## **Supporting Information**

## **Direct Electrochemical Imidation of Aliphatic Amines via**

## **Anodic Oxidation**

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**General Remarks:** <sup>1</sup>H NMR and <sup>13</sup>C NMR were recorded on a Bruker AC-300 FT (<sup>1</sup>H: 300 MHz, <sup>13</sup>C: 75 MHz) using TMS as internal reference. The chemical shifts ( $\delta$ ) and coupling constants (*J*) were expressed in ppm and Hz respectively. Infrared samples were recorded on a Perkin-Elmer 2000 FTIR spectrometer. HPLC analysis was carried out on an Agilent 1100 series HPLC with a multiple wavelength detector. Chiralpak OD columns was purchased from Daicel Chemical Industries, LTD. Hexane: 2-propanol = 80:20, flow rate = 0.45 mL/min, T = 23°C, UV = 254 nm.

## 1. Experimental Section.

**Instruments:** The instrument for electrolysis is dual display potentiostat (CJS-292) (made in China). Cyclic voltammetric (CV) experiments were carried out with a CHI400A system (USA CH Instruments) in a conventional three-electrode cell in the presence of supporting electrolyte. The anode electrode is Pt  $(1.0 \times 1.0 \text{ cm}^2)$  or a graphite (diameter 0.5 cm) while the cathode electrode is graphite (diameter 0.5 cm). A saturated calomel electrode (SCE) were used as the reference electrode. CH<sub>3</sub>CN and CH<sub>2</sub>Cl<sub>2</sub> were distilled from CaH<sub>2</sub>. Et<sub>3</sub>N, THF, hexane and 1, 4-dioxane were distilled from sodium/benzophenone. CCl<sub>4</sub> and CHCl<sub>3</sub> were dried over calcium chloride and distilled. *Caution: Azides and diazoalkanes may be hazardous and/or explosive.* 

**Representative procedures for imidation of tertiary amines:** An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In the electrolytic cell a solution of  $TsN_3$  (0.2 mmol), tertiary amine (0.4 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CH<sub>3</sub>CN (6 mL) was added. In addition, for the safety, a gas tube was installed in the electrolytic cell and the terminal of this gas tube was inserted in the excess solution of benzoic acid (1.22g, 10 mmol, benzoic acid dissolved in CH<sub>3</sub>CN) in a two-neck flask. One of the two necks was equipped with a buffer balloon for the safe. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 4 mA (electrode square 1.6 cm<sup>2</sup>)

at ambient temperature (25±1°C) for 3 h until the quantity of the electricity 2.2 F/mol was passed.

Upon completion of the reaction, the solvent was removed with a rotary evaporator. The residue was washed with anhydrous ether  $(3 \times 10 \text{ mL})$  and the insoluble Bu<sub>4</sub>NPF<sub>6</sub> was filtered and dried for next use. The filtrate was combined and the solvent was removed with a rotary evaporator. The residue was purified by column chromatography on silica gel. The schematic diagram is listed as follows:



Representative procedures for imidation of secondary amines: An undivided cell was

equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of  $TsN_3$  (0.2 mmol), secondary amine (0.6 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CH<sub>3</sub>CN (6 mL) was added. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 6 mA (electrode square 1.6 cm<sup>2</sup>) at ambient

temperature (25±1°C) for 3 h until the quantity of the electricity 3.4 F/mol was passed. Upon

completion of the reaction, the solvent was removed with a rotary evaporator. The residue was washed with anhydrous ether ( $3 \times 10$  mL) and the insoluble Bu<sub>4</sub>NPF<sub>6</sub> was filtered and dried for next use. The filtrate was combined and the solvent was removed with a rotary evaporator. The residue was purified by column chromatography on silica gel (Ditto for the handling of diazoalkanes).

**Representative procedures for imidation of primary amines:** An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of  $TsN_3$  (0.2 mmol), primary amine (1.0 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CHCl<sub>3</sub> (6 mL) was added. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 7 mA (electrode square 1.6 cm<sup>2</sup>) at ambient

temperature (25±1°C) for 3 h until the quantity of the electricity 3.9 F/mol was passed. Upon

completion of the reaction, the solvent was removed with a rotary evaporator. The residue was washed with anhydrous ether ( $3 \times 10$  mL) and the insoluble Bu<sub>4</sub>NPF<sub>6</sub> was filtered and dried for next use. The filtrate was combined and the solvent was removed with a rotary evaporator. The residue was purified by column chromatography on silica gel (Ditto for the handling of diazoalkanes).

### Experimental details for the capture of radical cation A:

An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of triethylamine (5 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CHCl<sub>3</sub> (6 mL) was added. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 4 mA (electrode square 1.6 cm<sup>2</sup>) at ambient

temperature (25±1°C) for 0.5 h. Then the electrolyte was frozen by liquid nitrogen. The EPR

measurements were performed with a Braker Elexsys X-band (9.7 GHz) E580 EPR spectrometer at room temperature (Ditto for the handling of diazoalkanes).

The X-band EPR measurement was conducted at room temperature and the result was shown in Figure 1. The characterized *g*- value of the radical was 2.0022. The EPR simulation demonstrated that the magnetic interactions between the unpaired electron and one <sup>14</sup>N and six equivalent <sup>1</sup>H (see **A** in Scheme 1) gave rise to the resolved isotropic hyperfine constants,  $A_{14N}$  (43.69 MHz) and  $A_{1H}$  (30.98 MHz) displayed in Figure 1.

#### Experimental details for the measurement of CV of Et<sub>3</sub>N:

The cyclic voltammetry (CV) of Et<sub>3</sub>N was measured in 0.05 M n-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN. As shown in

Figure S1 (Supporting Information, SI), the value of  $\pm 1.60$  V (vs SCE) in trace **b** (green) indicated the oxidation potential of Et<sub>3</sub>N in acetonitrile while trace **a** (red) showed that blank solution was not electroactive in the potential window of interest.



**Figure S1.** Cyclic voltammetry (CV) curves of a) 0.05 M n-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN (red), b) 1mmol Et<sub>3</sub>N, 0.05 M n-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN (green), recorded at a glassy carbon electrode (diameter 4.0 mm), Scan rate: 100 mV s<sup>-1</sup>, at room temperature.

### Experimental details for the optimization of the reaction conditions:

The optimization of the reaction conditions was achieved by screening different solvents and electrodes, as shown in Table S1. When the platinum anode was replaced with a graphite anode, the reaction yield was hexane, and H<sub>2</sub>O hardly gave the desired product and the use of MeOH and THF afforded the product with poor yields (Table S1, entries 8-14). In comparison with acetonitrile, the solvent CHCl<sub>3</sub> gave almost the same yield under the same condition while CH<sub>2</sub>Cl<sub>2</sub> gave a moderate yield (Table S1, entries 3-7). Therefore CH<sub>3</sub>CN and CHCl<sub>3</sub> should be the best solvents. Taking all factors into the consideration, the standard reaction condition was established as follows: CH<sub>3</sub>CN as the solvent, Bu<sub>4</sub>NPF<sub>6</sub> as the electrolyte, graphite as both anode and cathode.

Table S1. Synthesis of sulfonyl amidine under various conditions<sup>a</sup>



entry	solvent	anode	cathode	yield(%) <sup>b</sup>
1	CH <sub>3</sub> CN	Pt	С	67
2	CH <sub>3</sub> CN	С	С	96
3	$CH_2Cl_2$	Pt	С	45
4	$CH_2Cl_2$	С	С	68
5	$CH_2Cl_2$	Pt	Pt	39
6	CHCl <sub>3</sub>	Pt	С	73
7	CHCl <sub>3</sub>	С	С	94
8	1,4-dioxane	Pt	С	trace
9	1,4-dioxane	С	С	trace
10	$CCl_4$	С	С	N. D.
11	hexane	С	С	N. D.
12	THF	С	С	38
13	MeOH	С	С	33
14 <sup>c</sup>	$H_2O$	С	С	trace

<sup>*a*</sup> Reaction conditions: the mixture of 0.2 mmol of TsN<sub>3</sub>, 0.4 mmol of triethylamine, 0.3 mmol Bu<sub>4</sub>NPF<sub>6</sub> and 6 mL of solvent was stirred and electrolyzed with constant current of 4 mA at room temperature for 3 h. <sup>*b*</sup> Isolated yield based on TsN<sub>3</sub>. <sup>*c*</sup> Reaction conditions: the mixture of 0.2 mmol of TsN<sub>3</sub>, 0.4 mmol of triethylamine, 0.3 mmol KNO<sub>3</sub> and 6 mL of H<sub>2</sub>O was stirred and electrolyzed with constant current of 20 mA at room temperature for 3 h.

#### Experimental details for the investigation of reaction mechanism:

The electrolysis of  $TsN_3$  and  $Et_3N$  was performed separately. After electrolysis of  $TsN_3$  alone for 3 h,  $Et_3N$  was added and stirred for 1 h. Afterwards,  $TsN_3$  could be recovered with a yield of 98% and no product was detected. In the parallel electrolysis of  $Et_3N$  for 3 h, the solution color was changed from colorless into yellow. When  $TsN_3$  was added to this yellow solution and the mixture was stirred for 1 h, the corresponding sulfonyl amidine was obtained with a yield of 88%. These results showed that only  $Et_3N$  was electroactive substrate in this reaction.

#### Experimental details for the capture of enamine:



Scheme S1

An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of triethylamine (5 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CH<sub>3</sub>CN (6 mL) was electrolyzed. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 4 mA (electrode square 1.6 cm<sup>2</sup>) at ambient

temperature (25±1°C) for 2 h. Then 2, 4-dinitrobenzenehydrazine (0.100g, 0.5 mmol) was added

and the reaction was quenched with 0.1 N and extracted with ethyl acetate ( $3 \times 10$  mL). The combined organic layers were washed with brine, dried over sodium sulfate. The filtrate was concentrated under reduced pressure. The residue was washed with anhydrous ether ( $3 \times 10$  mL) and the insoluble Bu<sub>4</sub>NPF<sub>6</sub> was filtered and dried for next use. The filtrate was combined and the

solvent was removed with a rotary evaporator. The residue was purified by column chromatography on silica gel.

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 11.16 (s, 0.43 H), 11.03 (s, 0.95 H), 9.20-9.00 (m, 0.89 H), 8.40-8.20 (m, 1.22 H), 8.00-7.85 (m, 1.25 H), 7.70-7.40 (m, 1.05 H), 7.20-7.00 (m, 0.41 H), 2.14 (d, *J* = 5.4 Hz, 3 H), 2.08 (d, *J* = 5.7 Hz, 1.5 H).



Figure S2

Experimental details for the capture of CH<sub>2</sub>N<sub>2</sub>:



*Caution: Azides and diazoalkanes may be hazardous and/or explosive.* An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of  $TsN_3$  (2 mmol), tertiary amine (4 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CH<sub>3</sub>CN (6 mL) was added. In addition, for the safety, a gas tube was installed in the electrolytic cell and the terminal of this gas tube was inserted in the excess solution of benzoic acid (1.22g, 10 mmol, benzoic acid dissolved in CH<sub>3</sub>CN) in a two-neck flask. One of the two necks was equipped with a buffer balloon for the safe. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 4 mA (electrode square 1.6 cm<sup>2</sup>) at

ambient temperature (25±1°C) for 4 h. Upon completion of the reaction, the solvent was removed

with a rotary evaporator. The residue was washed with anhydrous ether  $(3 \times 10 \text{ mL})$ . The filtrate was combined and the solvent was removed with a rotary evaporator. The residue was purified by column chromatography on silica gel.

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.10-7.90 (m, 2 H), 7.60-7.50 (m, 1 H), 7.50-7.40 (m, 2 H), 3.92 (s, 3 H).





Figure S3

#### A representative mechanism for imidation of cyclic tertiary amine:

The mechanism of imidation of cyclic tertiary amine was also proposed as shown in Scheme S3. First, N-methylpiperidine is oxidized to form iminium ion **E** through the same route as triethylamine. It is easier for the cyclic hydrogen to be eliminated to produce iminium ion.<sup>7a,8h</sup> Then, iminium ion **E** is attacked by sulfonyl azide (nucleophile) with the release of one molecule of N<sub>2</sub> (Scheme S3, SI). At the cathode, the proton produced at the anode is reduced to hydrogen.



Scheme S3. Proposed mechanism for imidation of cyclic tertiary amine.

#### Experimental details for the measurment of CV of Et<sub>2</sub>NH:

The cyclic voltammetry (CV) of  $Et_2NH$  was measured in 0.05 M *n*-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN. As shown in Figure S4 (Supporting Information, SI), the value of +1.80 V (vs SCE) in trace **b** (green) indicated the oxidation potential of  $Et_2NH$  in acetonitrile while trace **a** (red) showed that blank solution was not electroactive in the potential window of interest.



**Figure S4.** Cyclic voltammetry (CV) curves of a) 0.05 M *n*-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN (red), b) 1mmol Et<sub>2</sub>NH, 0.05 M *n*-Bu<sub>4</sub>NPF<sub>6</sub>/CH<sub>3</sub>CN (green), recorded at a glassy carbon electrode (diameter 4.0 mm), Scan rate: 100 mV s<sup>-1</sup>, at room temperature.

#### Experimental details for the capture of ethylamine:

An undivided cell was equipped with a magnet stirrer, two graphite electrode both as the working electrode and the counter electrode respectively. In this cell a solution of benzaldehyde (0.5 mmol), diethylamine (5 mmol),  $Bu_4NPF_6$  (0.3 mmol) in CH<sub>3</sub>CN (6 mL) was electrolyzed. The electrolyte was allowed to stir and the electrolysis was carried out at a constant current of 6 mA (electrode square 1.6 cm<sup>2</sup>) at ambient temperature (25±1°C) for 4 h. Then the electrolyte was characterized by GC-MS.



Figure S5

## 2. Characterization data of all products.

**General Remarks:** <sup>1</sup>H NMR and <sup>13</sup>C NMR were recorded on a Bruker AC-300 FT (<sup>1</sup>H: 300 MHz, <sup>13</sup>C: 75 MHz) using TMS as internal reference. The chemical shifts ( $\delta$ ) and coupling constants (*J*) were expressed in ppm and Hz respectively. Infrared samples were recorded on a Perkin-Elmer 2000 FTIR spectrometer.

#### (E)-N, N-diethyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.14 (s, 1 H), 7.75 (d, *J* = 8.4 Hz, 2 H), 7.25 (d, *J* = 8.4 Hz, 2 H), 3.60-3.32 (m, 4 H), 2.39 (s, 3 H), 1.31-1.20 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.1, 142.4, 139.9, 129.4, 126.5, 47.2, 41.1, 29.8, 21.6, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2978, 2937, 1610, 1451, 1345, 1298, 1283, 1148, 1087, 955, 875, 817, 768, 674.

### (E)-N, N-diethyl-N'-(phenylsulfonyl)formimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.16 (s, 1 H), 8.00-7.80 (m, 2 H), 7.60-7.40 (m, 3 H),

3.60-3.32 (m, 4 H), 1.30-1.24 (t, J = 7.2 Hz, 3 H), 1.20-1.10 (t, J = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta = 158.3$ , 131.8, 128.9, 128.8, 126.5, 47.2, 41.1, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): v = 2978, 2938, 1612, 1447, 1345, 1299, 1149, 1088, 955, 875, 819, 770, 723, 690.

#### (E)-N, N-diethyl-N'-(4-methoxyphenylsulfonyl)formimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.14 (s, 1 H), 7.81 (d, *J* = 9.0 Hz, 2 H), 6.92 (d, *J* = 9.0 Hz, 2 H), 3.84 (s, 3 H), 3.53-3.30 (m, 4 H), 1.30-1.20 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.0, 128.5, 113.9, 55.6, 47.1, 41.0, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2977, 2939, 1612, 1499, 1452, 1346, 1290, 1257, 1146, 1090, 1025, 955, 875, 768, 676.

#### (E)-N'-(4-bromophenylsulfonyl)-N, N-diethylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.13 (s, 1 H), 7.75 (d, *J* = 8.4 Hz, 2 H), 7.59 (d, *J* = 8.4 Hz, 2 H), 3.53-3.30 (m, 4 H), 1.30-1.20 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.2, 132.0, 128.1, 47.3, 41.2, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2977, 2938, 1611, 1449, 1344, 1302, 1271, 1147, 1086, 877, 770, 740.

### (E)-N, N-diethyl-N'-(4-nitrophenylsulfonyl)formimidamide

$$O_2N \rightarrow O_2N \rightarrow$$

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  =8.31 (d, *J* = 9.0 Hz, 2 H), 8.16 (s, 1 H), 8.07 (d, *J* = 9.0 Hz, 2 H), 3.55-3.32 (m, 4 H), 1.33-1.25 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.5, 127.8, 124.1, 47.6, 41.4, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): v = 2980, 2940, 1613, 1528, 1449, 1345, 1295, 1150, 1086, 957, 881, 853, 732, 686.

#### (E)-N, N-diethyl-N'-(2-nitrophenylsulfonyl)formimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.30-8.20 (m, 1 H), 8.11 (s, 1 H), 7.80-7.60 (m, 3 H), 3.60-3.32 (m, 4 H), 1.39-1.29 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 160.0, 132.8, 132.1, 130.9, 124.1, 47.5, 41.4, 14.5, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2979, 2939, 1616, 1540, 1344, 1309, 1154, 1121, 880, 772, 607.

#### (E)-N,N-diethyl-N'-(3-nitrophenylsulfonyl)formimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.71 (s, 1 H), 8.40-8.34 (m, 1 H), 8.24 (d, *J* = 7.8 Hz, 1 H), 8.18 (s, 1 H), 7.73-7.60 (m, 1 H), 3.60-3.38 (m, 4 H), 1.35-1.27 (t, *J* = 7.2 Hz, 3 H), 1.20-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.5, 132.3, 130.2, 128.9, 126.4, 121.8, 47.5, 41.4, 14.5, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2979, 2939, 1613, 1530, 1351, 1156, 1118, 886, 664, 610, 587.

#### (E)-N'-(2,5-dibromophenylsulfonyl)-N,N-diethylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.39 (s, 1 H), 8.29 (s, 1 H), 7.56-7.40 (m, 2 H), 3.60-3.40 (m, 4 H), 1.40-1.29 (t, *J* = 7.2 Hz, 3 H), 1.25-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 160.2, 136.3, 135.9, 133.5, 121.6, 47.5, 41.5, 14.7, 12.1; IR (liquid film, cm<sup>-1</sup>): *v* = 2978, 2937, 1614, 1445, 1343, 1303, 1153, 1023, 956, 880, 882, 766, 611, 592.

(E)-N'-(((1R,4R)-7,7-dimethyl-2-oxobicyclo[2.2.1]heptan-1-yl)methylsulfonyl)-N,N-diethylfor mimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta = 8.05$  (s, 1 H), 3.55-3.41 (m, 4 H), 3.41-3.35 (m, 1 H), 3.10-2.90 (m, 1 H), 2.75-2.55 (m, 1 H), 2.40-2.20 (m, 1 H), 2.10-1.96 (m, 2 H), 1.95-1.80 (m, 1 H), 1.80-1.60 (m, 1 H), 1.50-1.35 (m, 1 H), 1.35-1.25 (t, J = 7.2 Hz, 3 H), 1.25-1.10 (t, J = 7.2 Hz, 3 H), 1.15 (s, 3 H), 0.85 (s, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta = 158.8$ , 58.6, 50.8, 48.1, 47.0, 42.8, 42.7, 40.8, 27.1, 24.8, 20.1, 19.8, 14.5, 12.1; IR (liquid film, cm<sup>-1</sup>): v = 2966, 2887, 1744, 1613, 1454, 1352, 1304, 1127, 955, 873, 766.

#### (E)-N,N-diethyl-N'-(thiophen-2-ylsulfonyl)formimidamide

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.15 (s, 1 H), 7.62-7.53 (m, 1 H), 7.50-7.45 (m, 1 H), 7.05-7.00 (m, 1 H), 3.60-3.40 (m, 4 H), 1.40-1.25 (t, *J* = 7.2 Hz, 3 H), 1.23-1.10 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.4, 130.6, 130.4, 127.0, 47.4, 41.3, 14.6, 12.2; IR (liquid film, cm<sup>-1</sup>): *v* = 2977, 2938, 1611, 1451, 1341, 1298, 1133, 1088, 1014, 876, 672.

#### (E)-N,N-dipropyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.15 (s, 1 H), 7.75 (d, *J* = 8.4 Hz, 2 H), 7.25 (d, *J* = 8.4 Hz, 2 H), 3.40-3.20 (m, 4 H), 2.39 (s, 3 H), 1.68-1.50 (m, 4 H), 1.00-0.80 (m, 6 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 159.0, 129.4, 126.5, 54.4, 48.0, 22.1, 21.6, 20.1, 11.3, 11.0; IR (liquid film, cm<sup>-1</sup>): *v* = 2964, 2931, 1607, 1451, 1343, 1297, 1283, 1147, 1087, 909, 877, 674.

#### (E)-N,N-diisopropyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.25 (s, 1 H), 7.75 (d, *J* = 8.4 Hz, 2 H), 7.25 (d, *J* = 8.4 Hz, 2 H), 4.60-4.40 (m, 1 H), 3.80-3.50 (m, 1 H), 2.39 (s, 3 H), 1.31 (d, *J* = 6.9 Hz, 6 H), 1.21 (d, *J* = 6.9 Hz, 6 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 156.5, 129.4, 126.4, 48.6, 48.0, 23.7, 21.6, 19.8; IR (liquid film, cm<sup>-1</sup>): *v* = 2976, 2933, 1602, 1341, 1282, 1146, 1088, 891, 839, 670.

#### (E)-4-methyl-N-(1-methylpyrrolidin-2-ylidene)benzenesulfonamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 7.81 (d, *J* = 8.4 Hz, 2 H), 7.24 (d, *J* = 8.4 Hz, 2 H), 3.50-3.40 (t, *J* = 7.2 Hz, 2 H), 3.10-3.00 (t, *J* = 7.8 Hz, 2 H), 2.97 (s, 3 H), 2.39 (s, 3 H), 2.20-2.00 (m, 2 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 170.0, 142.1, 140.7, 129.3, 126.6, 51.8, 32.1, 30.8, 21.6, 19.1; IR (liquid film, cm<sup>-1</sup>): *v* = 2927, 1600, 1492, 1301, 1281, 1144, 1091, 1007, 902, 813, 670, 606, 580, 558.

#### (E)-N-(1-ethylpiperidin-2-ylidene)-4-methylbenzenesulfonamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 7.81 (d, *J* = 8.4 Hz, 2 H), 7.26 (d, *J* = 8.4 Hz, 2 H), 3.60-3.40 (m, 2 H), 3.38-3.10 (t, *J* = 6.0 Hz, 2 H), 3.10-3.00 (t, *J* = 6.0 Hz, 2 H), 2.40 (s, 3 H), 1.80-1.70 (m, 4 H), 1.18-1.00 (t, *J* = 7.2 Hz, 3 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 165.3, 141.7, 129.8, 129.2, 126.6, 126.3, 48.3, 45.7, 28.7, 22.4, 21.5, 19.7, 11.5; IR (liquid film, cm<sup>-1</sup>): *v* = 2937, 2871, 1562, 1482, 1261, 1141, 1093, 1066, 935, 815, 671, 584, 554.

#### (E)-4-methyl-N-(1-methylpiperidin-2-ylidene)benzenesulfonamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 7.82 (d, *J* = 7.8 Hz, 2 H), 7.24 (d, *J* = 7.8 Hz, 2 H), 3.40-3.30 (m, 2 H), 3.03 (s, 3 H), 2.39 (s, 3 H), 1.90-1.60 (m, 6 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz,

ppm):  $\delta = 166.1$ , 141.8, 141.5, 129.6, 129.2, 126.6, 126.4, 50.9, 38.5, 28.7, 22.3, 21.4, 19.8; IR (liquid film, cm<sup>-1</sup>): v = 2950, 2870, 1571, 1483, 1273, 1142, 1089, 962, 823, 675.

#### (Z/E)-N,N-dipropyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.15 (s, 0.59 H), 7.81 (d, *J* = 8.4 Hz, 0.69 H), 7.75 (d, *J* = 8.4 Hz, 1.14 H), 7.30-7.20 (m, 1.83 H), 3.40-3.20 (m, 4 H), 2.39 (s, 3 H), 1.80-1.40 (m, 4 H), 1.00-0.70 (m, 6 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 169.0, 162.9, 159.0, 142.3, 139.9, 129.7, 129.4, 129.1, 129.0, 126.5, 126.4, 54.4, 50.7, 49.3, 47.9, 24.4, 23.4, 22.4, 22.2, 21.5, 20.1, 11.9, 11.4; IR (liquid film, cm<sup>-1</sup>): *v* = 2965, 2935, 2876, 1670, 1609, 1545, 1297, 1283, 1147, 1087, 878, 676.

#### (Z/E)-N,N-dibutyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.13 (s, 0.82 H), 7.82 (d, *J* = 8.4 Hz, 0.32 H), 7.75 (d, *J* = 8.4 Hz, 1.61 H), 7.30-7.20 (m, 1.94 H), 3.50-3.20 (m, 4 H), 2.36 (s, 3 H), 1.70-1.40 (m, 4 H), 1.40-1.10 (m, 4 H), 1.00-0.80 (m, 6 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.8, 142.3, 140.0, 129.3, 129.0, 126.4, 126.1, 52.4, 47.1, 46.0, 30.8, 28.8, 21.5, 20.2, 20.0, 19.7, 13.8, 13.7, 13.6; IR (liquid film, cm<sup>-1</sup>): *v* = 2958, 2932, 2872, 1672, 1608, 1458, 1348, 1298, 1147, 1088, 892, 814.

#### (Z/E)-N-ethyl-N-methyl-N'-tosylformimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.18 (s, 0.68 H), 8.09 (s, 0.36 H), 7.80-7.70 (m, 2 H), 7.25 (d, *J* = 7.2 Hz, 2 H), 3.50-3.30 (m, 2 H), 3.08 (s, 1 H), 2.98 (s, 2 H), 2.40 (s, 3 H),1.28-1.20 (t, 2 H), 1.20-1.10 (m, 1 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 158.7, 158.5, 142.5, 142.4, 139.8, 139.7, 129.4, 129.3, 126.6, 126.5, 49.6, 43.1, 38.9, 33.2, 21.6, 13.9, 11.3; IR (liquid film, cm<sup>-1</sup>): *v* = 2977, 2935, 1617, 1343, 1297, 1282, 1147, 1086, 905, 888, 673.

#### (Z/E)-N-propyl- N'-tosylpropionimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta$  = 8.25 (s, 0.39 H), 7.85-7.70 (m, 2.04 H), 7.30-7.20 (m, 2.22

H), 5.90 (s, 0.57 H), 3.35-3.20 (m, 2 H), 2.92-2.80 (m, 1 H), 2.40 (s, 3 H), 2.40-2.30 (m, 1 H), 1.70-1.50 (m, 2.19 H), 1.30-1.20 (m, 1.87 H), 1.20-1.10 (m, 1.37 H) 1.00-0.90 (m, 1.46 H) 0.90-0.80 (m, 1.85 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta$  = 169.7, 169.6, 142.7, 142.0, 141.3, 140.0, 129.4, 129.2, 126.4, 126.3, 45.5, 43.7, 27.5, 26.9, 23.1, 21.8, 21.6, 21.5, 11.5, 11.4, 11.2, 10.4; IR (liquid film, cm<sup>-1</sup>): v = 3313, 2966, 2934, 2877, 1558, 1271, 1142, 1088, 953, 694. HRMS calc.  $C_{13}H_{20}N_2O_2S$  (M<sup>+</sup>): 268.1245. Found: 268.1250.

#### (Z/E)-N-butyl- N'-tosylbutyrimidamide



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz, ppm):  $\delta = 8.20$  (s, 0.35 H), 7.85-7.70 (m, 1.87 H), 7.30-7.20 (m, 2.28 H), 5.67 (s, 0.43 H), 3.40-3.20 (m, 1.86 H), 2.80-2.60 (m, 0.97 H), 2.43 (s, 2.87 H), 2.30-2.20 (m, 0.88 H), 1.80-1.45 (m, 4.57 H), 1.45-1.35 (m, 0.96 H), 1.35-1.20 (m, 1.00 H) 1.10-0.80 (m, 5.97 H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75 MHz, ppm):  $\delta = 168.7$ , 168.5, 142.6, 141.9, 141.4, 140.0, 129.3, 129.2, 126.4, 43.7, 41.8, 36.2, 35.3, 31.9, 30.7, 21.6, 21.5, 20.8, 20.2, 19.9, 19.7, 13.9, 13.8, 13.7; IR (liquid film, cm<sup>-1</sup>): v = 3331, 2960, 2932, 2873, 1651, 1556, 1334, 1267, 1142, 1089, 814, 695. HRMS calc. C<sub>15</sub>H<sub>24</sub>N<sub>2</sub>O<sub>2</sub>S (M<sup>+</sup>): 296.1558. Found: 296.1553.

## 3. NMR Spectra of all products.

























![](_page_27_Figure_1.jpeg)

![](_page_28_Figure_1.jpeg)

![](_page_29_Figure_1.jpeg)

![](_page_30_Figure_1.jpeg)

![](_page_31_Figure_1.jpeg)

![](_page_32_Figure_1.jpeg)

![](_page_33_Figure_1.jpeg)

![](_page_34_Figure_1.jpeg)

# 4. HPLC Spectra of all products.

![](_page_35_Figure_2.jpeg)

![](_page_35_Figure_3.jpeg)

![](_page_36_Figure_1.jpeg)

![](_page_37_Figure_1.jpeg)

![](_page_37_Figure_2.jpeg)

![](_page_38_Figure_1.jpeg)

![](_page_38_Figure_2.jpeg)

![](_page_39_Figure_1.jpeg)

![](_page_40_Figure_1.jpeg)

![](_page_40_Figure_2.jpeg)

![](_page_41_Figure_1.jpeg)

![](_page_42_Figure_1.jpeg)

![](_page_43_Figure_1.jpeg)

![](_page_43_Figure_2.jpeg)

![](_page_44_Figure_1.jpeg)