

Electronic Supplementary Information

New Class of Rhodamine Luminophores: Design, Syntheses and Aggregation-Induced Emission Characteristics

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1. General remarks

Reagents were purchased from Wako, Nacalai Tesque, or TCI Japan. All other solvents were used without further purification. Experiments were conducted under an atmosphere of dry nitrogen or using a guard tube.

^1H -NMR and ^{13}C -NMR spectra were recorded using 300 or 500 MHz spectrometers (Varian UNITY INOVA). Solvents used for NMR measurements were CDCl_3 , $\text{DMSO-}d_6$ or $\text{TFA-}d_8$, with TMS as the internal standard. Mass spectra were acquired using a JMX-700 (2) (JEOL Co., Ltd.) MS instrument. Melting points were determined on a Yanagimoto micromelting point apparatus and are uncorrected. Liquid column chromatography was conducted over silica gel (Merck Silica Gel 60 mesh 70-230). Developed TLC plates were visualized under a short-wave UV lamp, by staining with an $\text{I}_2\text{-SiO}_2$ mixture, and/or by heating plates that were dipped in ammonium phosphomolybdate sulfate solution. Dry THF was distilled over sodium benzophenone ketyl under argon atmosphere.

UV-vis spectra were collected on a SHIMADZU UV-1700 spectrophotometer at RT using a 1 cm or a 0.1 cm quartz cuvette. Emission spectra were collected on a HITACHI F-4500 fluorescence spectrophotometer. To obtain an accurate spectrum, spectrum correction was carried out with rhodamine B concentrated solution and a secondary-standard light source. In surface photometry, a cut filter was utilized to eliminate multiple lights, such as secondary light, due to the effects of light scattering. Fluorescence spectra of all samples were measured with excitation with 365 nm. Average particle size was measured by dynamic light scattering (ELSZ-2, Otsuka Electronics).

2. Syntheses

2-1. General Procedure for the synthesis of **2**

To a solution of phthalic anhydride (1 equiv) and anhydrous AlCl_3 (1.1 equiv) in CH_2Cl_2 , **1** (1 equiv) was slowly added and the mixture was stirred under nitrogen at 0°C for 4 hrs. Then, the reaction mixture was poured into a mixture of water/6 M HCl, stirred for 10 min, and extracted with CH_2Cl_2 . The combined organic layers were dried over MgSO_4 and evaporated to give the crude product. Recrystallization from MeOH/ H_2O solution of **2a-2c** was carried out to obtain the pure product as a yellow solid, and **2d** was purified by silica gel column chromatography to obtain the pure product as yellow viscous oil.

2-[4-(Diethylamino)-2-methoxybenzoyl]benzoic acid 2a

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz): δ 8.00 (dd, 1H, $J = 7.8, 0.8$ Hz), 7.65 (d, 1H, $J = 8.9$ Hz), 7.53 (td, 1H, $J = 8.9$ Hz), 7.41 (td, 1H, $J = 7.8, 1.1$ Hz), 7.27 (d, 1H, $J = 7.3$ Hz), 6.24 (dd, 1H, $J = 8.9, 2.3$ Hz), 5.99 (sd, 1H, $J = 2.1$ Hz), 3.55 (s, 3H), 3.40 (q, 4H, $J = 7.1$ Hz), 1.20 (t, 6H, $J = 7.1$ Hz). $^{13}\text{C-NMR}$ (CDCl_3 , 500 MHz): δ 193.9, 170.7, 162.2, 153.1, 146.4, 134.4, 132.3, 130.4, 127.9, 127.4, 126.9, 114.3, 103.8, 93.6, 55.3, 44.6, 12.6. HRMS (EI) calcd for $\text{C}_{18}\text{H}_{19}\text{NO}_4$ (M^+): 327.1471, Found: 324.1471. Yield: 60 %.

2-[4-(Dipropylamino)-2-methoxybenzoyl]benzoic acid 2b

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz): δ 8.00 (dd, 1H, $J = 7.8, 0.92$ Hz), 7.63 (d, 1H, $J = 8.7$ Hz), 7.52 (td, 1H, $J = 7.3, 1.1$ Hz), 7.40 (td, 1H, $J = 7.6, 1.1$ Hz), 7.26 (d, 1H, $J = 6.6$ Hz), 6.23 (dd, 1H, $J = 9.2, 2.3$ Hz), 5.97 (sd, 1H, $J = 2.3$ Hz), 3.54 (s, 3H), 3.28 (brs, 4H), 1.68-1.60 (m, 4H), 0.94 (t, 6H, $J = 7.3$ Hz). $^{13}\text{C-NMR}$ (CDCl_3 , 500 MHz): δ 193.8, 170.9, 162.1, 153.5, 146.5, 134.3, 132.3, 130.3, 127.9, 127.4, 126.8, 114.3, 104.0, 93.9, 55.2, 52.8, 20.5, 11.4. HRMS (EI) calcd for $\text{C}_{21}\text{H}_{25}\text{NO}_4$ (M^+): 355.1779, Found: 355.1786. Yield: 24 %.

2-[4-(Dibutylamino)-2-methoxybenzoyl]benzoic acid 2c

¹H-NMR (CDCl₃, 500 MHz): δ 8.01 (dd, 1H, J = 7.8, 0.92 Hz), 7.63 (brs, 1H), 7.53 (td, 1H, J = 7.3, 1.1 Hz), 7.41 (td, 1H, J = 7.8, 1.1 Hz), 7.26 (d, 1H, J = 7.3 Hz), 6.22 (dd, 1H, J = 9.2, 2.3 Hz), 5.98 (sd, 1H, J = 1.8 Hz), 3.55 (s, 3H), 3.31 (brs, 4H), 1.63-1.57 (m, 4H), 1.40-1.32 (m, 4H), 0.96 (t, 6H, J = 7.3 Hz). ¹³C-NMR (CDCl₃, 500 MHz): δ 193.9, 170.6, 164.7, 162.2, 153.5, 146.4, 134.4, 132.3, 130.4, 127.9, 127.4, 126.9, 114.2, 103.9, 93.8, 55.2, 50.8, 29.4, 20.2, 13.9. HRMS (EI) calcd for C₂₃H₂₉NO₄ (M⁺): 383.2097, Found: 383.2093. Yield: 23 %.

2-[4-(Dihexylamino)-2-methoxybenzoyl]benzoic acid 2d

¹H-NMR (CDCl₃, 500 MHz): δ 8.02 (dd, 1H, J = 7.8, 0.92 Hz), 7.63 (brs, 1H), 7.53 (td, 1H, J = 7.6, 1.4 Hz), 7.42 (td, 1H, J = 7.8, 1.4 Hz), 7.27 (d, 1H, J = 8.7 Hz), 6.21 (dd, 1H, J = 8.9, 2.3 Hz), 5.97 (sd, 1H, J = 1.8 Hz), 3.56 (s, 3H), 3.23 (t, 4H, J = 7.8 Hz), 1.62-1.58 (m, 4H), 1.40-1.32 (m, 12H), 0.89 (t, 6H, J = 7.1 Hz). ¹³C-NMR (CDCl₃, 500 MHz): 194.0, 170.4, 162.2, 153.5, 146.3, 134.4, 132.3, 131.9, 130.8, 130.5, 129.9, 128.6, 128.0, 127.5, 127.0, 114.2, 103.95, 93.8, 55.2, 52.8, 51.1, 31.6, 27.2, 26.7, 22.6, 14.0. HRMS (EI) calcd for C₂₇H₃₇NO₄ (M⁺): 439.2723, Found: 439.2730.

2-2. General Procedure for the synthesis of 3

To a stirred solution of **2** (1.0 equiv) in dry CH₂Cl₂ was added a solution of BBr₃ (1.9 equiv) in CH₂Cl₂ at -78 °C. After 1 hrs, the mixture was warmed to -25 °C. After completion of the reaction, the mixture was quenched with H₂O and evaporated to give the crude product. Recrystallization from MeOH/H₂O solution of **3a-3c** yielded the pure product as a yellow solid, and **3d** was purified by silica gel column chromatography to obtain the pure product as yellow viscous oil.

2-[4-(Diethylamino)-2-hydroxybenzoyl]benzoic acid 3a

¹H-NMR (CDCl₃, 500 MHz): δ 12.65 (s, 1H), 8.07 (dd, 1H, J = 7.8, 0.9 Hz), 7.58 (td, 1H, J = 7.6, 1.4 Hz), 7.51 (td, 1H, J = 7.8, 1.4 Hz), 7.34 (dd, 1H, J = 7.6, 1.1 Hz), 6.91 (d, 1H, J = 8.9 Hz), 6.11 (sd, 1H, J = 2.5 Hz), 6.03 (dd, 1H, J = 9.2, 2.5 Hz), 3.37 (q, 4H, J = 7.1 Hz), 1.18 (t, 6H, J = 7.1 Hz). ¹³C-NMR (CDCl₃, 500 MHz): δ 199.0, 167.5, 165.2, 153.6, 140.6, 134.6, 131.6, 130.4, 129.5, 128.9, 127.6, 110.1, 103.5, 96.7, 44.5, 12.5. HRMS (EI) calcd for C₁₉H₂₁NO₄ (M⁺): 313.1314, Found: 313.1313. Yield: 65 %.

2-[4-(Dipropylamino)-2-hydroxybenzoyl]benzoic acid 3b

Yield: 1.0 g (52 %); $^1\text{H-NMR}$ (CDCl_3 , 500 MHz): δ 12.53 (s, 1H), 8.10 (d, 1H, $J = 7.1$ Hz), 7.62 (t, 1H, $J = 7.3$ Hz), 7.53 (t, 1H, $J = 7.3$ Hz), 7.36 (dd, 1H, $J = 7.8, 1.1$ Hz), 6.87 (d, 1H, $J = 9.2$ Hz), 6.12 (s, 1H), 6.03 (d, 1H, $J = 8.9$ Hz), 3.26 (t, 4H, $J = 7.6$ Hz), 1.65-1.59 (m, 4H), 0.92 (t, 6H, $J = 7.3$ Hz). $^{13}\text{C-NMR}$ (CDCl_3 , 500 MHz): δ 198.1, 169.8, 165.4, 154.4, 141.2, 134.5, 132.7, 131.1, 129.1, 128.1, 127.6, 109.9, 104.0, 97.3, 52.8, 20.5, 11.3. HRMS (EI) calcd for $\text{C}_{20}\text{H}_{23}\text{NO}_4$ (M^+): 341.1630, Found: 341.1626.

2-[4-(Dibutylamino)-2-hydroxybenzoyl]benzoic acid 3c

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz): δ 12.54 (s, 1H), 8.12 (d, 1H, $J = 7.8$ Hz), 7.63 (t, 1H, $J = 7.3$ Hz), 7.54 (t, 1H, $J = 7.6$ Hz), 7.36 (dd, 1H, $J = 7.6, 0.92$ Hz), 6.87 (d, 1H, $J = 8.9$ Hz), 6.11 (s, 1H), 6.03 (d, 1H, $J = 8.5$ Hz), 3.29 (t, 4H, $J = 7.3$ Hz), 1.58 (brs, 4H), 1.37-1.30 (m, 4H), 0.94 (t, 6H, $J = 7.3$ Hz). $^{13}\text{C-NMR}$ (CDCl_3 , 500 MHz): δ 198.1, 169.5, 165.5, 154.4, 141.1, 134.5, 132.7, 131.1, 129.2, 128.1, 127.6, 109.8, 103.9, 97.2, 50.9, 29.4, 20.2, 13.9. HRMS (EI) calcd for $\text{C}_{22}\text{H}_{27}\text{NO}_4$ (M^+): 369.1940, Found: 369.1932. Yield: 49 %.

2-[4-(Dihexylamino)-2-hydroxybenzoyl]benzoic acid 3d

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz): δ 12.55 (s, 1H), 8.10 (dd, 1H, $J = 8.0, 0.92$ Hz), 7.62 (dd, 1H, $J = 7.6, 1.4$ Hz), 7.53 (td, 1H, $J = 7.8, 1.4$ Hz), 7.34 (dd, 1H, $J = 7.1, 0.92$ Hz), 6.87 (d, 1H, $J = 9.2$ Hz), 6.11 (sd, 1H, $J = 2.5$ Hz), 6.02 (dd, 1H, $J = 9.2, 2.5$ Hz), 3.27 (brs, 4H), 1.59 (brs, 4H), 1.33-1.30 (m, 12H), 0.90-0.88 (m, 6H). $^{13}\text{C-NMR}$ (CDCl_3 , 500 MHz): δ 198.1, 169.0, 165.5, 154.4, 141.1, 134.5, 132.6, 131.1, 129.1, 128.1, 127.7, 109.8, 103.9, 97.2, 51.1, 31.59, 27.3, 26.7, 22.6, 14.0. HRMS (EI) calcd for $\text{C}_{26}\text{H}_{35}\text{NO}_4$ (M^+): 425.2563, Found: 425.2569.

2-3. General Procedure for the synthesis of **4**

3 (2.0 equiv) and resorcinol (1.0 equiv) were combined in methanesulfuric acid (2.0 mL) in a sealed tube and heated at 90 °C for 2 hrs. The reaction was poured into stirred ice water, its pH was adjusted to 11~12 with 1.0 M sodium hydroxide aqueous solution, and the mixture was stirred for 20 min. Then, the mixture was extracted with CH₂Cl₂ three times. The organic layers were dried over MgSO₄ and evaporated to give the crude product. This was purified by silica gel column chromatography and further recrystallized from acetonitrile solution to obtain the pure product as a white solid.

3',3''-Bis(oxospiroisobenzofuran)-3,7-bis(diethylamino)benzopyranoxanthene (ABPX01) 4a

¹H-NMR (THF-d₈, 500 MHz): δ 7.73-7.72 (m, 1H), 7.70 (dt, 1H, J = 7.6, 1.1 Hz), 7.61 (td, 1H, J = 7.6, 0.92 Hz), 7.52 (td, 1H, J = 7.3, 0.9 Hz), 7.47-7.43 (m, 2H), 7.16 (dt, 1H, J = 7.6, 0.9 Hz), 7.11 (s, 1H), 6.94-6.92 (m, 1H), 6.50-6.49 (m, 2H), 6.47 (d, 1H, J = 7.6 Hz), 6.45 (d, 1H, J = 7.6 Hz), 6.37-6.34 (m, 2H), 6.07 (s, 1H), 3.30-3.25 (m, 8H), 0.92 (t, 12H, J = 7.6 Hz). ¹³C-NMR (CDCl₃, 500 MHz): δ 168.4, 168.3, 154.0, 153.9, 153.8, 153.7, 153.1, 153.0, 151.0, 150.9, 135.5, 134.8, 130.3, 130.0, 129.7, 129.4, 129.3, 128.8, 128.4, 128.1, 125.0, 124.9, 124.8, 124.4, 118.00, 117.8, 109.6, 106.6, 106.5, 104.4, 104.3, 98.6, 98.5, 83.0, 82.8, 53.4, 11.5. ¹³C-NMR (CDCl₃, 500 MHz): δ HRMS (FAB) calcd for C₄₂H₃₇N₂O₆ (M+H): 665.2651, Found: 665.2654. Yield: 72 %.

3',3''-Bis(oxospiroisobenzofuran)-3,7-bis(dipropylamino)benzopyranoxanthene (ABPX02) 4b

¹H-NMR (THF-d₈, 500 MHz): δ 7.73-7.72 (m, 1H), 7.70 (dt, 1H, J = 7.6, 1.1 Hz), 7.61 (td, 1H, J = 7.6, 0.92 Hz), 7.52 (td, 1H, J = 7.3, 0.9 Hz), 7.47-7.43 (m, 2H), 7.16 (dt, 1H, J = 7.6, 0.9 Hz), 7.11 (s, 1H), 6.94-6.92 (m, 1H), 6.50-6.49 (m, 2H), 6.47 (d, 1H, J = 7.6 Hz), 6.45 (d, 1H, J = 7.6 Hz), 6.37-6.34 (m, 2H), 6.07 (s, 1H), 3.30-3.25 (m, 8H), 1.65-1.57 (m, 8H), 0.92 (t, 12H, J = 7.6 Hz). ¹³C-NMR (CDCl₃, 500 MHz): δ 168.4, 168.3, 154.0, 153.9, 153.8, 153.7, 153.2, 153.0, 151.0, 150.9, 135.5, 134.8, 130.3, 130.0, 129.7, 129.4, 129.3, 128.8, 128.4, 128.1, 125.0, 124.9, 124.8, 124.4, 118.0, 117.8, 109.6, 106.6, 106.5, 104.4, 104.3, 98.6, 98.5, 83.0, 82.8, 53.4, 21.1, 11.5. HRMS (FAB) calcd for C₄₆H₄₅N₂O₆ (M+H): 721.3278, Found: 721.3276. Yield: 56 %.

3',3''-Bis(oxospiroisobenzofuran)-3,7-bis(dibuthylamino)benzopyranoxanthene

(ABPX03) 4c

¹H-NMR (THF-d₈, 500 MHz): δ 7.74-7.72 (m, 1H), 7.70 (dt, 1H, J = 7.6, 0.92 Hz), 7.62 (td, 1H, J = 7.6, 1.1 Hz), 7.52 (td, 1H, J = 7.3, 0.92 Hz), 7.47-7.43 (m, 2H), 7.16 (dt, 1H, J = 7.6, 0.92 Hz), 7.13 (s, 1H), 6.94-6.92 (m, 1H), 6.50-6.49 (m, 2H), 6.47 (d, 1H, J = 7.8 Hz), 6.45 (d, 1H, J = 7.8 Hz), 6.37-6.34 (m, 2H), 6.07 (s, 1H), 3.33-3.30 (m, 8H), 1.60-1.54 (m, 8H), 1.40-1.32 (m, 8H), 0.95 (t, 12H, J = 7.3 Hz). ¹³C-NMR (CDCl₃, 500 MHz): δ 168.4, 168.3, 154.0, 153.9, 153.8, 153.7, 153.2, 153.0, 151.0, 150.9, 135.4, 134.8, 130.3, 130.0, 129.7, 130.0, 129.4, 128.8, 128.5, 128.1, 125.0, 124.9, 124.8, 124.4, 118.0, 117.8, 109.6, 106.6, 106.5, 104.5, 104.4, 98.6, 98.5, 83.0, 82.8, 51.4, 30.2, 21.0, 14.3. HRMS (FAB) calcd for C₅₀H₅₃N₂O₆ (M+H): 777.3903, Found: 777.3911. Yield: 52 %.

3',3''-bis(oxospiroisobenzofuran)-3,7-bis(dihexylamino)benzopyranoxanthene

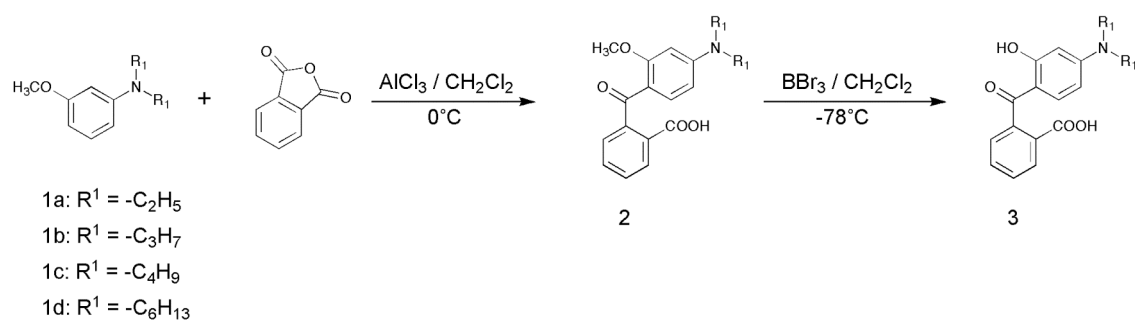
(ABPX04) 4d

¹H-NMR (THF-d₈, 500 MHz): δ 7.74-7.72 (m, 1H), 7.70 (d, 1H, J = 7.8 Hz), 7.62 (td, 1H, J = 7.6, 0.92 Hz), 7.52 (td, 1H, J = 7.6, 0.92 Hz), 7.47-7.44 (m, 2H), 7.16 (d, 1H, J = 7.6 Hz), 7.12 (s, 1H), 6.94-6.92 (m, 1H), 6.49-6.48 (m, 2H), 6.47 (d, 1H, J = 8.7 Hz), 6.45 (d, 1H, J = 8.0 Hz), 6.36-6.34 (m, 2H), 6.06 (s, 1H), 3.31 (brs, 8H), 1.59 (brs, 8H), 1.33 (brs, 24H), 0.90 (brs, 12H). ¹³C-NMR (CDCl₃, 500 MHz): δ 168.4, 168.3, 154.0, 154.0, 153.9, 153.8, 153.7, 153.2, 153.0, 151.0, 150.9, 150.8, 135.4, 134.8, 130.3, 130.0, 129.7, 129.4, 129.3, 128.7, 128.5, 128.1, 125.0, 125.9, 124.8, 124.4, 118.0, 117.8, 109.6, 106.6, 106.5, 104.5, 104.4, 98.6, 98.5, 83.0, 82.8, 51.7, 32.7, 28.0, 27.6, 23.6, 14.4. HRMS (FAB) calcd for C₅₈H₆₉N₂O₆ (M+H): 889.5155, Found: 889.5165

3. Crystal data and experimental data of ABPX01⁰

Crystal and experimental data: C₄₂H₃₇N₂O₆, M = 665.76, orthorhombic, Fdd2, $a = 17.5837$ (11), $b = 23.2759$ (11), $c = 18.2237$ (9) Å, $\alpha, \beta, \gamma = 90^\circ$, $V = 7458.5$ (7) Å³, $Z = 8$, $D_x = 1.186$ g cm⁻³, $F(000) = 2808$, μ (Mo K α) = 0.079 mm⁻¹. The data were collected on a Rigaku RAXIS-RAPID area detector using graphite-monochromated MoK α radiation at 293.1 K. A total of 17,587 reflections were measured up to $\theta_{\max} = 27.46^\circ$ (0.85 Å resolution) and merged to 14,495 reflections with $R_{\text{int}} = 0.034$. The structure was solved using CrystalStructure 3.8 and refined with CrystalStructure 3.8. Non-hydrogen atoms were refined anisotropically. Hydrogen atoms were calculated at the ideal positions and isotropically included in the calculations of the structure factors. The structure was converted at goodness of fit = 1.232, $(\Delta/\sigma)_{\max} = 0.0088$, $\Delta\rho_{\max} = 2.50$ eÅ⁻³, $\Delta\rho_{\min} = -2.27$ eÅ⁻³, $R = 0.0589$, and $wR = 0.0772$. Crystallographic data (excluding structure factors) for the structure of ABPX01⁰ reported in this paper are deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC-779022. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

4. Scheme and figures



Scheme S1 Synthetic routes of benzophenone derivatives.

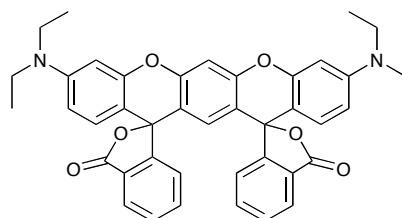


Fig. S1 Chemical structure of ABPX01⁰.

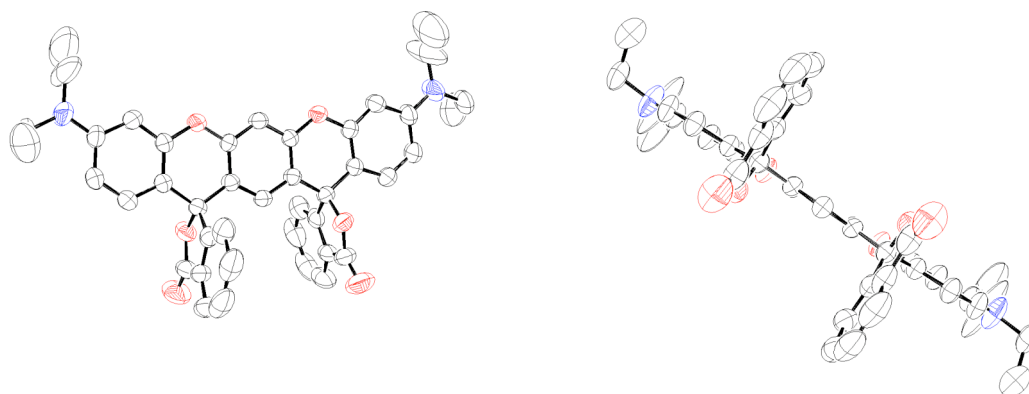


Fig. S2 ORTEP drawing of ABPX01⁰. The trans-ABPX01⁰ was confirmed in solids, of which five rings had a high degree of planarity.

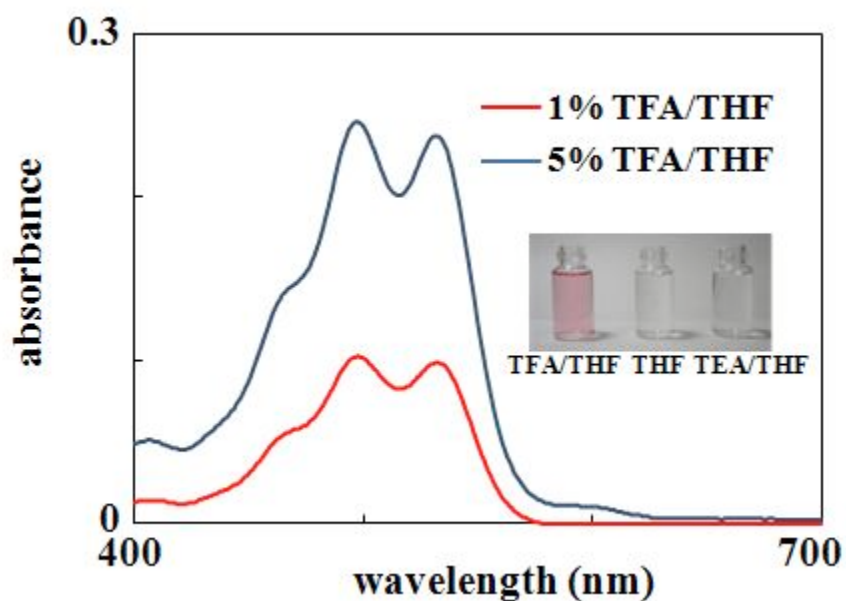


Fig. S3 Effect of protolytic reaction on absorption spectra of ABPX01 in TFA/THF solutions. TFA: trifluoroacetic acid, TEA: triethylamine. Inset: photograph of the solutions under room light. In strong acid solution, the absorbance spectra having two peaks at 490 nm and 540 nm should be attributable to the cationic species ABPX01^+ . In THF and alkaline solutions, the spectra had no peaks that were attributable to the neutral lactonoid species ABPX01^0 .

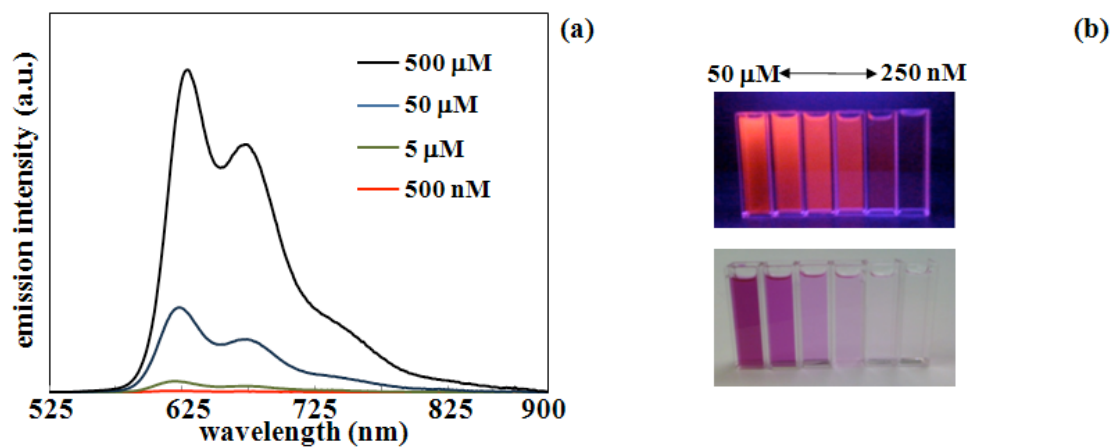


Fig. S4 (a) Emission at various concentrations of ABPX01⁺. (b) photograph of the solutions, top: under 365 nm irradiation, bottom: under room light irradiation.

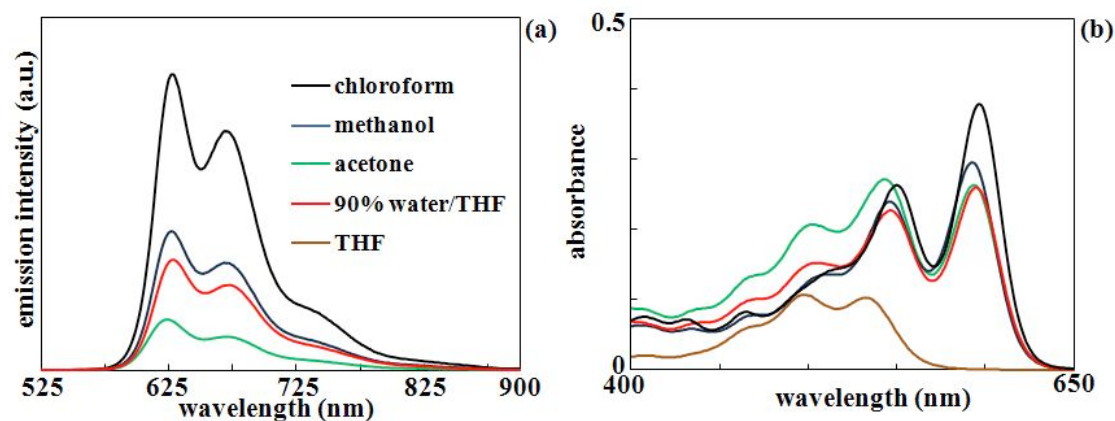


Fig. S5 (a) Emission spectra of 500 μM of ABPX01⁺ and (b) absorption spectra of 5 μM of ABPX01⁺ in various solvents. The emission spectrum in THF was not observed. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

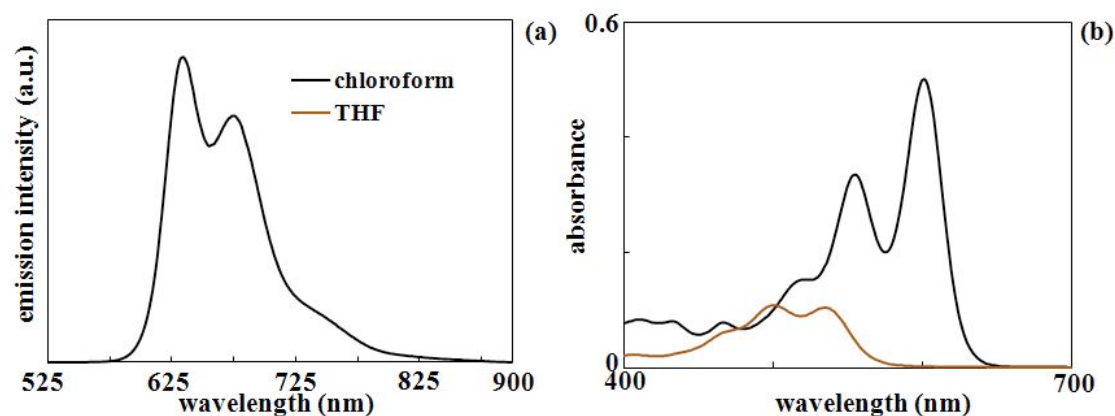


Fig. S6 (a) Emission spectra of 500 μM of ABPX02⁺ and (b) absorption spectra of 5 μM of ABPX02⁺ in various solvents. The emission spectrum in THF was not observed. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

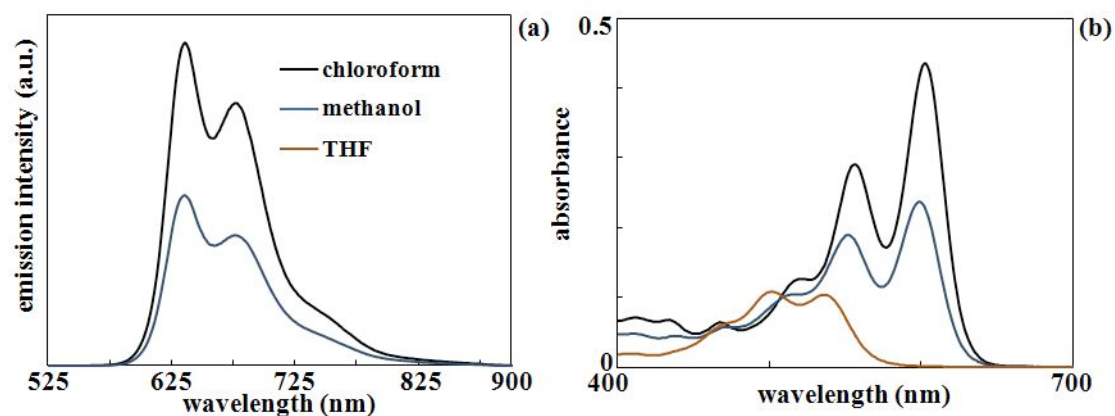


Fig. S7 (a) Emission spectra of 500 μM of ABPX03⁺ and (b) absorption spectra of 5 μM of ABPX03⁺ in various solvents. The emission spectrum in THF was not observed. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

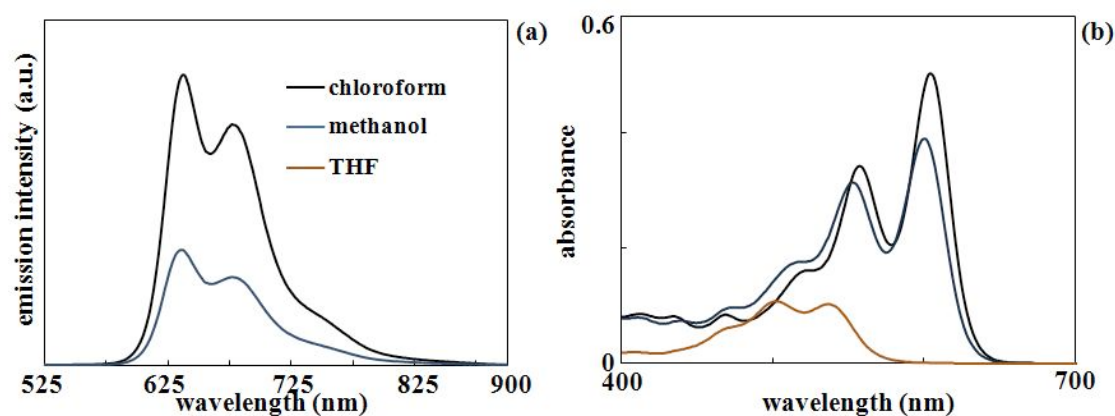


Fig. S8 (a) Emission spectra of 500 μM of ABPX04⁺ and (b) absorption spectra of 5 μM of ABPX04⁺ in various solvents. The emission spectrum in THF was not observed. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

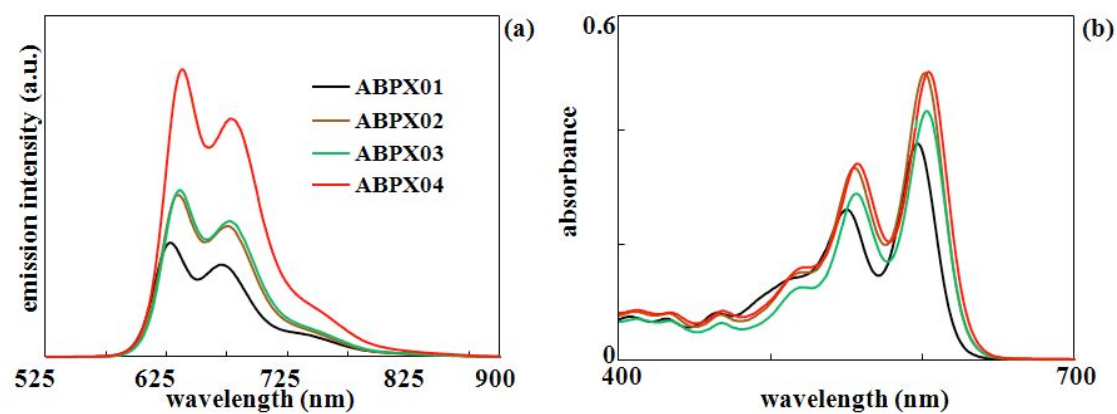


Fig. S9 (a) Emission and (b) absorption spectra of ABPX01⁺-04⁺ in chloroform. A slightly continuous red shift was observed with an increase in *N*-alkyl chain length. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

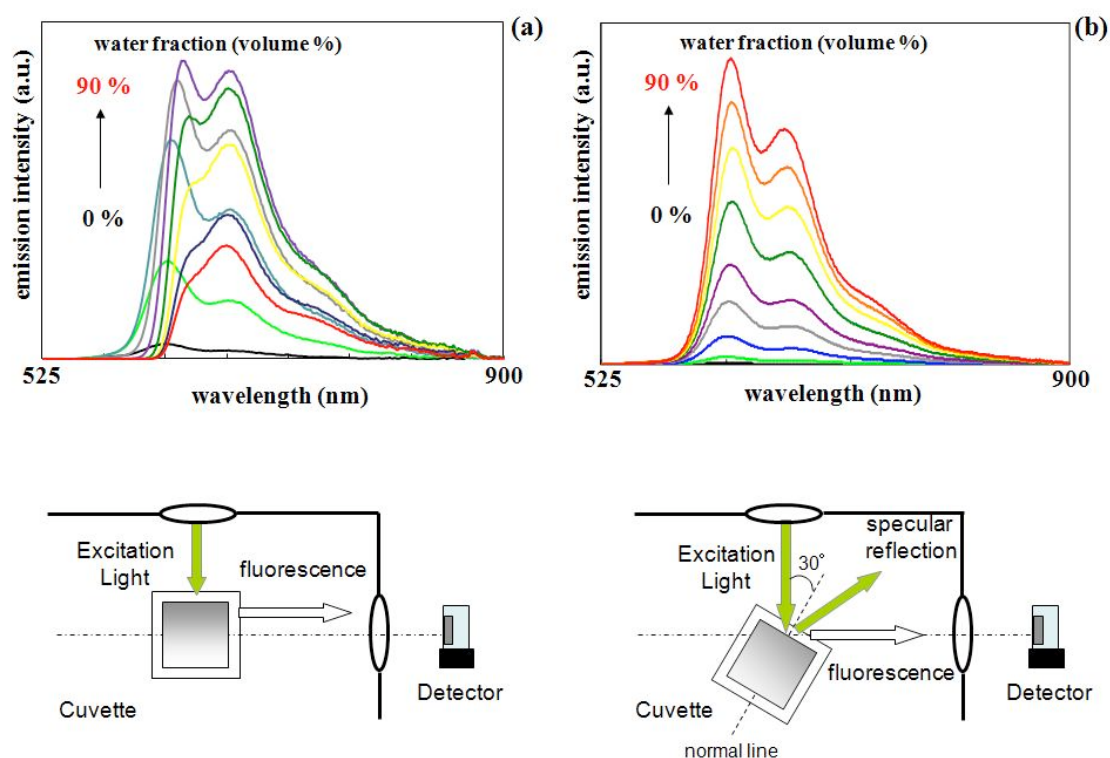


Fig. S10 Emission spectra of ABPX01 by (a) conventional photometric method and (b) surface photometric method. Shown below are the respective equipment configurations. The emission spectra by the conventional photometric method showed random shifts of the maximum wavelengths, which were caused by the internal filter effect when measurement of ABPX01⁺ was conducted at a high concentration. The problem was solved by using the surface photometric method. The sample holder was set at an angle of 30° with respect to the excitation light. However, attention should be paid to the linearity of the emission intensity against the concentration for the accurate evaluation of high-concentration sample solutions, such as AIEE molecules. In addition, the relative quantum yields of ABPX in solution could not be determined because of the lack of a standard compound. We are now developing an apparatus for the measurement of the absolute fluorescence quantum yields of high-concentration solutions.

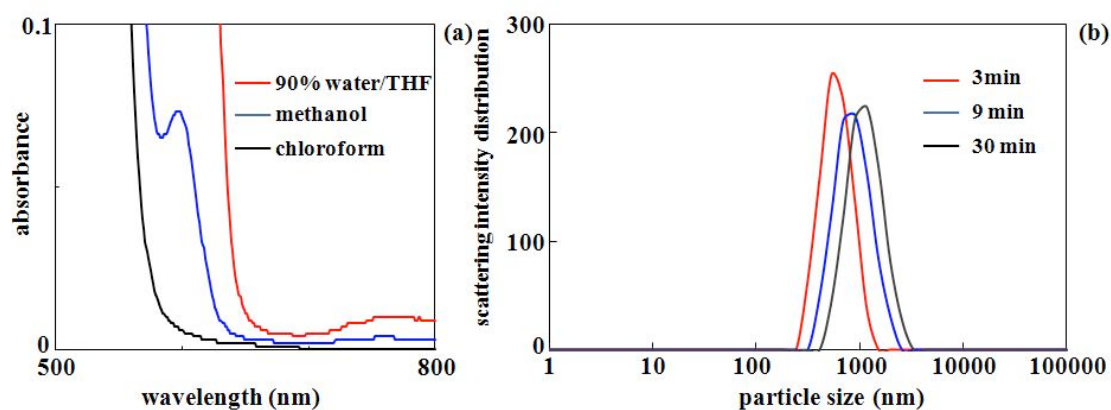


Fig. S11 (a) Level-off tails of 500 μM of ABPX01⁺ were observed in chloroform, methanol, and 90% water/THF mixture solutions. In comparison, level-off tails of 500 μM of ABPX01⁺ in chloroform could not be observed. (b) Dynamic laser scattering (time dependence of particle size distributions of 500 μM of ABPX01⁺ in 90% water/THF mixtures). The nanoaggregate size of ABPX01⁺ in solutions was increased as a time-dependent change. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

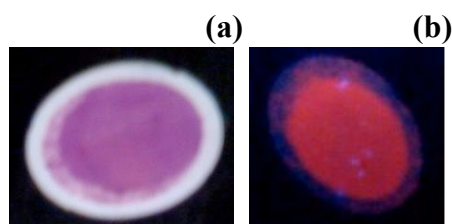


Fig. S12 Membrane filter photographs of ABPX01⁺ in 90% water/THF mixtures (containing 1% TFA) under (a) room light and (b) 365 nm irradiation.

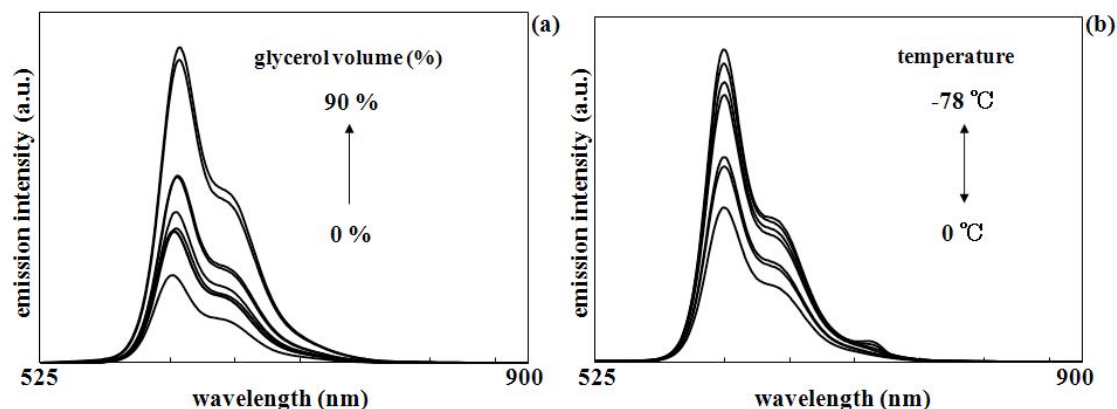
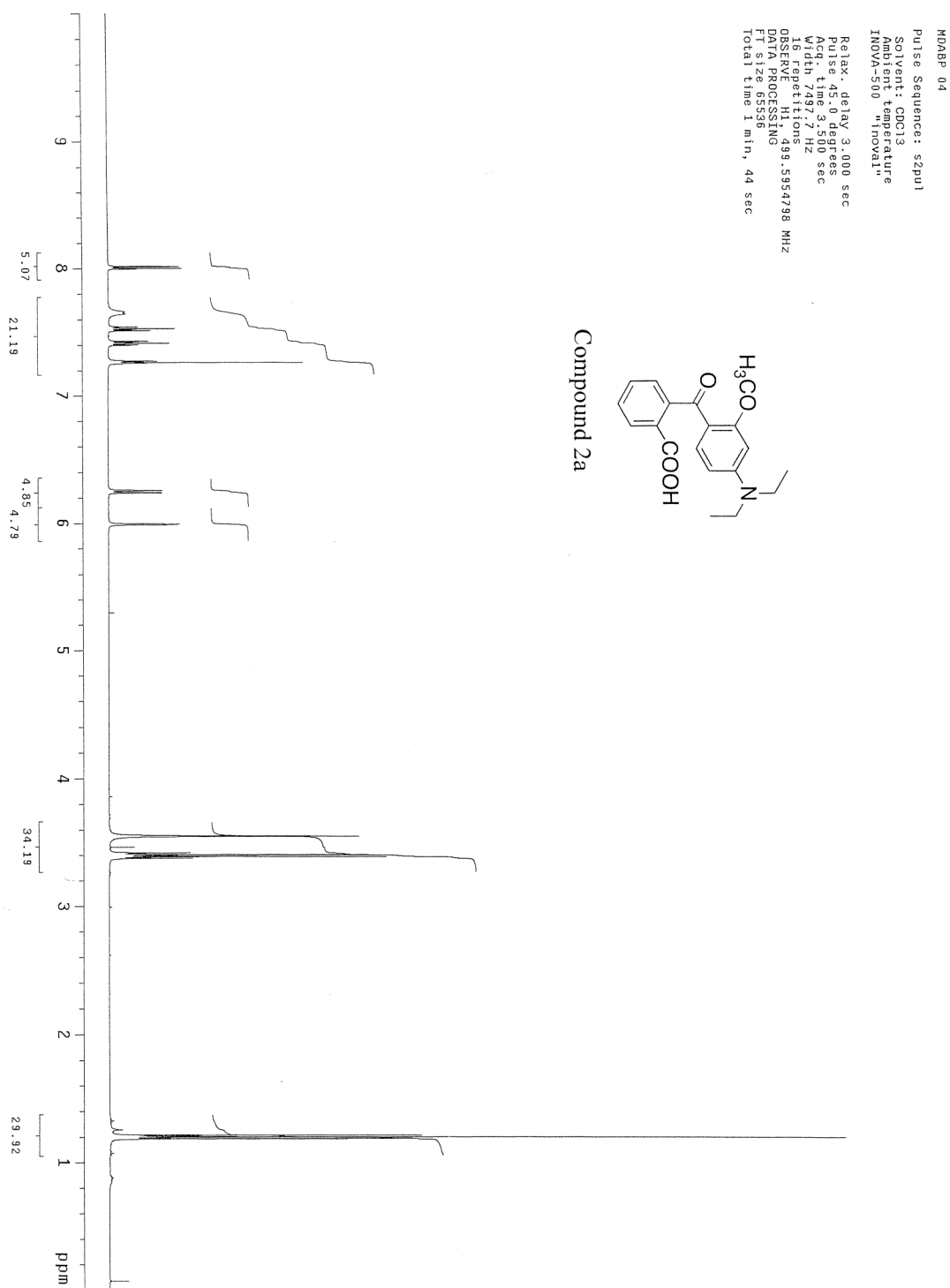
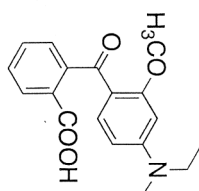


Fig S13. (a) Effects of composition of glycerol-methanol mixture on emission intensity of ABPX01. In the mixtures with glycerol fractions > 80%, the emission intensity rapidly increased due to the viscosity effect (viscchromism). (b) Effects of temperature on emission intensity of ABPX01 in methanol. When a methanol solution of ABPX01 was cooled, its emission intensity was increased due to thermochromism. The above results suggested that RIR caused AIEE. 1.0% TFA was added to ensure protonation and prevent pseudobase formation during measurement.

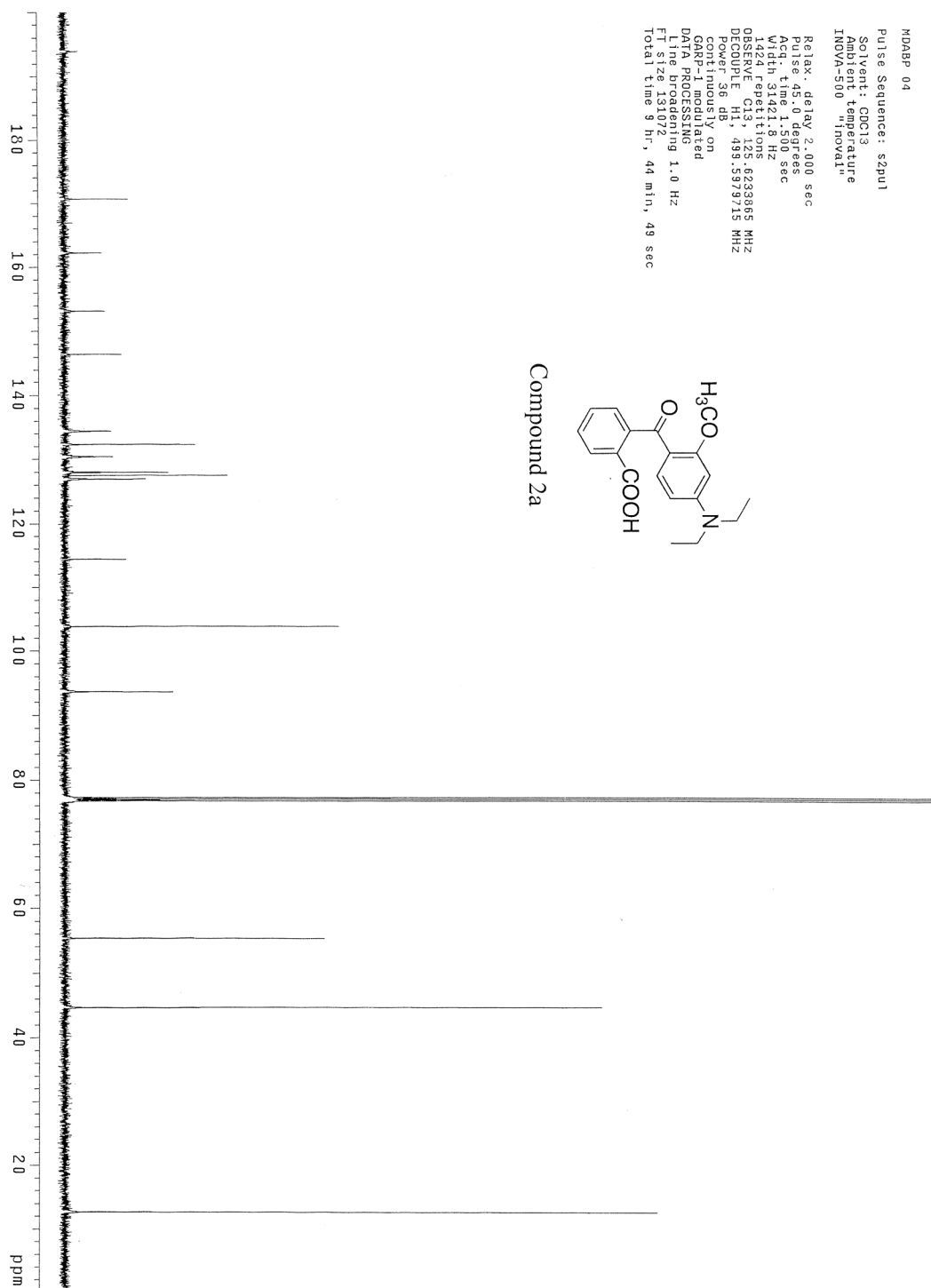
5. NMR data of benzophenone derivatives and ABPX01-04



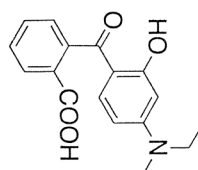
MDAP 04
Pulse Sequence: szpu1
Solvent: CDCl3
Ambient temperature
INOVA-500 "nova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
1424 repetitions
OBSERVE C13, 125.623365 MHz
P1 11.435.9379715 MHz
PC 1.56 dB
continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



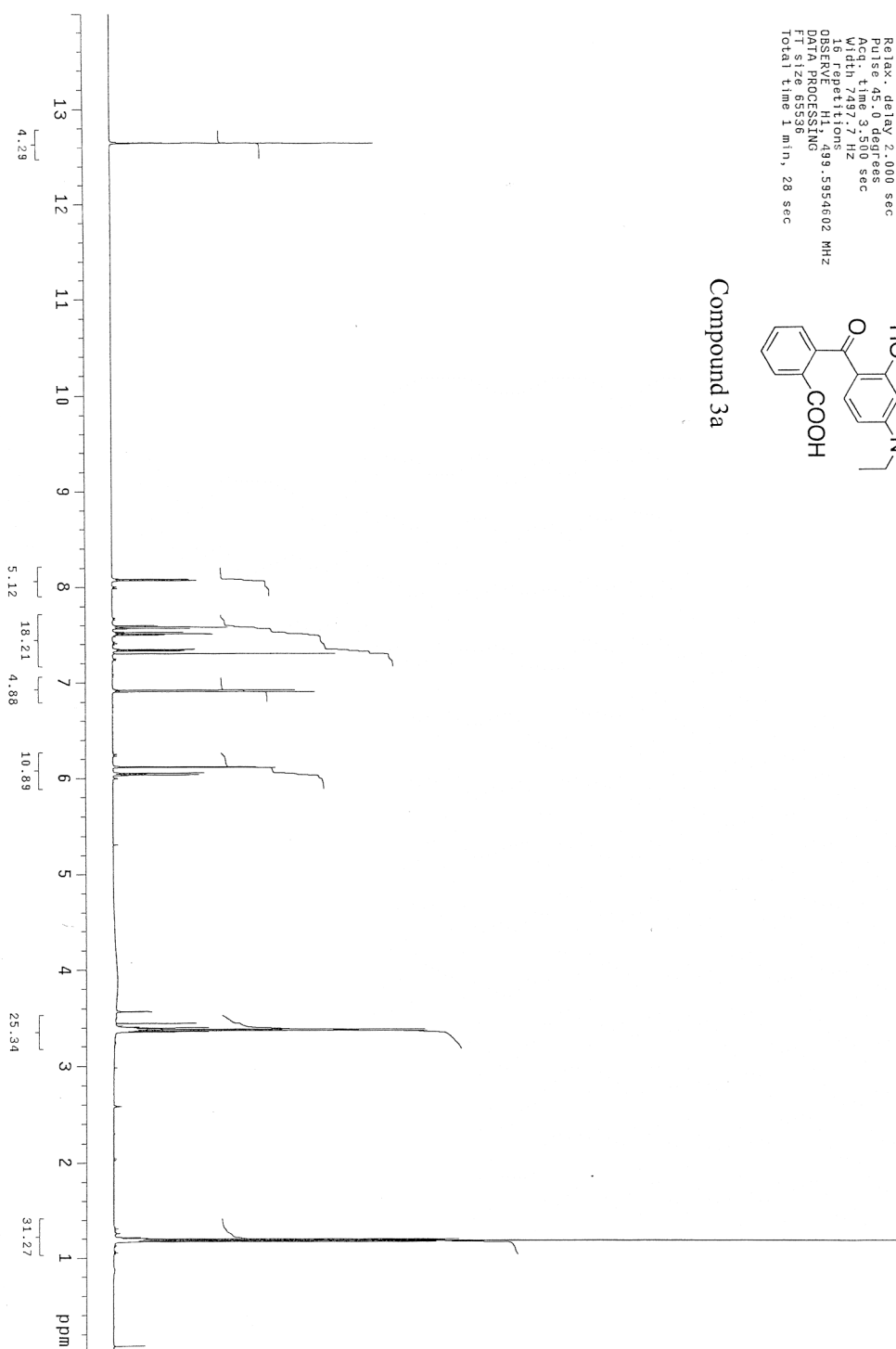
Compound 2a



HDABP 04
Pulse Sequence: s2pul
Solvent: CDCl3
Ambient temperature
INDVA-500 "nova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 3.500 sec
Width 7497.7 Hz
Spectrum 1099.5954602 MHz
03SEVY4118
DATA PROCESSING
FT size 65536
Total time 1 min, 28 sec

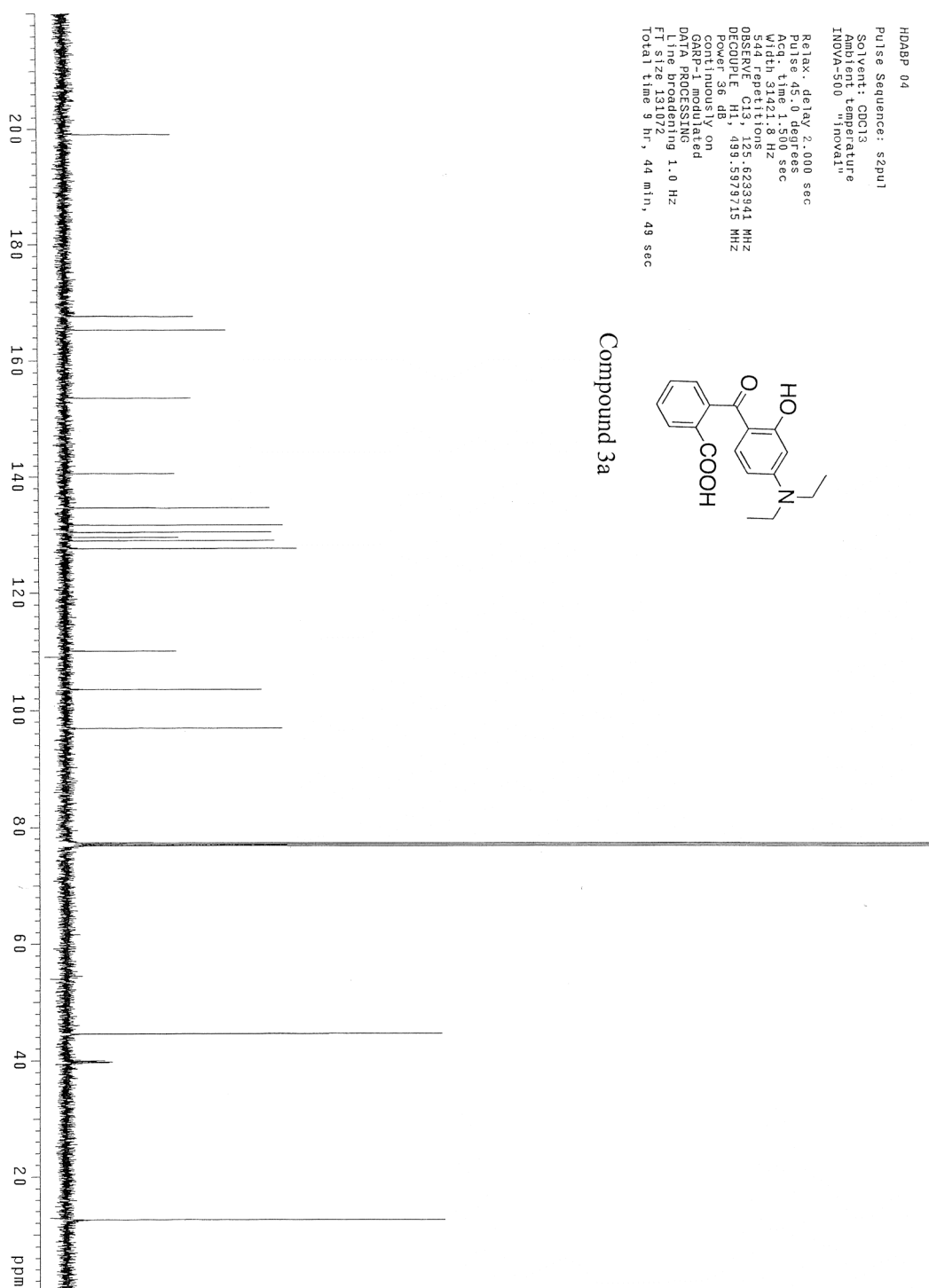
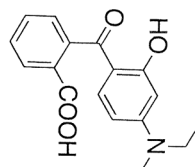


Compound 3a

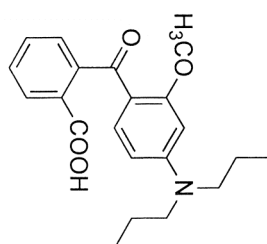


HDABP 04
Pulse Sequence: s2pu1
Solvent: CDCl3
Ambient temperature
INDVA-500 "jnova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
344 repetitions
633840.000 Hz
DECOUPLE CH1 439.5979715 MHz
Power 36 db
continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 3 hr, 44 min, 49 sec

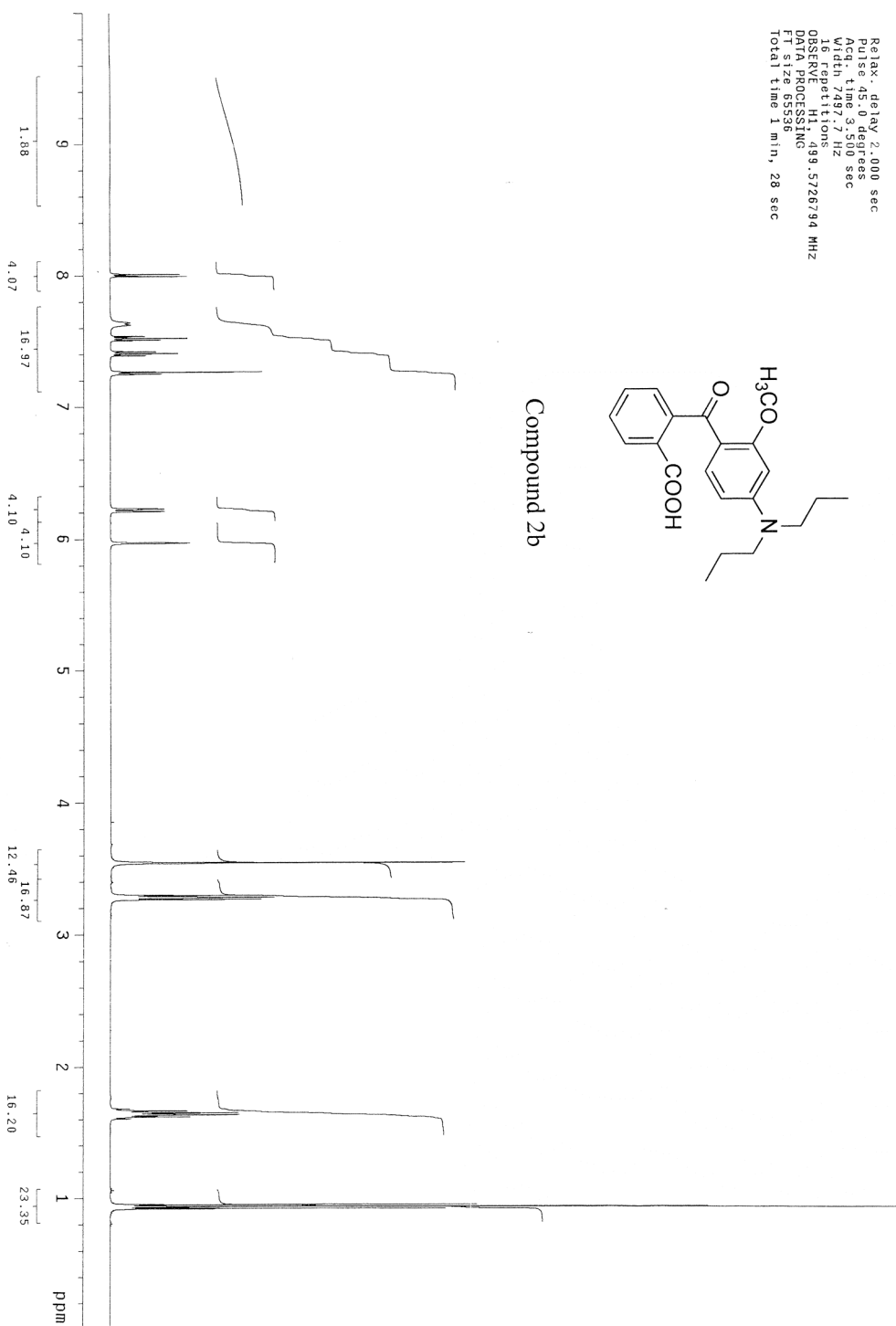
Compound 3a



OLM02-C3
Pulse Sequence: szpu1
Solvent: CDCl3
Ambient temperature
INOVA-500 "Inovair"
Relax. delay 2.000 sec
Pulse time 0.000 sec
Acq. time 3.550 sec
Width 7497.7 Hz
16 repetitions
OBSERVE H1, 499.5726794 MHz
DATA PROCESSING
FT size 65536
Total time 1 min, 28 sec



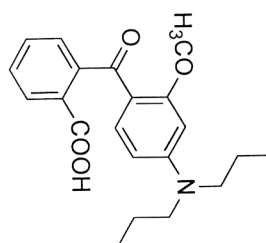
Compound 2b



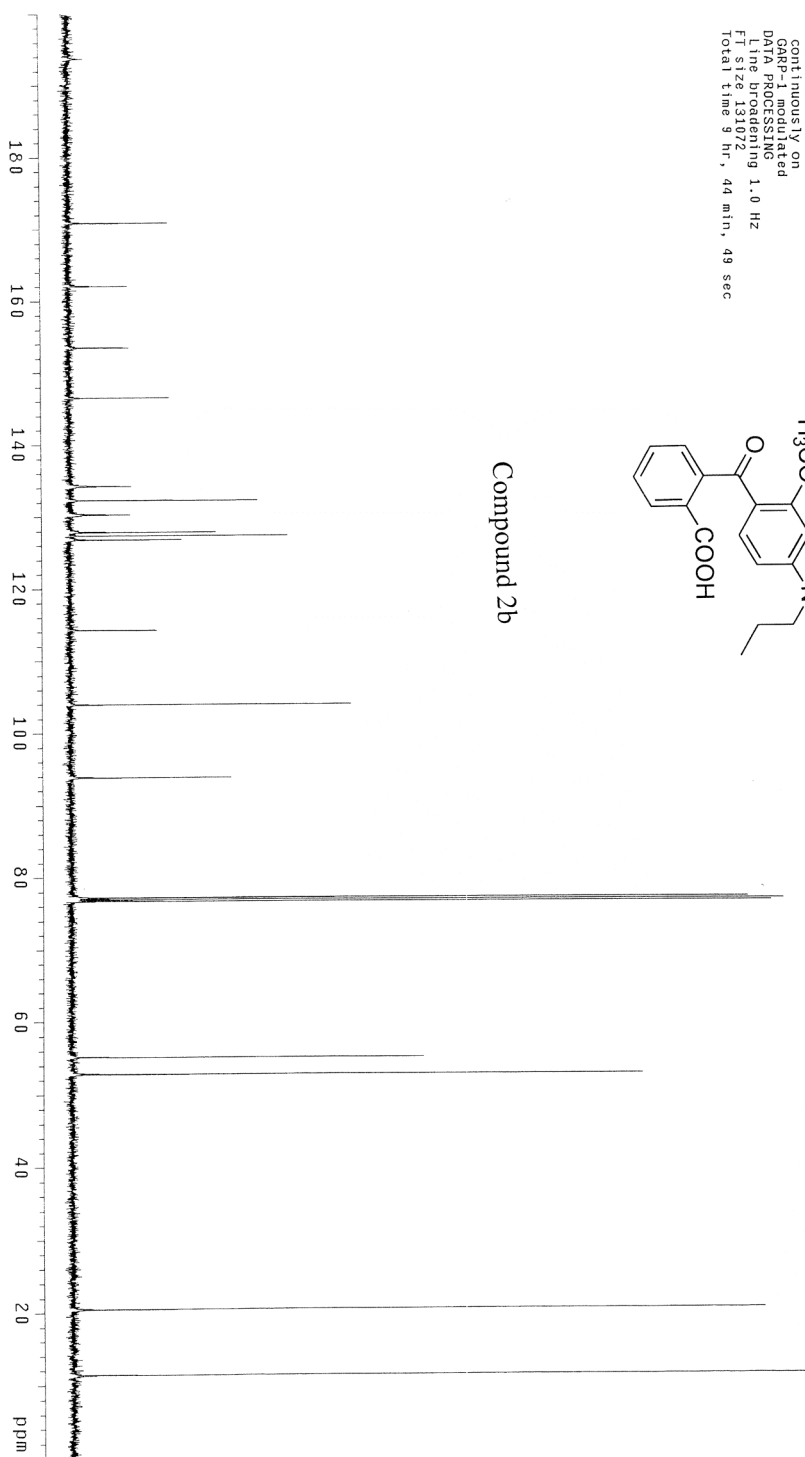
OLM02-C3

Pulse Sequence: s2pul
Solvent: CDCl3
Ambient temperature
INNOVA-500 "nova1"

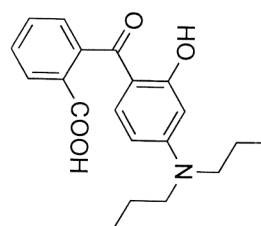
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.500 sec
SOLVENT CDCl3
592 repetitions
OBSERVE C13, 125.6176562 MHz
DECOUPLE H1, 499.5751711 MHz
Power 36 db
continuously on
GARP-1 modulated
DATA PROCESSING
1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



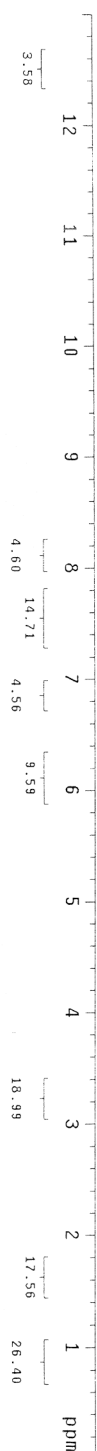
Compound 2b



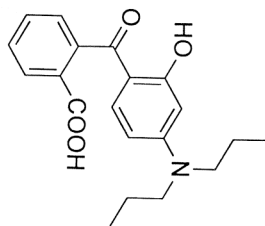
OLM02-C4
Pulse Sequence: s2pul
Solvent: CDCl3
Ambient temperature
INNOVA-500 "INNOVA1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acquisition 3.50 sec
Width 7487.2 Hz
32 repetitions
OBSERVE H1: 499.5726803 MHz
DATA PROCESSING
F1 size 65536
Total time 2 min, 56 sec



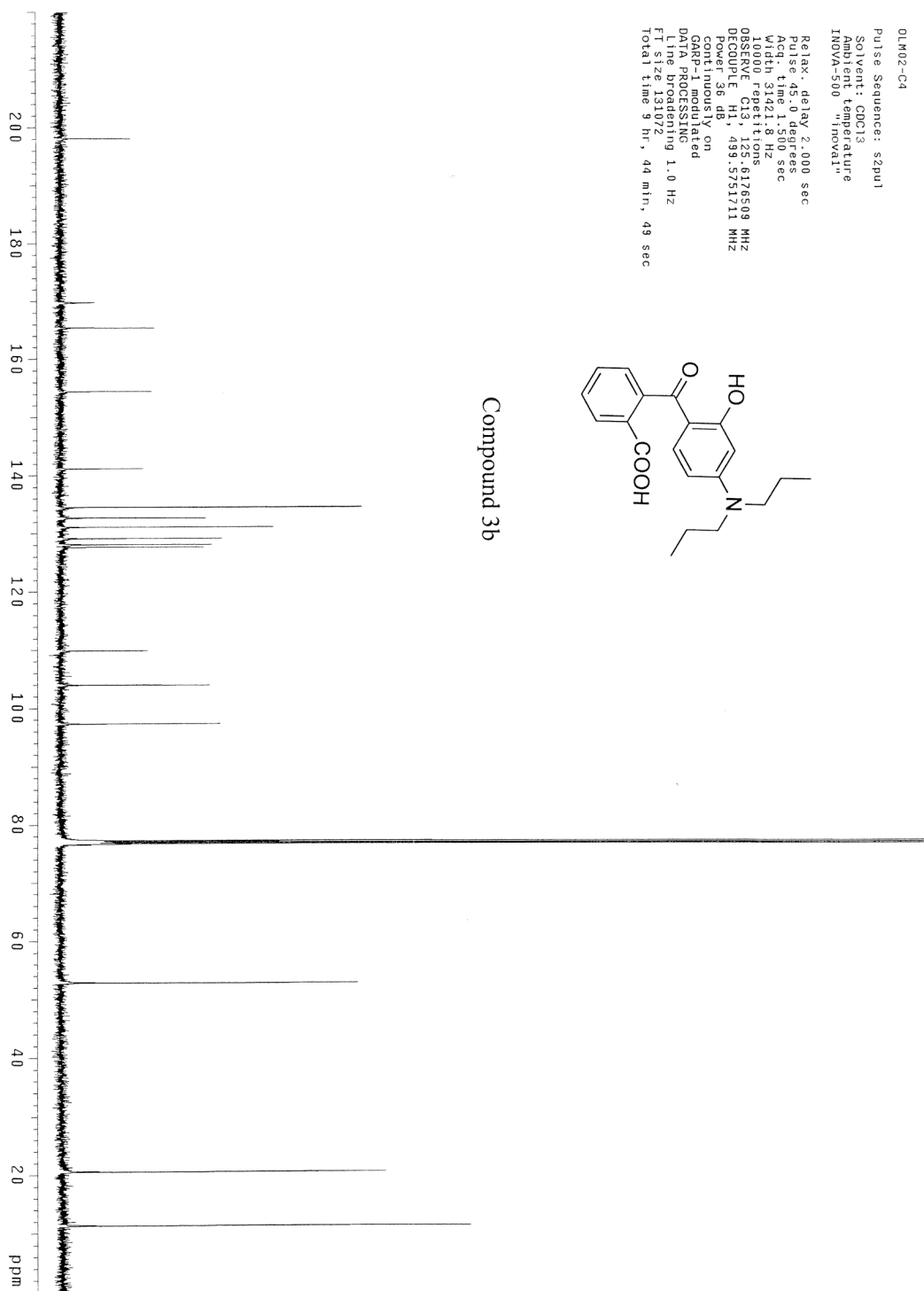
Compound 3b



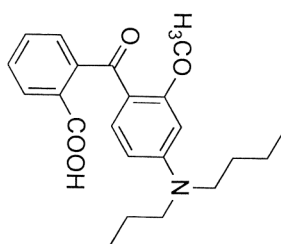
OLM02-C4
Pulse Sequence: s2pul
Solvent: CDCl3
Ambient temperature
INOVA-500 "Inova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Pulse time 1.500 sec
Width 31621.8 Hz
10000 repetitions
OBSERVE C13, 125.6176509 MHz
DECOUPLE H1, 499.5751711 MHz
Power 36 db
continuously on
GARP modulated
D1 1.00000000
line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



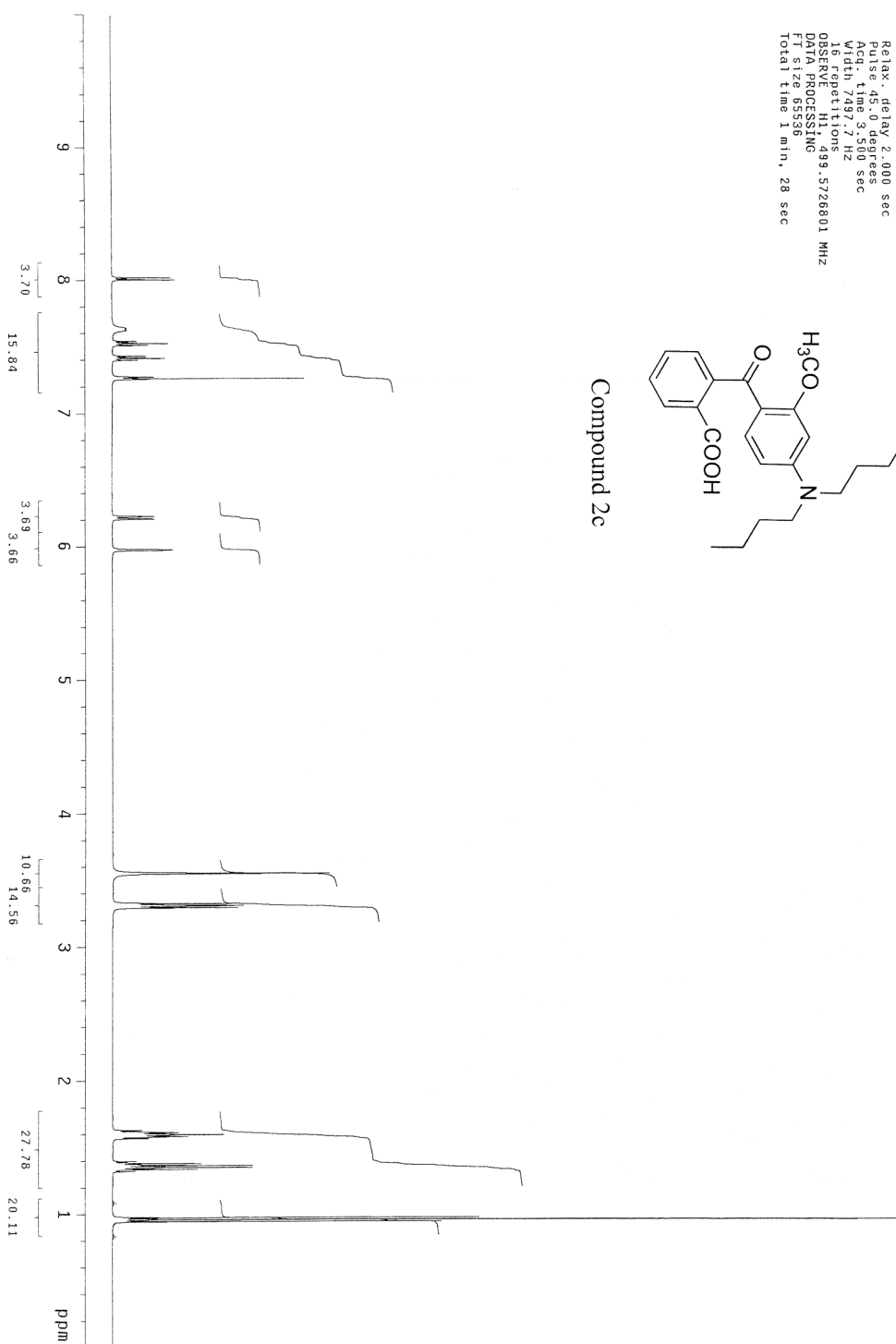
Compound 3b



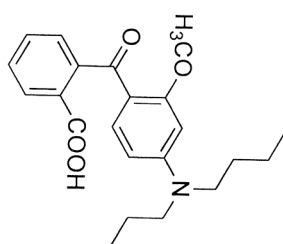
OLM03-C3
Pulse Sequence: zgpg30
Solvent: CDCl3
Ambient Temperature
INOVA-500 100VAT
Relax. delay 2.000 sec
Acq. se 45.000 sec
Acq. time 3.500 sec
Width 7497.7 Hz
16 repetitions
OBSERVE H1, 499.5726801 MHz
DATA PROCESSING
FT size 65536
Total time 1 min, 28 sec



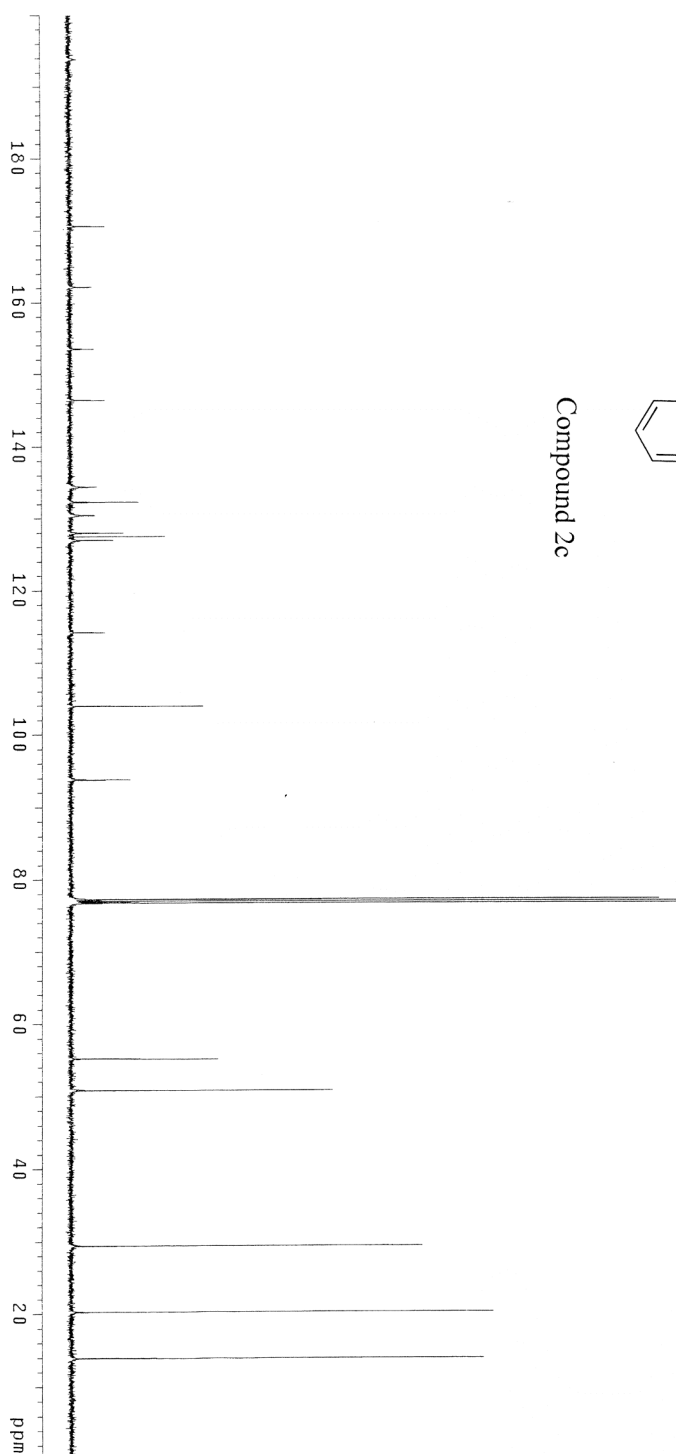
Compound 2c



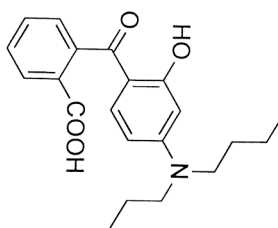
OLM03-C3
Pulse Sequence: szpu1
Solvent: CDCl3
Ambient temperature
INOVA-500 "Inova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq time 1.500 sec
Width 31421.8 Hz
944 repetitions
OBSERVE C13, 125.617653 MHz
DECOUPLE H1, 499.5751711 MHz
Power 36 db
continuously on
GARP1 modulated
DARRP crosspolarization
line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



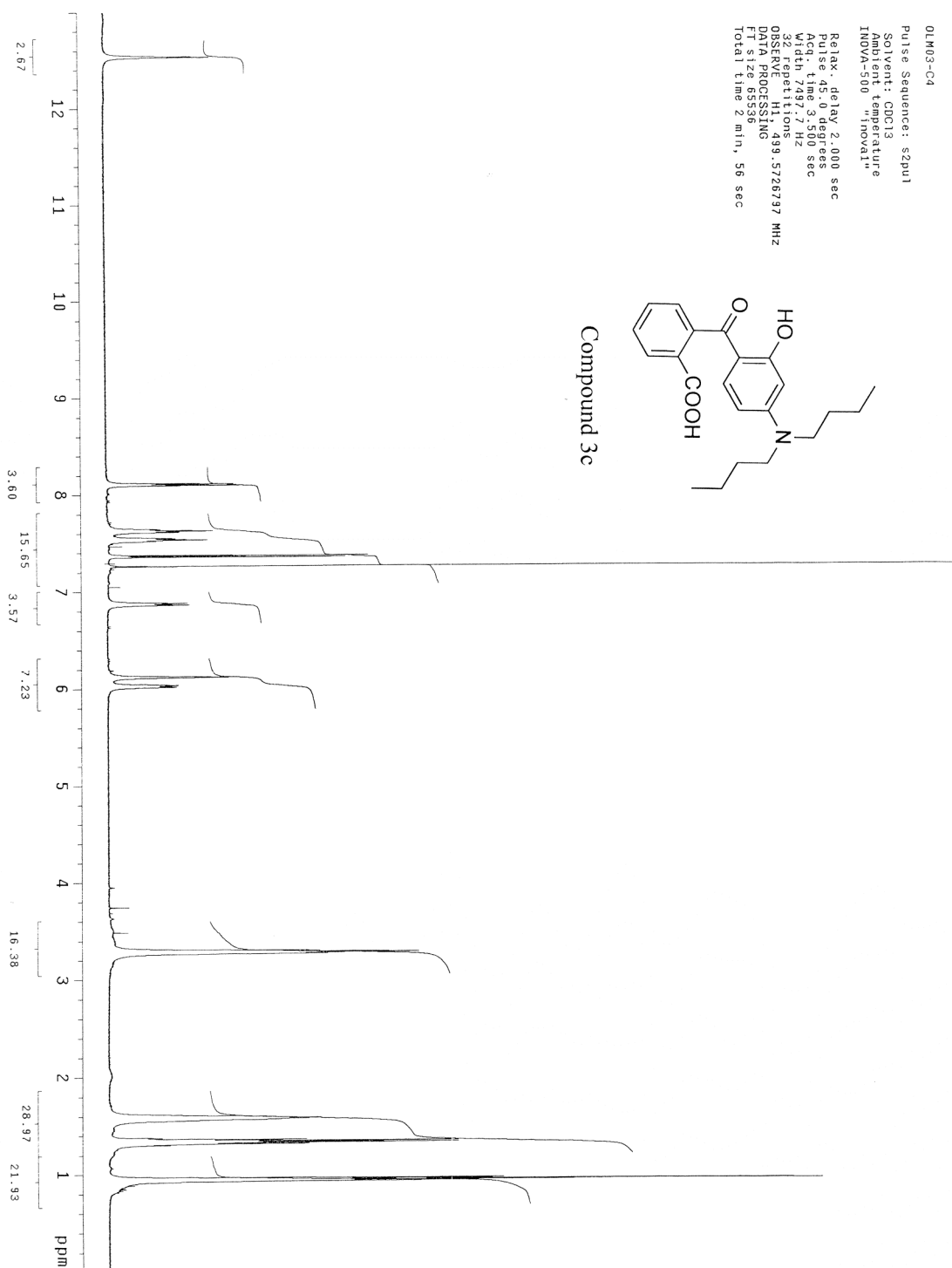
Compound 2c



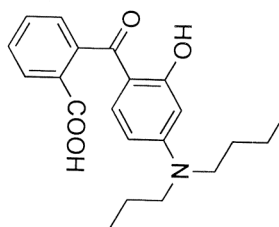
OLM03-C4
Pulse Sequence: s2pul
Solvent: CDCl3
Ambient temperature
INOVA-500 "Inova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 3.500 sec
Nuc1 13C 101.3 MHz
Nuc2 1H 499.5726797 MHz
OBSERVE HI 499.5726797 MHz
DATA PROCESSING
FT size 65536
Total time 2 min, 56 sec



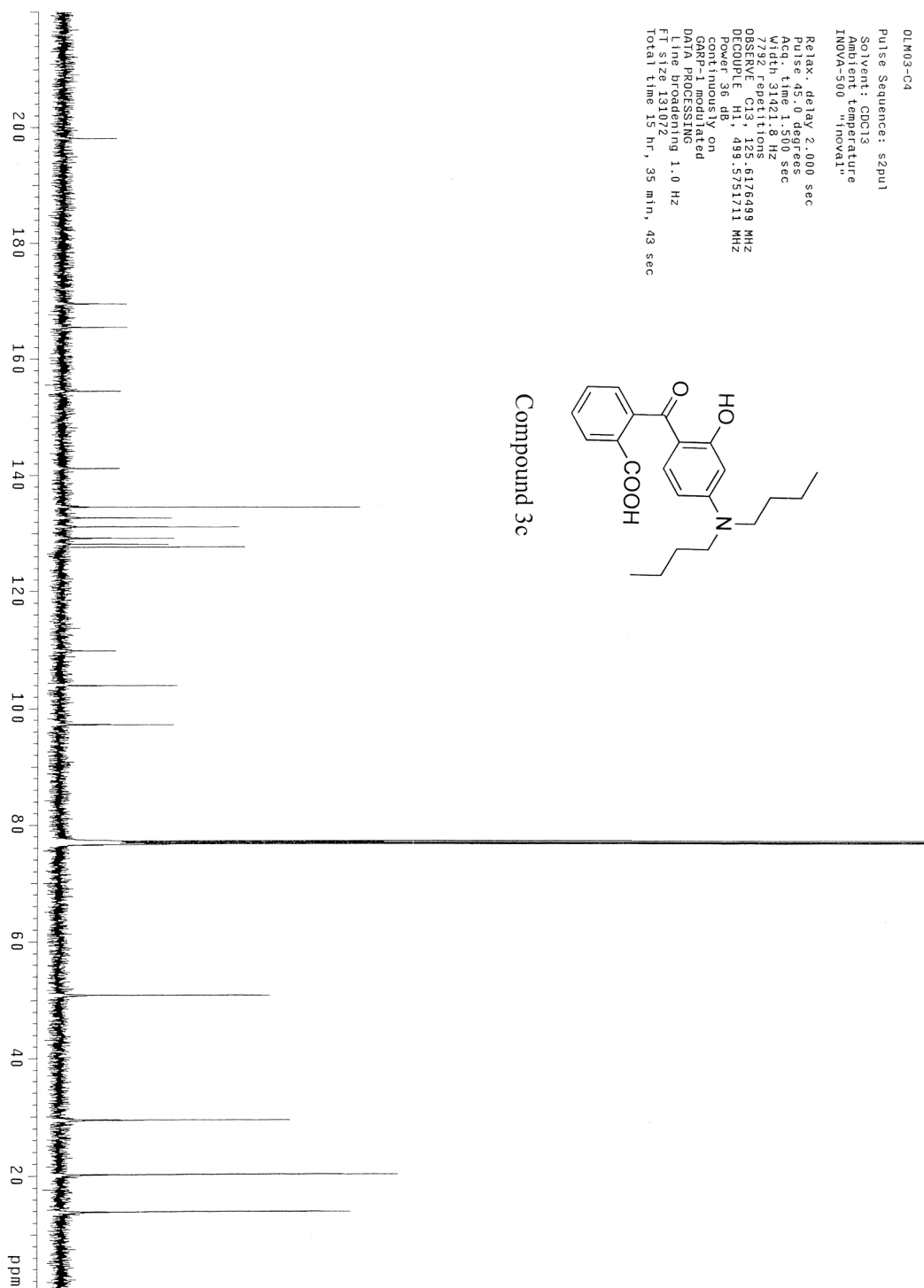
Compound 3c

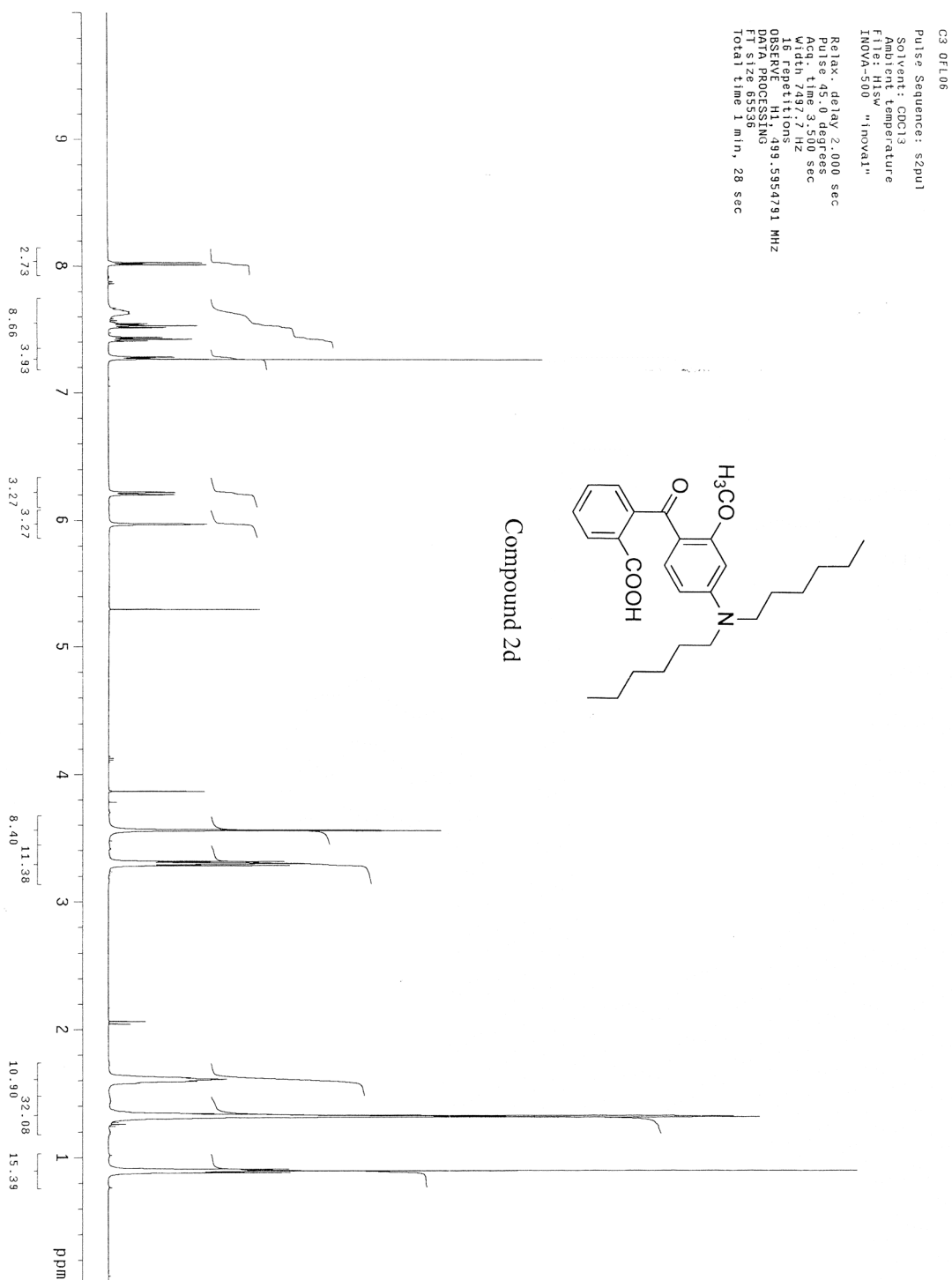


OLM03-C4
Pulse Sequence: szpu1
Solvent: CDCl3
Sample Temperature
INOVA-500 "1HNOVA1"
Relax. delay 2.000 sec
Pulse: 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
7792 repetitions
OBSERVE: C13, 125.617649 MHz
DECOUPLE: H1, 499.575171 MHz
Coupling 36.48
Continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 15 hr, 35 min, 43 sec

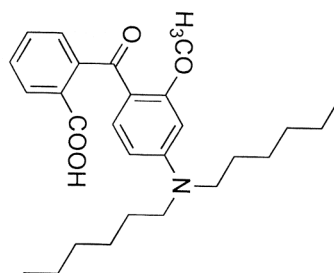


Compound 3c

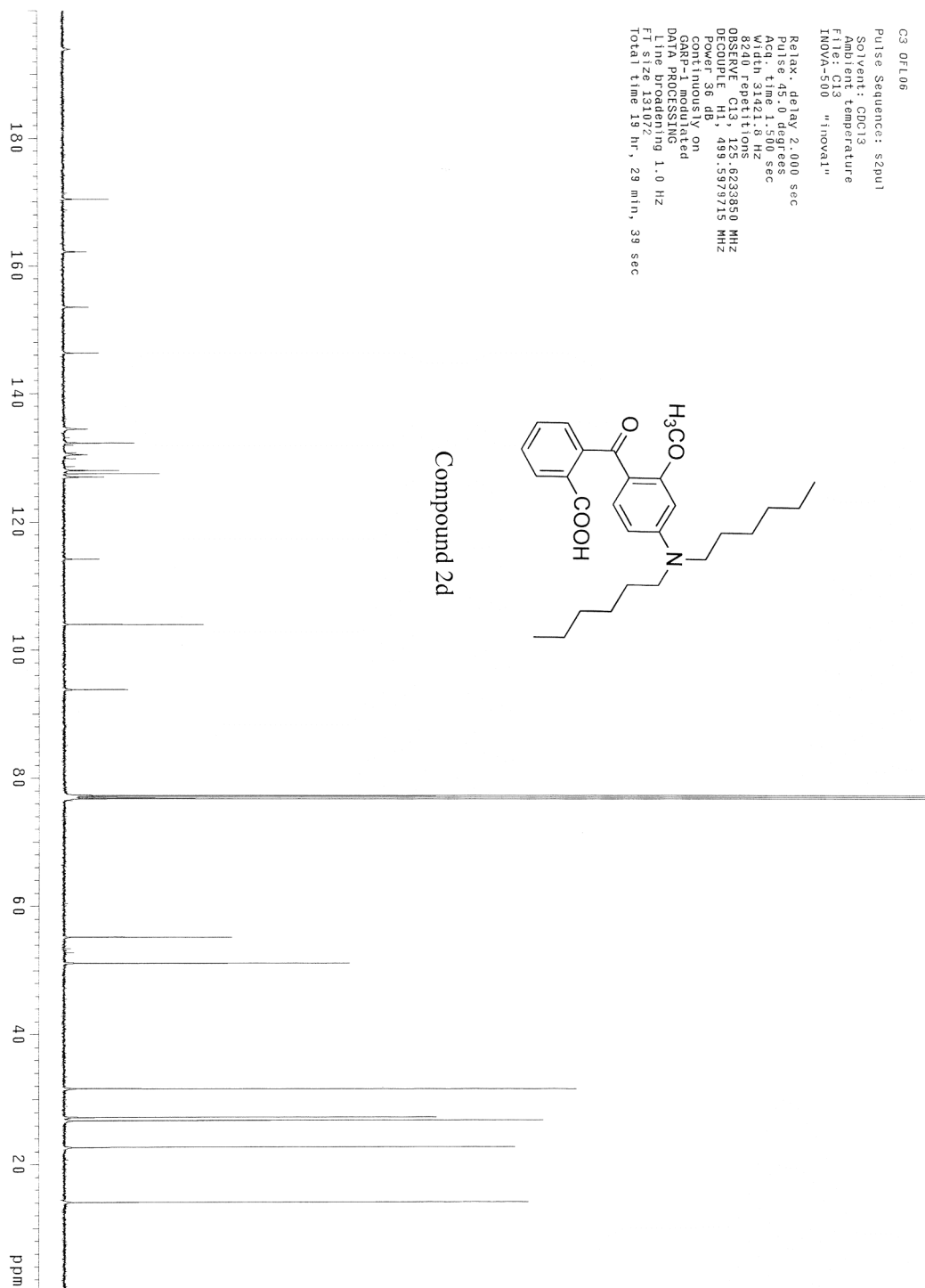


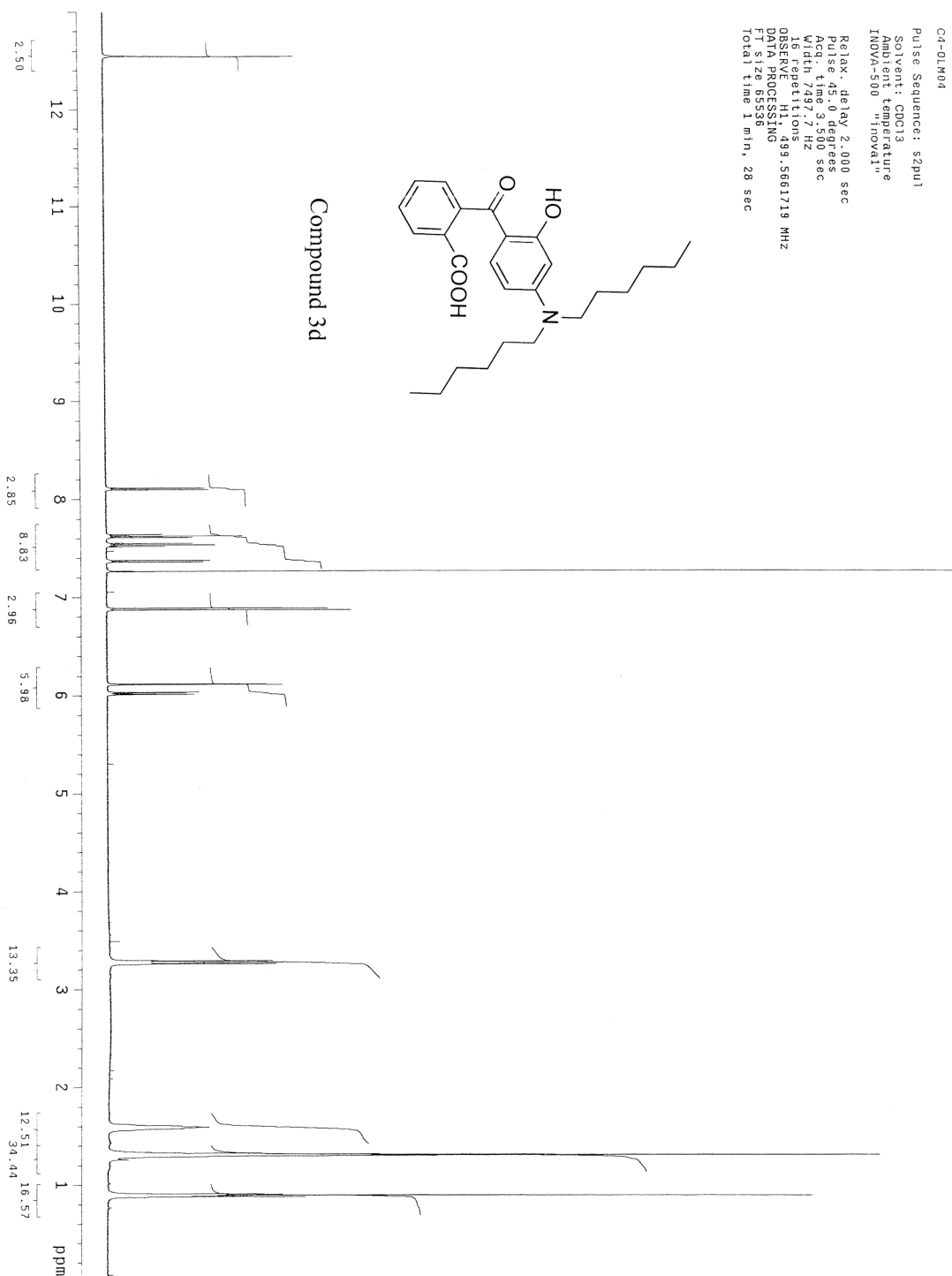


C3 OFL06
Pulse Sequence: s2pul1
Solvent: CDCl3
Ambient temperature
File: C13
INOVA-500 "nova1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
0.840 repetitions
0.840 scans
F2 size 623850 MHz
DECUPLE H1, 499.5979715 MHz
Power 36 dB
continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 19 min, 29 sec

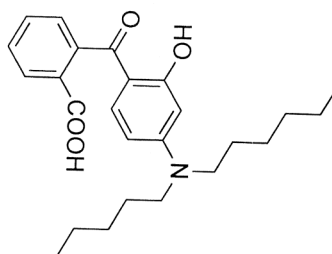


Compound 2d

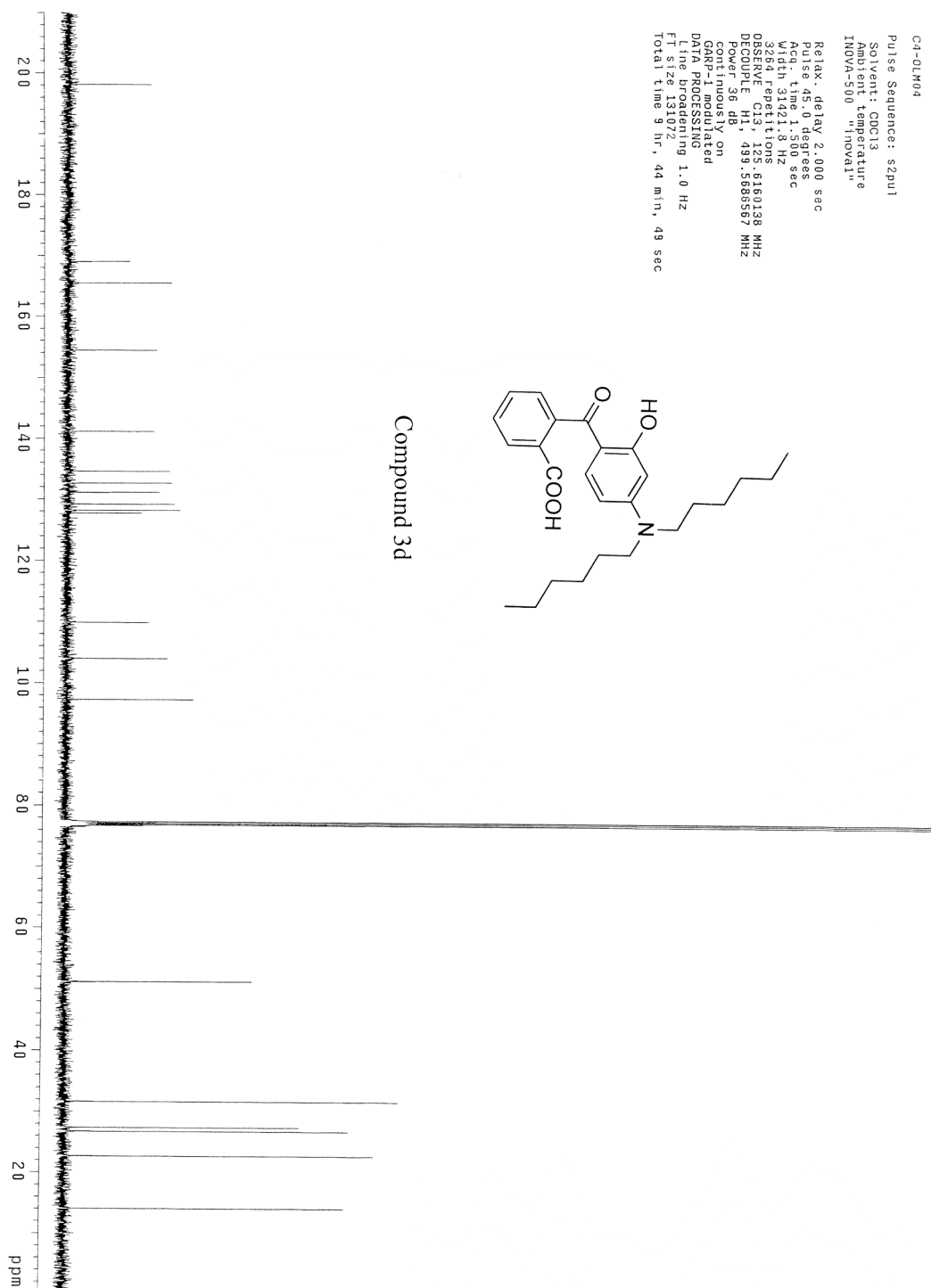




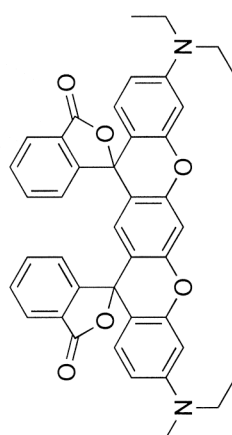
C4-OLM04
Pulse Sequence: szpul
Solvent: CDCl3
Acquisition Temperature
INOVA-500 "INOVAT1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
3264 repetitions
OBSERVE C13, 125.6160138 MHz
DECOUPLE H1, 499.5685567 MHz
Power 3.000 dB
Sensitivity on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



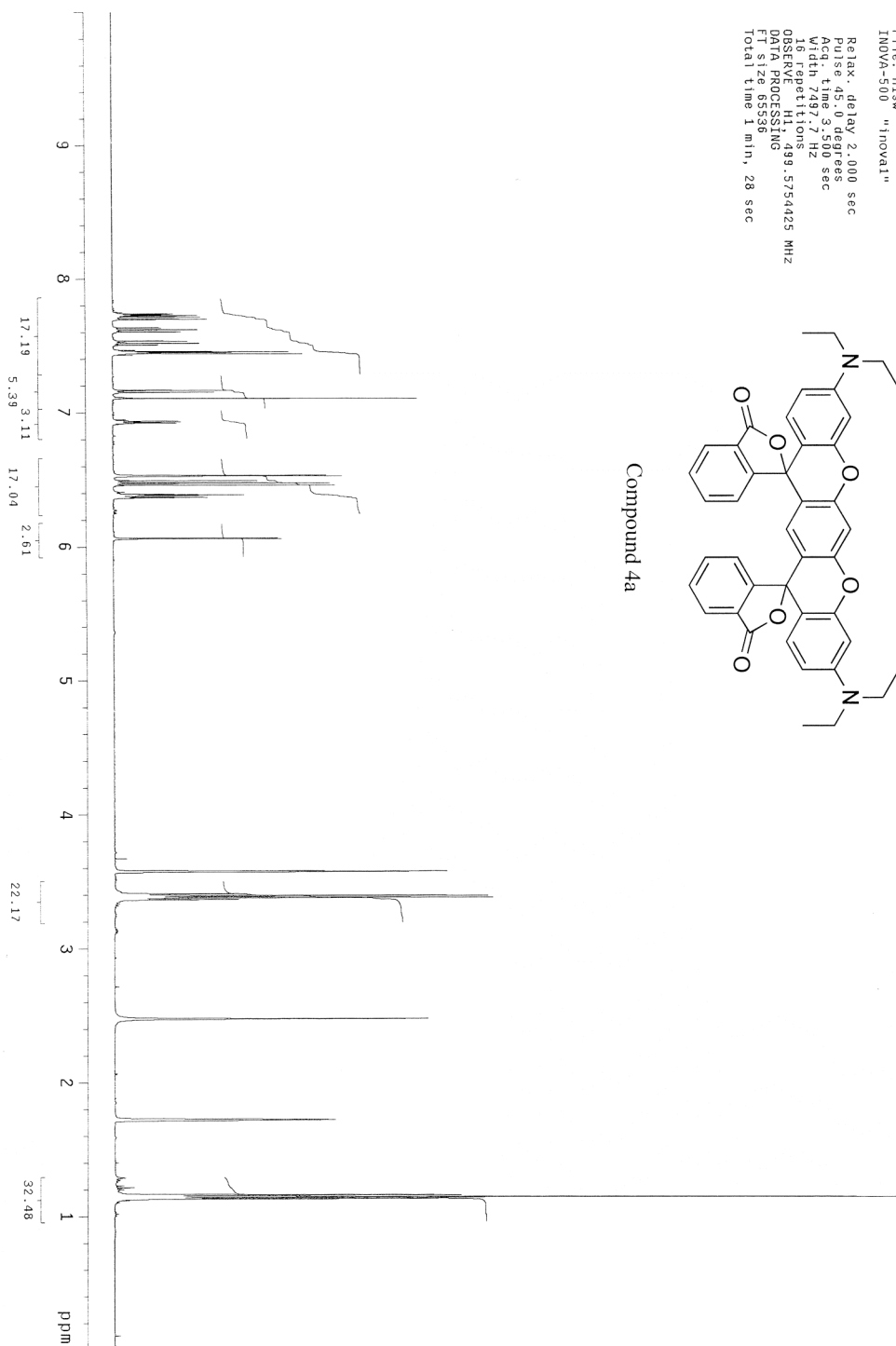
Compound 3d



OLM01
in THF
100624
Pulse Sequence: zgpg30
Solvent: Acetone
Sample Temperature
F1: 125.130 MHz
INNOVA-500 "nova1"
Relax. delay: 2.000 sec
Pulse: 45.0 degrees
Acq. time: 3.500 sec
Width: 7497.7 Hz
16 repetitions
OBSERVE H1: 499.5754425 MHz
DATA PROCESSING
F2: 125.130 MHz
Total time: 1 min, 28 sec

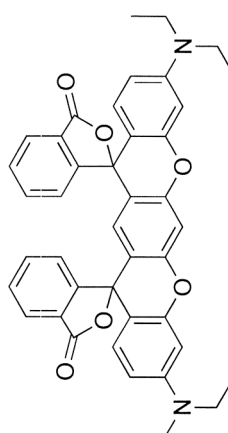


Compound 4a

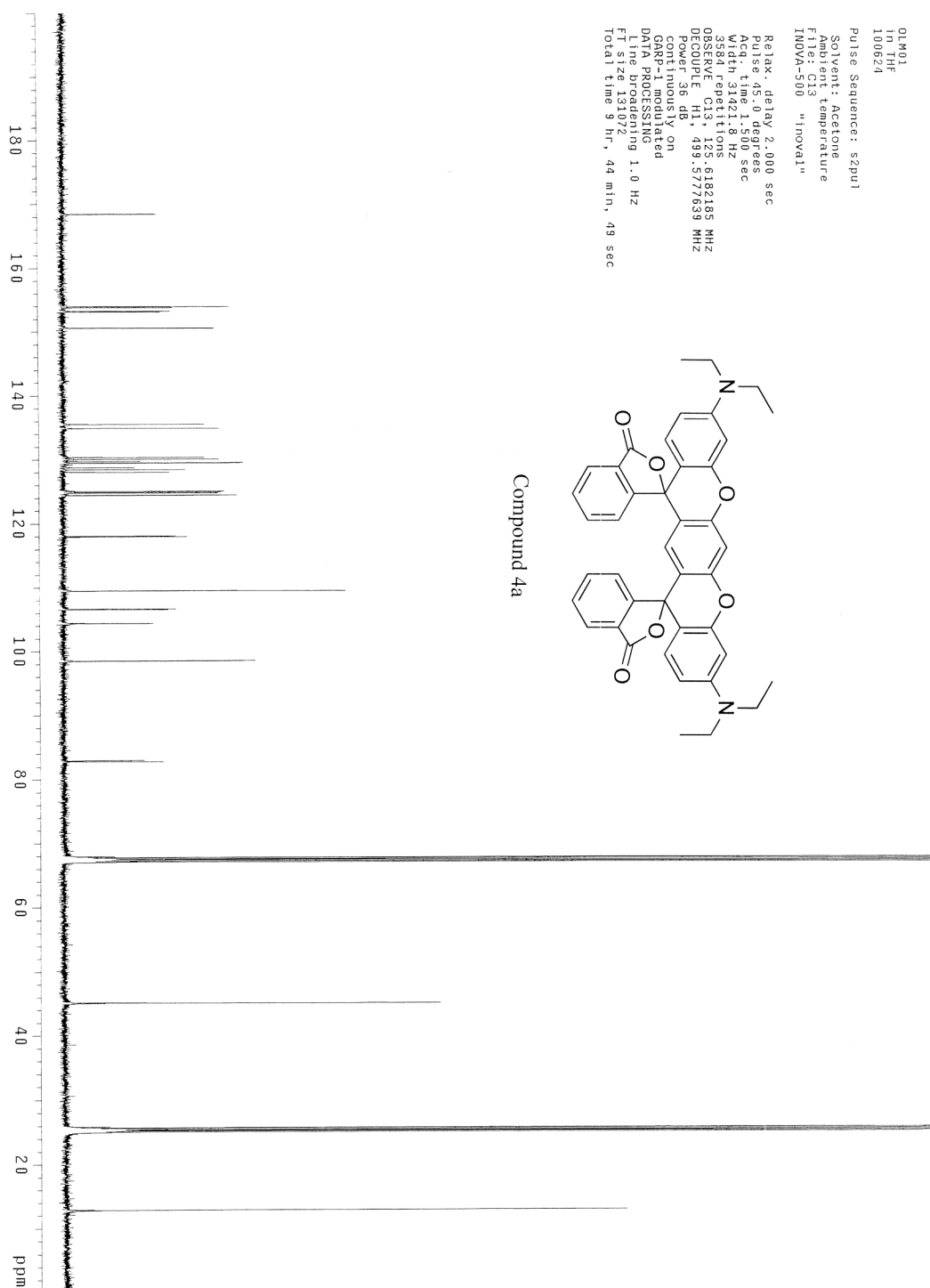


OLM01
in THF
100624
Pulse Sequence: szpul
Solvent: Acetone
Sample temperature
F2: 113
INOVA-500 "nova1"

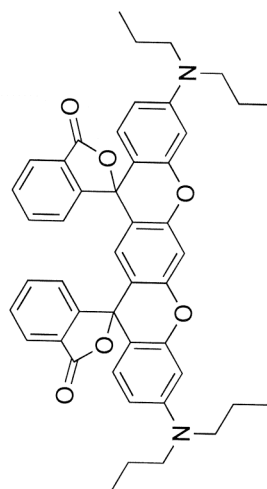
Relax: delay 2.000 sec
Pulse: 45.0 degrees
Acq. time 1.500 sec
Width 31421.8 Hz
3584 repetitions
OBSERVE C13, 125.6182185 MHz
DECOUPLE H1, 499.577653 MHz
Pulse program
Continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



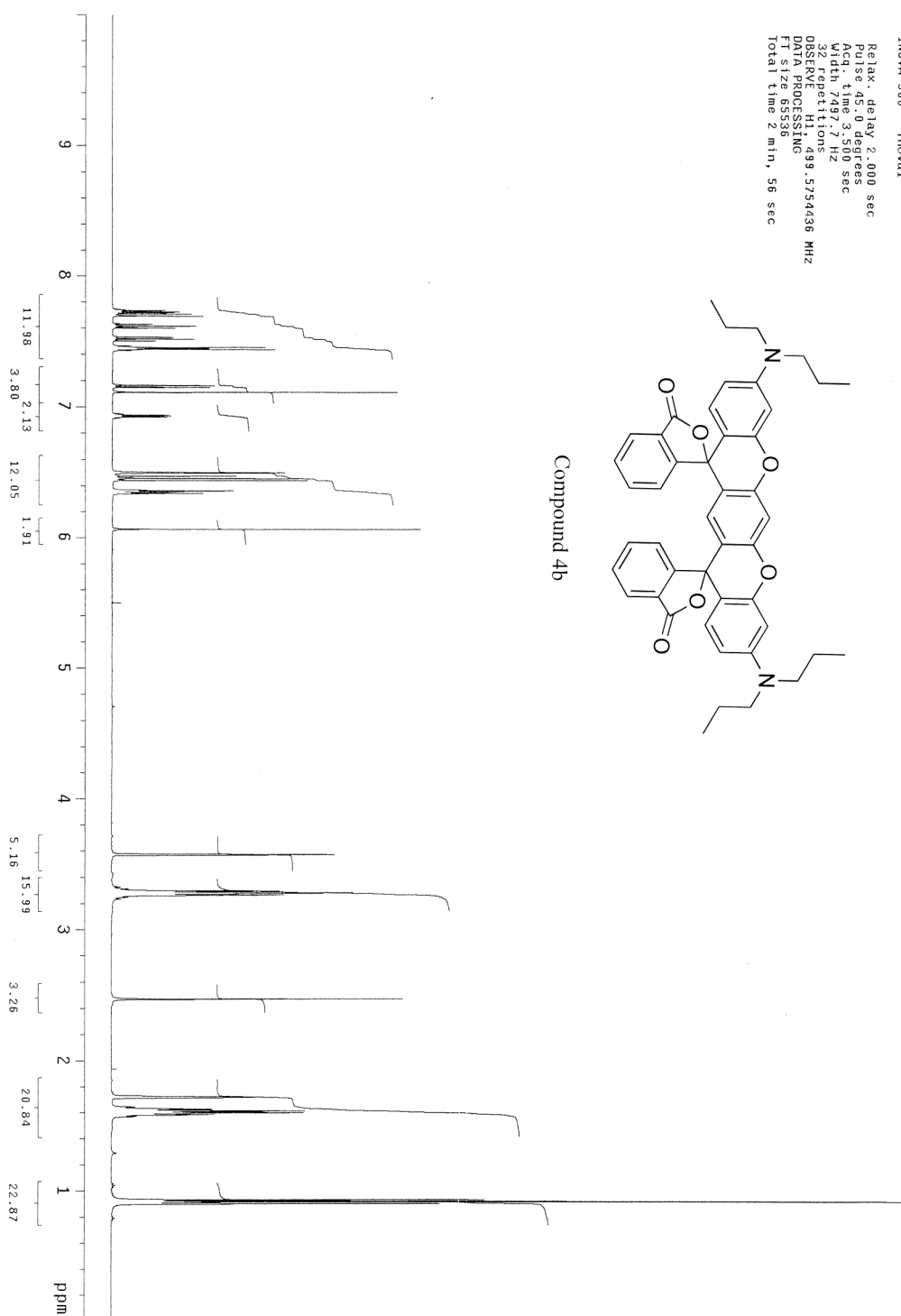
Compound 4a



0LM02
Pulse Sequence: s2pul
Solvent: Acetone
Ambient temperature
INOVA-500 "INOVAT"
Relax. delay 2.000 sec
Acq. time 3.50 sec
Width 7497.7 Hz
32 repetitions
OBSERVE H1, 499.579436 MHz
DATA PROCESSING
FT size 65536
Total time 2 min, 56 sec



Compound 4b



Pulse Sequence: s2pu1

Solvent: Acetone
Ambient temperature
INOVA-500 "inova1"

Relax. delay 2.000 sec
Pulse 45.0 degrees

Acq. time 1.500 sec
Width 31421.8 Hz
1000 points

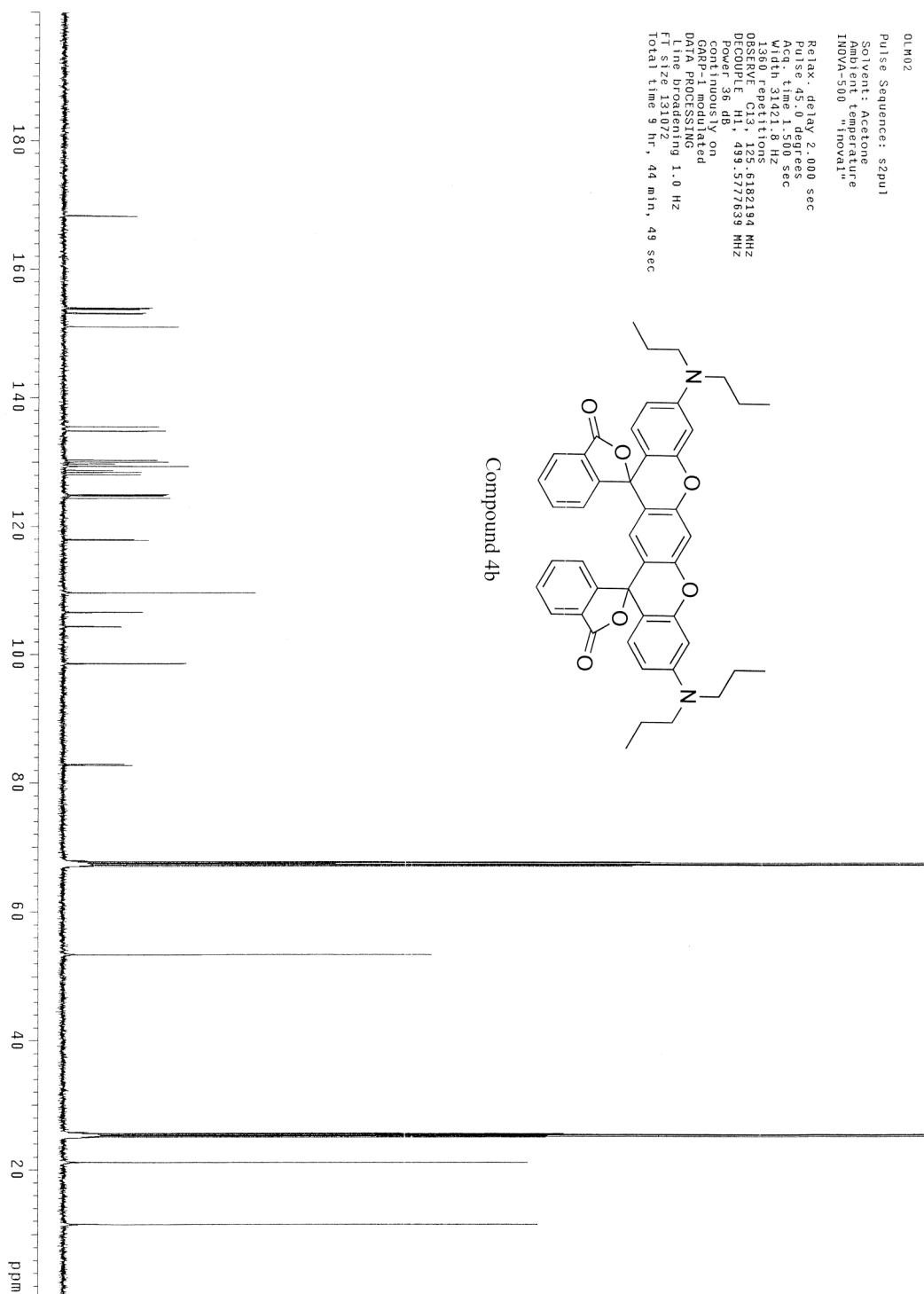
1360 repetitions
OBSERVE C13, 125.6182194 MHZ
DECOUPLE H1, 499.5727639 MHZ

DECOUPLE H1, 499.5777639 MHz
Power 36 dB
continuously on

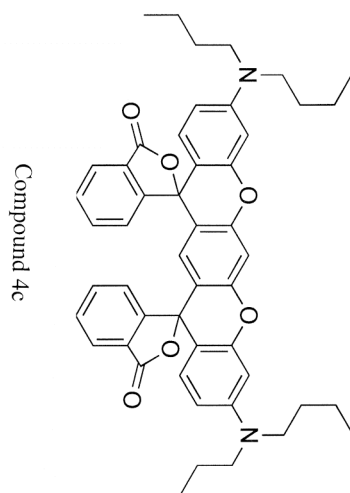
continuously on
GARP-1 modulated
DATA PROCESSING

Line broadening 1.0 Hz
FT size 131072

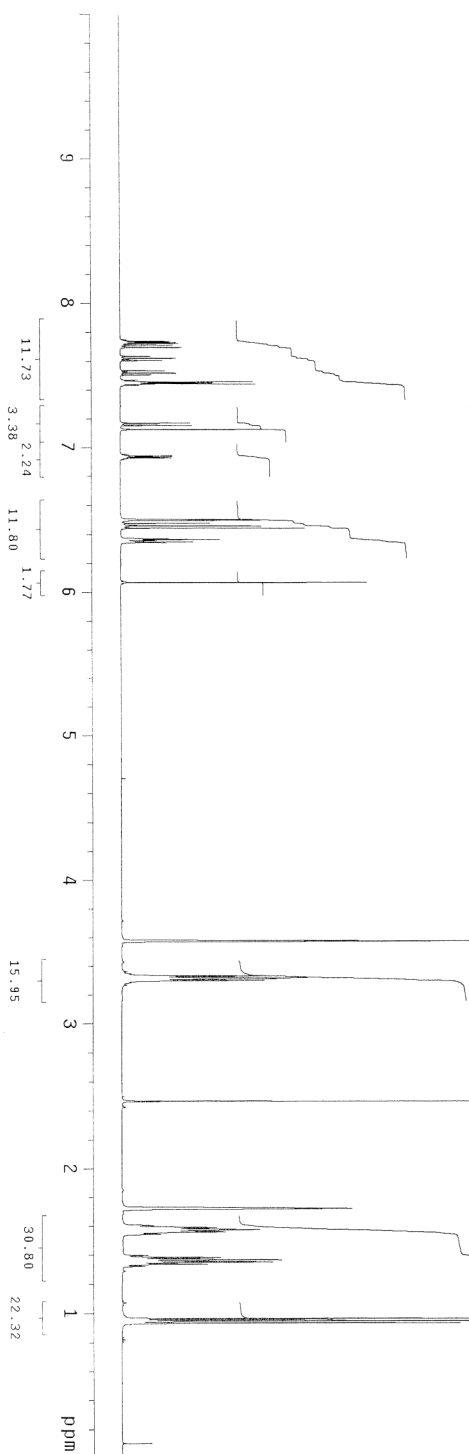
Total time 9 hr, 44 min, 49 sec



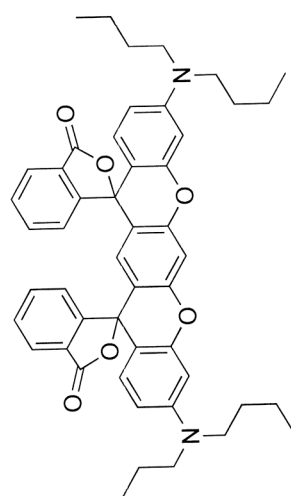
01M03
Pulse Sequence: zgpg30
Solvent: Acetone
Acq. Temp: 300.2 K
INNOVA-500 "1H"NOVA11"
Relax. delay: 2.000 sec
Pulse: 45.0 degrees
Acq. time: 3.500 sec
Width: 7497.7 Hz
32 repetitions
OBSERVE: H1, 499.575427 MHz
DATA PROCESSING
FT size: 65536
Total time: 2 min, 56 sec



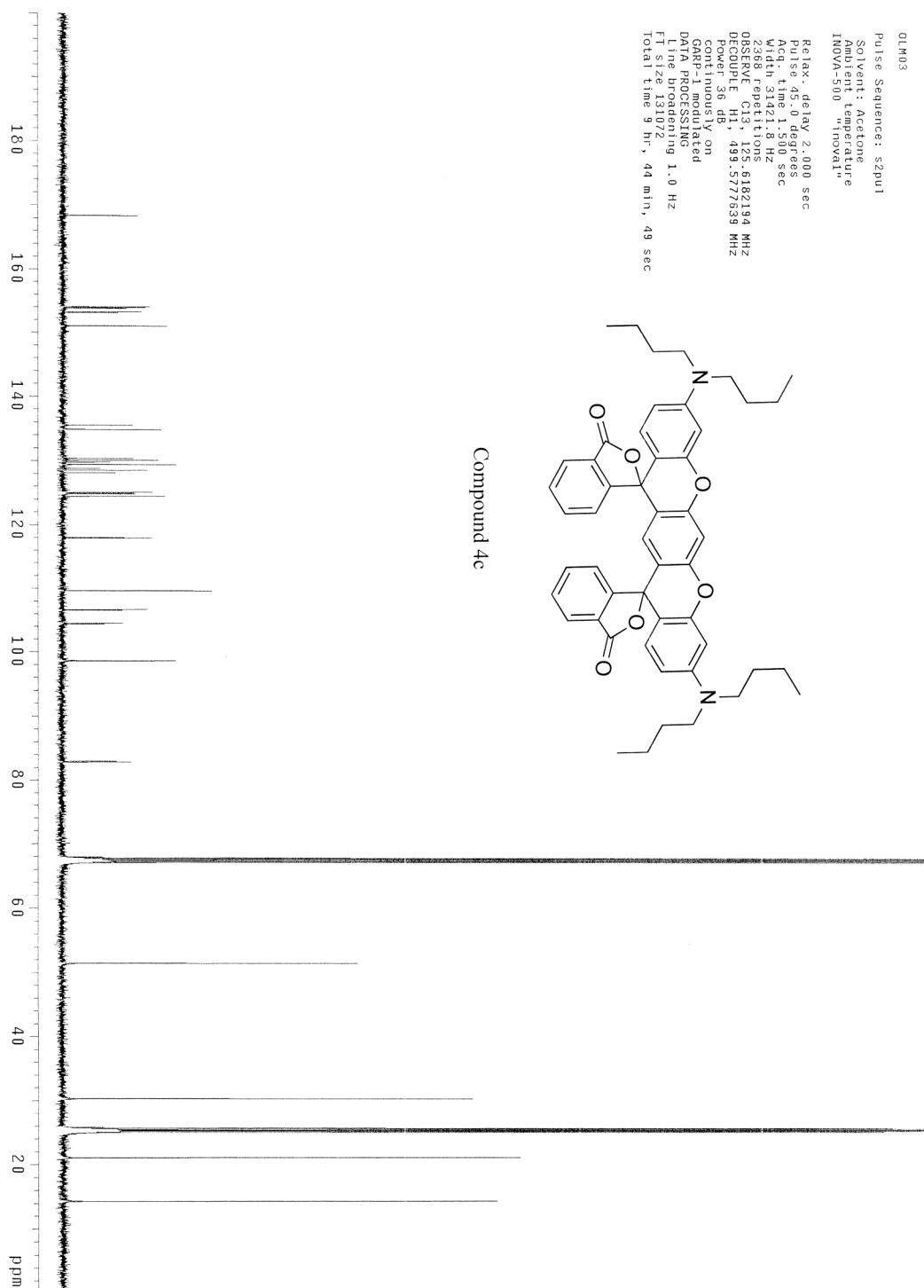
Compound 4c



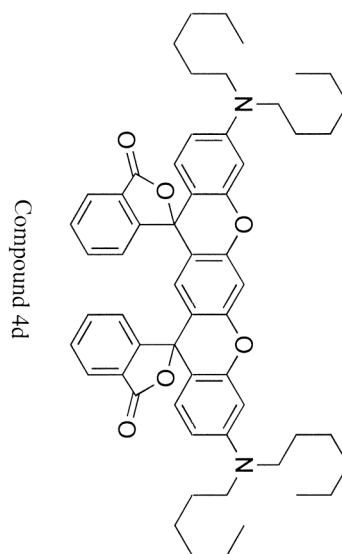
01.M03
Pulse Sequence: zgpg30
Solvent: Acetone
Acquisition temperature
INNOVA-500
"nova1"
Relax: delay 2.000 sec
Pulse: 45.0 degrees
Acq: time 1.500 sec
Width: 31421.8 Hz
2368 repetitions
OBSERVE: C13, 125.6182194 MHz
DECOUPLE: H1, 499.577653 MHz
NUC1: 13C, 56.480
Continuously on
GARP-1 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 131072
Total time 9 hr, 44 min, 49 sec



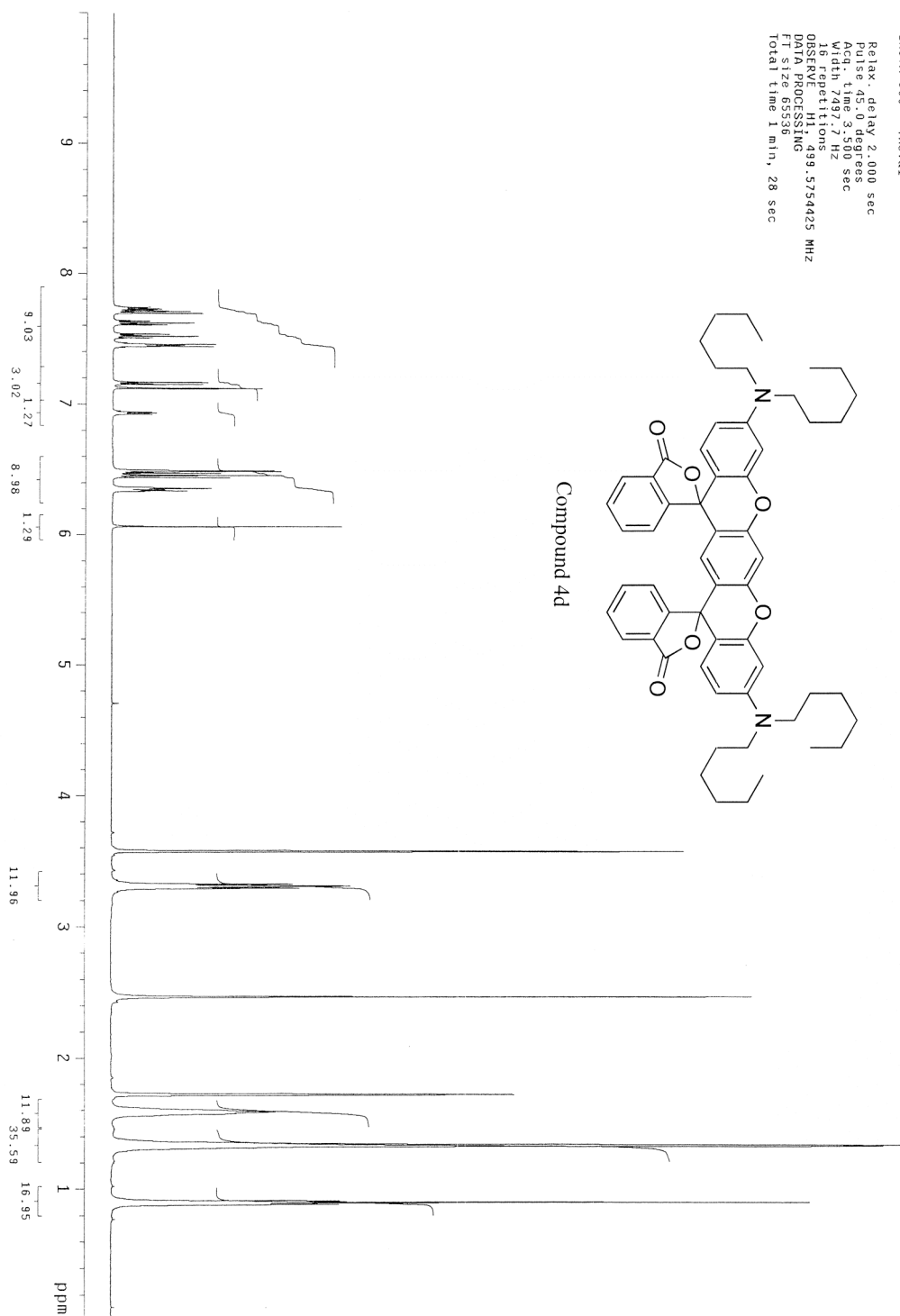
Compound 4c



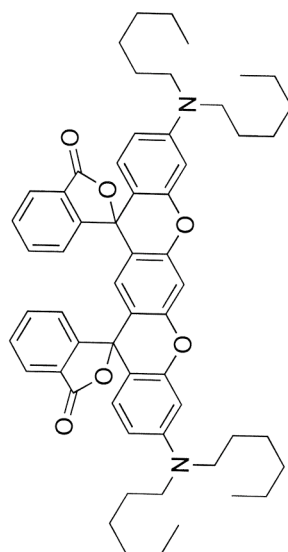
OLM04
Pulse Sequence: szpul
Solvent: Acetone
Acquisition Temperature
INOVA-500 "INOVAT1"
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 3.500 sec
Width 7497.7 Hz
16 repetitions
OBSERVE NH: 499.5754425 MHz
DATA PROCESSING
F2 size 65536
Total time 1 min, 28 sec



Compound 4d



01M04
Pulse Sequence: s2pul
Solvent: Acetone
Modent: temperature
INVA-500 110xvxl
Relax: delay 2.000 sec
Pulse: 45.0 usef100
Acq: time 1.500 sec
Width 31421.8 Hz
10000 repetitions
OBSERVE C13, 125.6182194 MHz
DECOUPLE H1, 499.577639 MHz
Power 36 dB
Pulse program: s2pul
DATA PROCESSING
Line broadening 1.0 Hz
Ft size 131072
Total time 9 hr, 44 min, 49 sec



Compound 4d

