#### **Electronic Supporting Information for the Article:**

# A Graphene Oxide-based Sensing Platform for Lead (II) Ions

# Analysis with Tunable Dynamic Range by using Pb<sup>2+</sup>-dependant

## DNAzyme

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### **Experimental Section**

#### Materials

The DNA/RNA oligonucleotides were synthesized and purified by TaKaRa Biotechnology Co. (Dalian, China). The following oligonucleotide sequences were used:

17S: 5'- Cy3-ACTCACTAT*rA*GGAAGAGATGTCTGT-3' (The bold italic letter is the cleavage site).

## 17E: 5'-ACAGACATCTCT TCTCCG AGCCGGTCGAAATAGTGAGT-3'

## 17E<sub>C</sub>: 5'-ACAGACATCTCTTCCCCGAGCCGGTCGAAATAGTGA GT-3'

Pb(NO<sub>3</sub>)<sub>2</sub> was purchased from Aldrich and used as received. Cu(NO<sub>3</sub>)<sub>2</sub>, Zn(NO<sub>3</sub>)<sub>2</sub>, Cd(NO<sub>3</sub>)<sub>2</sub>, Co(NO<sub>3</sub>)<sub>2</sub>, Mn(CH<sub>3</sub>COO)<sub>2</sub>, Ni(NO<sub>3</sub>)<sub>2</sub> and Hg(NO<sub>3</sub>)<sub>2</sub> were of analytical grade and used as received. All solutions were prepared with Milli-Q water (18 M $\Omega$  cm<sup>-1</sup>) from a Millipore system. Environmental water sample was taken from a nearby river and centrifuged to remove the insoluble impurities.

## **Preparation of GO**

GO was synthesized from graphite powder based on the Hummer's method. Briefly, graphite power (4 g) was oxidized in a hot solution (80°C) of concentrated  $H_2SO_4(24 \text{ mL})$  containing  $K_2S_2O_8$  (8 g), and  $P_2O_5$  (8 g). The resulting dark blue mixture was

thermally isolated and slowly cooled to room temperature over a period of 6 h. The mixture was diluted to 300 mL, and then filtrated with a filter membrane of 0.22  $\mu$ m (Generay Biotech Co., Ltd., Shanghai, China) and dried overnight at 60°C. These preoxidized graphite powder (2 g) was added to 92 mL of cold H<sub>2</sub>SO<sub>4</sub> (0°C), to which KMnO<sub>4</sub> (12 g) was gradually added under continuous stirring in ice-bath. After 15 min, NaNO<sub>3</sub> (2 g) was added to the mixture. The solution was further stirred for 2 h at 35 °C and distilled water (200 mL) was added. The reaction was stopped with the addition of a mixture of 560 mL of distilled water and 10 mL of H<sub>2</sub>O<sub>2</sub> (30 %). The product was washed with HCl (1:10) and then with water, and then suspended in distilled water. The brown dispersion was extensively dialyzed to remove residual metal ions and acids, and then exfoliated via sonication for 1.5 h (300 W). Unexfoliated graphite oxide was removed by centrifugation (3000 rpm, 5 min) using Centrifuge himac-CF 16RX (Hitachi, Japan).

#### Fluorescence assay for Pb(II) Ions

Pb (II) ions of different concentrations were incubated in Tris-HCl buffer (50 mM, pH 7.4) containing 50 mM of NaCl and 20 nM of DNAzyme for 30 min at room temperature. Then 4  $\mu$ L of GO (2.5  $\mu$ g/ $\mu$ L) was added to this mixture and the fluorescence measurement was carried out 1 min after the GO addition.

#### Instruments

The fluorescence spectra were measured using a Hitachi F-4500 spectrophotometer equipped with a Xenon lamp excitation source. The excitation wavelength was  $\lambda$ =545 nm, and the fluorescence measurements were carried out at 23°C. The time-resolved fluorescence was measured by using the time-correlated single photon counting (TCSPC) technique. The experiments were conducted on a Fluorolog HORIBA JOBNYVON instrument with instrument response function (IRF) of 260 ps. A HORIBA JobinYvon pulsed laser diode at 488 nm was used as the excitation source for all samples.



**Figure S1**. a) AFM tapping-mode image of the as-prepared GO sheets and the height profile along the dashed line in panel. b) HR-TEM image of as-prepared GO.



**Figure S2**. Fluorescence quenching of Cy3 (30 nM) in the absence (black) and presence of a series of amounts of GO (top to bottom: 0.25, 0.50, 1.00, 1.25, 1.50, 1.75 and  $2.00 \mu g$ ).

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**Figure S3**. a) The fluorescence spectra of Cy3-tagged 17S upon incubation with different amounts of GO. b) The Stern–Volmer plots of the Cy3-labeled 17S quenched by GO. The  $K_{sv}$  was calculated as 0.63 L mg<sup>-1</sup> for Cy3.



**Figure S4.** The GO concentration-dependent fluorescence quenching of FITC (a), FAM (b) and ROX (c)-labeled ss-DNA (all of 20 nM). Insets are the Stern–Volmer

plots of these dye-labeled SS-DNA quenched by GO. The  $K_{sv}$  was calculated as 1.12 L mg<sup>-1</sup>, 0.97 L mg<sup>-1</sup> and 1.46 L mg<sup>-1</sup> for FITC, FAM and ROX, respectively.



**Figure S5.** Fluorescence spectra of the Cy3-labeled 17S strand upon incubation with different concentrations of  $Pb^{2+}$ .



Figure S6. Photoluminescence spectrum of GO.  $\lambda_{ex} = 545$  nm

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**Figure S7.** Fluorescence spectra of DNAzyme upon incubation with 10 nM  $Pb^{2+}$  for different time intervals at room temperature and then mixed with 10 µg GO. All other experimental conditions are identical to those in Scheme 1.



**Figure S8**. Optimization of the amount of GO addition. 100 nM of Pb<sup>2+</sup> of lead (II) was incubated with DNAzyme for 30 min, then different volume of GO ( $2.5\mu g/\mu L$ ) was added to the mixture. The fluorescence spectra were recorded after 30, 60 or 90s of the addition. The y-axis of this figure of the ratio of fluorescence intensity of background signal (F<sub>o</sub>) to that of 100 nM of Pb<sup>2+</sup> (F<sub>100</sub>).



Figure S9. Selectivity of the present assay towards  $Pb^{2+}$  ions over other 10 interference ions. The concentration of all the metal ions was 1  $\mu$ M.



**Figure S10.** Selectivity of the present assay towards  $Pb^{2+}$  ions (1  $\mu$ M)over Ca<sup>2+</sup> and Mg<sup>2+</sup> of higher concentrations (1 mM) since the concentration of Ca<sup>2+</sup> and Mg<sup>2+</sup> in real environmental samples are at mM range.

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**Figure S11.** Comparison of fluorescence responses of this GO-based sensor for different concentrations of  $Pb^{2+}$  dissolved in real environmental water samples.