

SUPPORTING INFORMATION

for

New azaborine–thiophene heteroacenes

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Experimental Part

Materials and Methods

All reactions were carried under dry nitrogen or argon atmosphere (specified in procedure). The reagents *n*-butyllithium (2.0 M in hexanes), tributyltin chloride, palladium (II) chloride (Alfa Aesar), tin (II) chloride, glyoxal, phenyl dichloroborane, triethylamine were obtained commercially and used as received. Fuming nitric acid was prepared in the laboratory. All solvents were of at least reagent grade and dried if necessary.

¹H NMR and ¹³C NMR spectra were run on a Varian Mercury 300 or 400 MHz NMR spectrometer. Low-resolution mass spectra (70 eV, EI and ESI) were run on a Kratos 7525 RFA or Finnigan LCQ DUO mass spectrometer. UV-vis spectra were measured with a Varian Cary 5000 spectrometer in 1 cm cuvettes (in CH₂Cl₂ solutions). Fluorescence spectra were measured with a Varian Eclipse spectrofluorometer, in 1 cm cuvettes (in CH₂Cl₂ solutions). Quantum yield was determined relative to 9,10-diphenylanthracene ($\Phi_{\text{PL}} = 90\%$).

Electrochemical Measurements: All electrochemical measurements were performed at room temperature in a three-electrode cell using a CHI-770 Electrochemical Workstation. Electrochemical investigations were conducted in anhydrous CH₂Cl₂ with a Pt disk (d=1.6 mm) as the working electrode, platinum wire as the auxiliary electrode and Ag/AgCl reference electrode. Bu₄ClO₄ (0.1 M) was used as a supporting electrolyte. The electrolyte solution was purged with Ar gas before and between electrochemical measurements. The redox potential of Fc/Fc⁺ couple in our conditions occurred at 0.50 V vs. Ag/AgCl.

X-ray Measurements. Needles of **1a** and **1b** was mounted on a nylon loop with paratone. X-ray data were collected at 100 K using omega scans with a Bruker APEX I CCD detector on a D8 3-circle goniometer and Mo KR (λ) 0.71073 Å) radiation. The data were scanned using Bruker's SMART program and integrated using Bruker's SAINT software.¹ The structure was solved by direct methods using SHELXS-97² and refined by a least-squares methods on F₂ using SHELXL-97 incorporated in the SHELXTL⁴³³ suite of programs.

Description of Density Functional Theory Calculations (DFT). All calculations were performed on neutral closed shell molecules with density functional theory at B3LYP/6-311G(d,p) level implemented in GAUSSIAN 03W.⁴ Default convergence criteria and no symmetry constrains were used.

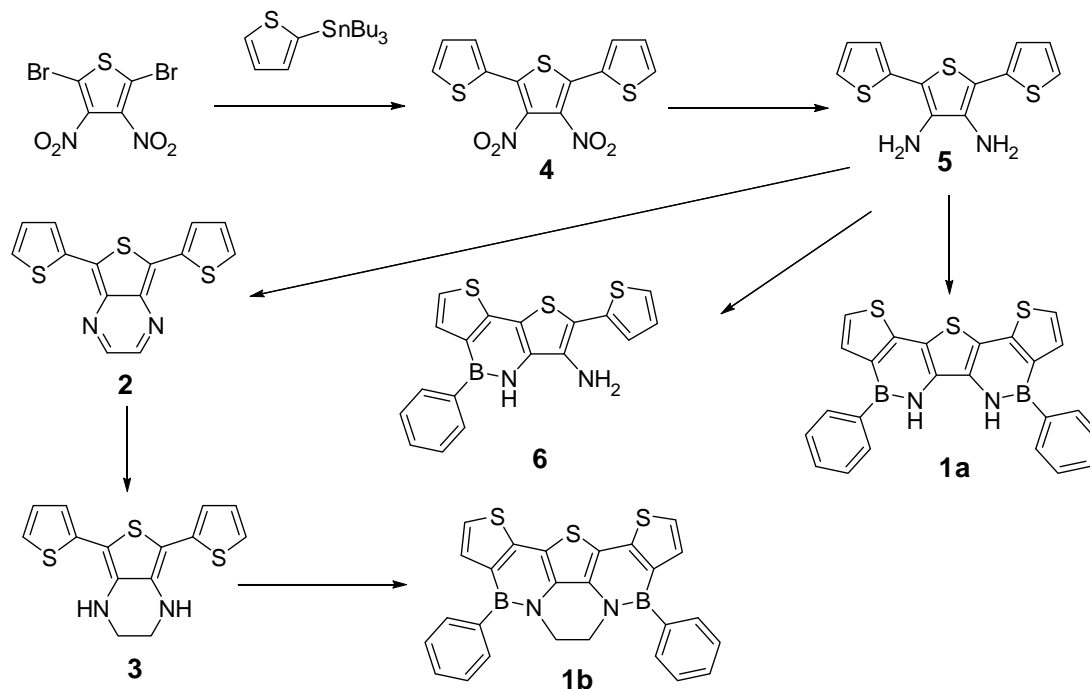
Reorganization energy λ was calculated as the difference between the total energy of radical cation in the optimized geometry and in the geometry of a neutral molecule (λ_+) plus the difference between the total energy of a neutral molecule in the optimized geometry and in the geometry of a radical cation (λ_0).

¹ Bruker (2006). SAINT Release 7.34A. Integration Software for Single Crystal Data. Bruker AXS Inc., Madison, USA.

² Sheldrick, G. M. (1997). SHELXS97. Program for Crystal Structure solution. University of Göttingen, Germany.

³ Bruker (1997). SHELXTL (1997). Release 5.10; The Complete Software Package for Single Crystal Structure Determination. Bruker AXS Inc., Madison, USA.

⁴ Gaussian 03, Revision C.02, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, J. A. Montgomery, Jr., T. Vreven, K. N. Kudin, J. C. Burant, J. M. Millam, S. S. Iyengar, J. Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G. A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J. E. Knox, H. P. Hratchian, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, P. Y. Ayala, K. Morokuma, G. A. Voth, P. Salvador, J. J. Dannenberg, V. G. Zakrzewski, S. Dapprich, A. D. Daniels, M. C. Strain, O. Farkas, D. K. Malick, A. D. Rabuck, K. Raghavachari, J. B. Foresman, J. V. Ortiz, Q. Cui, A. G. Baboul, S. Clifford, J. Cioslowski, B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. L. Martin, D. J. Fox, T. Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, M. Challacombe, P. M. W. Gill, B. Johnson, W. Chen, M. W. Wong, C. Gonzalez, and J. A. Pople, Gaussian, Inc., Wallingford CT, 2004.



2,5-Bis(2-thienyl)-3,4-dinitrothiophene (4). This was synthesized by Stille coupling of 2,5-dibromo-3,4-dinitrothiophene⁵ with tributyl-2-thienylstannane as described previously.⁶ ¹H NMR (400 MHz, CDCl₃, ppm): 7.61 (dd, *J* = 5.2, 1.2 Hz, 2H), 7.55 (dd, *J* = 4.0, 1.2 Hz, 2H), 7.18 (dd, *J* = 5.2, 4.0 Hz, 2H).

2,5-Bis(2-thienyl)-3,4-diaminothiophene (5). This was synthesized by modified published procedure⁷ with an improved yield. To a solution of 2,5-bis(2-thienyl)-3,4-dinitrothiophene (5.0 g, 14.8 mmol) in ethanol (200 mL) and concentrated HCl (100 mL) under argon was added SnCl₂×2H₂O (60 g, 267 mmol). The reaction mixture was refluxed for 15 h. Then, the mixture was poured into aqueous KOH (25 %) to basify the solution. The solution was filtered and the product was extracted with toluene (3×150 mL). The organic phase was dried over MgSO₄ and the solvent was evaporated. The product was purified by chromatography on silica gel to give the desired product as a yellow powder (3.12 g, 76 %), ¹H NMR (300 MHz, CDCl₃, ppm): 7.27 (dd, *J* = 4.5, 1.5 Hz, 2H), 7.09 (m, 4H), 3.74 (s, 4H).

⁵ D. D. Kenning, K. A. Mitchell, T. R. Calhoun, M. R. Funfar, D. J. Sattler, S. C. Rasmussen *J. Org. Chem.* **2002**, 67, 9073

⁶ Y. Xia, J. Luo, X. Deng, X. Li, X. Zhu, W. Yang, Y. Cao, *Macromol. Chem. Phys.*, **2006**, 207, 511

⁷ C. Kitamura, S. Tanaka, Y. Yamashita, *Chem. Mater.*, **1996**, 8, 570

5,7-bis(2-thienyl)thieno[3,4-*b*]pyrazine (2). This was synthesized according to procedure.⁷ To a solution of **5** (1.026 g, 3.70 mmol) in ethanol (20 mL) under argon was added glyoxal (40 % in water) (0.29 mL, 1.98 mmol) and Na₂CO₃ (3.92 g, 37.0 mmol). The reaction mixture was refluxed for 2h, the reaction turned purple. Then, water was added (50 mL) and the product was extracted by 3 × 50 mL of diethyl ether. The organic phase was dried over MgSO₄ and the solvent was evaporated. The product was purified by chromatography on silica gel to give product pyrazine **2** as a purple powder (1.10 g, 99 %). ¹H NMR (400 MHz, CDCl₃, ppm): 8.54 (s, 2H), 7.65 (dd, *J* = 4.0 Hz, *J* = 1.2 Hz, 2H), 7.42 (dd, *J* = 5.2 Hz, *J* = 1.2 Hz, 2H), 7.14 (dd, ³*J* = 5.2 Hz, ³*J* = 4.0 Hz, 2H).

Ethylenediaminoterthiophene (3) To a solution of **2** (0.38 g, 1.26 mmol) in ethanol (10 mL) under argon was added NaBH₄ (2.4 g, 63 mmol). The reaction mixture was refluxed for 1h, the solution turned orange. Then water was added (20 mL) and the product was extracted by 3 x 50 mL of ethyl acetate. The organic phase was dried over MgSO₄ and the solvent was evaporated. Quick filtration through a silica gel plug gave intermediate **3** as yellow powder (0.39 g, 99 %) which was of sufficient purity for further reaction (>95% by NMR). ¹H NMR (300 MHz, CDCl₃, ppm): 7.21 (m, 2H), 7.06 (m, 4H), 4.30 (br. s, 2H), 3.42 (s, 4H). We note that the product should be isolated/purified rapidly and stored under nitrogen as prolonged exposure to air leads to oxidation.

Azaborine (1a). To a solution of 2,5-bis(2-thienyl)-3,4-diaminothiophene **5** (1.00 g, 3.60 mmol) in chlorobenzene (50 mL) under argon was added phenyldichloroborane (1.71 g, 10.8 mmol) and triethylamine (2 mL, 14 mmol). The reaction mixture was refluxed for 48h. Then the solvent was evaporated and the product was purified by chromatography on silica gel to give product **1a** as yellow powder (1.40 g, 86 %), m.p. 216 °C. ¹H NMR (300 MHz, CDCl₃, ppm): 8.37 (br. s, 2H), 7.91 (m, 4H), 7.77 (d, ³*J* = 5.0 Hz, 2H), 7.53 (m, 6H), 7.36 (d, ³*J* = 5.0 Hz, 2H). ¹³C NMR (125.8 MHz, CDCl₃, ppm): 148.6, 133.4, 131.9, 130.2, 129.5, 128.6, 123.4, 117.0.⁸ MS (ESI) *m/z* = 449.52 (100 %) [M-H]⁺. C₂₄H₁₆S₃B₂N₂: Calcd. C 64.03, H 3.58, N 6.22; Found C 64.12, H 3.72, N 6.20.

Azaborine (1b). To a solution of ethylenediaminoterthiophene **3** (0.392 g, 1.29 mmol) in chlorobenzene (50 mL) under argon was added phenyldichloroborane (0.43 mL, 3.22 mmol) and triethylamine (0.6 mL, 3.9 mmol). The reaction mixture refluxed overnight, then the solvent was evaporated in vacuo and the product was purified by chromatography on silica gel to give product **1b** as yellow powder (0.33 g, 54 %), m.p. 224-226 °C. ¹H NMR (300 MHz, CDCl₃, ppm): 7.66 (m, 4H),

⁸ ¹³C NMR signal for the carbon attached to boron is not observed due to coupling with adjacent ¹¹B.

7.48 (m, 8H), 7.27 (d, $J = 5.1$ Hz, 2H), 4.31 (s, 4H). ^{13}C NMR (125.8 MHz, CDCl_3 , ppm): 146.7, 139.8 (br), 138.4 (br), 133.7, 132.7, 131.0, 128.4, 128.1, 122.7, 116.2, 46.2. MS (ESI) $m/z = 476.0$ $[\text{M}]^+$. $\text{C}_{26}\text{H}_{18}\text{S}_3\text{B}_2\text{N}_2$: Calcd. C 65.57, H 3.81, N 5.88, S 20.20; Found C 65.11, H 3.74, N 5.95, S 19.91. The reaction was repeated with 0.75 g of **3** without heating (stirring overnight); the purification as above gave the same product **1b** (0.51 g, 42%).

Azaborine (6). To a solution of 2,5-bis(2-thienyl)-3,4-diaminothiophene **4** (106 mg, 0.381 mmol) in chlorobenzene (50 mL) under argon was added phenyldichloroborane (60 μL , 0.45 mmol). The reaction mixture was refluxed for 2 h and the solvent was evaporated. Excess of triethylamine (3 drops) was added and the product was purified by chromatography on silica gel (EtOAc/hexane) to give product **6** as a yellow-brownish solid which rapidly decomposes on storage (45 mg, 32 %). ^1H NMR (400 MHz, CDCl_3 , ppm): 8.01 (br. s, 1H, NH), 7.88 (dd, $J = 7.6$ Hz, 2H, *o*-H), 7.75 (d, $J = 5.2$ Hz, 1H, *th*-H), 7.50 (m, 3H, *m*-,*p*-H), 7.35 (d, $J = 4.8$ Hz, 1H, *th*-H), 7.31 (d, $J = 5.2$ Hz, 1H, *th*-H), 7.21 (d, $J = 3.6$ Hz, 1H, *th*-H), 7.13 (dd, $J = 4.8$ and 3.6 Hz, 1H, *th*-H), 3.5 (br, 2H, NH_2). MS (EI) $m/z = 364$ $[\text{M}]^+$.

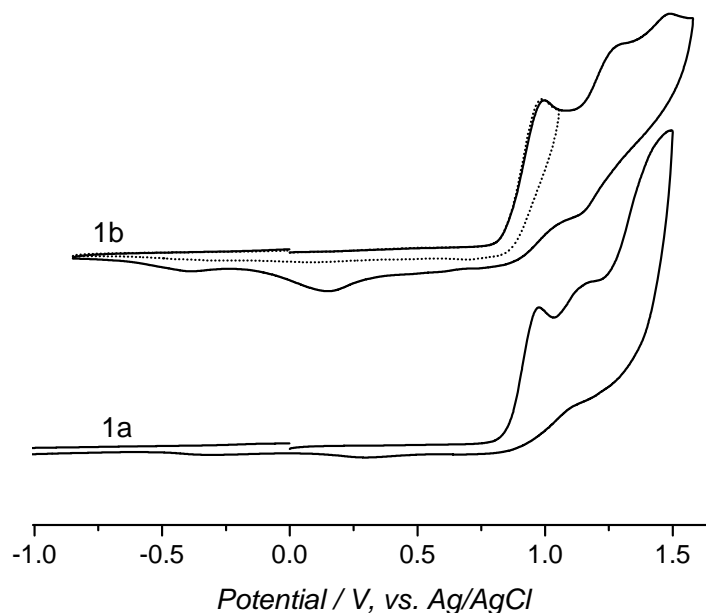


Fig. S1. CVs of **1a** (bottom curve) and **1b** (top curves) in 0.1 M $\text{Bu}_4\text{NClO}_4/\text{CH}_2\text{Cl}_2$, scan rate 0.1 V s^{-1} .

Fluoride titration experiments

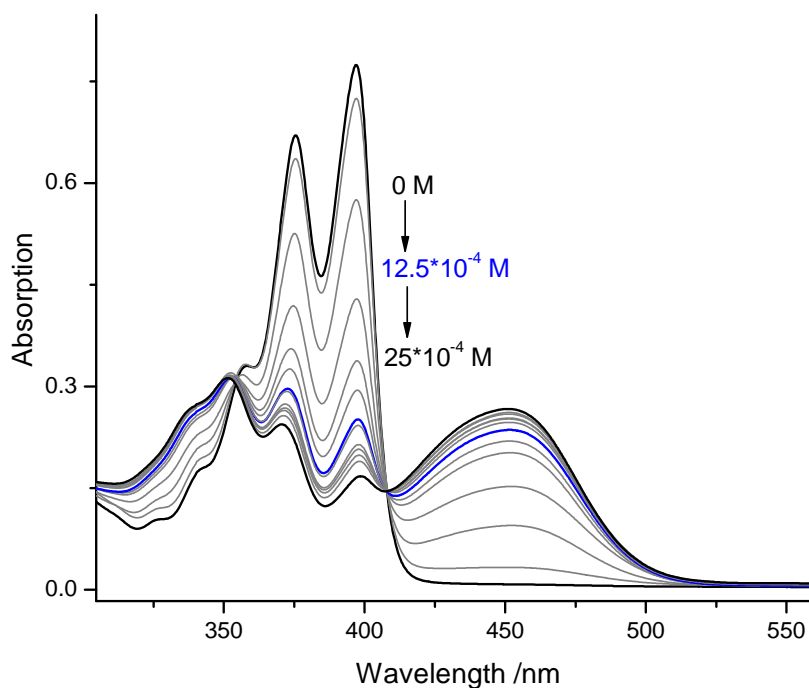


Fig. S2. UV-Vis of titration of **1b** (3.1×10^{-5} M, 2mL) with TBAF in DCM (see also Table S1).

Table S1. UV-Vis titration of **1b** (3.1×10^{-5} M, 2mL) with TBAF in DCM. Extinction coefficients are:

1b: $\epsilon_{397\text{nm}} = 33,500 \text{ cm}^{-1} \text{ M}^{-1}$; $\epsilon_{453\text{nm}} = 0$; **1b:F⁻:** $\epsilon_{397\text{nm}} = 13,300 \text{ cm}^{-1} \text{ M}^{-1}$; $\epsilon_{453\text{nm}} = 4,000$

[TBAF] ₀	Abs ₃₉₇	Abs ₄₅₃
0	0.7742	0.00776
0.000125	0.72405	0.03256
0.00025	0.5752	0.09388
0.0005	0.4288	0.1523
0.00075	0.337	0.2018
Abs0.001	0.29443	0.2189
0.00125	0.2512	0.2356
0.0015	0.242	0.2466
0.00175	0.214	0.2532
0.002	0.207	
0.00225	0.1975	0.259
0.0025	0.1893	

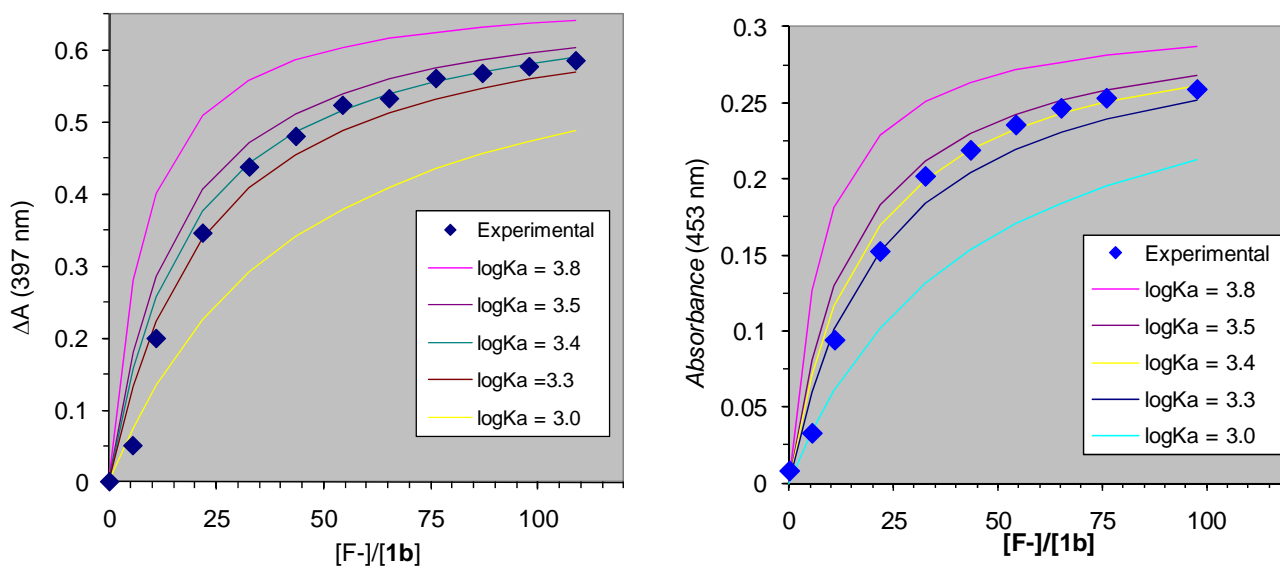
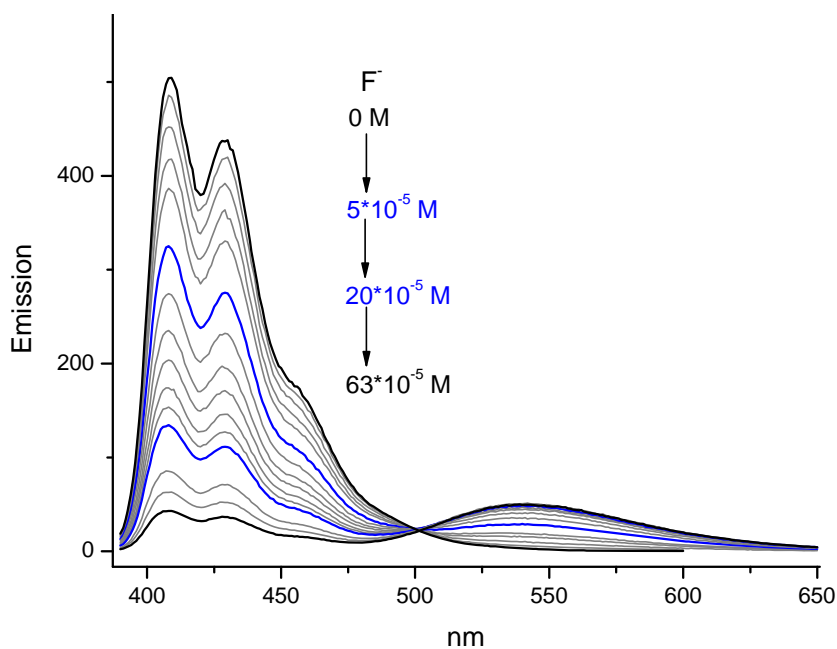


Fig. S3. Binding isotherms for **1b**:F⁻ based on UV-Vis titration (Fig. S2, Table S1) measured by depletion of the **1b** absorption (397 nm, left) and appearance of the **1b**:F⁻ absorption (453 nm, right).



Fig, S4. Fluorescence titration of **1b** (0.85×10^{-5} M, 2 mL) with TBAF in DCM.

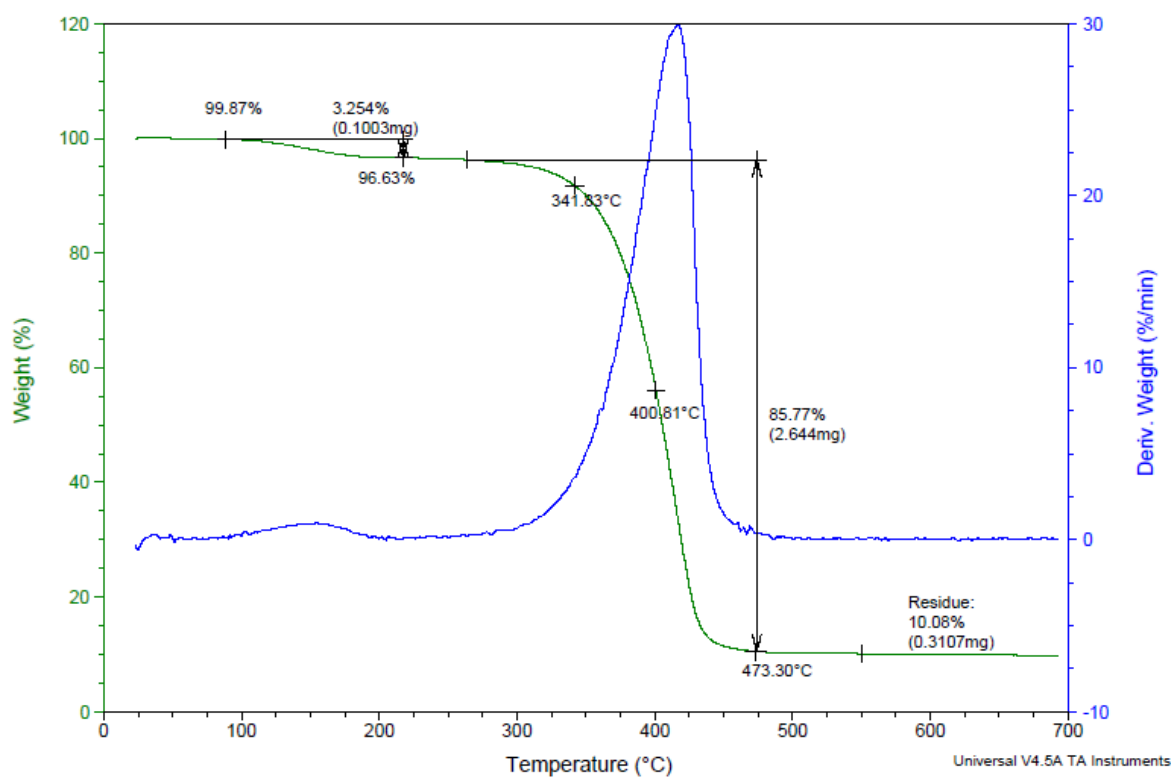


Fig. S5. Thermogravimetric analysis of azaborine **1a**. 3% mass loss at 100–150 °C is due to specifically bound molecule of water (hydrogen bonding to two N-H fragments; see X-ray data for **1a**). $T_{\text{dec}} \sim 300^\circ\text{C}$ (based on 5% mass loss).

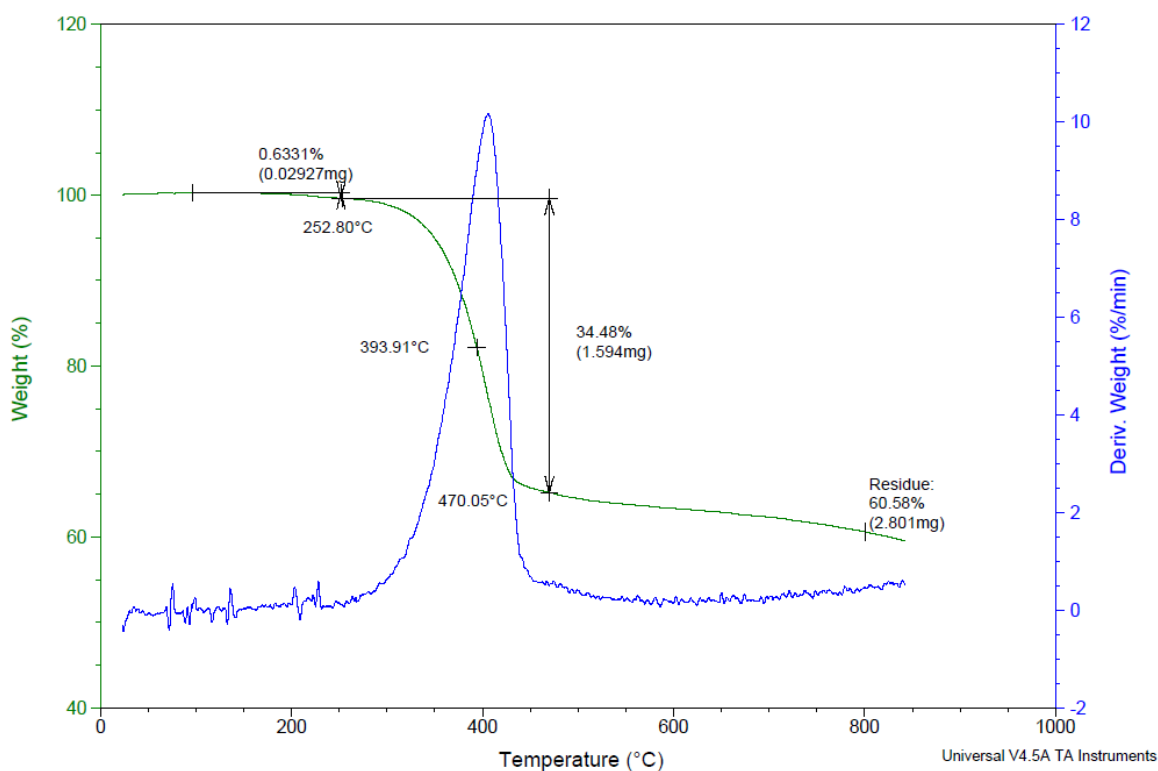
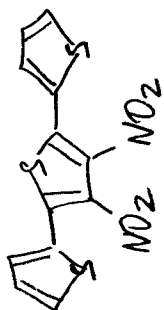


Fig. S6. Thermogravimetric analysis of azaborine **1b**. $T_{\text{dec}} \sim 300^\circ\text{C}$ (based on 5% mass loss).

17L375-1

OK



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

Mercury-300 "m300"

Relax. delay 1.000 sec

Pulse 58.9 degrees

Acq. time 1.995 sec

Width 4506.5 Hz

12 repetitions

OBSERVE H1 300.0549900 MHZ

DATA PROCESSING

FT size 32768

Total time 0 min, 0 sec

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7.165
7.175
7.178
7.181
7.191
7.194

7.618
7.615
7.605
7.602
7.598
7.558
7.555
7.551

7.622

7.545
7.542
7.539
7.260
7.257

ppm

1

2

3

4

5

6

7

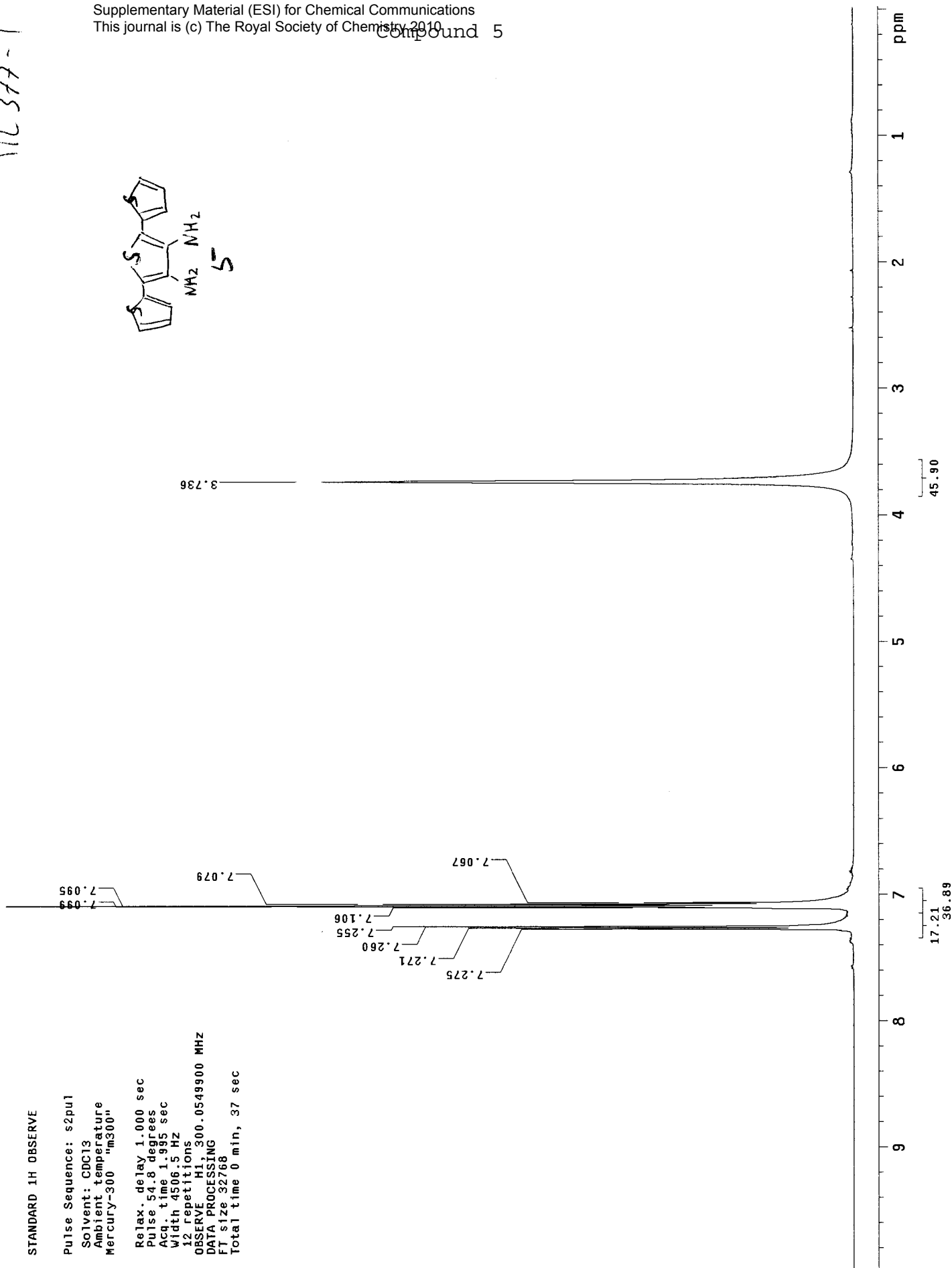
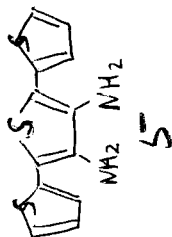
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9

31.29 36.31
32.39

Compound 5

ML 377-1



STANDARD 1H OBSERVE

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Ambient temperature

Mercury-300 "m300"

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Width 4506.5 Hz

12 repetitions

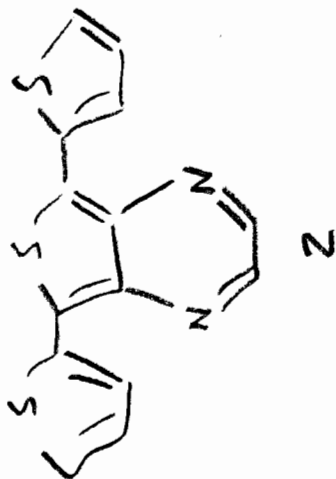
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Compound 2



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Sample directory:

File: PROTON

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Solvent: CDC13

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Pulse 45.0 degrees
Acq. time 1.995 sec
width 6410.3 Hz
12 repetitions

OBSERVE H1, 400.1219604 MHz
DATA PROCESSING
FT size 32768
Total time 0 min

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0.54

7.653
7.650
7.643
7.640
7.427
7.424
7.148
7.139
7.135
7.126

7.414
7.411
7.263

ppm

1

2

3

4

5

6

7

8

9

25.51

23.73 24.06 26.70

26.70

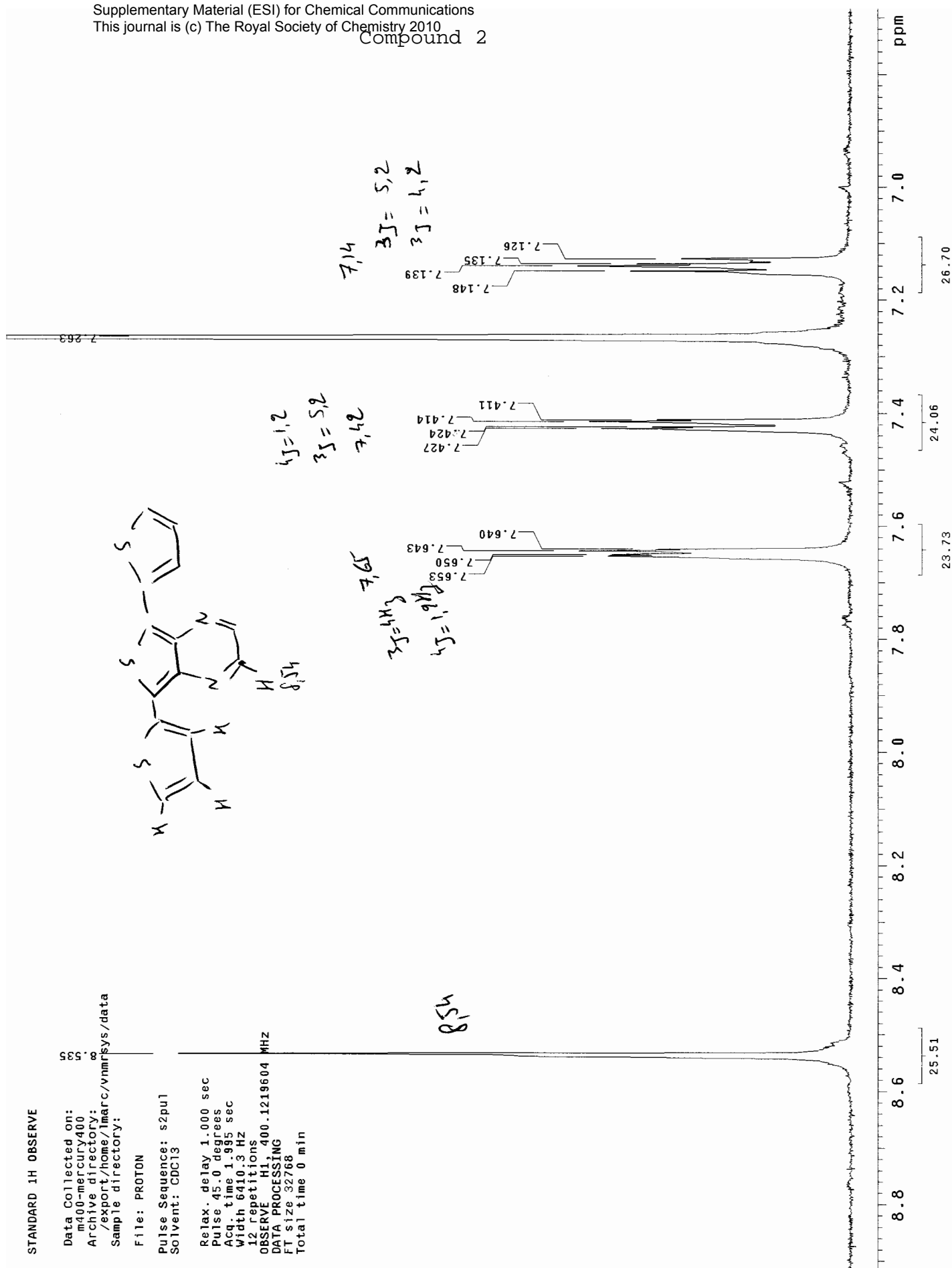
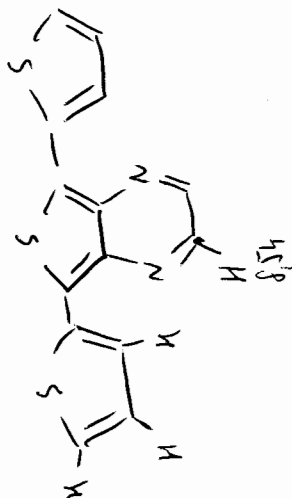
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sample directory:

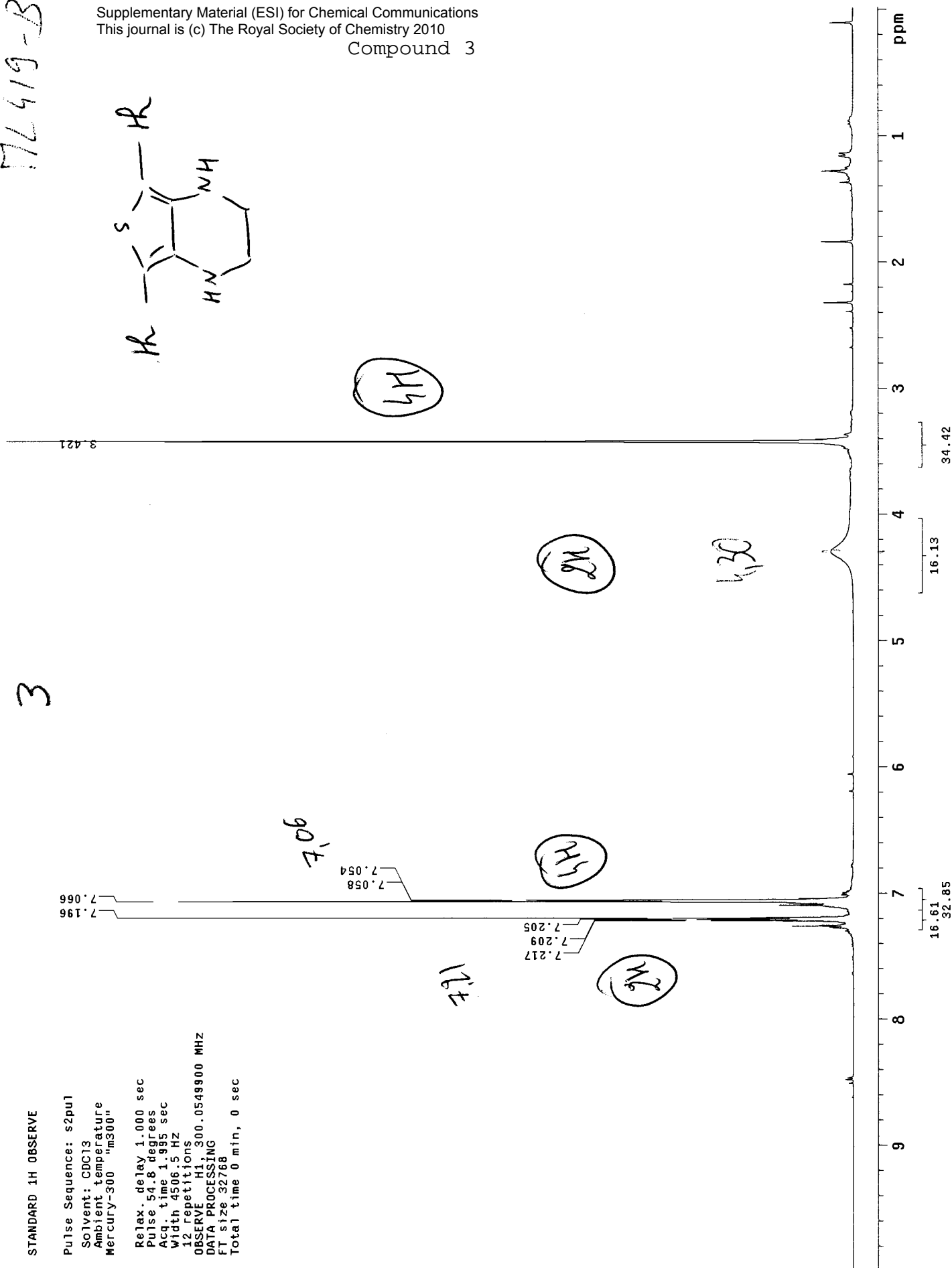
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Solvent: CDC13

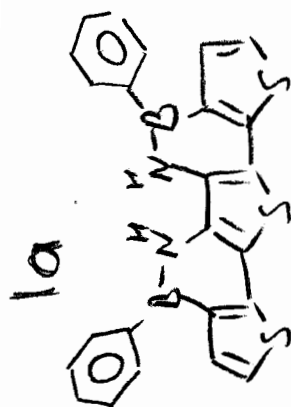
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Wd 6410.3 Hz
12 repetitions
OBSERVE H1, 400.121960
DATA PROCESSING
FT size 32768
Total time 0 min



3

Rc1c(R)sc2c1[nH]c[nH]2

Compound 1a



STANDARD 1H OBSERVE

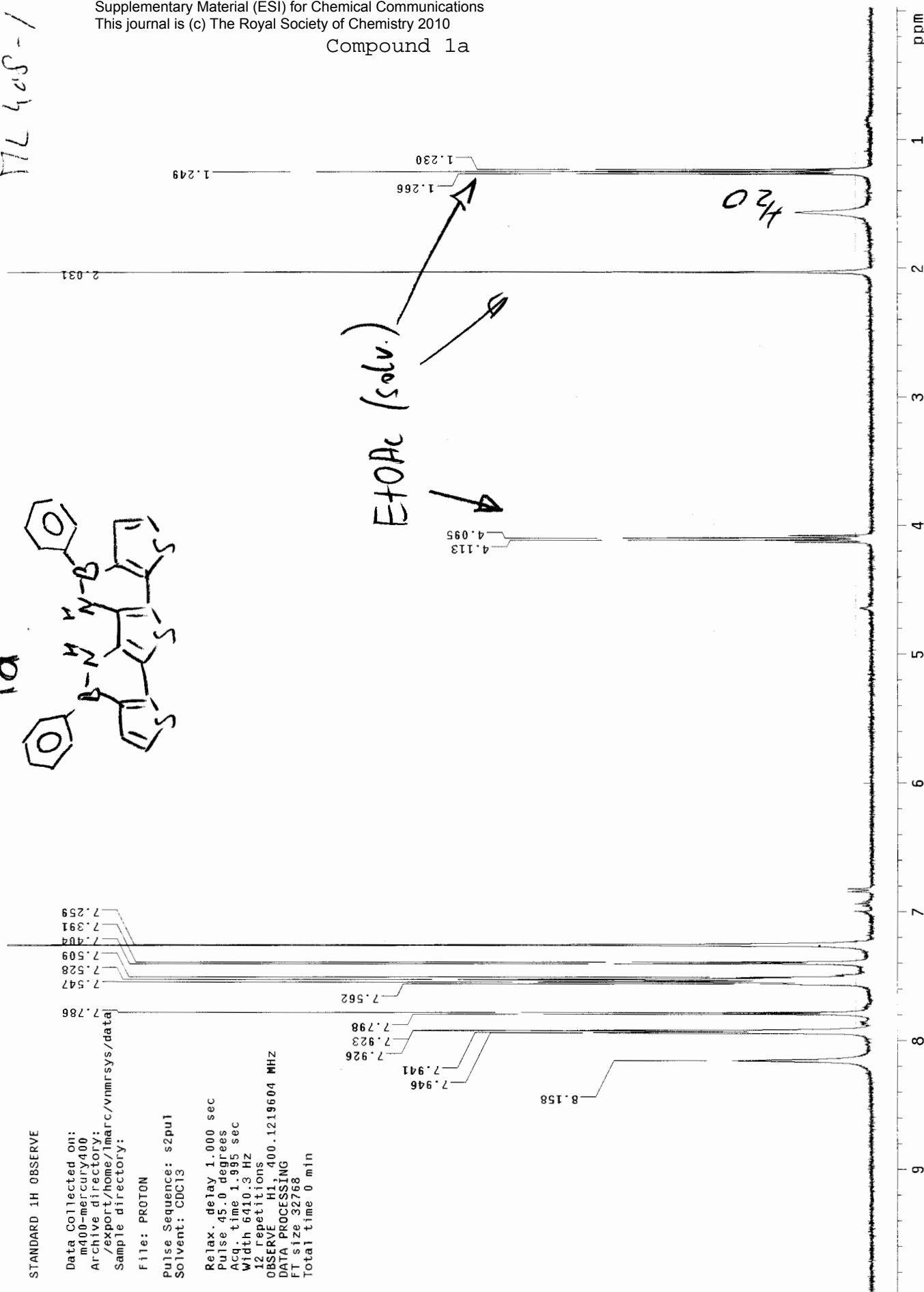
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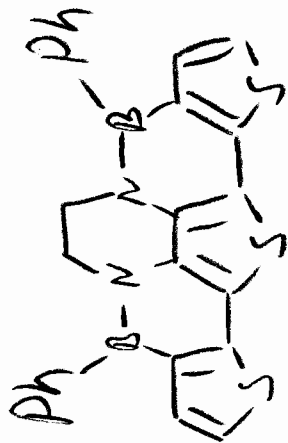
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Pulse 45.0 degrees
Acq. time 1.995 sec
Width 6410.3 Hz
12 repetitions

OBSERVE H1, 400.1219604 MHZ
DATA PROCESSING
FT size 32768
Total time 0 min



8.158
7.946
7.941
7.926
7.923
7.798
7.562
7.786
7.547
7.528
7.509
7.494
7.391
7.259
4.113
4.095
1.266
1.230
1.249
2.031

Compound 1b



STANDARD 1H OBSERVE

Data Collected only:
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Archive directory:
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Sample directory:

File: PROTON

Pulse Sequence: s2pu1

Solvent: CDCl3

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.999 sec

Width 6410.8 Hz

8 repetitions

OBSERVE H1, 400.1219604 MHz

DATA PROCESSING

FT size 65536

Total time 0 min

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7.480

7.262
7.260

ppm

2

3

4

5

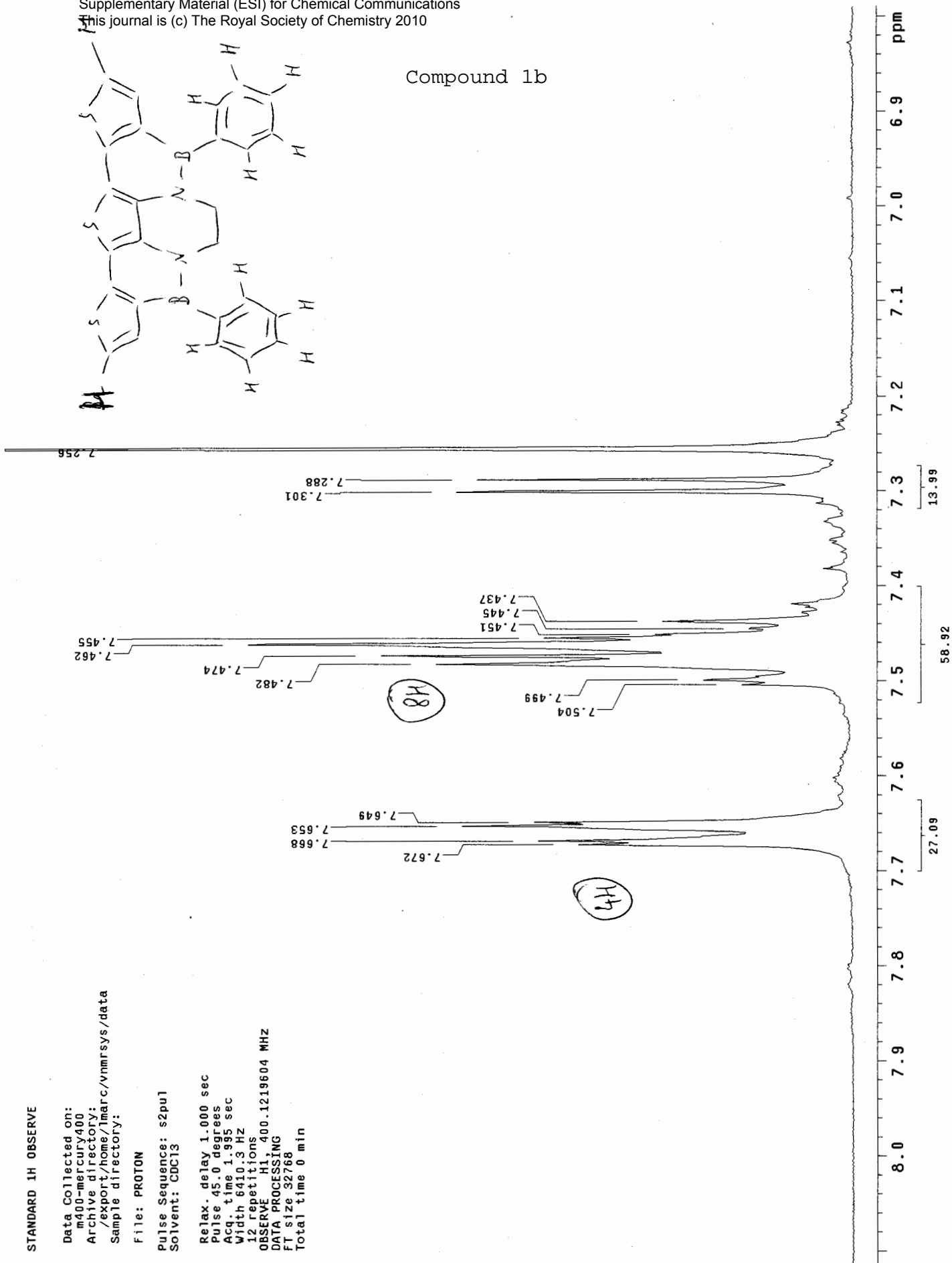
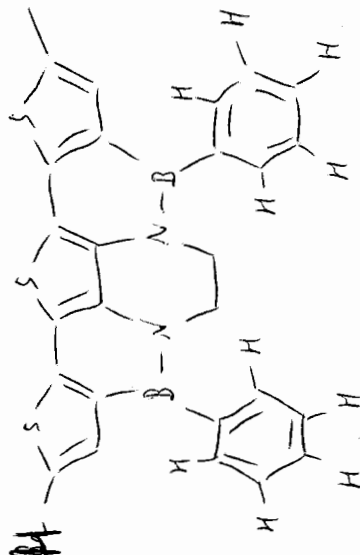
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7

8

16-cdc13- 400 MHz

Compound 1b



STANDARD 1H OBSERVE

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 Sample directory:

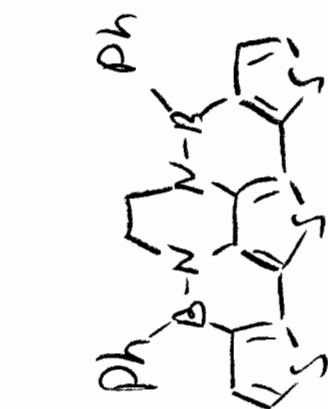
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 Solvent: CDCl3

Relax. delay 1.000 sec
 Pulse 45.0 degrees
 Acq time 1.995 sec
 Width 6410.3 Hz
 12 repetitions

OBSERVE H1, 400.1219604 MHz
 DATA PROCESSING
 FT size 32768
 Total time 0 min

Compound 1b

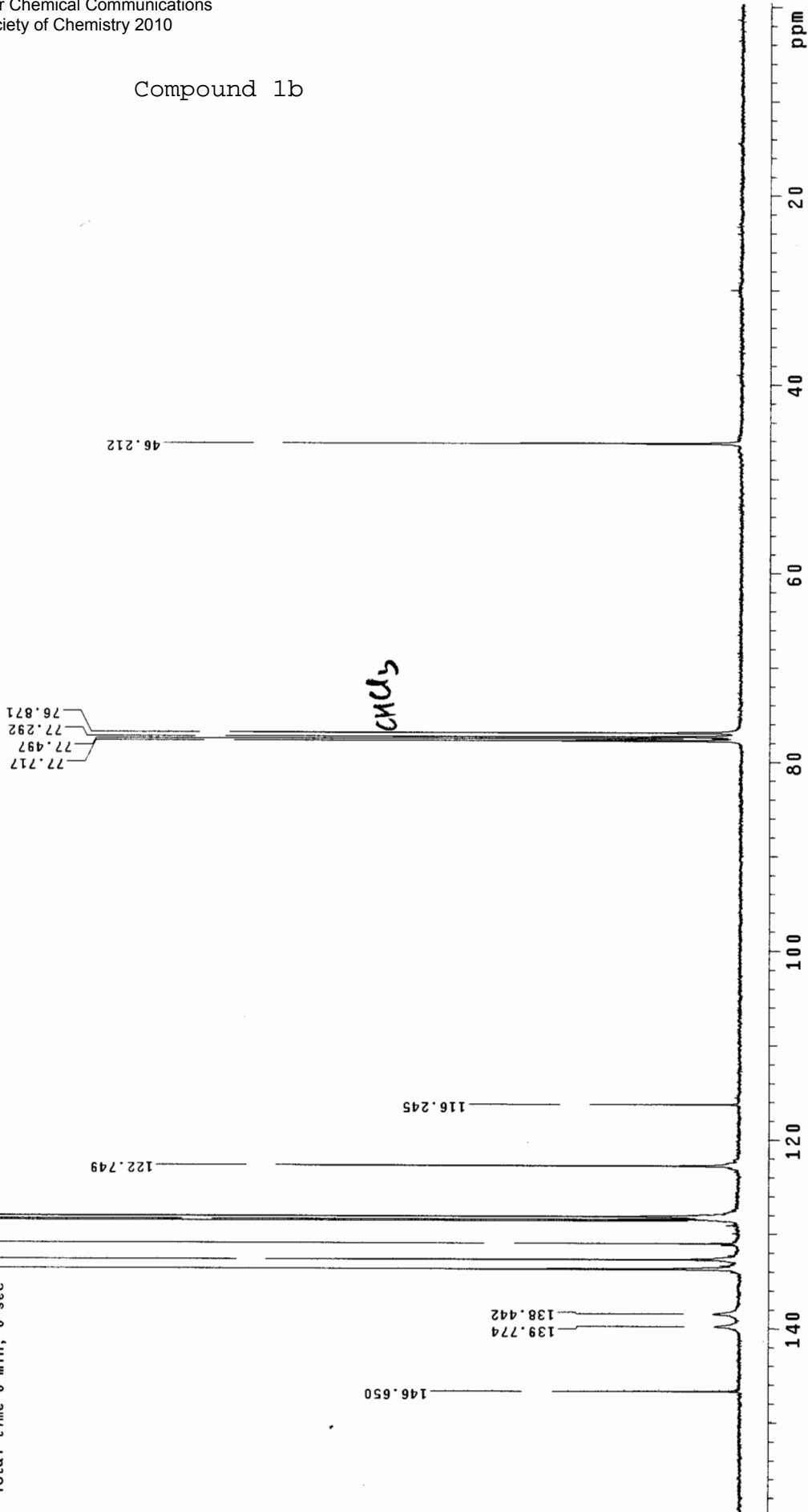


¹³C, 300MHz

¹³C OBSERVE

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Solvent: CDCl₃
Ambient temperature
Mercury-300 "m300"

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Acq. time 1.815 sec
Width 18761.7 Hz
15000 repetitions
OBSERVE C13, 75.4488692 MHz
DECOUPLE H1, 300.0564325 MHz
Power 34 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 0 min, 0 sec



ML406 °C

Compound 1b

