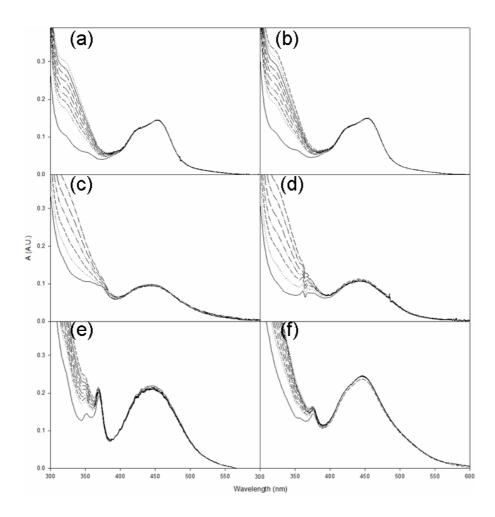
## Supporting Information

## Lack of Quenching by [Fe(CN)<sub>6</sub>]<sup>4-</sup> is Not Proof of DNA Intercalation

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The electronic absorption spectra of 1-3 were invariable with to  $[Fe(CN)_6]^4$ -concentration, indicating that pre-association in solution is minimal (Fig. S1), and the luminescence was independent of ferrocynide and ionic strength (Figs. S2 – S4). Figure S5 shows the Stern-Volmer plots for 1-3 in the presence of 100  $\mu$ M DNA upon addition of  $[Fe(CN_6]^4$  in 5 mM Tris (pH = 7.5) with 5 mM NaCl.



**Figure S1.** Electronic absorption spectra of 10 μM compound **1 (a)** in absence and **(b)** presence of 100 μM DNA, 10 μM compound **2 (c)** in absence and **(d)** presence of 100 μM

S1

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DNA and 10  $\mu$ M compound **3 (e)** in absence and **(f)** presence of 100  $\mu$ M DNA at different [[Fe(CN)<sub>6</sub>]<sup>4-</sup>] (0-6 mM) in 50 mM NaCl buffer.

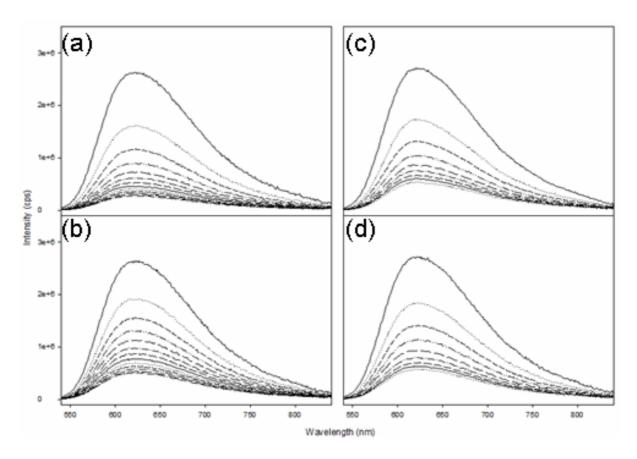
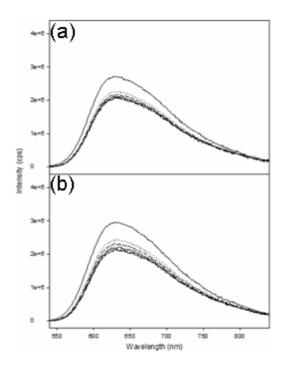


Fig. S2. Emission quenching of  $10~\mu M$  1 by 0-6~mM ferrocyanide in 5 mM Tris buffer (pH = 7.5) in the absence of DNA and (a) 5 mM NaCl and (b) 50 mM NaCl and in the presence of  $100~\mu M$  DNA in (c) 5 mM NaCl and (d) 50 mM.

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**Figure S3.** Emission quenching of  $10 \,\mu\text{M}$  **2** by  $0-6 \,\text{mM}$  ferrocyanide in the presence of  $100 \,\mu\text{M}$  DNA in 5 mM Tris buffer (pH = 7.5) and (a) 5 mM NaCl and (b) 50 mM NaCl.

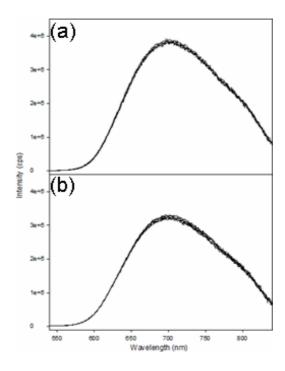


Fig. S4. Emission quenching of 10  $\mu$ M 3 by 0 – 6 mM ferrocyanide in the presence of 100  $\mu$ M DNA in 5 mM Tris buffer (pH = 7.5) and (a) 5 mM NaCl and (b) 50 mM NaCl.

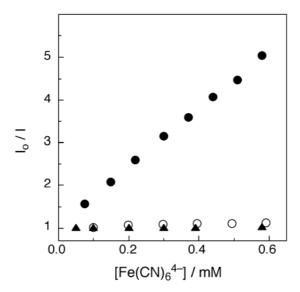


Fig. S5. Stern-Volmer plots of 10  $\mu$ M 1 (closed circles), 2 (open circles), and 3 (closed triangles) in the presence of 100  $\mu$ M DNA with 5 mM NaCl (5 mM Tris buffer, pH = 7.5).

## **Experimental Details**

Sodium chloride and Tris (tris(hydroxymethyl) aminomethane) were purchased from Sigma-Aldrich and used as received. Potassium ferrocyanide was purchased from Baker and Adamson and HCl was purchased from Curtin Matheson. Calf-thymus DNA was purchased from Sigma and equilibrium dialysis, with a Sigma cellulose membrane, was performed to remove low molecular weight proteins. Electronic absorption measurements were performed on an HP diode array spectrophotometer with HP 8453 *WinSystem* software. Steady-state emission measurements were performed on a SPEX Fluormax-2 spectrometer with DataMax for Windows software. Lifetime measurements were performed with a home-built instrument excited using the 532 nm output from a pulsed Nd:YAG laser (fwhm  $^{\sim}$  8 ns). Each solution was bubbled for 10 min with N2 prior to steady-state or lifetime measurement.