

Supplementary Material (ESI) for Chemical Communications

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Supplementary Information

New strategy for controlling the oligothiophene aggregation mode utilizing the gel-to-sol phase transition induced by crown-alkali metal interactions

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Measurements details

General: All chemical reagents were obtained from Aldrich, TCI, Kanto Chemicals or Wako were used without further purification. ^1H and ^{13}C NMR spectra were measured on JOEL ECS400 (400 MHz) spectrometers. Chemical shifts are recorded in ppm downfield from tetramethylsilane as the internal standard. FT-IR spectra were obtained using a Perkin–Elmer 100 instrument. Mass spectral data were obtained using a MALDI TOF Bruker autoflex II TOF/TOF mass spectrometer. Samples were prepared with dithianol matrix. UV/Vis spectra were measured on a Shimadzu UV-2500PC spectrometer. Solution samples were measured in 1 cm length cell, gel samples were measured in 0.10 cm length cell. Fluorescence emission spectra were measured on a Perkin-Elmer LS55 instrument.

Gelation test of organic fluids: The gelator and the solvents were put in a capped test-tube sonicated about 3 min and heated until the solid was dissolved. The sample vial was cooled to 25 °C and left for 12 h at the ambient conditions. The state was evaluated by the “stable to inversion of a test tube” method.

Gelation test of organic fluids with alkali metal salts: The gelator, conc. solution of alkali metal salts in ethanol and the solvents were put in a capped test-tube sonicated about 3 min and heated until the solid was dissolved. The sample vial was cooled to 25 °C and left for 12 h at the ambient conditions. The state was evaluated by the “stable to inversion of a test tube” method.

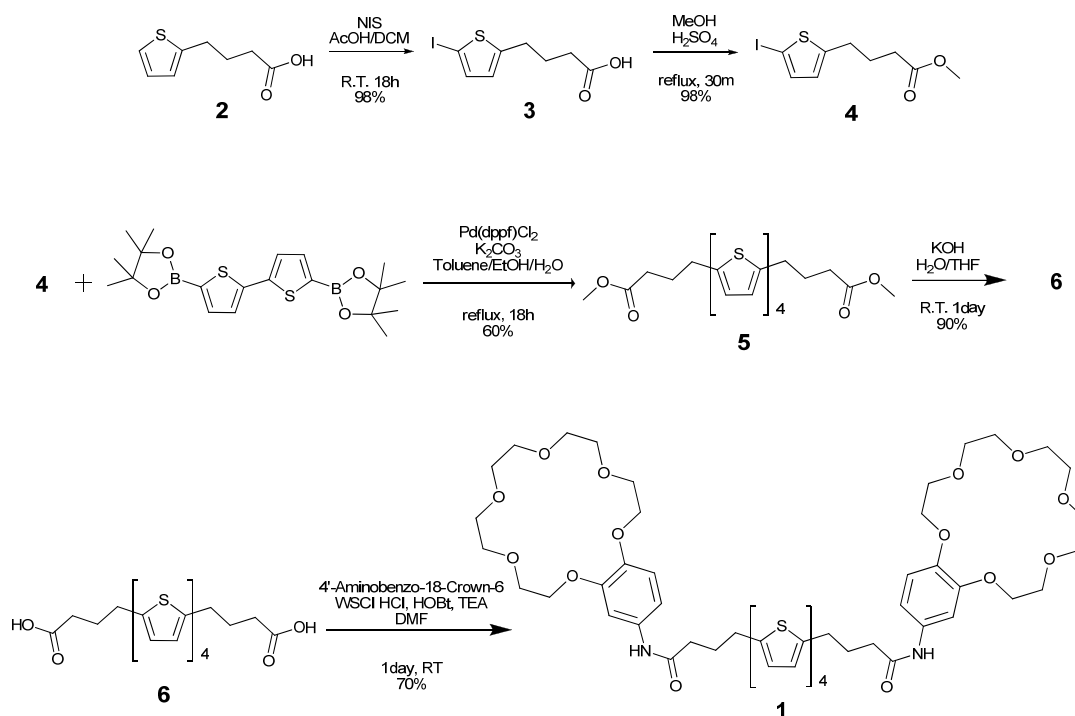
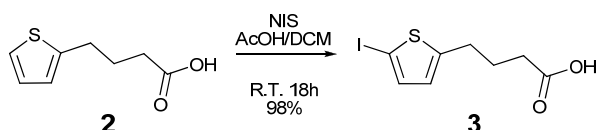
Gel-sol transition temperature (T_{gel}): T_{gel} was measured by the test-tube-tilting method where a test tube containing the gel was immersed inversely in a thermostatted bath and the temperature was raised at 0.5°C/min. The T_{gel} was considered as the temperature when the mass was started to flow. The error involved in measuring T_{gel} was ± 1 °C.

TEM and SEM measurements: The gel was put on a carbon-coated copper grid and dried in vacuo for 12 h at room temperature. Then, the xerogel sample was subjected to TEM observations with a FEI-TEM Philips TECNAI F20. For SEM observations, the xerogels were shielded by Pt and examined with a Keyence VE-9800 scanning electron microscope. The accelerating voltage of SEM was 20 kV.

AFM measurements: The gel was cast on a splintered mica plate with spin-coating (5000 rpm, 30 s). The sample was examined by S II NanoNavi/E-swap (non-contact mode).

Synthesis details

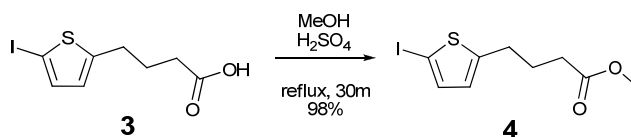
General synthetic route.

**Reaction details**

N-Iodosuccinimide (4.50g, 20.0mmol) was added under N₂ to a CH₂Cl₂ (dry) (50 mL) and glacial AcOH (50 mL) solution of 4-(thiophen-2-yl)butanoic acid **2** (3.40g, 20.0mmol), in a round-bottom flask, covered with aluminum foil. The mixture was stirred for 18h under N₂ at room temperature. The reaction was monitored by TLC Hex/AcOEt (7:3). After completion, the reaction mixture was poured to aqueous Na₂S₂O₃, organic phase was separated and evaporated under reduced pressure. Residue was dissolved in CHCl₃ (100ml), and was extracted by 1M NaOH (100mL x 2). The combined alkaline phases were acidified with 6M HCl to pH 2 and was extracted by CHCl₃ (100 mL x 3). The combined organic extract was dried over anhydrous MgSO₄, and evaporated to dryness to give a red oil of **3** (5.80g, 98%).

¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.97 (q, 2H, CH₂), 2.41 (t, 2H, CH₂), 2.89 (t, 2H, CH₂), 6.50 (d, 1H, CH), 7.05 (d, 1H, CH), 10.1 (br. s, 1H, COOH).

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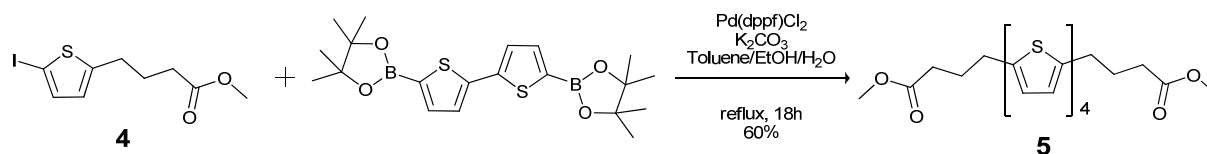


4-(5-Iodothiophen-2-yl)butanoic acid **3** (5.80g, 19.6mmol) was added to a MeOH (dry) (100 mL) and concentrated H₂SO₄ (150 μ L) solution, in a round-bottom flask, under N₂. The mixture was stirred under reflux for 1h under N₂. The reaction was monitored by TLC Hex/AcOEt (7:3). After completion, the reaction mixture was evaporated under reduced pressure. After evaporation residue was dissolved in CHCl₃ (100ml) and was washed with saturated aqueous NaHCO₃ (100mL), dried over anhydrous MgSO₄, and evaporated to dryness to give a purple solid. The residue was chromatographed on silica with gradient Hex/AcOEt (95:5 – 9:1) as the eluent. Compound **4** was obtained as colourless oil (5.90g, 98%).

¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.95 (q, 2H, CH₂), 2.34 (t, 2H, CH₂), 2.84 (t, 2H, CH₂), 3.65 (s, 3H, CH₃), 6.47 (d, 1H, CH), 7.02 (d, 1H, CH).

¹³CNMR (100 MHz, CDCl₃): δ (ppm) 26.6, (CH₂), 29.4 (CH₂), 33.0 (CH₂), 51.7 (CH₃), 70.1 (C-I), 126.5 (CH), 136.8 (CH), 150.5 (C-alkyl), 173.6 (C=O).

MALDI-TOF MS m/z 310.95, calc. for C₉H₁₂IO₂S (M+H⁺) found 311.10.

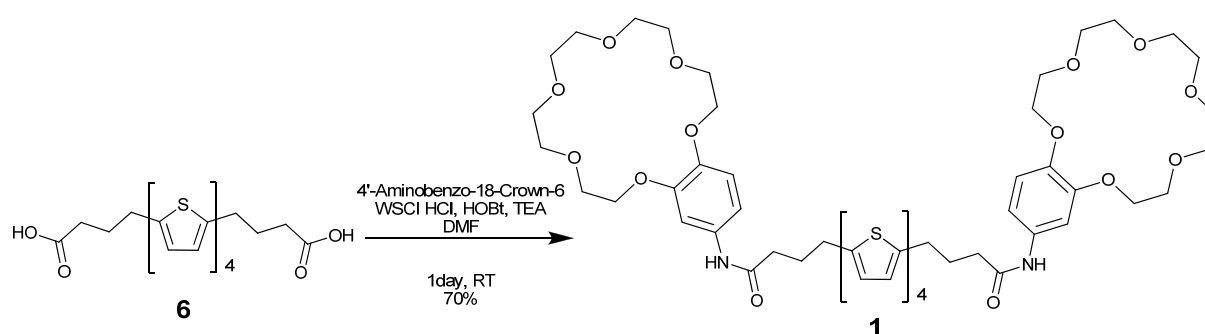


After sedimentation of orange solid product, THF was removed. Orange solid of 5 potassium salt was dissolved in water (250ml). Aqueous solution was acidified to pH-2 with 6M HCl. After filtration and drying under vacuum dark orange solid of product 6 was obtained in 90% yield (0.51g).

^1H NMR (400 MHz, d_6 -DMSO): δ (ppm) 1.85 (q, 2H, CH_2), 2.29 (t, 2H, CH_2), 2.81 (t, 2H, CH_2), 6.83 (d, 1H, CH), 7.16 (d, 1H, CH), 7.19 (d, 1H, CH), 7.26 (d, 1H, CH), 12.13 (bs, 1H, COOH).

^{13}C NMR (100 MHz, d_6 -DMSO $_3$): δ (ppm) 26.9, (CH_2), 29.2 (CH_2), 33.3 (CH_2), 124.6 (CH), 124.9 (CH), 125.5 (C-H), 126.5 (C-H), 134.1, 134.2, 136.3, 144.8, 174.6 (C=O).

MALDI-TOF MS m/z 502.04, calc. for $\text{C}_{24}\text{H}_{22}\text{O}_4\text{S}_4$ (M^+) found 502.19.



WSCI HCl (0.5g, 3.06mmol) and HOBT (0.42g, 3.06mmol) were added to a DMF solution of 6 (0.51g, 1.02mmol) with 4'-aminobenzo-18-crown-6 (1.0g, 3.06mmol) and TEA (1.3ml, 10.2mmol). Reaction was stirred under N_2 flow in room temperature during 1 day. The reaction mixture was evaporated to dryness under reduced pressure, and the residue was dissolved in CHCl_3 (100ml) and treated with 1 M aqueous HCl (200ml), washed with aqueous NaHCO_3 (200ml) and brine (200ml), dried over anhydrous MgSO_4 , and then evaporated to dryness. The residue was chromatographed on alumina with $\text{CHCl}_3/\text{MeOH}$ (97:3) as the eluent; a orange fraction of 1 was isolated with 70% yield (0.8g).

^1H NMR (400 MHz, CDCl_3): δ (ppm) 2.12 (q, 2H, CH_2), 2.37 (t, 2H, CH_2), 2.90 (t, 2H, CH_2), 3.64-3.78 (bm, 12H, - $\text{OCH}_2\text{CH}_2\text{O}$ -), 3.89 (m, 4H CH_2), 4.10-4.18 (bm, 4H, PhO-CH_2), 6.72 (d, 1H, CH), 6.77-6.87 (m, 2H, CH), 6.98 (t, 2H, CH), 7.03 (d, 1H, CH), 7.14 (bs, 1H, NH), 7.32 (d, 1H, CH).

^{13}C NMR (100 MHz, CDCl_3): δ (ppm) 26.9, (CH_2), 29.4 (CH_2), 36.3 (CH_2), 68.9, 69.5, 69.6, 70.8, 70.9, 107.0, 112.2, 114.7, 123.5, 123.9, 124.2, 125.7, 132.1, 135.1, 135.6, 136.6, 143.9, 145.7, 149.3, 170.4 (C=O).

MALDI-TOF MS m/z 1120.35, calc. for $\text{C}_{56}\text{H}_{68}\text{N}_2\text{O}_{14}\text{S}_4$ (M^+) found 1120.87; 1143.34, calc. for $\text{C}_{56}\text{H}_{68}\text{N}_2\text{O}_{14}\text{S}_4\text{Na}$ ($\text{M}+\text{Na}^+$) found 1143.88; 1159.32, calc. for $\text{C}_{56}\text{H}_{68}\text{N}_2\text{O}_{14}\text{S}_4\text{K}$ ($\text{M}+\text{K}^+$) found 1159.88.

IR of the dry gel (cm^{-1}): 3280 ($\nu_{\text{N-H}}$), 3067 ($\nu_{\text{C-H}}$), 1444, 1410 ($\nu_{\text{C=C}}$), 1370, 1258 ($\nu_{\text{C-H}}$), 1013-1130 ($\nu_{\text{C-C}}$), 845-863 ($\nu_{\text{C-O-C}}$), 790 ($\nu_{\text{C-H}}$, $\nu_{\text{C-S-C}}$).

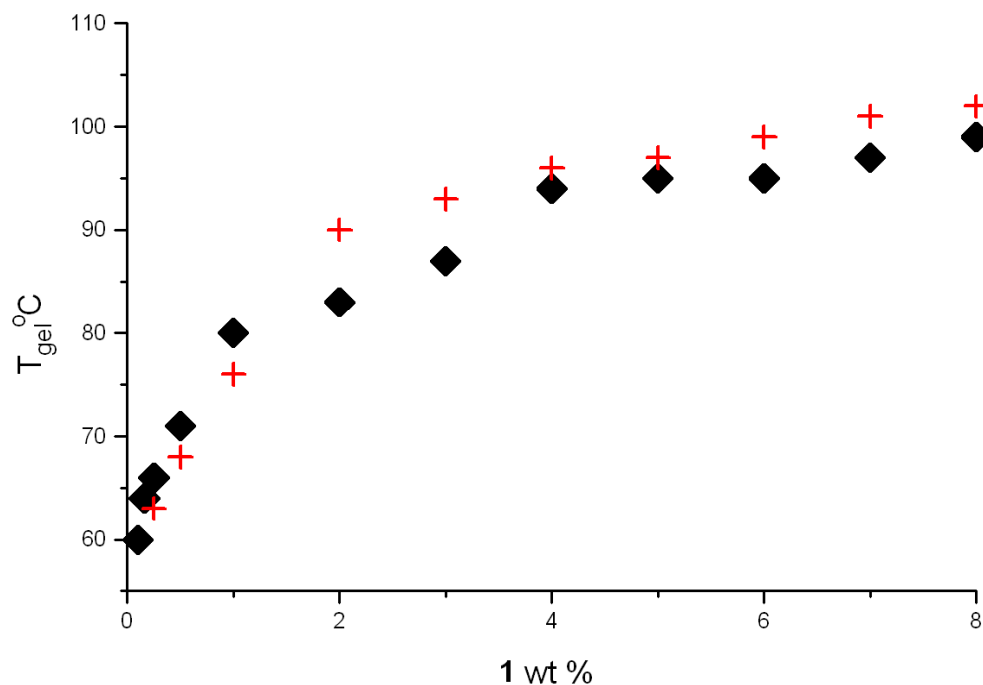


Fig. S1 T_{gel} changes with concentration of **1** from: EtOH/CHCl₃ (8:2) – (square) and *n*-PrOH/CHCl₃ (8:2) – (cross).

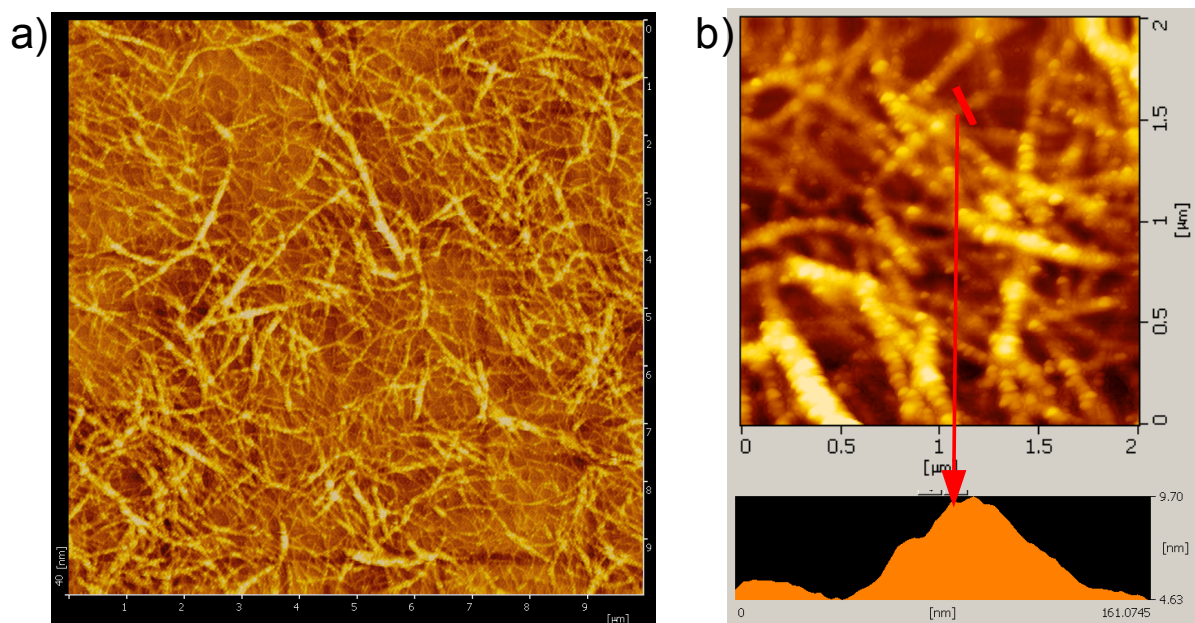


Fig. S2 (a) AFM image of nano-fibre structure of the gel of **1** 0.25 wt% - EtOH/CHCl₃ (8:2); (b) Nano-fibre height measurements.

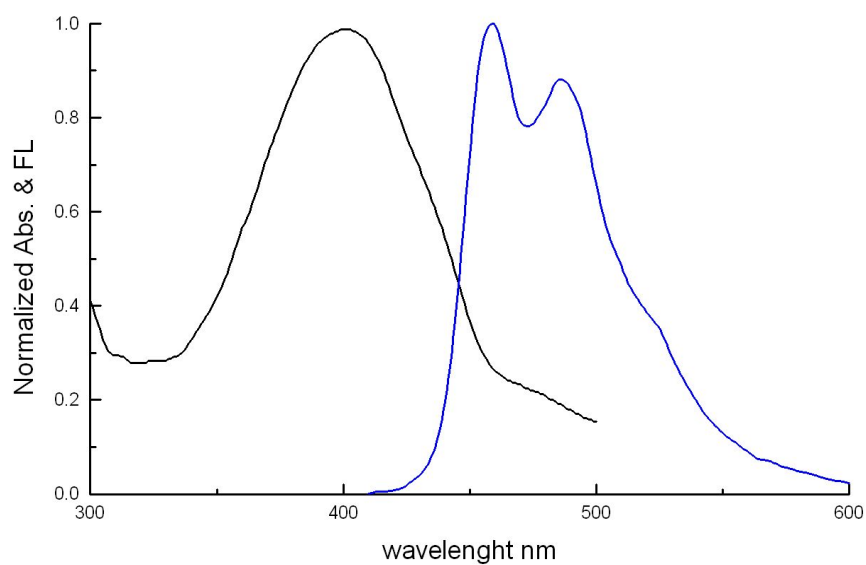


Fig S3 Normalized absorption (black) and fluorescence (blue) spectra in EtOH ($8.6 \mu\text{mol dm}^{-3}$).

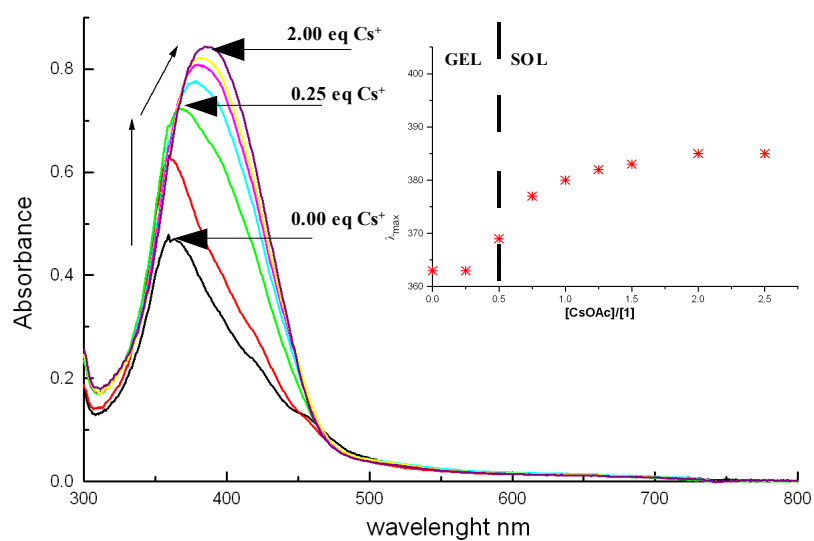


Fig. S4 UV/Vis titration of the gel of **1** 0.25%wt by CsOAc in EtOH/CHCl₃ (8:2). Inset: λ_{max} changes with increasing concentration of CsOAc and the gel-to-sol phase transition.