Electronic Supplementary Information

Surfactant-mediated control of CBPQT⁴⁺- dialkoxynaphthalene complexation.

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1.General Information

Materials. All reagents were purchased from Aldrich and were used as received unless otherwise mentioned.

Instrumentation and Measurements. ¹H and ¹³C NMR spectra were recorded on a Bruker 400 MHz spectrometer. Mass spectrometry was carried out on a Micromass Platform II spectrometer with an electrospray ionization source. UV/vis turbidity measurements were carried out on a Varian Cary 50 Scan UV/vis spectrophotometer equipped with a single cell Peltier temperature controller. IR spectra were recorded using a Spectrum One PerkinElmer spectrometer. Isothermal titration calorimetry (ITC) experiments were performed at 20 °C using a nano-ITC titration calorimeter from TA Instruments with a standard sample cell volume of 1mL, following standard procedures. A 250 μ L injection syringe was used with stirring at 400 rpm. Samples were dissolved in deionised water and the solutions were degassed gently under vacuum before use. Each titration comprised an initial 1 μ l pre-injection followed by 25 x 10 μ l injections of (3.5 mM) into host solution (0.06 mM). Control experiments with identical injections of CBPQT(4 CI) into water alone were used to correct titration data.

2. Synthesis and characterization

Compound $\mathbf{3}^{[1]}$ was prepared according to the literature methods.

Synthesis of 4

Triphenylphosphine (1.56 g, 5.96 mmol, 1 eq.) was added to a solution of carbon tetrabromide (1.98 g, 5.96 mmol, 1eq.) and **3** (2 g, 5.96 mmol, 1eq.) in dry acetonitrile (200 mL). The mixture was stirred at room temperature under nitrogen for 24h. The solvent was evaporated to afford a crude product which was subjected to column chromatography (SiO₂: Dichloromethane / Ethyl Acetate, 3:1). The product was obtained as a white solid in 45 % yield.

¹H NMR (400 MHz, CDCl₃): δ = 7.79 (d, J=8.4 Hz, 2H, H-4,8), 7.29 (t, J=8 Hz, 2H, H-3,7), 6.77 (d, J=7.6 Hz, 2H, H-2,6), 4.23 (t, J=4.6 Hz, 4H, 2×CH₂O), 3.95-3.87 (m, 6H, 3×CH₂O), 3.68 (m, 4H, 2×CH₂O, 3.45 (t, J=6 Hz, 2H, CH₂Br).

¹³C NMR (100 MHz, CDCl₃): δ = 154.30, 154.28 (aromatic C-1,5), 126.78 (Aromatic C-9,10), 125.24, 125.18 (Aromatic (C-7,3), 114.74, 114.63 (Aromatic C-4,8), 105.83, 105.79 (Aromatic C-2,6), 72.64, 71.51, 69.79, 69.77, 67.96, 67.92, 61.86 (CH₂O), 30.47 (CH₂Br).

Synthesis of 1:

To a solution of **4** (500 mg, 1.25 mmol, 1 eq.) in methanol (70 mL) was added dropwise trimethylamine 31-35 % in ethanol (4.5 mL, 18.8 mmol, 15 eq.). The mixture was stirred at reflux overnight. The mixture was concentrated under reduce pressure to a volume of 10 mL and diethyl ether (250 mL) was slowly added. The precipitate was filtered and washed with diethyl ether. The product was obtained as a white solid in 95 % yield. Mpt. 51-53 °C (dec).

¹H NMR (300 MHz, D₂O): δ = 7.77 (d, J= 8.4 Hz, 1H, H-4), 7.71 (d, J=8.4 Hz, 1H, H-8), 7.34 (t, J= 8.1 Hz, 2H, H-3,7), 6.95 (d, J= 8.1 Hz, 1H, H-6), 6.92 (d, J= 8.1 Hz, 1H, H-2), 4.30-4.19 (m, 4H, CH₂O), 3.86 (m, 6H, CH₂O), 3.61 (m, 4H, CH₂O), 3.38 (t, J = 4.8 Hz, 2H, N-CH₂), 2.90 (s, 9H, N-CH₃).

¹³C NMR (100 MHz, D₂O): δ = 153.7, 153.6, 126.2, 125.9, 125.8, 114.4, 114.1, 107.1, 106.8, 72.6, 71.5, 69.8, 69.77, 68.0, 67.9, 61.9, 30.5, 53.7, 53.6, 53.6

MS (ESI⁺): 378 (M⁺, 100).



Fig S2. Partial ¹H- ¹H COSY spectrum of compound **1** recorded in D_2O .



Fig S3. Partial ¹H- ¹H COSY spectra of compound **1** recorded in D_2O .



Fig S4. Partial ¹H NMR spectrum of 1 (red curve) and 3 (dark curve) recorded in D_2O

153.7268 153.5885	126.1666 125.8948 125.8262	114.4324 114.1182 107.0703 106.8134	71.8918 69.2071 69.2071 67.63934 67.63934 65.1263 65.1263 65.1263 53.5774 53.5774 53.5767
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[1] P. R. Ahston, R. Ballardini, V. Balzani, S. E. Boyd, A. Credi, *Chem. Eur. J.* 1997, **3**, 152-170.

<u>3. Complexation studies</u>



Fig. S6. UV-vis spectra of **1.2** (~10⁻⁴ M). Recorded in D_2O solution at 25°C.



Fig S7. Isothermal titration calorimetry data for the addition of aliquots of 1 to 2. Recorded in H_2O at 25°C.

4. Micellization studies.



Fig S8. Graph showing the intensity of fluorescence emission of 1 and 3 as a function of the SMS and SDS concentration, respectively. λ_{ex} = 295nm. λ_{em} = 329 nm. Recorded in H₂O at 25°C



Fig S9. Partial ¹H NMR spectrum of **1** (3.6 mM) with various concentrations of SDS recorded in D_2O .



Fig S10. Partial ¹H NMR spectrum of 1 (3.6 mM) with various concentrations of SDS recorded in D_2O



Fig S11. Partial ¹H-¹H COSY spectrum of **1** (3.6 mM) with SDS (17.28 mM) recorded in D_2O .



Fig S12. Incorporation of **1** into micelles of SDS: Graphs showing the effect of SDS concentration on the chemical shift of protons $H_{4/8}$, $H_{3/7}$ and $H_{2/6}$.



Fig S13. Partial ¹H NMR spectrum of **1** (3.6 mM) recorded in the presence of various concentrations of SDS. Recorded in D_2O .



Fig S14. Incorporation of 1 into micelles of SDS: Graphs showing the effect of SDS concentration on the chemical shift of protons $H_{a/b}$, H_i .

5. Control of the complexation between 1.2



Fig S15. UV-vis spectra of: **1.2** (black curve), **1.2** +SDS (supernatant, red curve), **1.2** +SMS (blue curve). ($\sim 10^{-4}$ M) at 25°C. Recorded in H₂O.



2.3 +SDS 1.2 +SDS

(after centrifugation) (after centrifugation) **Fig S16.** Photographs of the solids recovered (following centrifugation) upon the addition of SDS to **2.3** and **1.2**.



Fig S17. Partial ¹H NMR spectra of: (a)CBPQT⁴⁺,4DS (1mM); (b) CBPQT⁴⁺,4Cl⁻/**3** + SDS (precipitate) and (c) **3** in DMSO- d_6 . Recorded at 25°C.



Fig S18. UV-vis spectrum of **2.3**, $(4DS^{-})$ (~10⁻⁴ M). Recorded in DMSO at 25°C



Fig S19. UV-vis spectra of **1.2** (~5 mM) (black curve) and **1.2** + **SDS** + HCl _{conc} in excess (red curve). (final concentration of **1.2**, $(4DS^{-}) \sim 2.7$ mM). Recorded in H₂O at 25°C