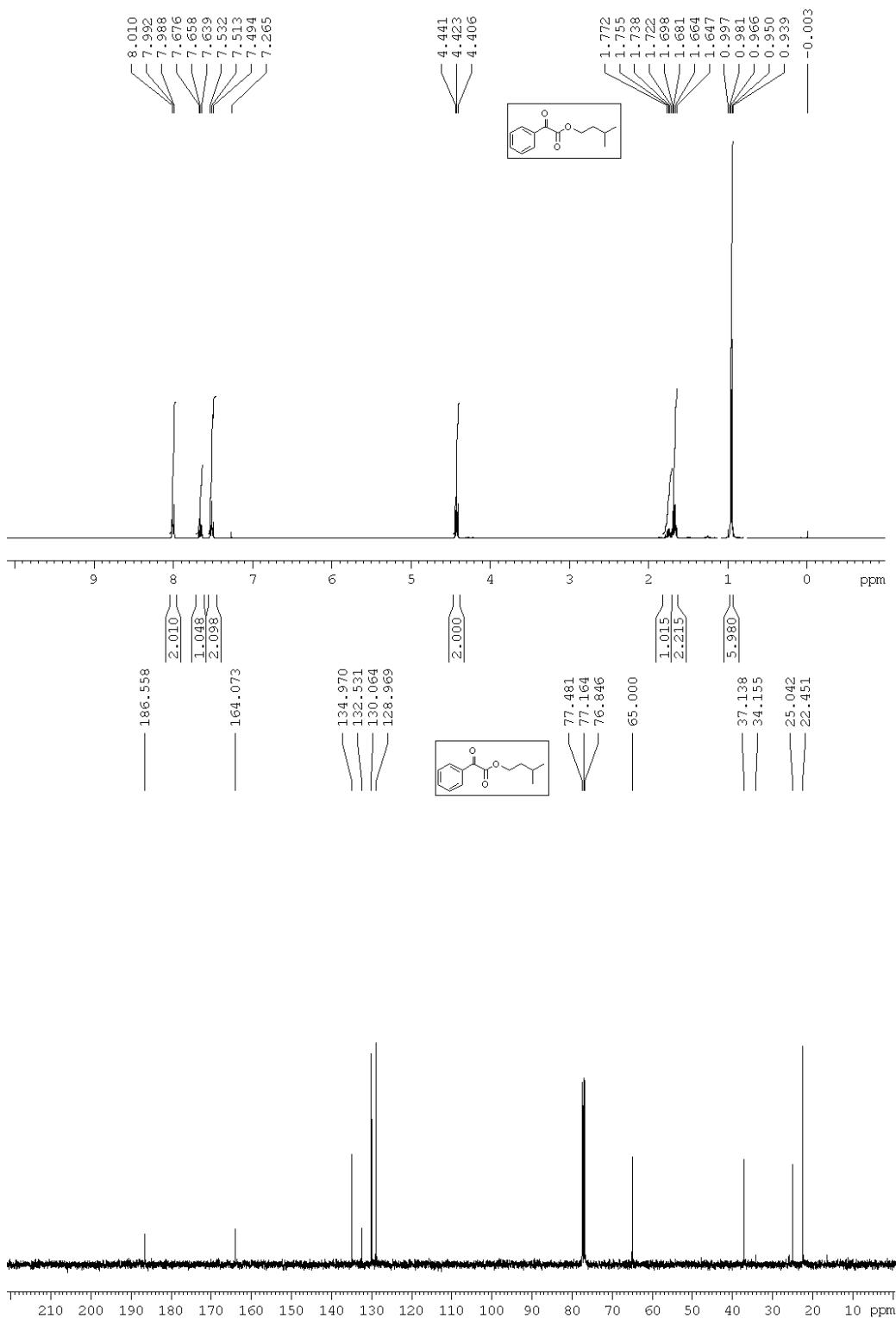


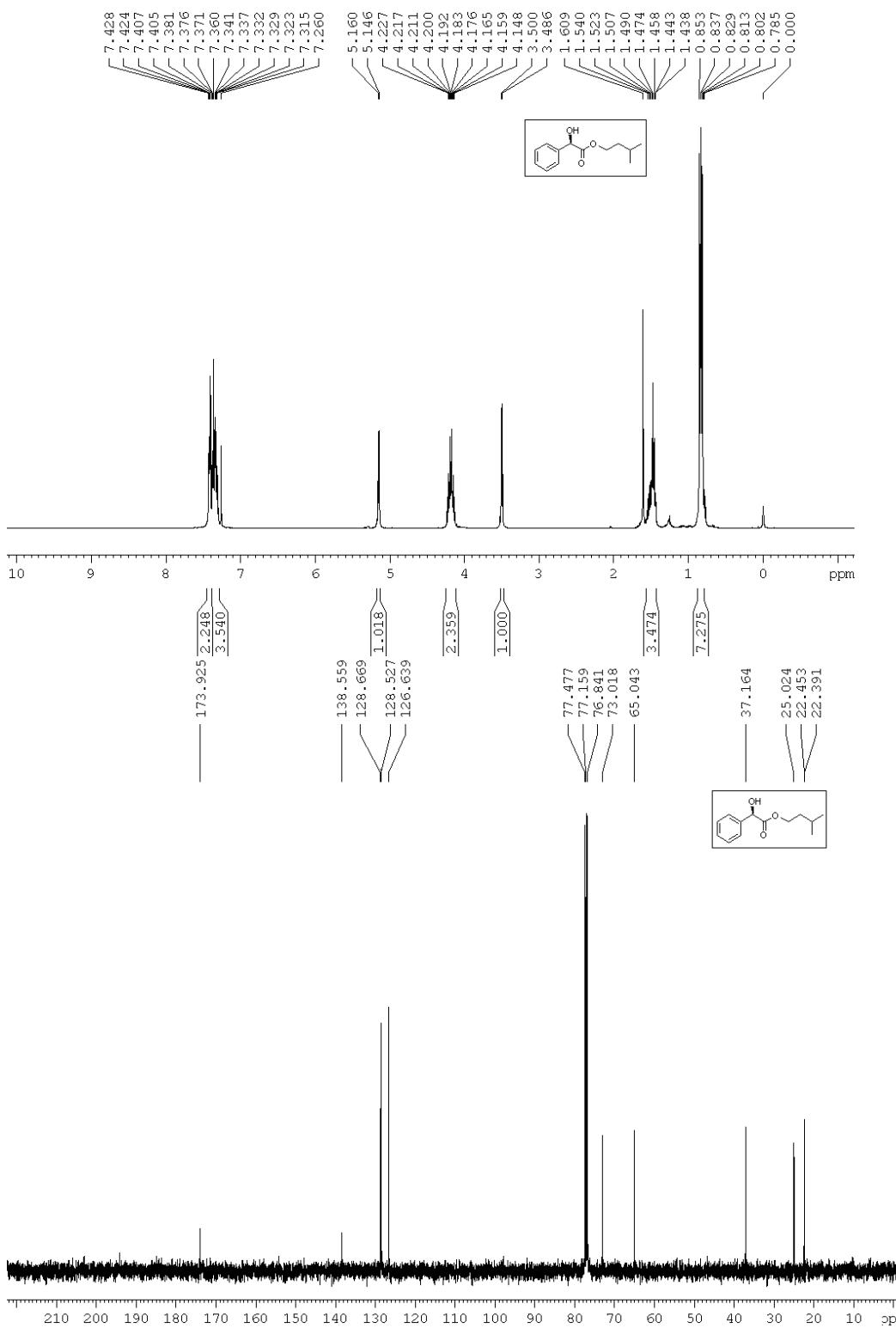


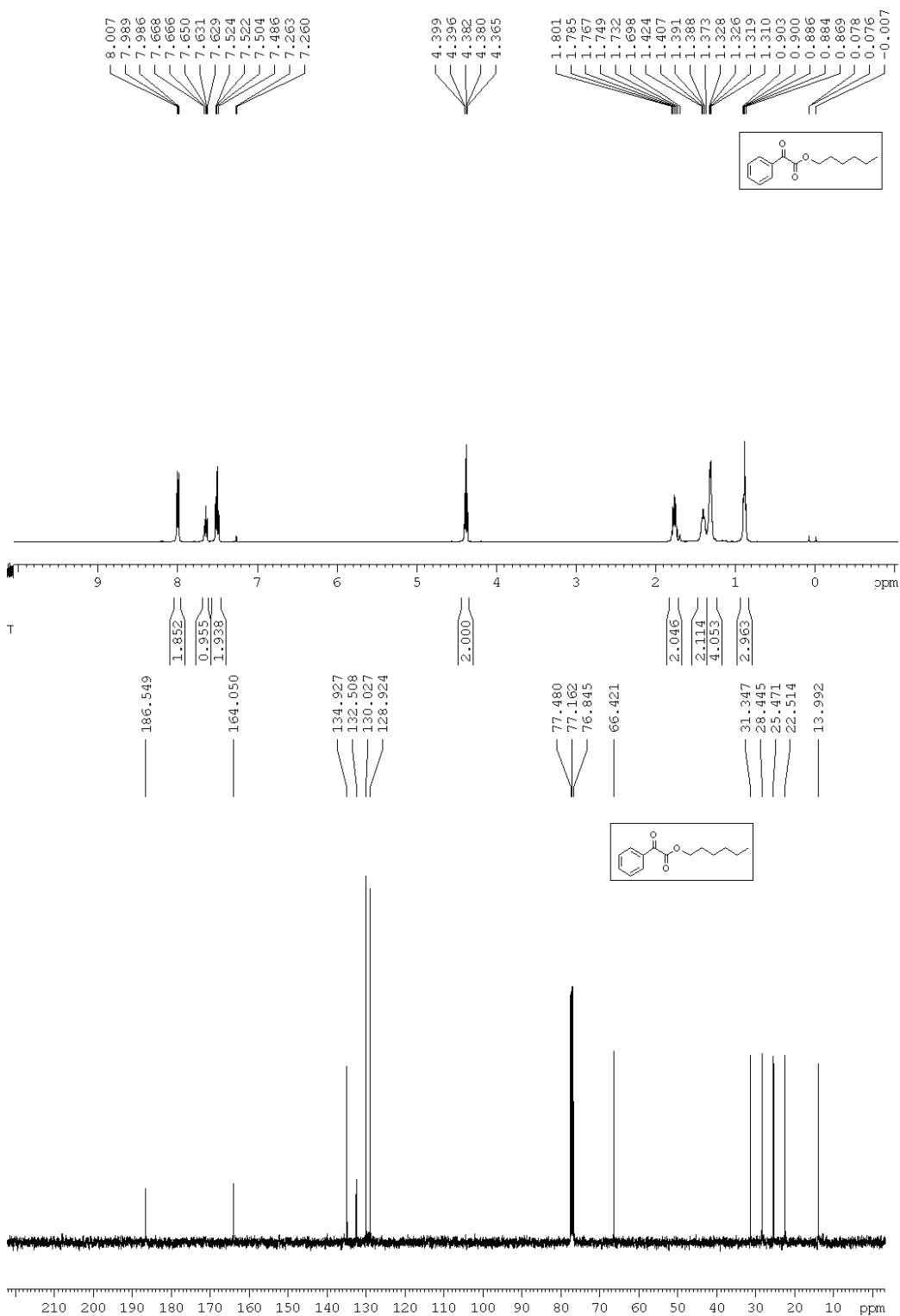


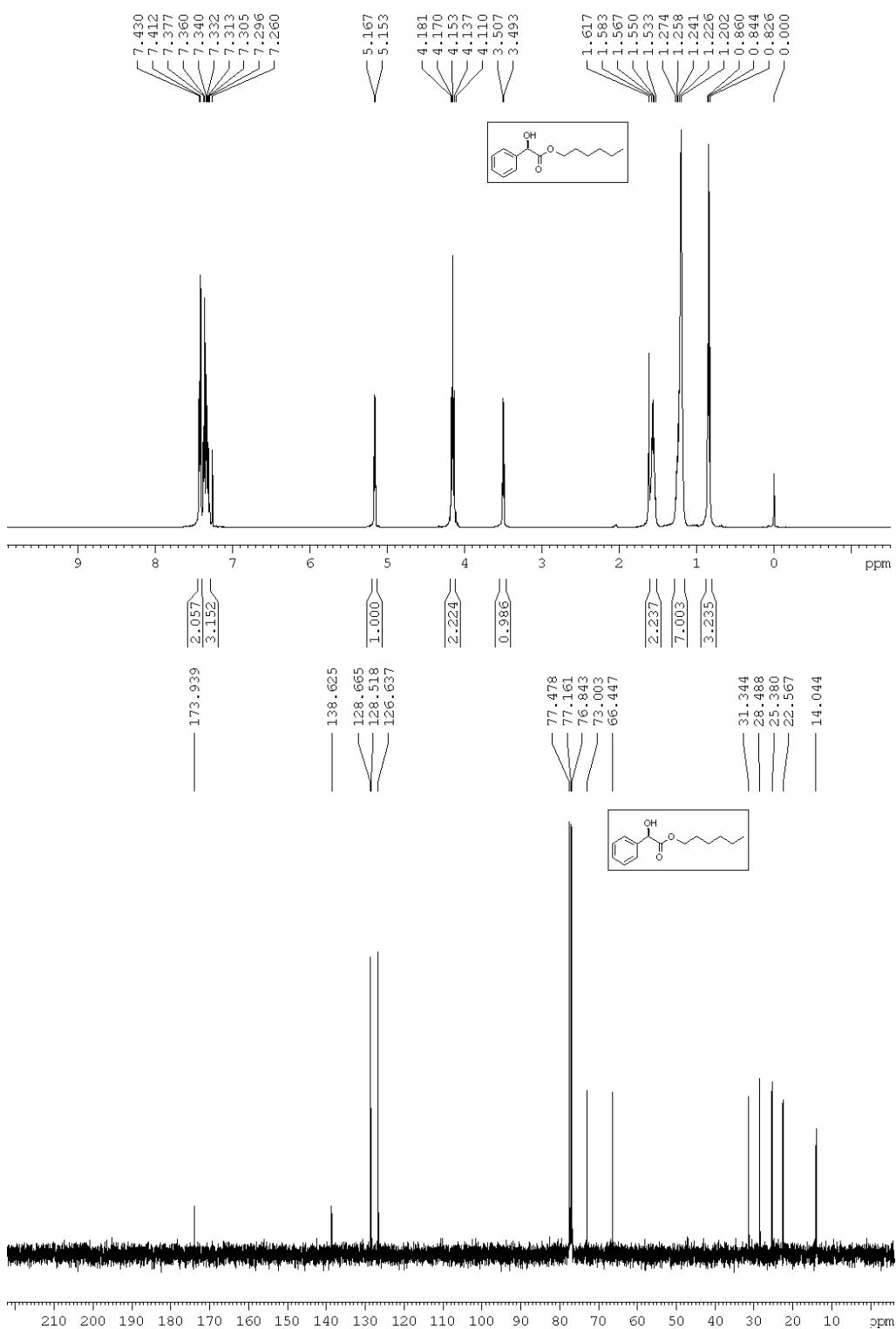


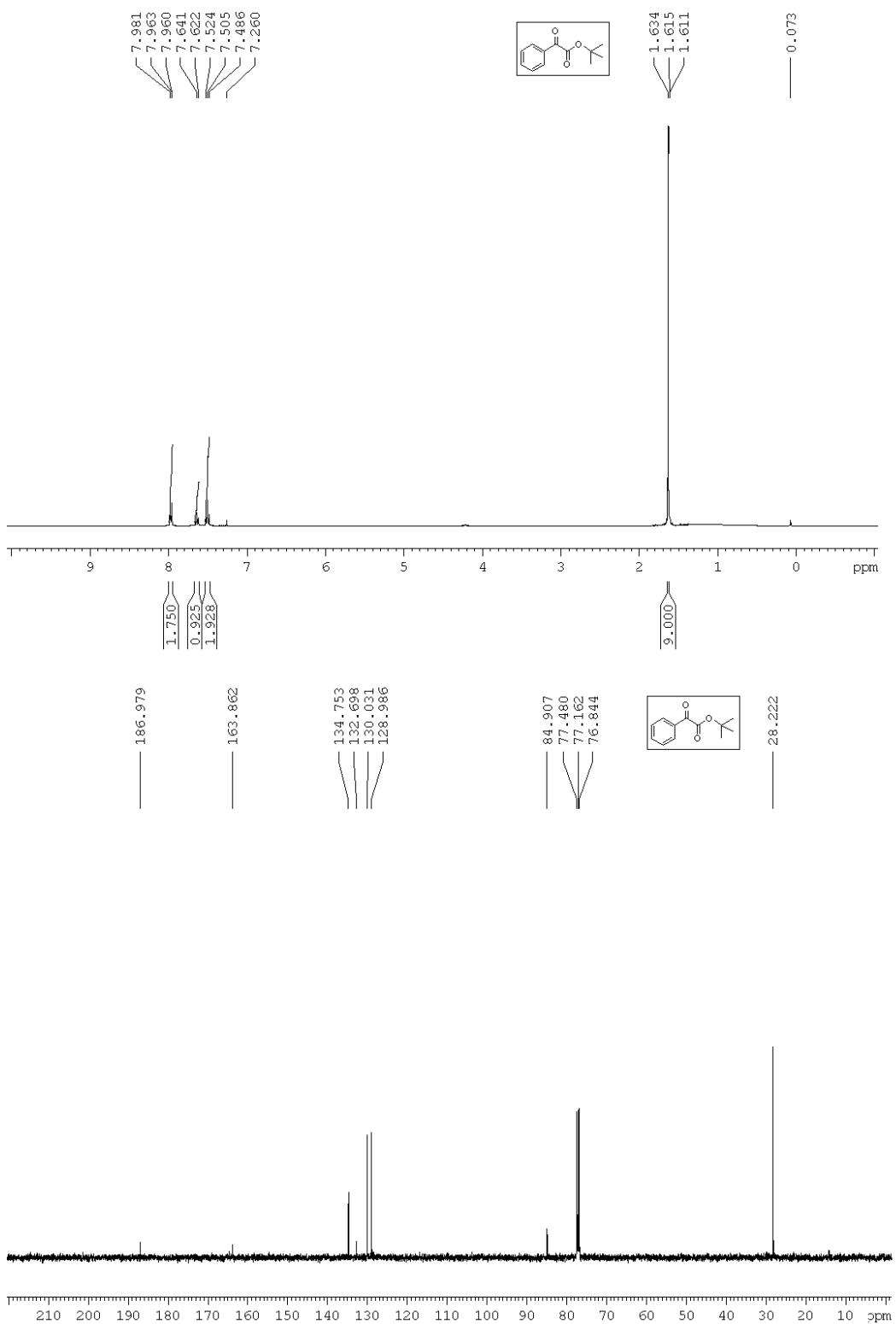


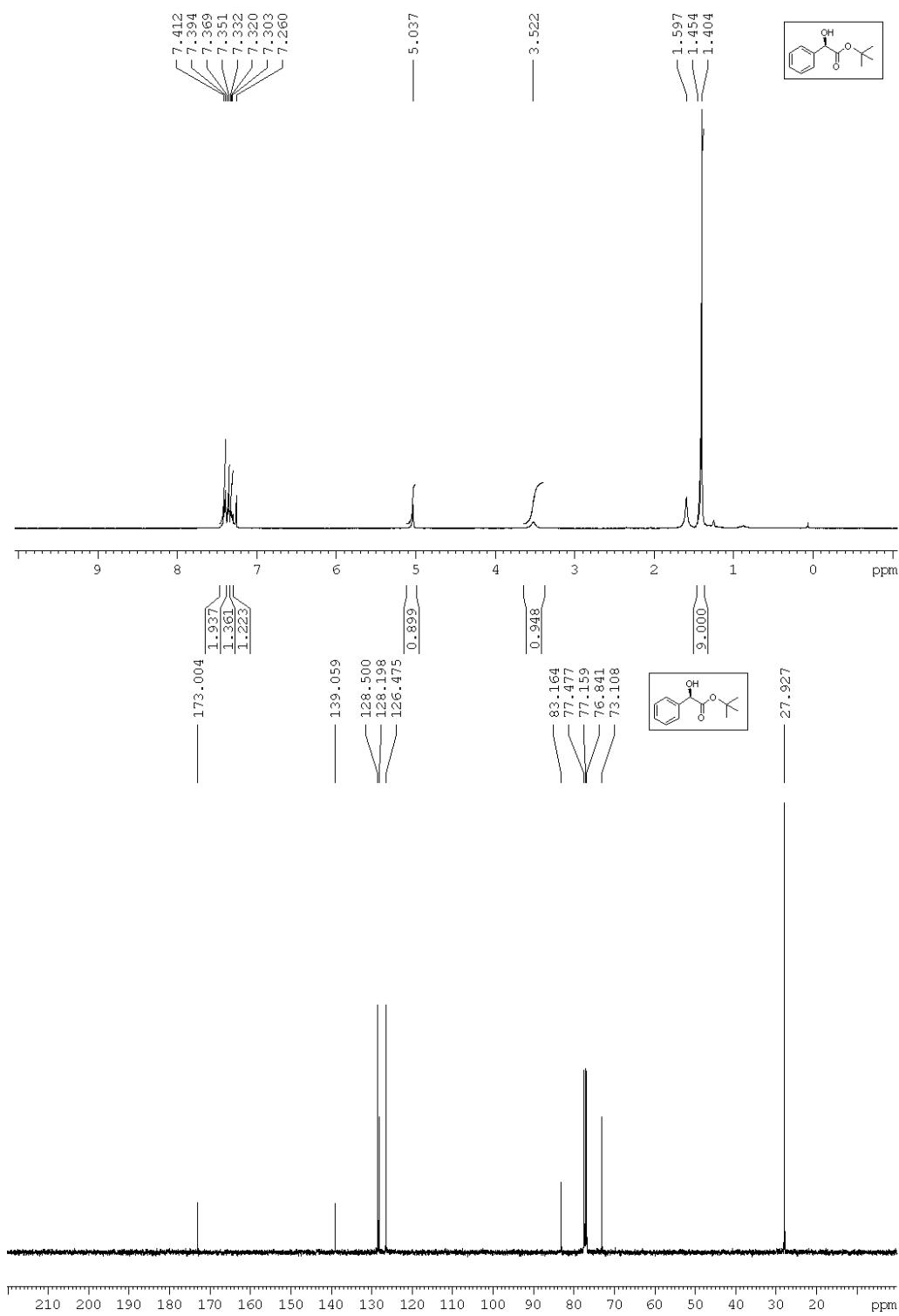


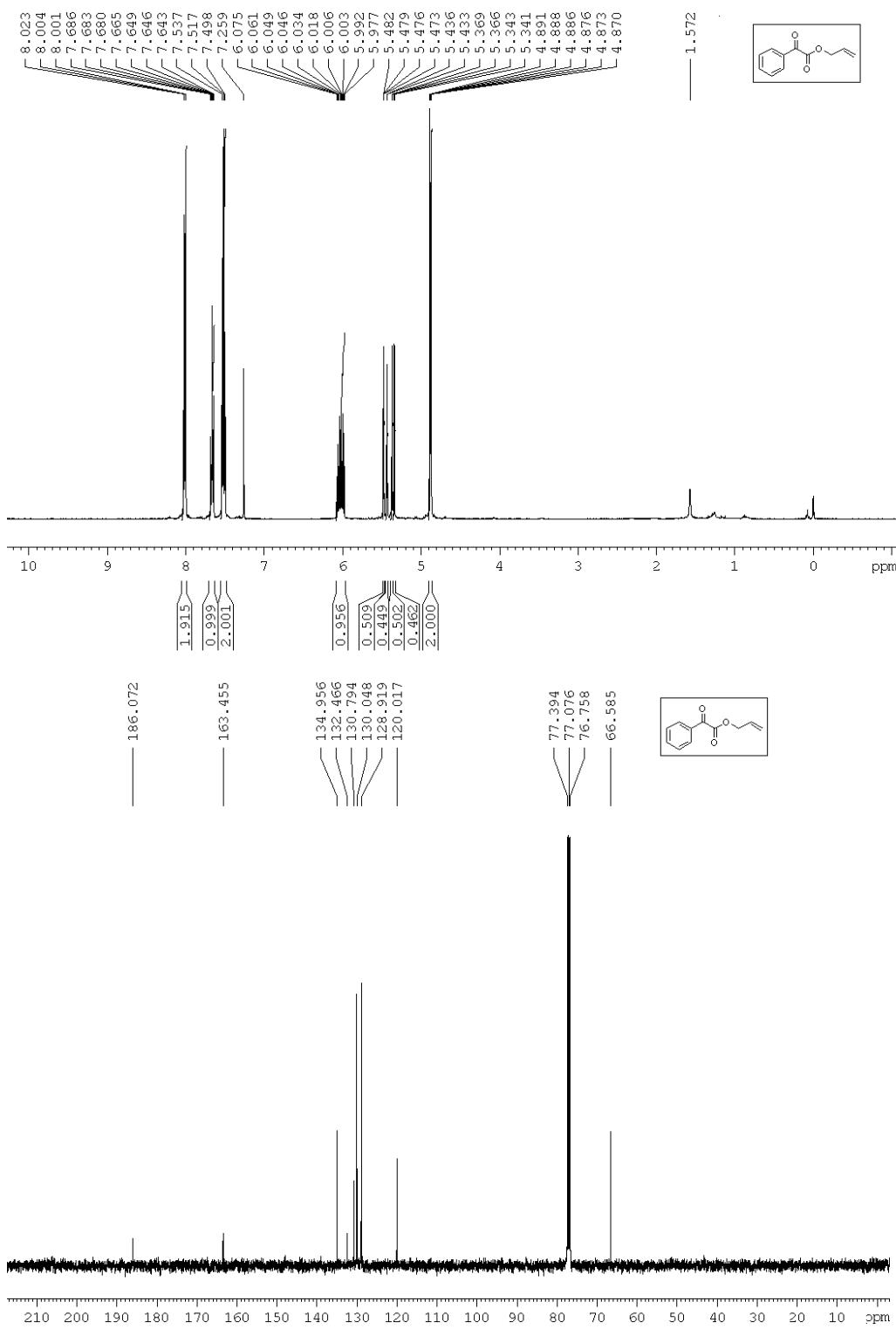


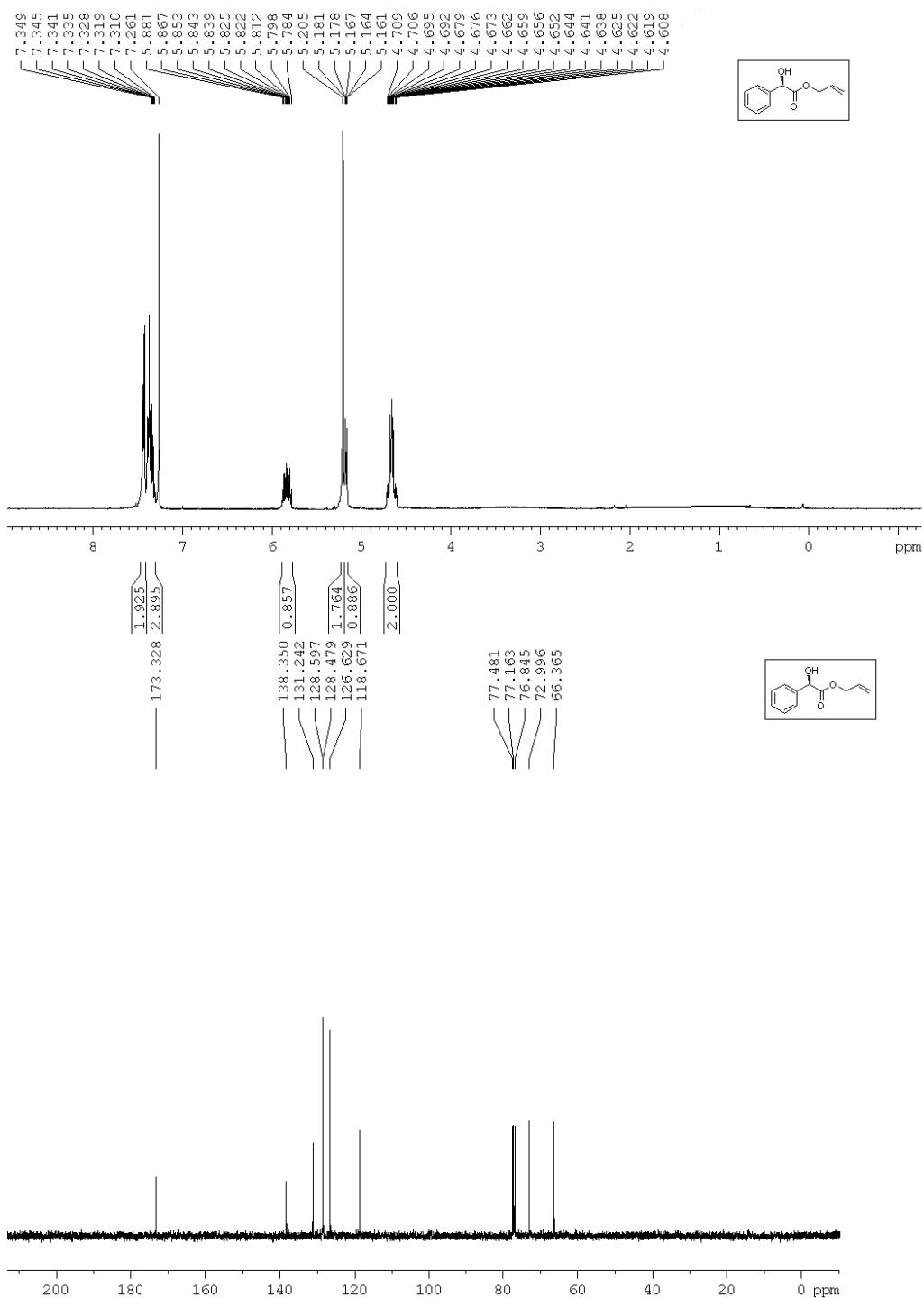


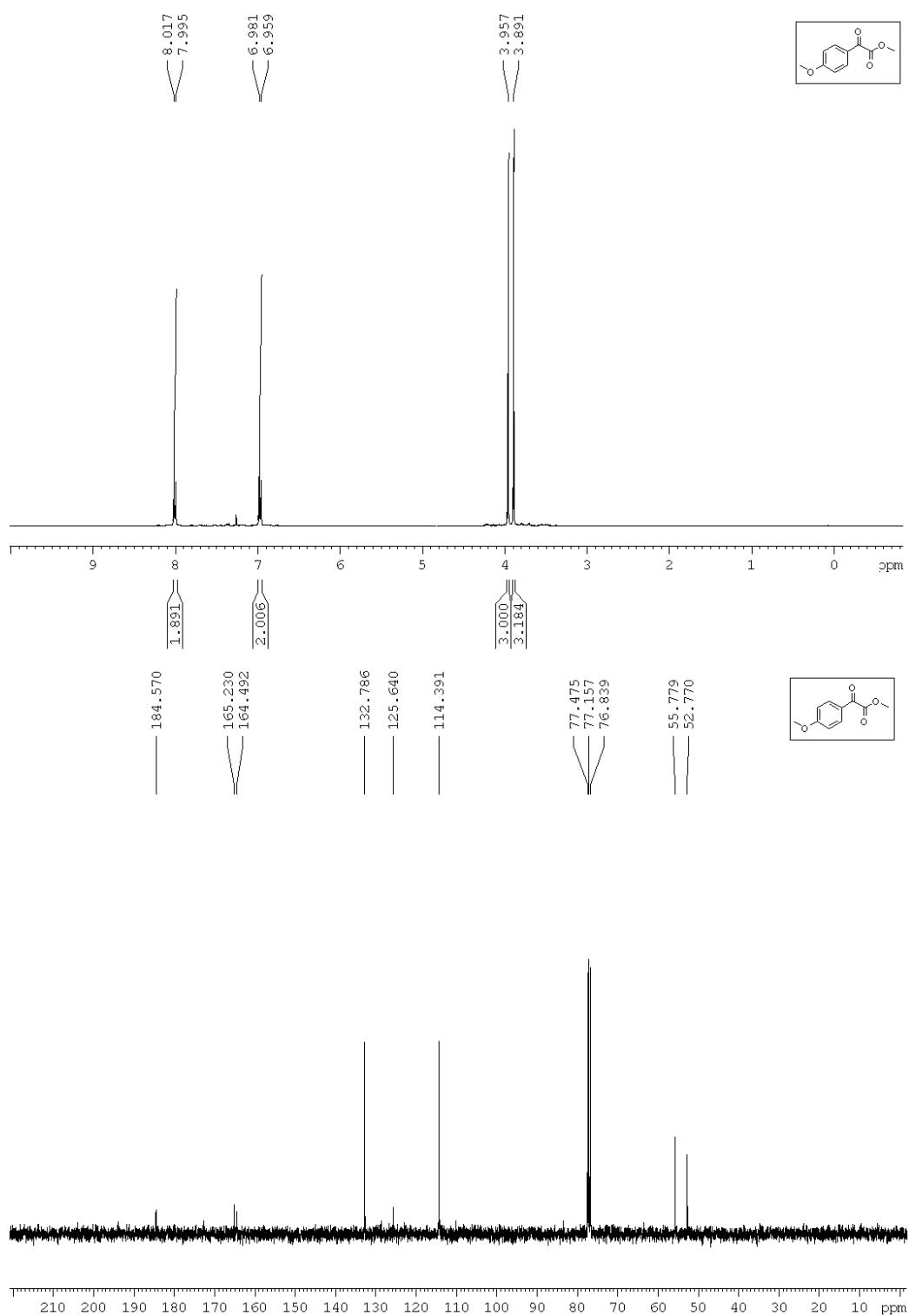


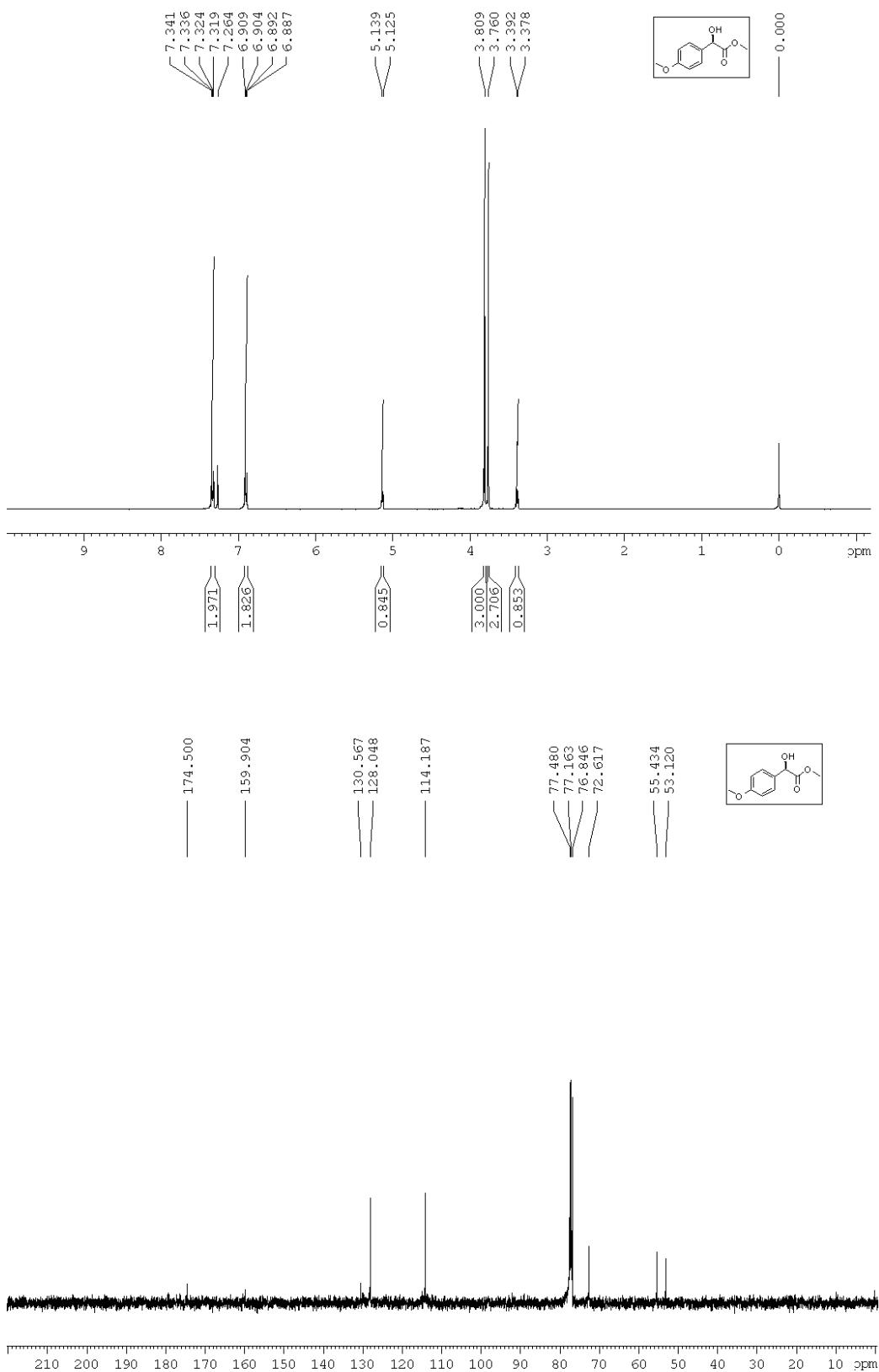


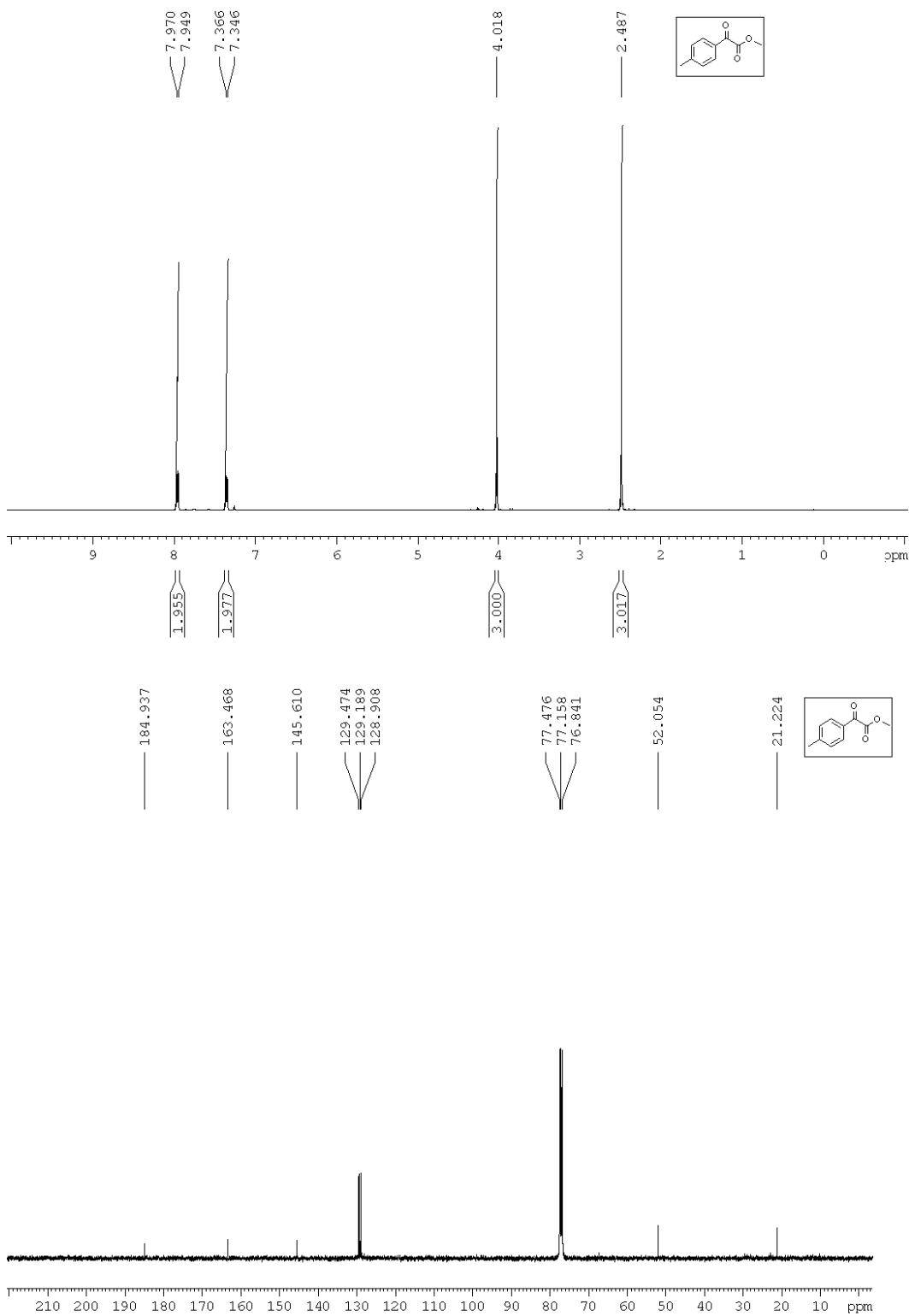


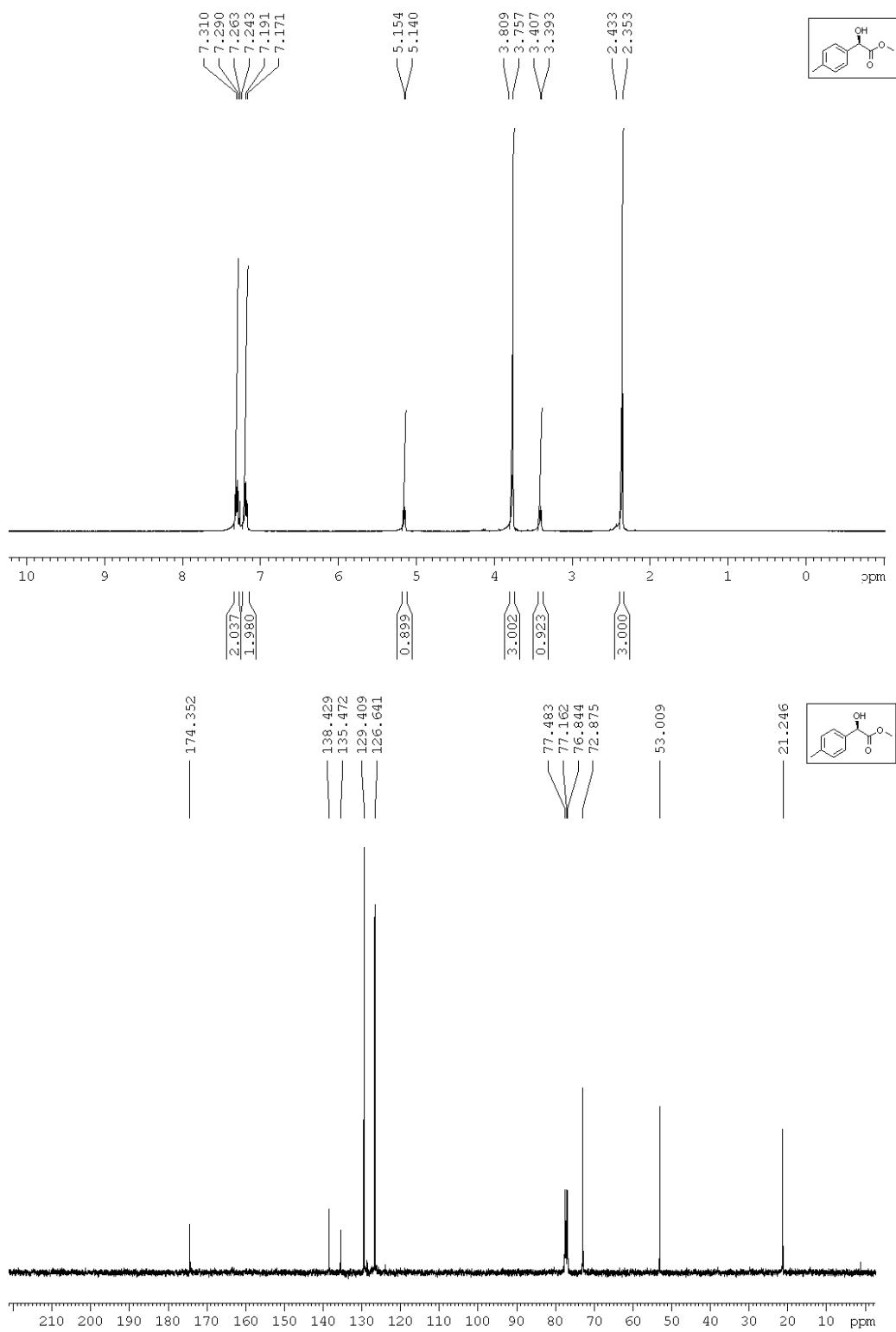


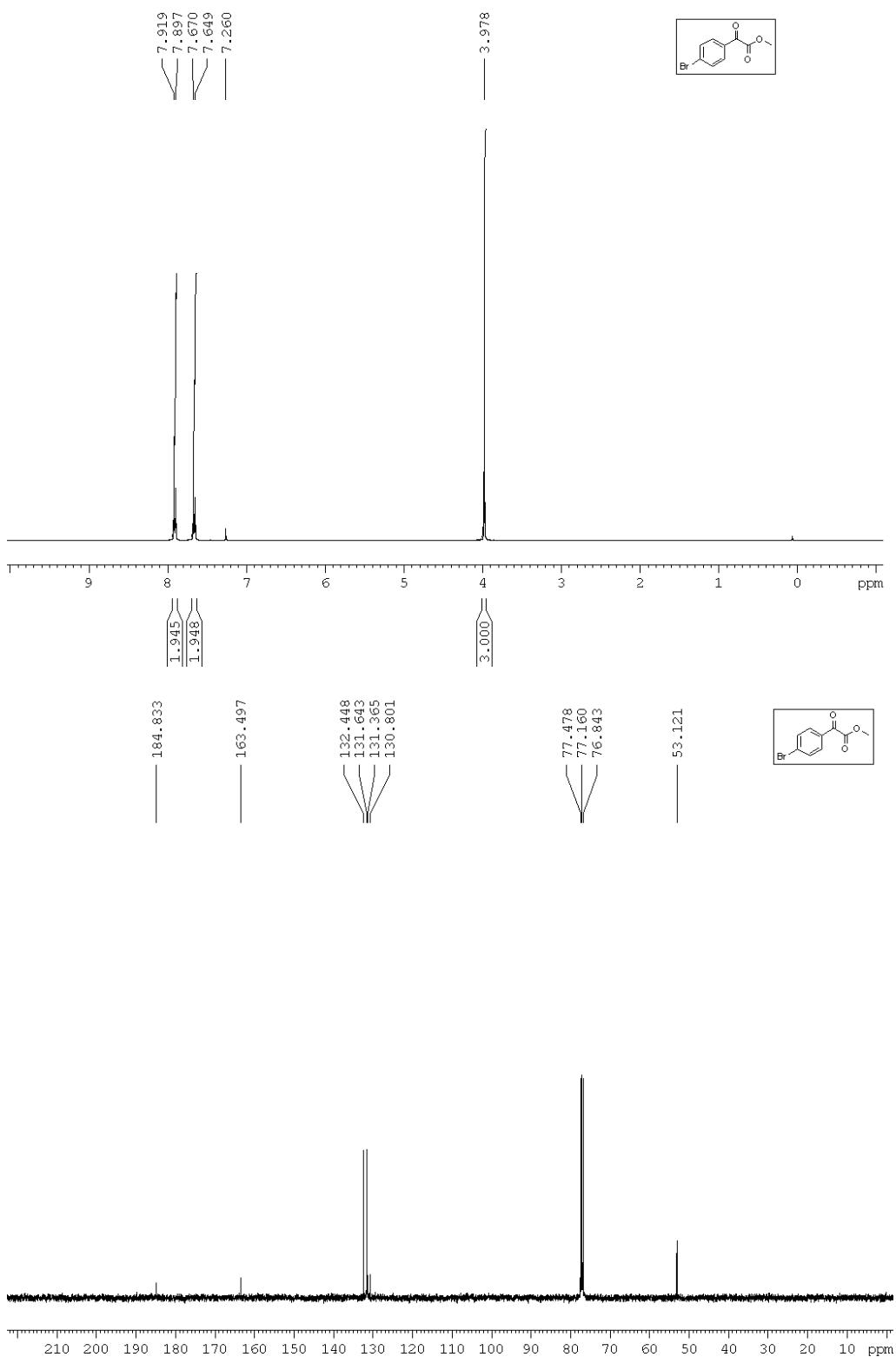


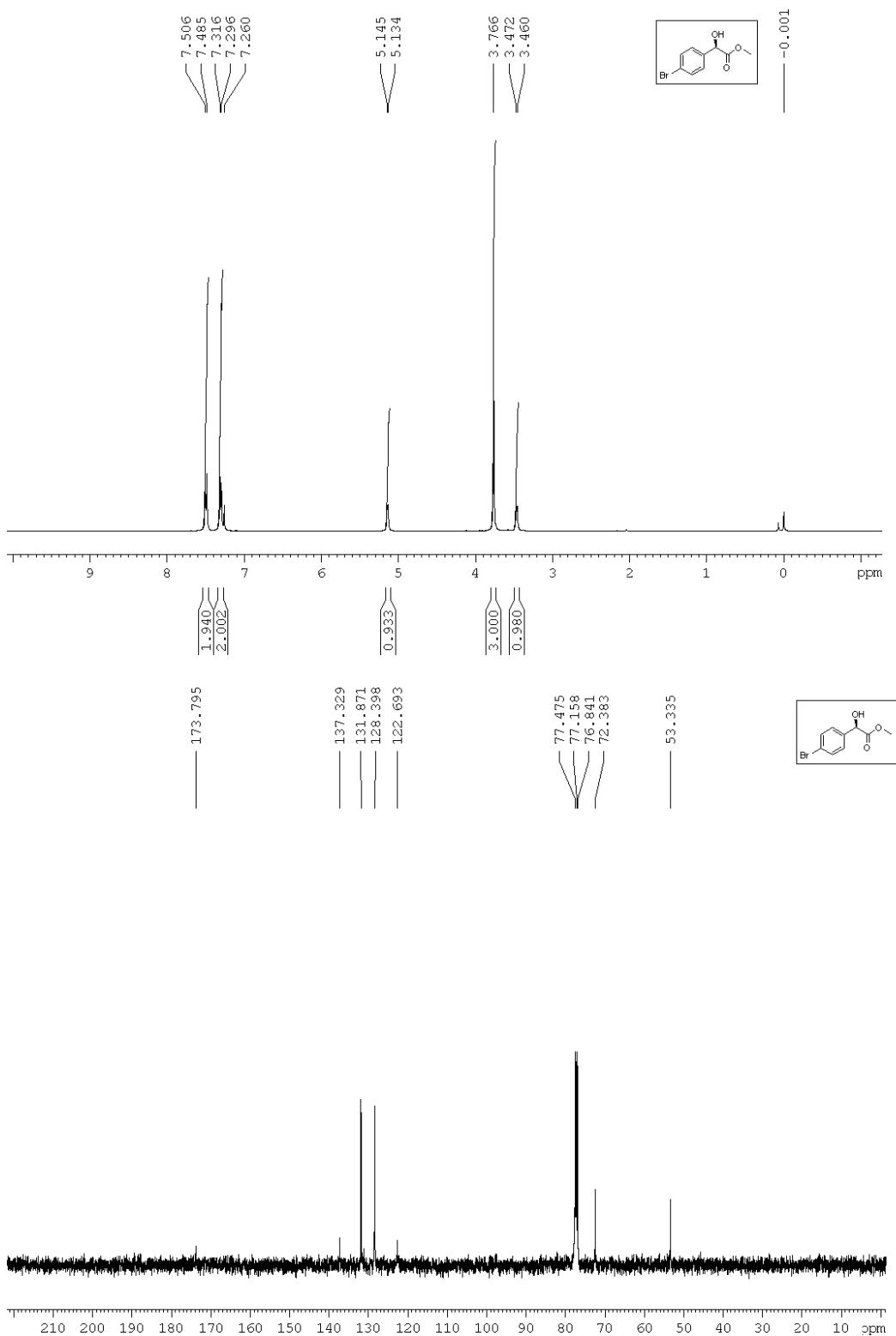


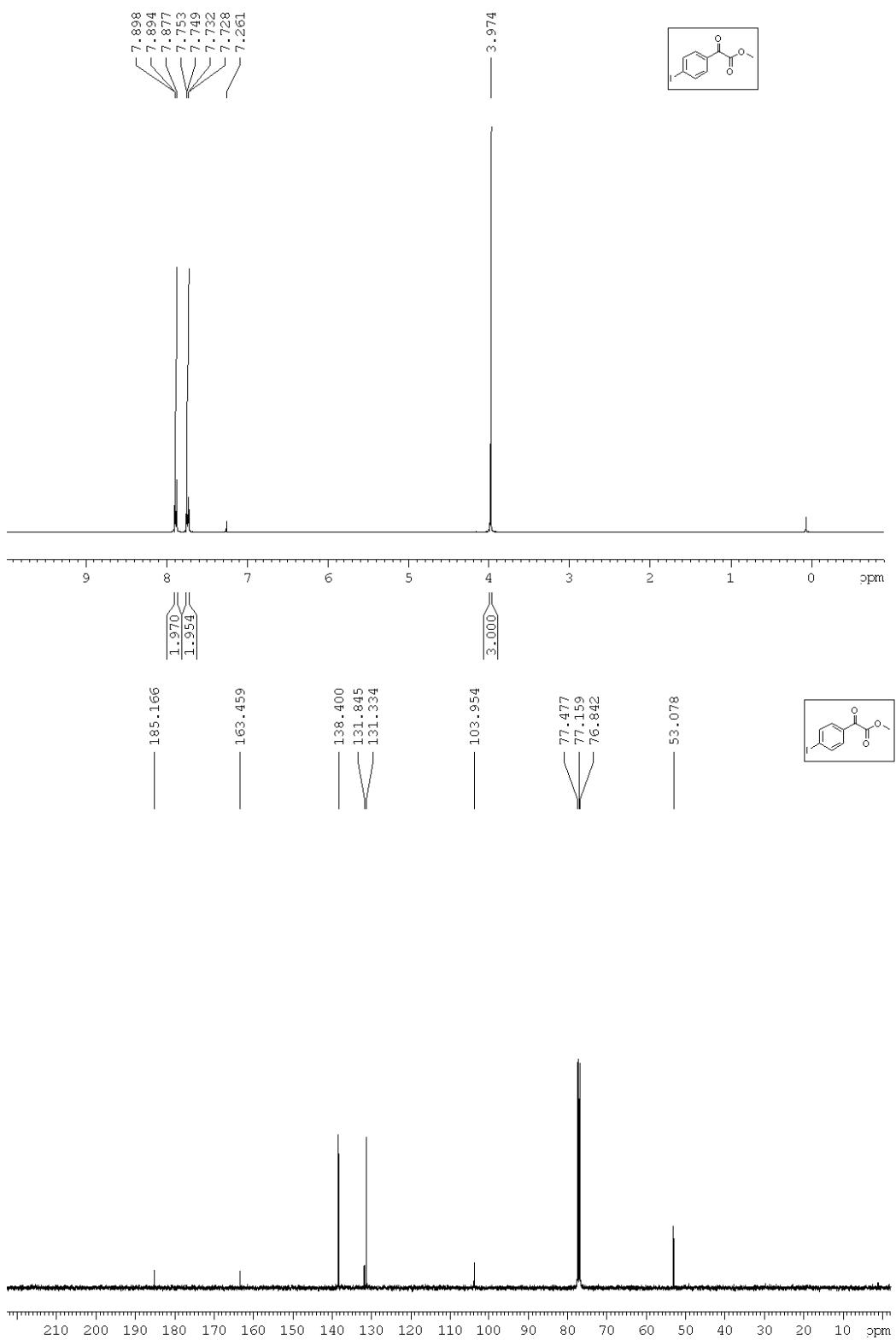


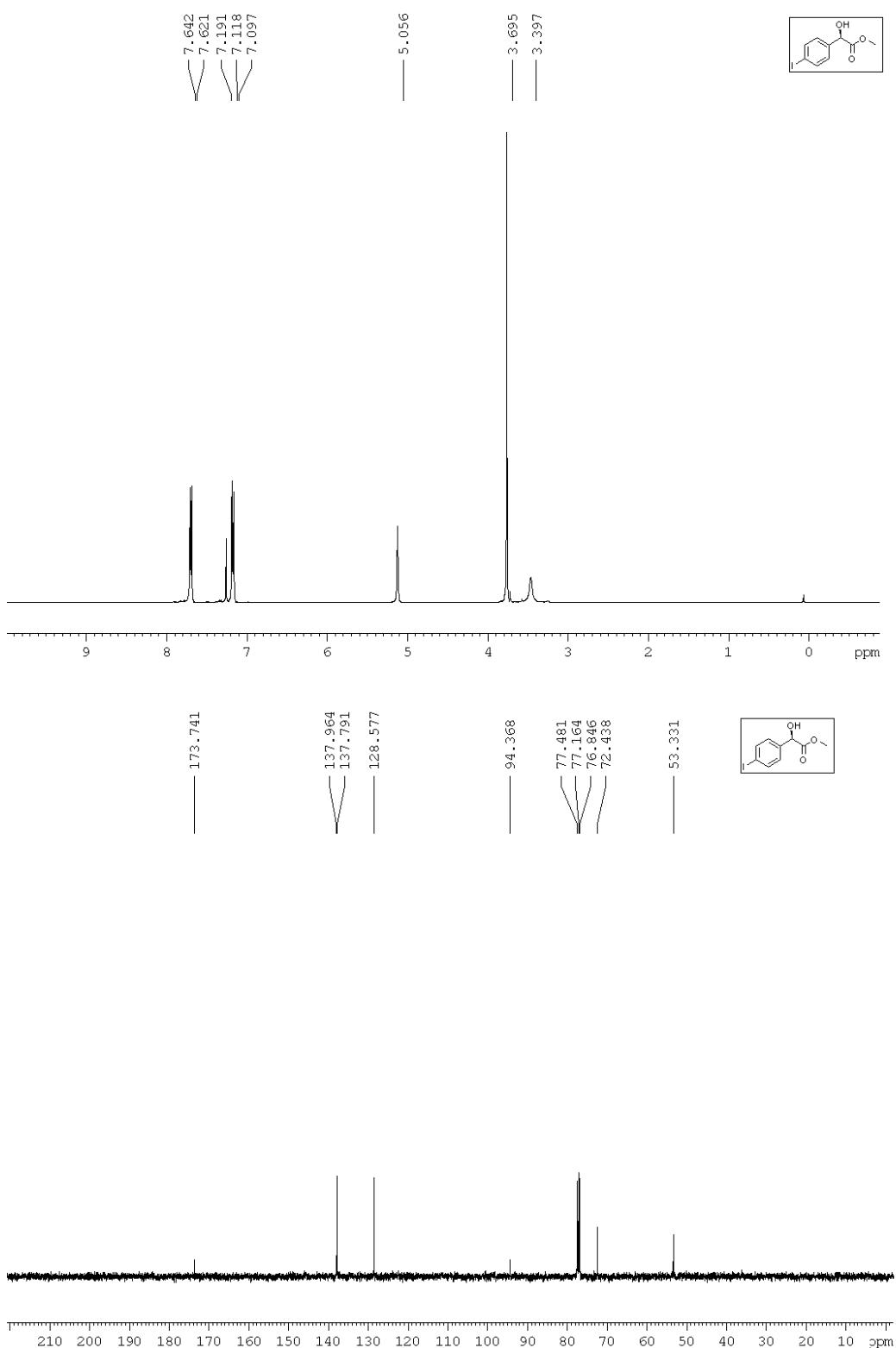












Reference

1. M. Shi, C.-J. Wang, *Tetrahedron: Asymmetr*, 2002, **13**, 2161.
2. E. R. Albert, P. M. Steven, P. M. James, *J. Org. Chem.*, 2000, **65**, 8381.
3. M. K. Margaret, D. M. Marko, K. Jeff, F. Anton, D. S. Jon, *J. Org. Chem.*, 1999, **64**, 6603.
4. E. R. Albert, P. M. Steven, P. M. James, *J. Org. Chem.*, 2000, **65**, 8381.
5. H. S. Bevinakatti, A. A. Banerji, R. V. Newadkar, *J. Org. Chem.*, 1989, **54**, 2453.
6. Y. S. Tsantrizos, K. O. Kelvin, *Can. J. Chem.*, 1991, **69**, 772.
7. M. K. Margaret, M. D. Marko, K. Jeff, F. Anton, S. D. Jon, *Org. Chem.*, 1999, **64**, 6603.
8. W. Yang, J. Xu, Y. Xie, Y. Xu, G. Zhao, G. Lin, *Tetrahedron: Asymmetry*, 2006, **17**, 1769.
9. S. Xianfeng, L. Wei, Z. Le, Z. Xumu, *Chem. Euro. J.*, 2009, **15**, 7302.
10. S. Toshiyasu, Y. Hiroshi, K. Toshi-aki, H. Teruyuki, T. Masato, *J. Org. Chem.*, 1987, **52**, 5733.
11. Y. Izawa, K. Ishiguro, H. omioka, *Bulletin of the Chemical Society of Japan*, 1983, **56**, 1490.
12. N. J. A. Martin, X. Cheng, B. List, *J. Am. Chem. Soc.*, 2008, **130**, 13862.
13. B. E. Howard,; K. A. Woerpel, *Org. Lett.*, 2007, **9**, 4651.
14. C. W. Laura. D. Hongbo, L. S. Marc. H. H. Amir. *J. Am. Chem. Soc.*, 2005, **127**, 15453.