

## Supporting Information

### Using Boronolectin in MALDI-MS Imaging for the Histological Analysis of Cancer Tissue Expressing the Sialyl Lewis X Antigen

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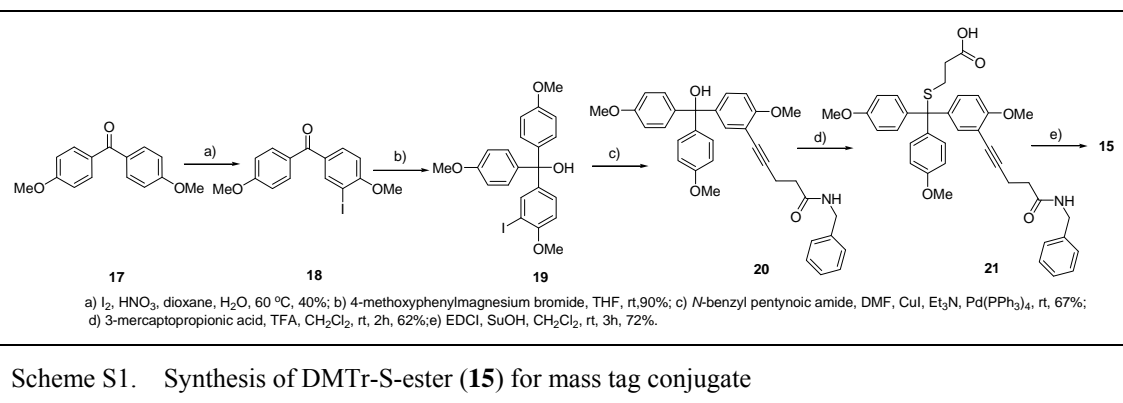
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#### General Information

Solvents and reagents were purchased from VWR, Acros, or Aldrich and used without purification unless specified otherwise. When necessary, solid reagents were dried under high vacuum. Reactions with compounds sensitive to air or moisture were performed under argon. Solvent mixtures are indicated as volume/volume ratios. Thin layer chromatography (TLC) was run on Sorbtech W/UV254 plates (0.25 mm thick), and visualized under UV-light or by a Ce-Mo staining solution (phosphomolybdate, 25 g; Ce(SO<sub>4</sub>)<sub>2</sub>·4H<sub>2</sub>O, 10 g; H<sub>2</sub>SO<sub>4</sub>, 60 mL, conc.; H<sub>2</sub>O, 940 mL) with heating. Flash chromatography was performed using Fluka silica gel 60 (mesh size 0.040-0.063 mm) using a weight ratio of ca. 30:1 for silica gel over crude compound. <sup>1</sup>H, <sup>13</sup>C NMR spectra were recorded on a Bruker 400 MHz NMR spectrometer in deuterated chloroform (CDCl<sub>3</sub>), methanol-*d*<sub>4</sub> (CD<sub>3</sub>OD), or DMSO-*d*<sub>6</sub> with either tetramethylsilane (TMS) (0.00 ppm) or the NMR solvent as the internal reference.

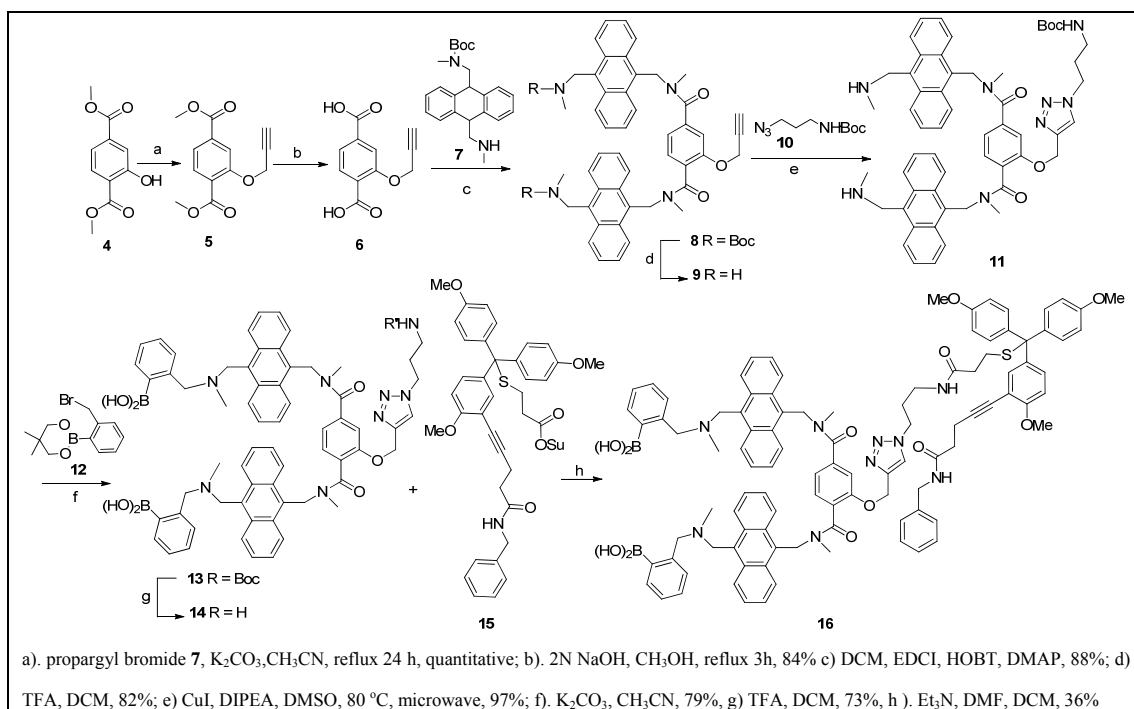
## Experimental Section



### 1. Synthesis of DMTr-S-ester (**15**) for mass tag conjugation

As shown in Scheme S1, iodination of diarylketone compound **17** afforded monoiodo compound **18**. Subsequent Grignard addition gave trityl compound **19**. A side arm was installed through Sonogashira reaction to give **20**, which was reacted with 3-mercaptopropionic acid in the presence of trifluoroacetic acid (TFA) to give DMTr-S-acid (**21**). The carboxyl group was then activated through conversion to its *N*-hydroxysuccinimide ester (DMTr-S-ester, **15**).

### 2. Synthesis of boronolactin-DMTr mass tag conjugate



By reacting **4** with propargyl bromide, the alkyne side chain was introduced to give **5** in quantitative yield. The subsequent hydrolysis of **5** under basic conditions gave diacid linker **6** with an alkyne handle in 84% yield. Compound **7** was coupled with diacid linker **6** using 1-(2-dimethylamino-propyl)-3-ethyl carbodiimide hydrochloride (EDCI) as the activating reagent to furnish compound **8** with a di-anthracene group and an alkyne handle in 88% yield. Deprotection of **8** with TFA in dichloromethane (DCM) gave **9**, which was reacted with azido compound **10** under microwave conditions to give triazole compound **11**. The boronic acid moiety was attached through alkylation using potassium carbonate as the base to give the bisboronic acid compound **13**. Deprotection of the Boc group with TFA in DCM gave free amine **14**, which was then conjugated with DMTr-S-ester **15** to give the final mass spectrometric tag conjugate **16**.

## Synthesis

### 3-Iodo-4,4'-dimethoxybenzophenone (**18**)

To a solution of 4,4'-dimethoxybenzophenone (**17**) (1.8 g, 7.5 mmol) in dioxane (10 mL), iodine (1.0 g, 3.9 mmol) was added at 60 °C. After the mixture was stirred for 15 min, water (2 mL) was added followed by the addition of 58% HNO<sub>3</sub> (4.1 mL) in a drop wise fashion within 1 h. The reaction was kept stirring at 60 °C for about 6 h until iodine color disappeared. Nitrogen was bubbled through the reaction mixture until the cessation of brown gas emission. Then the mixture was diluted with water (100 mL) to afford pale yellow precipitates, which were collected and washed with 5% NaHCO<sub>3</sub> aqueous solution and water. Re-crystallization from EtOH (20 mL), followed by column chromatography purification with hexane/EtOAc (10:1) gave white needles (1.1 g, 40% yield). <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 8.11 (s, 1H), 7.73 (m, 3H), 7.12 (m, 3H), 3.95 (s, 3H), and 3.87 (s, 3H) ppm. MS-ESI: [M+H]<sup>+</sup> (m/z, 369.0).

### 3-Iodo-4-methoxyphenyl-bis(4-methoxyphenyl)methanol (**19**)

To a solution of 3-iodo-4,4'-dimethoxybenzophenone (**18**) (1.0 g, 2.7 mmol) in dry THF (20 mL), 1M solution of 4-methoxyphenylmagnesium bromide in THF (5.5

ml, 5.5 mmol) was added in one portion at 0 °C. After stirring at RT overnight, the reaction solution was diluted with 150 mL of saturated NaHCO<sub>3</sub> solution and extracted with EtOAc (50 mL × 3). The EtOAc layer was washed with water and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography with hexane/EtOAc (10:1) to give white crystals (1.2 g, 90% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>): 7.77 (s, 1H), 7.16 (m, 5H), 6.87 (m, 4H), 6.74 (d, *J* = 5.6 Hz 1H), 3.89 (s, 3H), and 3.82 (s, 6H) ppm. MS-ESI: [M+H]<sup>+</sup> (*m/z*, 477.3).

***N*-Benzyl-3-{2-methoxy-5-[hydroxyl-bis(4-methoxyphenyl)methyl]-phenyl}-4-pentynoic amide (20)**

To the solution of compound **19** (530 mg, 1.11 mmol) and *N*-benzyl pentynoic amide (219 mg, 1.17 mmol) in DMF (6 mL), Pd(PPh<sub>3</sub>)<sub>4</sub> (116 mg, 0.1 mmol), CuI (38 mg, 0.2 mmol), and Et<sub>3</sub>N (235 μL, 1.68 mmol) were added in sequence under N<sub>2</sub> protection. The reaction mixture was stirred at room temperature overnight, then diluted with water (100 mL) and extracted with EtOAc (60 mL × 3). The EtOAc extract was washed with water (50 mL), 0.1 M solution of (NH<sub>4</sub>)<sub>2</sub>EDTA (50 mL), and brine (50 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified on a silica gel column with hexane/EtOAc 1:1 as the solvent to give a white solid (400 mg, 67% yield). <sup>1</sup>H NMR (CD<sub>3</sub>OD): 7.24 (dd, *J* = 3.6, 0.8 Hz, 2H), 7.14 (m, 5H), 7.08 (m, 4H), 6.97 (dd, *J* = 6.8, 2.0 Hz, 1H), 6.81 (m, 4H), 4.34 (s, 2H), 3.79 (s, 3H), 3.76 (s, 6H), 2.72 (t, *J* = 6.8 Hz, 2H), and 2.48 (t, *J* = 6.8 Hz, 2H) ppm. MS-ESI: [M+H]<sup>+</sup> (*m/z*, 536.4).

**DMTr-S-acid (21)**

To a solution of compound **20** (340 mg, 0.64 mmol) in 6 mL of DCM under N<sub>2</sub> were added 3-mercaptopropionic acid (56 μL, 0.64mmol) and TFA (67 μL, 0.83 mmol). The mixture was stirred at RT for 2 h. At that point, TLC indicated that compound **20** had disappeared. The volatiles in the reaction solution were removed under vacuum. The residue was purified by silica gel chromatography with DCM/MeOH (10:1) to give a colorless needle-like product (248 mg, 62% yield). <sup>1</sup>H

NMR (CDCl<sub>3</sub>): 7.64 (d,  $J$  = 2.2 Hz, 1H), 7.27-7.11 (m, 10H), 6.78 (d,  $J$  = 8.8 Hz, 4H), 6.71 (d,  $J$  = 8.8 Hz, 1H), 6.50 (brs, 1H), 4.45 (d,  $J$  = 5.6 Hz, 2H), 3.77 (s, 6H), 3.72 (s, 3H), 2.77 (t,  $J$  = 6.8 Hz, 2H), 2.52 (t,  $J$  = 6.6 Hz, 2H), 2.40 (t,  $J$  = 6.8 Hz, 2H), 2.29 (t,  $J$  = 6.8 Hz, 2H) ppm. MS-ESI:  $[M+H]^+$  ( $m/z$ , 624.0).

#### DMTr-S-ester (15)

Compound **21** (248 mg, 0.4 mmol) and *N*-hydroxysuccinimide (SuOH, 46 mg, 0.4 mmol) were dissolved in 8 mL of DCM. Then EDCI (77 mg, 0.4 mmol) was added at RT with stirring. After stirring at RT for 3 h, the reaction solution was diluted with DCM (60 mL), washed with water (5 mL) and brine (5 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography with DCM/MeOH (20:1) to give white solid (204 mg, 72% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  7.43 (d,  $J$  = 2.5 Hz, 1H), 7.26 (m, 5H), 7.22 (m, 5H), 6.82 (m, 4H), 6.46 (brs, 1H), 3.80 (s, 6H), 3.74 (s, 3H), 2.78 (m, 6H), 2.53 (m, 4H), 2.47 (m, 2H) ppm. MS-ESI:  $[M+H]^+$  ( $m/z$ , 721.2).

#### 2-Prop-2-ynyloxy-terephthalic acid dimethyl ester (5)

To a solution of compound **4** (24 g, 114 mmol) in 250 mL of CH<sub>3</sub>CN was added K<sub>2</sub>CO<sub>3</sub> (18.9 g, 137 mmol) and propargyl bromide (16.38 mL, 137 mmol). The reaction mixture was refluxed for 24 h under nitrogen. After cooling down to RT, reaction mixture was then poured into a mixture of EtOAc (100 mL) and 10 % HCl aqueous solution (10 mL). The organic phase was separated and washed with saturated NaHCO<sub>3</sub>, brine and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Removal of solvent to provide a wax-like light white solid (28.25 g, quantitative yield): <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.84-7.82 (m, 1H), 7.71-7.69 (m, 1H), 7.782-7.779 (m, 1H), 4.85 (d,  $J$  = 2.0 Hz, 2H), 3.95 (s, 3H), 3.92 (s, 3H), 2.56 (t,  $J$  = 2.0 Hz, 1H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  166.0, 165.9, 156.6, 134.3, 131.6, 125.2, 122.3, 115.1, 77.6, 76.5, 56.9, 52.5, 52.4. MS (+ESI)  $m/z$  249.1  $[M+H]^+$ .

#### 2-Prop-2-ynyloxy-terephthalic acid (6)

To a solution of compound **5** (3.0 g, 143 mmol) in 15 mL of CH<sub>3</sub>OH was added 25 mL of sodium hydroxide solution (2M), the reaction mixture was refluxed for 3 h.

After removal of solvent by vacuum, the residue was acidified to pH 2 with 10% HCl solution. Solid was collected, washed with water, dried on vacuum to afford a white solid (2.11 g, 84%):  $^1\text{H}$  NMR ( $\text{DMSO-}d_6$ )  $\delta$ 13.22 (brs, 2H), 7.71-7.73 (m, 2 H), 7.61-7.63 (m, 1H), 4.96 (d,  $J=2$  Hz, 3H), 3.65 (s, 1H);  $^{13}\text{C}$  NMR ( $\text{DMSO-}d_6$ )  $\delta$ 167.3, 167.0, 155.8, 134.7, 130.9, 126.9, 122.2, 114.7, 79.4, 79.2, 56.6. MS (-ESI)  $m/z$  219.1  $[\text{M-H}]^-$ .

***N*<sup>1</sup>, *N*<sup>4</sup>-Bis((9-(*N*-Boc-methylamino)methyl)anthracen-10-yl)methyl)-*N*<sup>1</sup>, *N*<sup>4</sup>-dimethyl-2-(prop-2-ynyloxy)terephthalamide (8)**

To a solution of **7** (1.62 g, 4.4 mmol) and **6** (488 mg, 2.2 mmol) in 320 mL of dried DCM was added EDCI (820 mg, 9.0 mmol), HOBt (1.2 g, 8.9 mmol) and DMAP (108 mg, 0.9 mmol). After stirring overnight at room temperature under nitrogen atmosphere, the reaction mixture was washed with water and dried over  $\text{MgSO}_4$ . After removal of the solvent, the residue was purified by silica gel chromatography (DCM/ $\text{CH}_3\text{OH}$  100/1) to afford a light yellow powder (1.76 g, 88%):  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$ 8.50 (s, 8 H), 7.58 (s, 8H), 7.28 (s, 1H), 7.12 (s, 1H), 7.02 (s, 1H), 5.86 (s, 4H), 5.55 (s, 4H), 4.55 (s, 2H), 2.54 (s, 3H), 2.51 (s, 6H), 2.32 (s, 3H), 1.57 (s, 18H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$ 170.3, 168.0, 153.2, 138.1, 130.9, 130.8, 130.1, 128.4, 128.1, 128.04, 126.2, 126.0, 125.9, 125.0, 124.5, 124.2, 120.2, 111.4, 79.8, 77.5, 76.1, 55.8, 53.3, 41.4, 38.5, 35.5, 33.9, 28.4. HRMS (+ESI): Calc. for  $[\text{C}_{57}\text{H}_{61}\text{N}_4\text{O}_7]^+ [\text{M+H}]^+$   $m/z$  913.4540, Found 913.4566. MS (+ESI)  $m/z$ : 913.4  $[\text{M+H}]^+$ .

***N*<sup>1</sup>, *N*<sup>4</sup>-Dimethyl-*N*<sup>1</sup>, *N*<sup>4</sup>-bis((9-((methylamino)methyl)anthracen-10-yl)methyl)-2-(prop-2-ynyloxy)terephthalamide (9)**

A mixture of compound **8** (1.0 g, 1.1 mmol) and TFA (3 mL) in 12 mL of DCM was stirred at room temperature in the dark for 3 h. After removal of solvent, a mixed solvent of EtOAc/hexane 1:1 (20 mL) was added to the residue. Precipitate was generated and the solid was collected, washed with saturated  $\text{NaHCO}_3$  solution and water, dried under vacuum to provide **9** as a light yellow solid (615 mg, 82%):  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  8.43-8.39 (m, 8H), 7.57-7.52 (m, 8H), 7.29-7.26 (m, 1H), 7.09 (s, 1 H), 6.99-6.97 (m, 1H), 5.83-5.78 (m, 4H), 4.69 (s, 4H), 4.53 (s, 2H), 2.69 (s, 6H),

2.51 (s, 3H), 2.39 (s, 3H), 2.33 (s, 1H), 1.94 (s, 2H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 168.6, 153.7, 138.6, 133.5, 131.6, 131.5, 130.5, 128.6, 128.0, 126.7, 126.5, 126.3, 125.5, 125.4, 125.4, 125.0, 124.7, 120.7, 111.9, 78.1, 76.8, 56.4, 48.5, 42.4, 41.9, 37.6, 36.1, 34.5. HRMS (+ESI): Calc. for  $[\text{C}_{47}\text{H}_{45}\text{N}_4\text{O}_3]^+ [\text{M}+\text{H}]^+$   $m/z$  713.3492, Found 713.3512. MS (-ESI)  $m/z$  711.4  $[\text{M}-\text{H}]^-$ .

### Compound 11

To a mixture of compound **9** (105 mg, 0.14 mmol) and compound **10** (84 mg, 0.42 mmol) in 0.5 mL of DMSO was added DIPEA (0.12 mL, 0.7 mmol) and CuI (11 mg, 0.056 mmol). The reaction mixture was microwave-irradiated at 80 °C for 30 min under nitrogen atmosphere. To the reaction mixture, water was slowly added in (5 mL). Then the mixture was extracted with DCM. The combined DCM phase was washed with brine and dried over  $\text{MgSO}_4$ . Solvent was removed under vacuum and the residue was purified by silica gel chromatography (DCM/ $\text{CH}_3\text{OH}$  10/3) to provide a light yellow solid (130 mg, 97%).  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  8.44-8.43 (m, 8H), 7.58-7.53 (m, 9H), 7.08-6.96 (m, 3H), 5.87-5.69 (m, 4H), 5.10 (s, 2H), 4.76 (s, 4H), 4.20 (s, 2H), 2.94 (s, 2H), 2.84 (s, 6H), 2.40 (s, 6H), 1.78 (s, 2H), 1.44 (s, 9H); MS (ESI)  $m/z$  913.4  $[\text{M}+\text{H}]^+$

### Compound 13

To a solution of compound **11** (200 mg, 0.11 mmol) in 12 mL of  $\text{CH}_3\text{CN}$  were added boronate **16** (124 mg, 0.44 mmol),  $\text{K}_2\text{CO}_3$  (152 mg, 1.1 mmol), and NaI (4 mg, 0.022 mmol). The reaction mixture was stirred at room temperature for 16 h under nitrogen atmosphere in the dark. After filtering out the solid, the organic solvent was removed, and the residue was re-dissolved in DCM, washed with 5%  $\text{NaHCO}_3$  solution and brine, and dried over  $\text{MgSO}_4$ . Solvent evaporation gave a crude product, which was re-crystallized with DCM/ $\text{Et}_2\text{O}$  to provide a light yellow solid (205 mg, 79% yield).  $^1\text{H}$  NMR ( $\text{DMSO}-d_6$ )  $\delta$  6.80-9.15 (m, 28 H), 5.60-5.85 (m, 4 H), 5.15 (s, 2H), 4.53 (s, 4H), 4.25 (s, 2H), 3.95 (s, 4H), 3.31 (s, 6H), 2.46 (s, 6H), 2.31 (s, 2H), 1.34 (s, 9H). MS (ESI):  $m/z$  1163.3  $[\text{M}-\text{H}_2\text{O}+\text{H}]^+$ . HRMS (ESI): Calc. for  $[\text{C}_{69}\text{H}_{73}\text{B}_2\text{N}_8\text{O}_8]^+ [\text{M}-\text{H}_2\text{O}+\text{H}]^+$   $m/z$  1163.5737, Found 1163.5760.

### Compound 14

A mixture of compound **13** (240 mg, 0.2 mmol) and TFA (1.0 mL) in 10 mL of DCM was stirred for 4 h at room temperature under nitrogen atmosphere in the dark. After removal of solvent, the residual oil was dissolved in 3 mL of EtOAc. This was followed by the slow addition of 50 mL of Et<sub>2</sub>O. The precipitate was collected and washed with saturated K<sub>2</sub>CO<sub>3</sub> solution and water. Further purification by flash chromatography provided a white solid (133 mg, 73%): <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>) δ 6.80-9.40 (m, 30 H), 4.40-6.00 (m, 12 H), 3.40 (s, 6 H), 3.17-3.10 (m, 2 H), 2.82-2.74 (m, 2 H), 2.39 (s, 6 H). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>) δ: 158.9, 158.6, 136.2, 130.9, 130.6, 127.6, 127.0, 126.8, 125.5, 125.0, 119.0, 116.0, 67.8, 51.3, 47.1, 36.8, 28.3, 21.9, 21.7. MS (ESI) *m/z* 1063.4 [M-H<sub>2</sub>O+H]<sup>+</sup>, 1081.5 [M+H]<sup>+</sup>. HRMS (+ESI): Calc. for [C<sub>64</sub>H<sub>67</sub>B<sub>2</sub>N<sub>8</sub>O<sub>7</sub>]<sup>+</sup> [M+H]<sup>+</sup> *m/z* 1081.5319, Found 1081.5363; Calc. for [C<sub>64</sub>H<sub>65</sub>B<sub>2</sub>N<sub>8</sub>O<sub>6</sub>]<sup>+</sup> [M-H<sub>2</sub>O+H]<sup>+</sup> *m/z* 1063.5213, Found 1063.5253.

### Compound 16

In a 10-mL flask, compound **14** (60 mg, 0.05 mmol) and DMTr-S-ester (**15**) were dissolved in a mixture of 0.5 mL DMF and 1 mL DCM. Then Et<sub>3</sub>N was added at RT in the dark. The mixture was stirred at RT overnight and then solvent was evaporated. To the resulting viscous residue was slowly added 10 mL of Et<sub>2</sub>O to afford a suspension. The solid was filtered and washed with EtOAc to give crude product, which was purified by silica gel chromatography with DCM/MeOH (15:1) to give a white solid (30 mg, 36%). <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>) 9.0 (m, 3H), 8.64-8.41 (m, 9H), 7.94-6.83 (m, 32H), 5.76-5.10 (m, 5H), 4.53-4.32 (m, 2H), 4.42 (s, 3H), 3.76 (s, 3H), 3.69 (s, 6H), 3.40 (s, 12H), 3.39-3.01 (m, 18H), 2.55-2.0 (m, 8H), 1.23 (m, 2H). MS-ESI: [M-H-H<sub>2</sub>O]<sup>-</sup> (*m/z*, 1666.6) and [M+H-H<sub>2</sub>O]<sup>+</sup> (*m/z*, 1669.2).

### Tissue preparation and MALDI-IMS

Frozen tissue was sectioned and slides stored at -80 °C. For data collection, slides were removed from the freezer and rinsed in PBS for 5 min, followed by a rinse in water. The slides were then air-dried and overlaid with the boronolactin-trityl conjugate solution diluted in 100% methanol and incubated at room temperature for 2



h. Slides were then rinsed with 100% methanol to remove any unbound boronolactin-trityl conjugate, washed in PBS for 5 min, followed by a final water rinse. The slides were then allowed to air-dry and dessicated for 1 h before reading in the Ultraflex III MALDI-TOF (Bruker Daltonics), operated in reflectron mode with a raster width of 200  $\mu\text{m}$ , to detect the trityl tags ( $m/z$  500-600).

#### **Immunostaining for Sialyl Lewis X**

Immunostaining of frozen specimens was performed by the avidin-biotin peroxidase complex method using a Vectastain Elite ABC kit (Vector, Burlingame, CA). Frozen tissue was first treated with 0.3% hydrogen peroxide to block endogenous peroxidase activity for 15 minutes. Sections were incubated in normal serum (provided in the kit) to block nonspecific binding and incubated with mouse monoclonal antibody to sialyl Lewis X, (clone KM93, Millipore, Billerica, MA) diluted 1:50 in 5% serum diluted in PBS for 1 h at room temperature. After washing in PSA, sections were then treated with biotinylated goat anti-mouse immunoglobulin G (IgG), followed by treatment with avidin-biotin-peroxidase complex, and stained with IMPACT DAB peroxidase substrate (Vector Labs) according to the supplier's protocol. Counterstaining was performed with Mayer's hematoxylin.

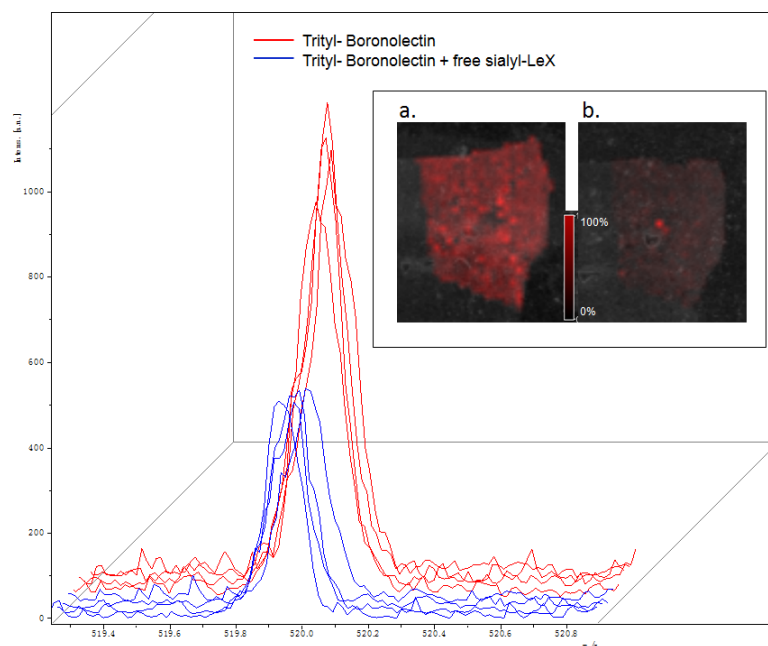
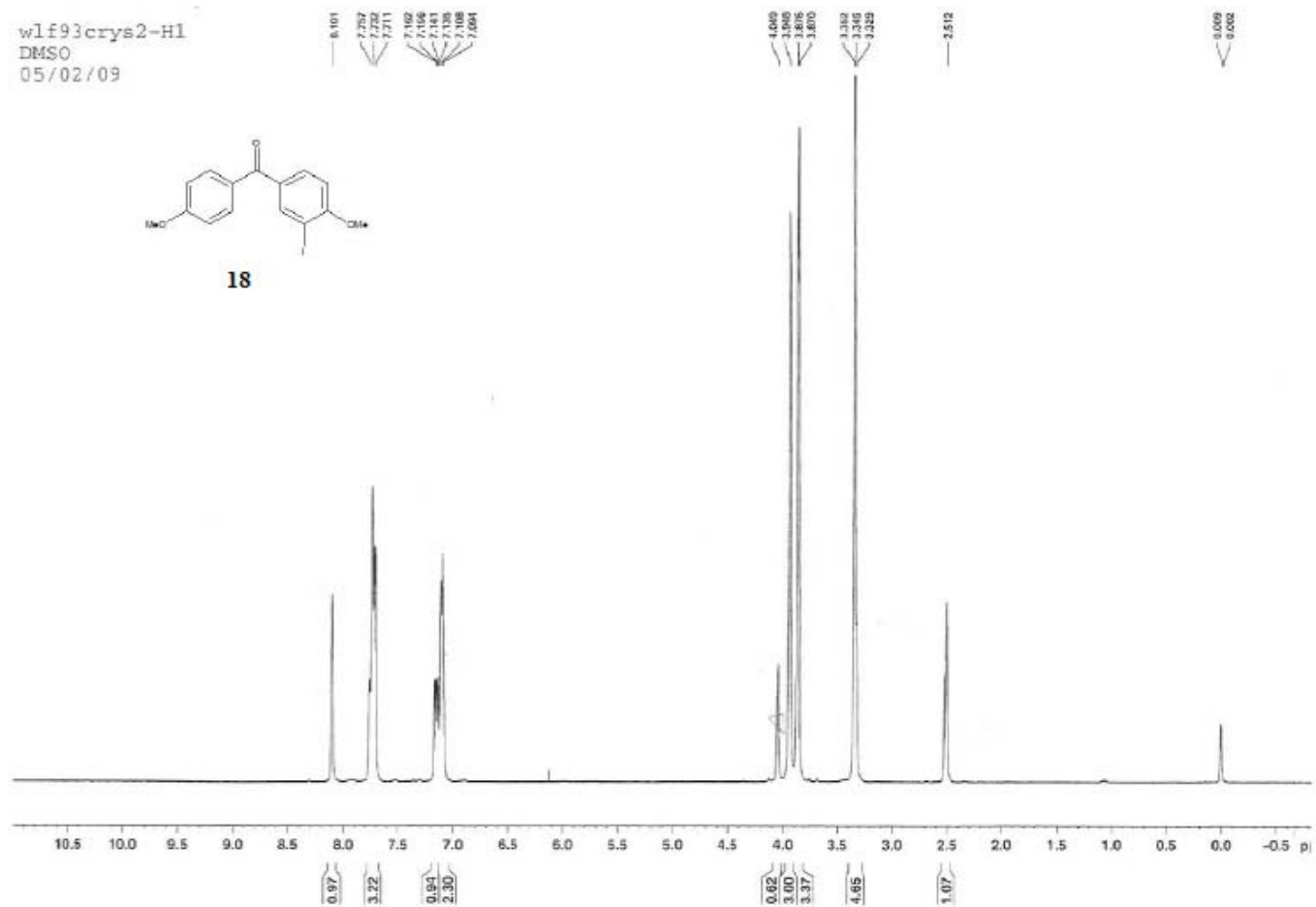
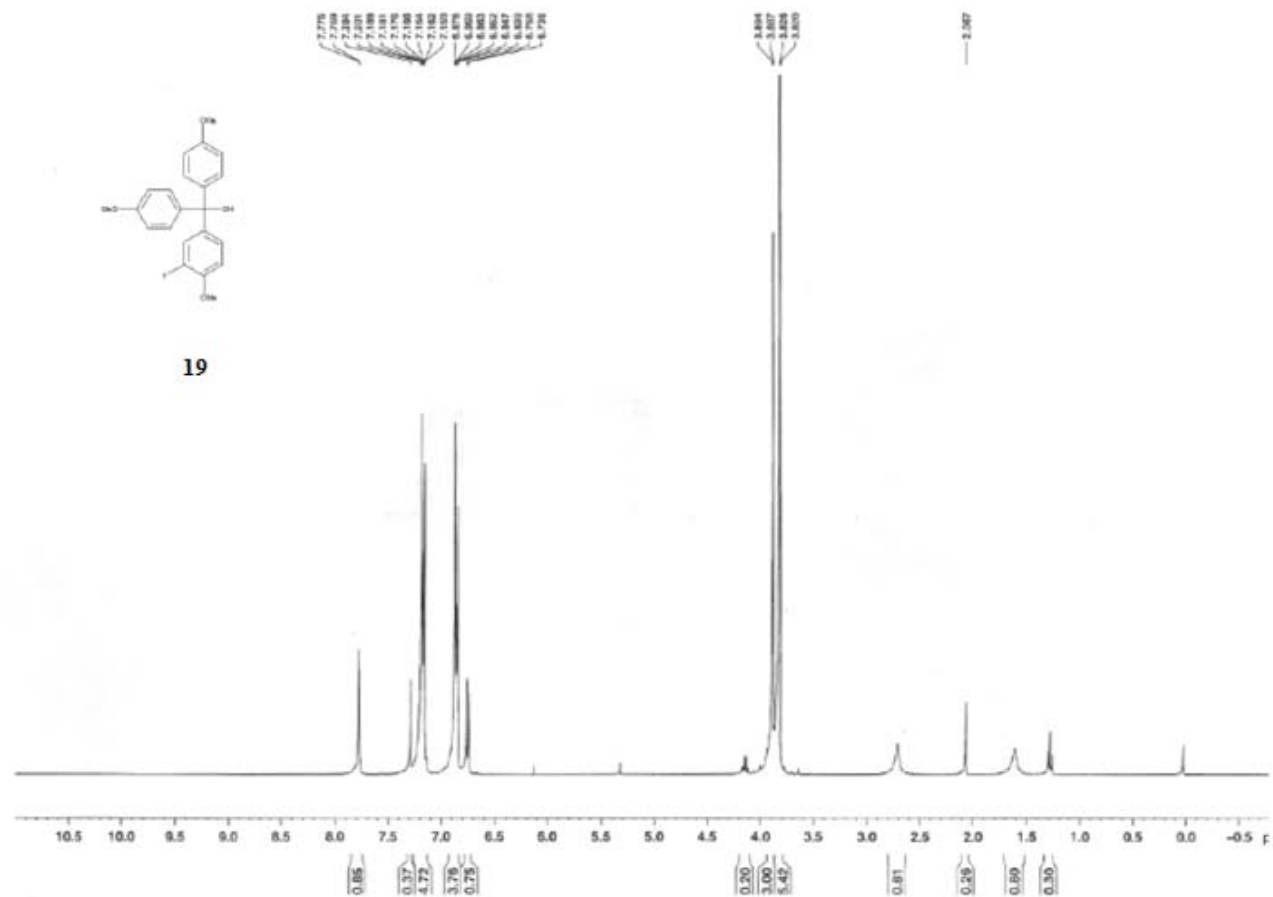
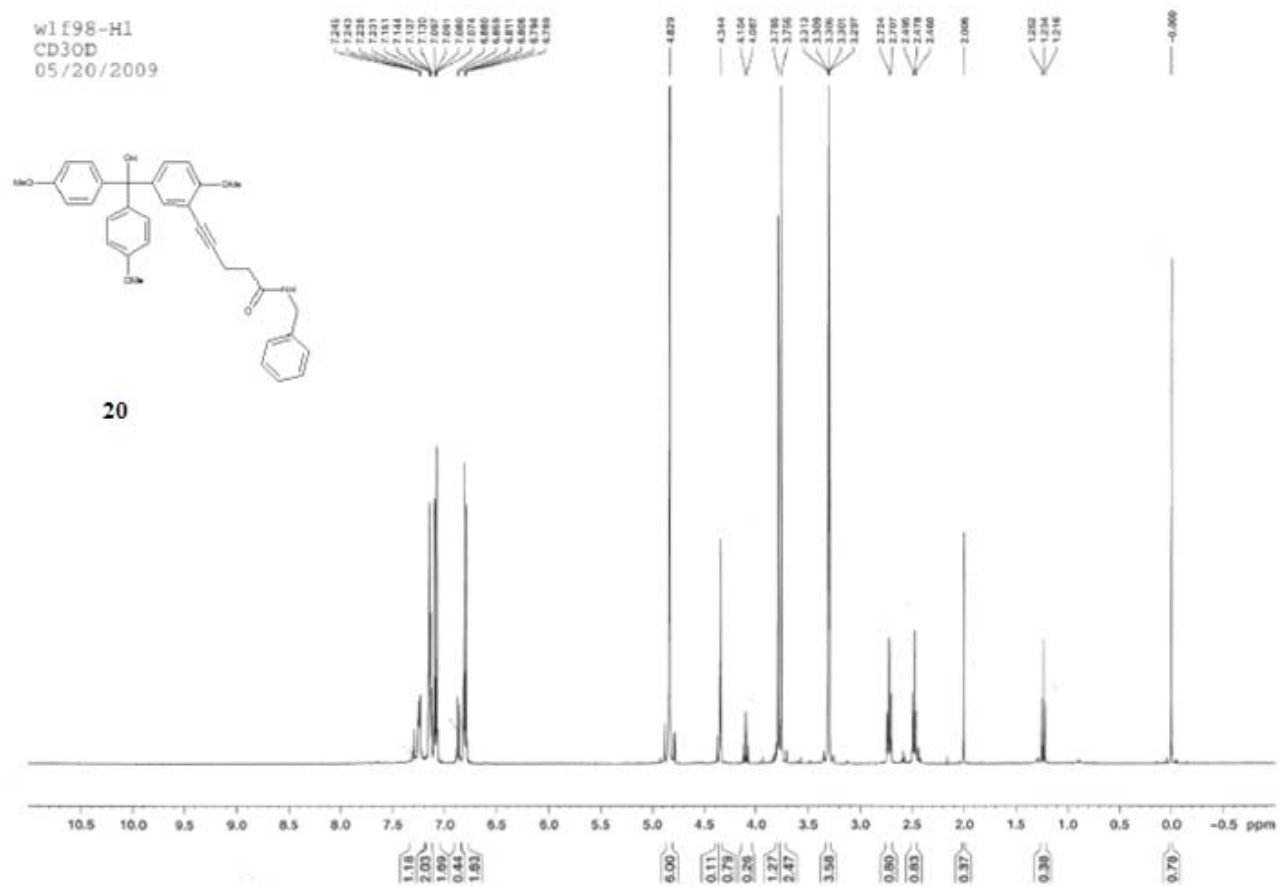


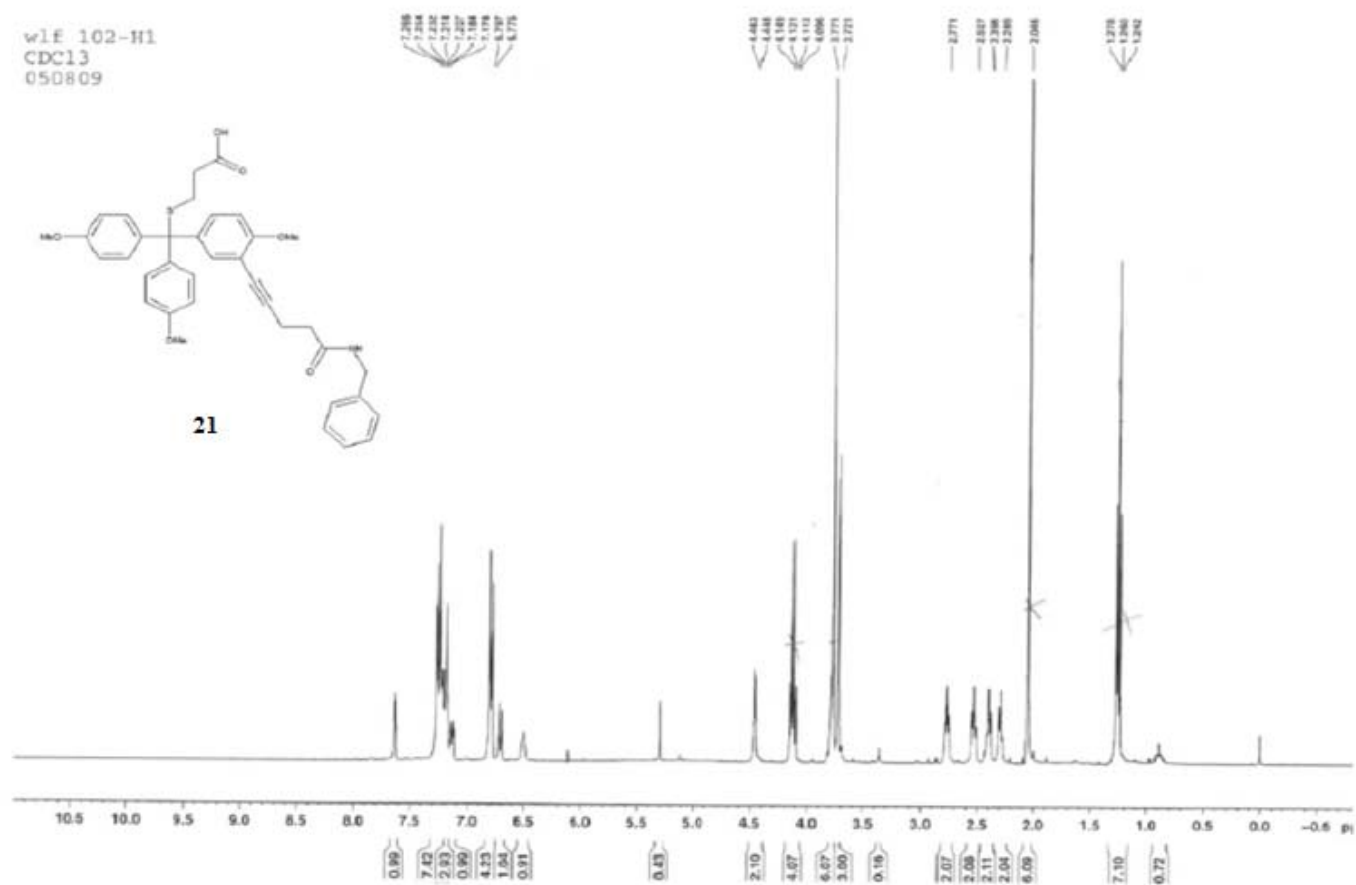
Figure S1. Borono-lectin trityl is blocked by carbohydrate.

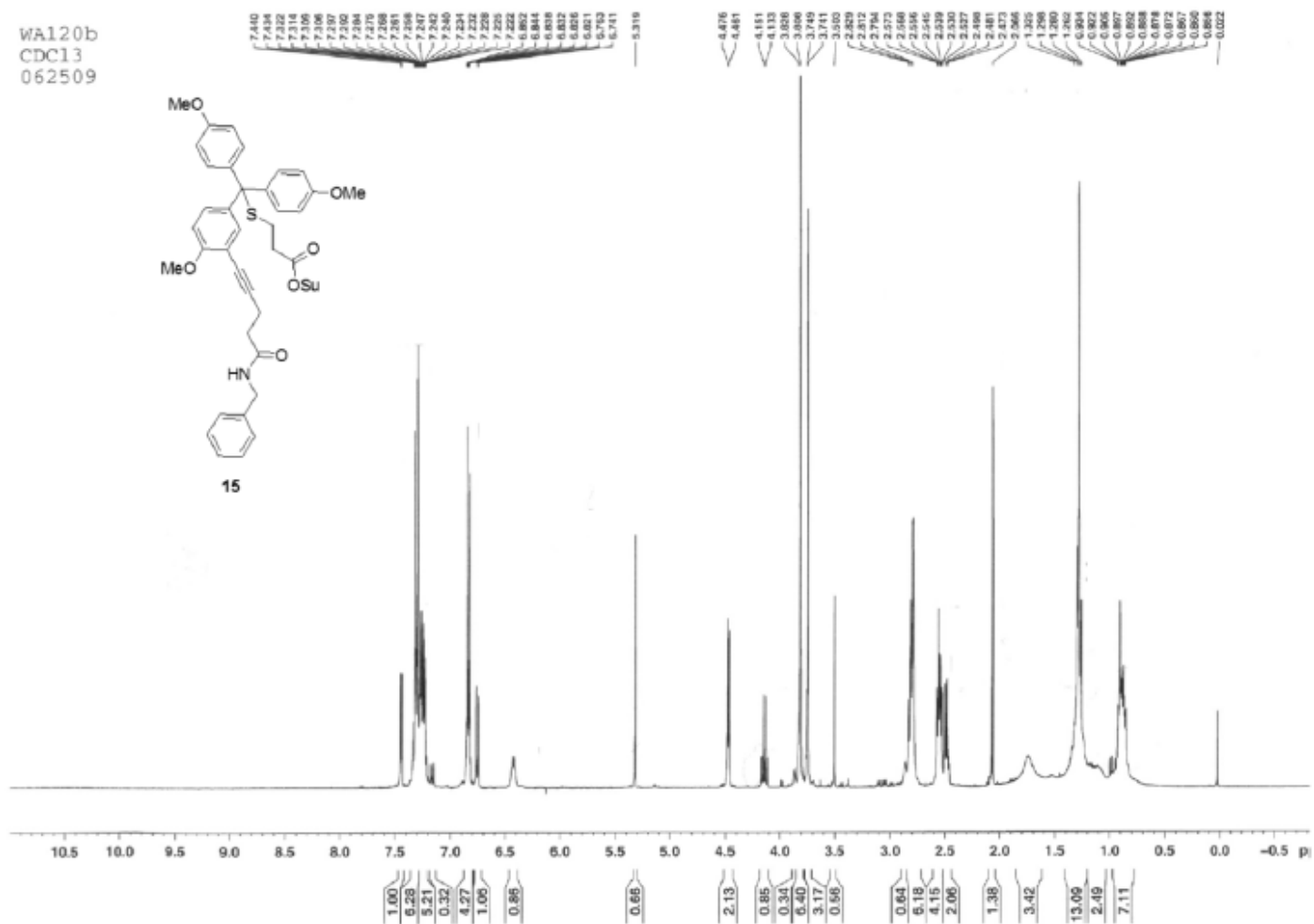
Representative spectra exported from RCC regions after incubation overnight in the presence (blue) or absence (red) of 7  $\mu\text{M}$  of sialyl Lewis X, followed by trityl-boronlectin-sialyl-LeX (1.5  $\mu\text{M}$ ) labeling and TIMS showing reduction in peak intensity with the carbohydrate was present. Inset) Images generated from RCC issue subjected to TIMS with the boronlectin-trityl alone (a) and with sialyl Lewis X (b), depicting the differences in signal intensity.











No title

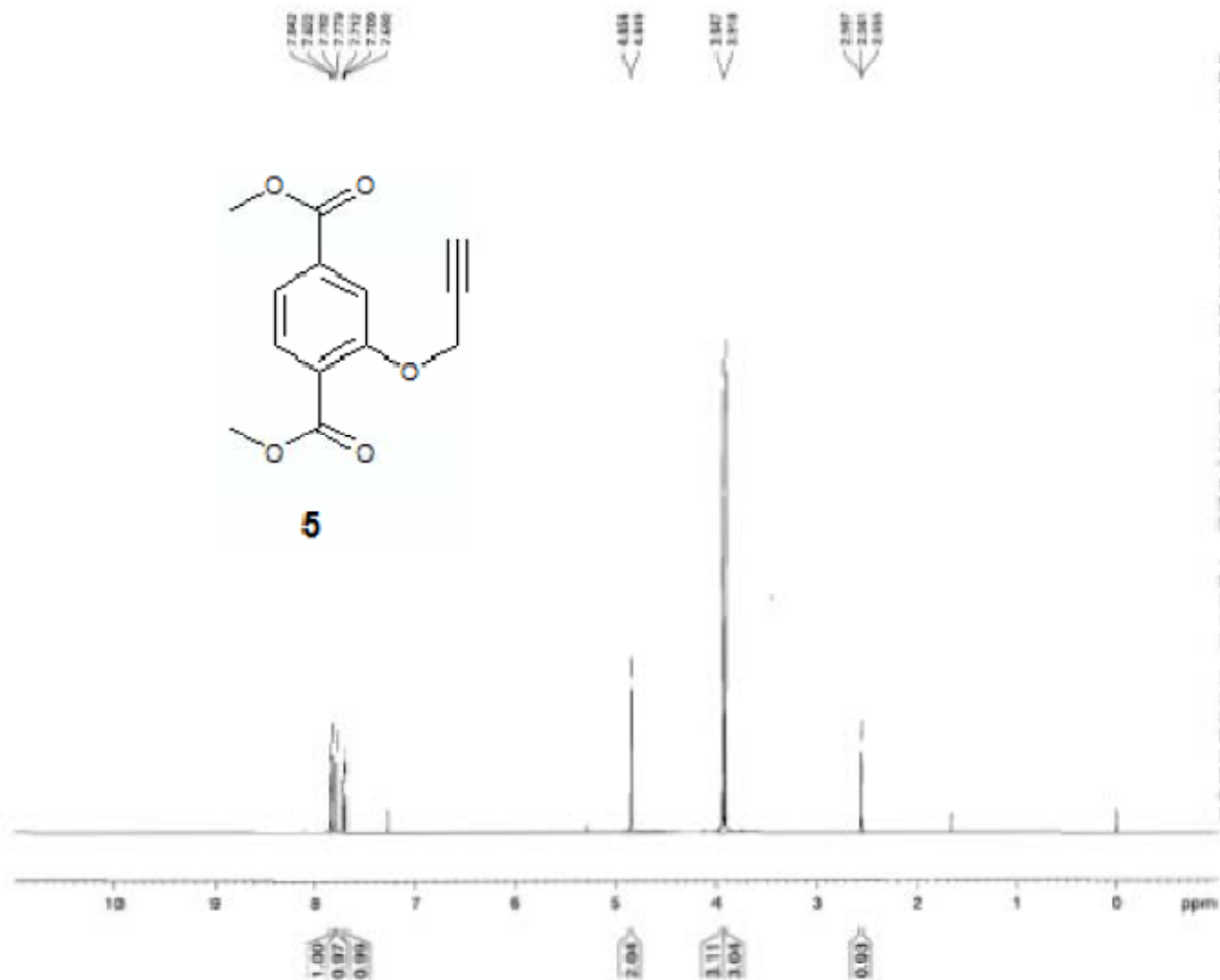


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DE: 7.00 usec  
TE: 298.2 K  
D1: 1.00000000 sec  
MCHST: 0.00000000 sec  
MCHNK: 0.01500000 sec

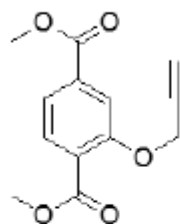
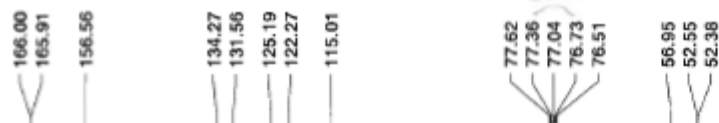
----- CHANNEL f1 -----  
NUC1: 1H  
P1: 12.80 usec  
PL1: 0.00 dB  
SFO1: 400.1324710 MHz

F2 - Processing parameters  
SI: 32768  
SF: 400.1300046 MHz  
WDW: EM  
SSB: 0  
LB: 0.30 Hz  
GB: 0  
PC: 1.40

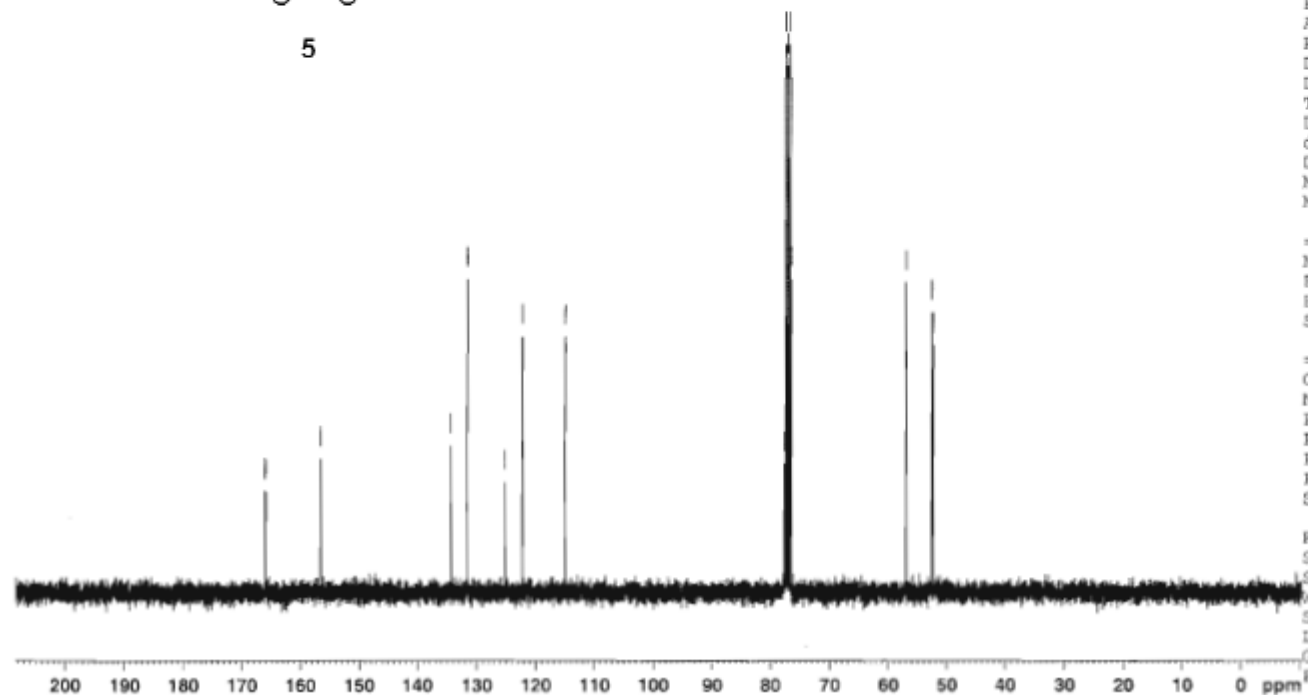




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5



Current Data Parameters  
NAME IV153  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20070221  
Time 17.57  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT cdcl3  
NS 94  
DS 4  
SWH 23980.814 Hz  
FIDRES 0.365918 Hz  
AQ 1.3664756 sec  
RG 32768  
DW 20.850 usec  
DE 7.00 usec  
TE 298.5 K  
D1 2.00000000 sec  
d11 0.03000000 sec  
DELTA 1.89999998 sec  
MCREST 0.00000000 sec  
MCWRK 0.01500000 sec

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.00 usec  
PL1 -3.00 dB  
SFO1 100.6226298 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 70.00 usec  
PL2 -1.00 dB  
PL12 14.00 dB  
PL13 14.00 dB  
SFO2 400.1316005 MHz

F2 - Processing parameters  
SI 32768  
SF 100.6127690 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.00

No title

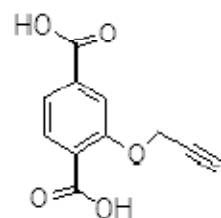


Current Data Parameters  
NAME CY14-012A01-H  
EXPNO 1  
PROCNO 1

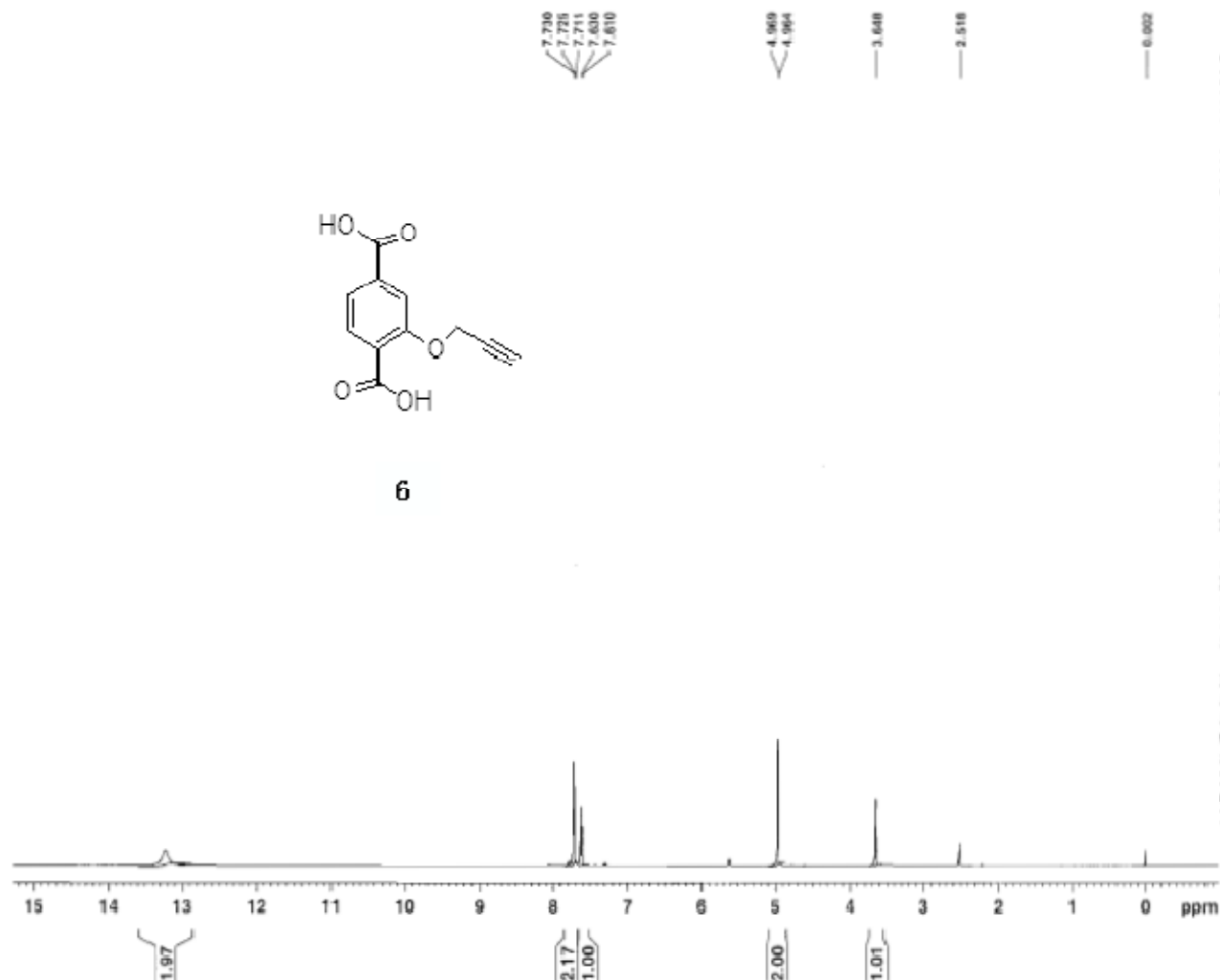
F2 - Acquisition Parameters  
Date\_ 20080702  
Time 15.58  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 16  
DS 2  
SWH 8278.146 Hz  
FIDRES 0.126314 Hz  
AQ 3.9584243 sec  
RG 161.3  
DW 60.400 usec  
DE 7.00 usec  
TE 297.6 K  
D1 1.00000000 sec  
MCREST 0.00000000 sec  
MCWRK 0.01500000 sec

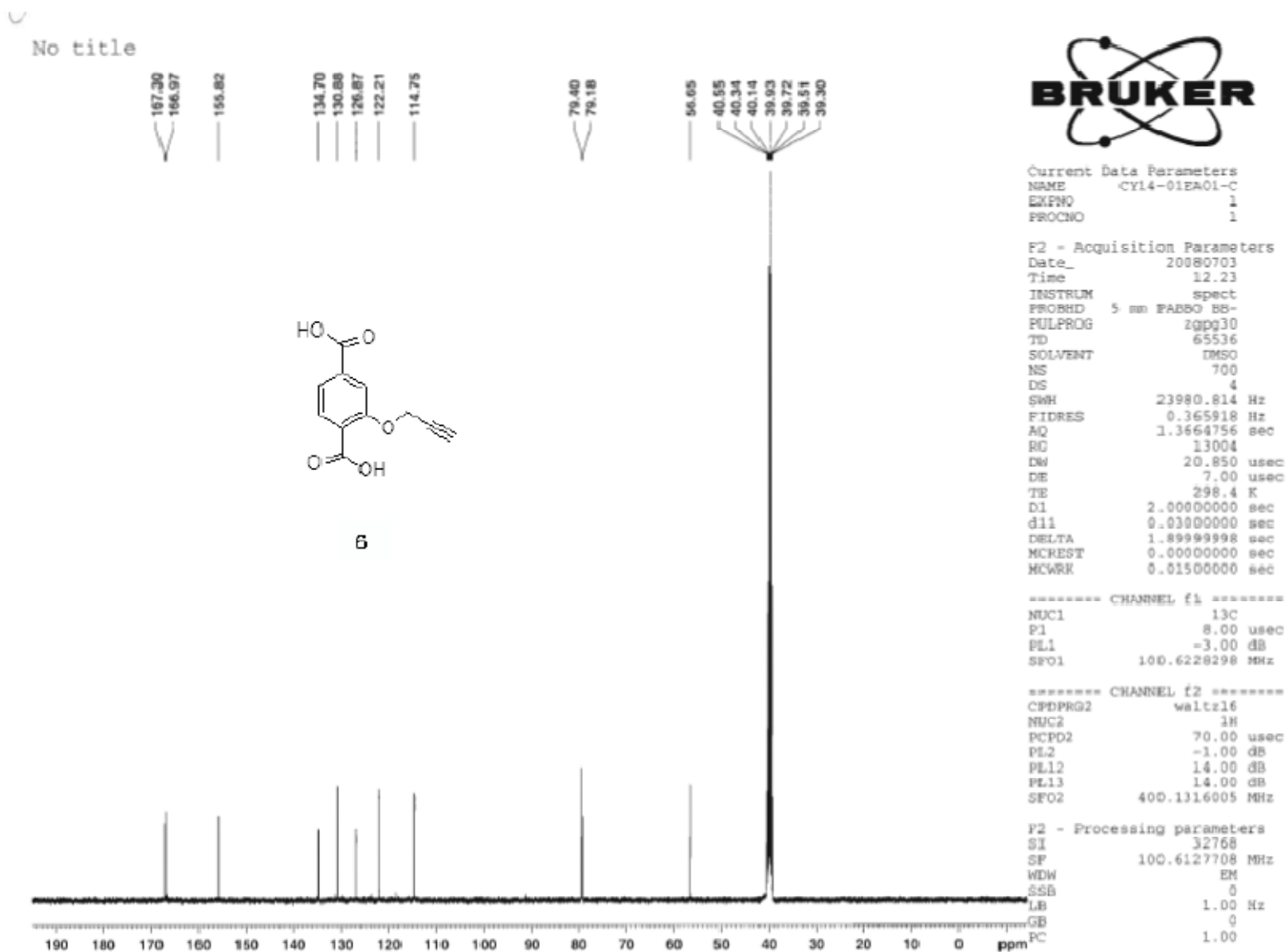
===== CHANNEL f1 =====  
NUC1 1H  
P1 12.80 usec  
PL1 0.00 dB  
SFO1 400.1324710 MHz

F2 - Processing parameters  
SI 32768  
SF 400.1299962 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

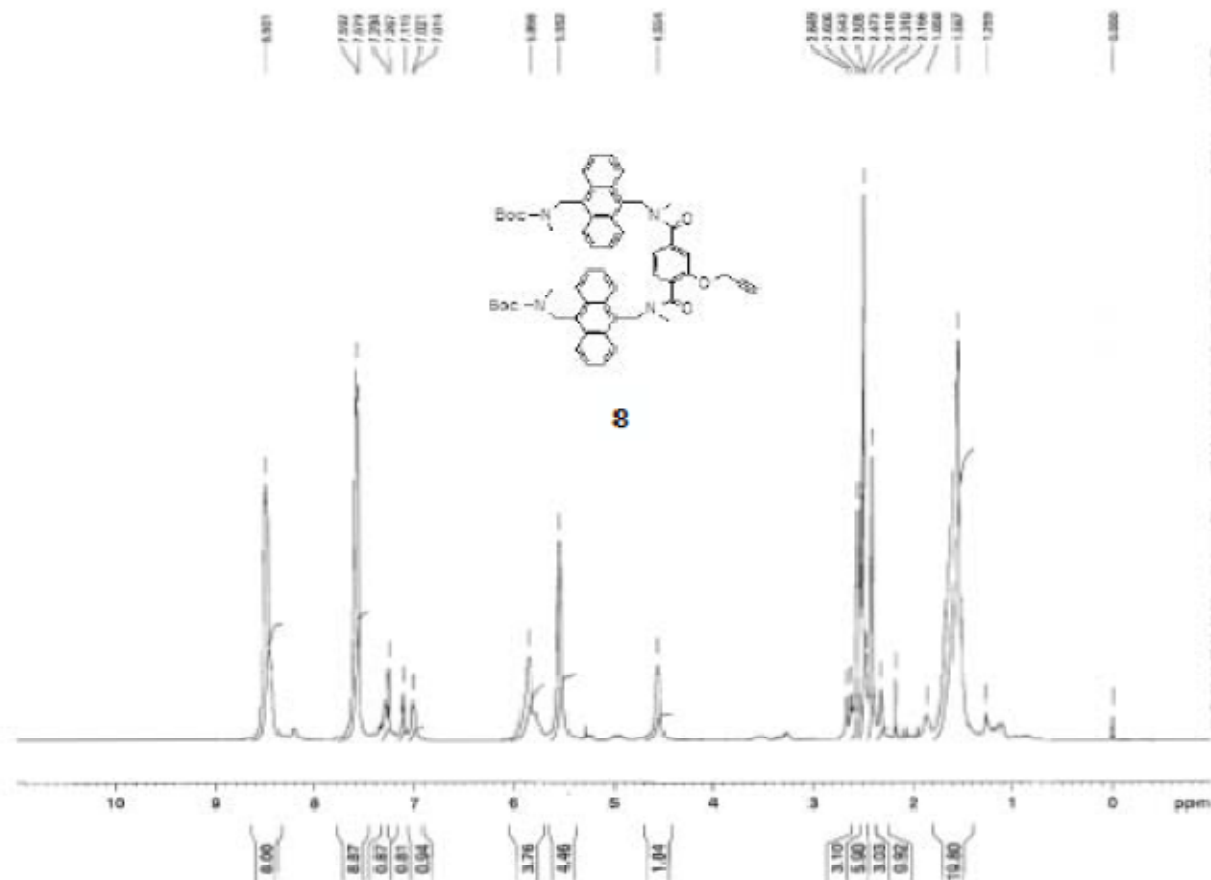


6





No title



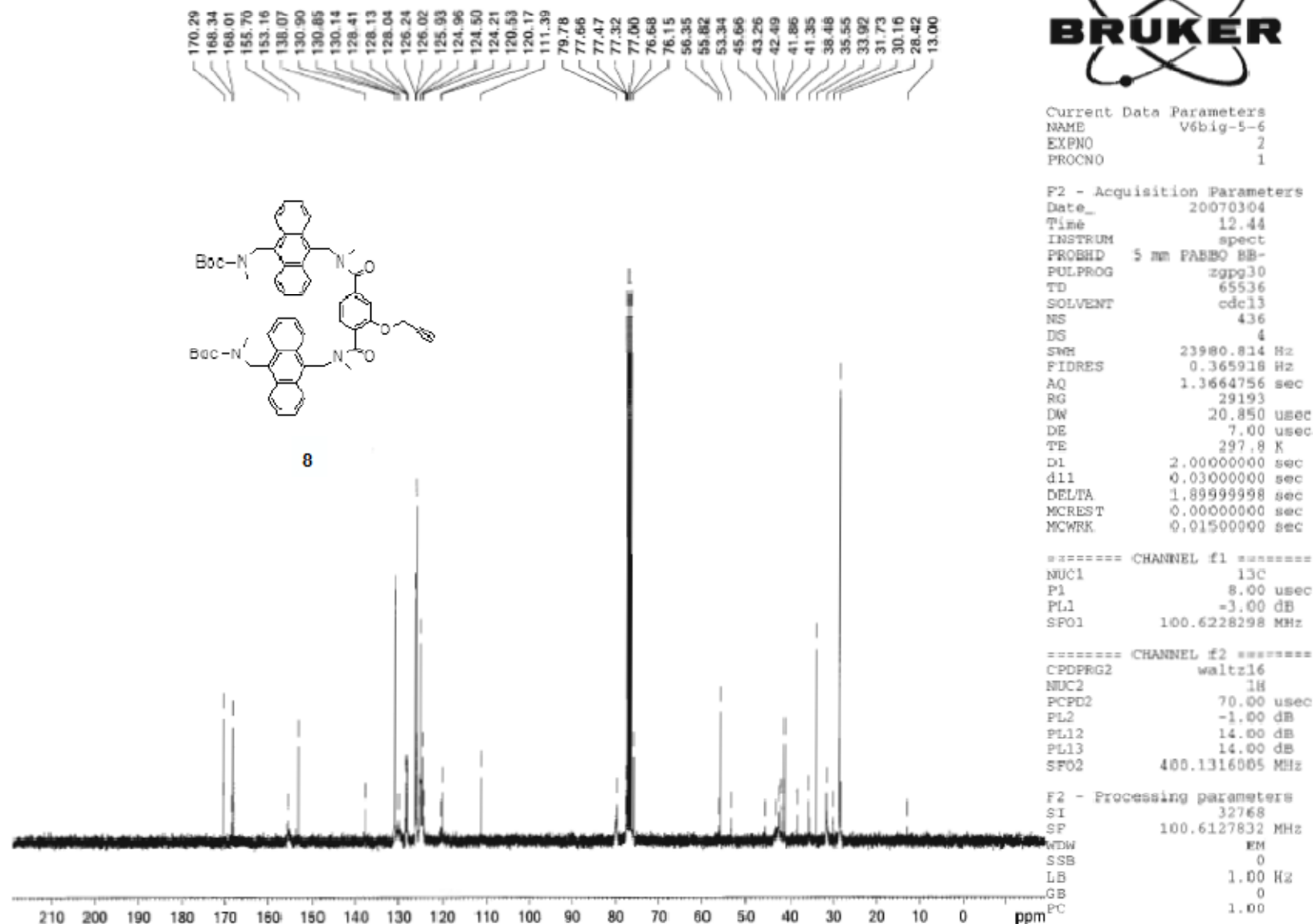
Current Data Parameters  
NAME V15-10-15  
EXPNO 1  
PROCNO 1

F1 - Acquisition Parameters  
Date\_ 20070322  
Time 9.21  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT cdc13  
NS 16  
DS 2  
SWH 8278.146 Kz  
FIDRES 0.126314 Kz  
AQ 3.9584243 sec  
RG 64  
DW 60.400 usec  
DE 7.00 usec  
TE 297.6 K  
D1 1.00000000 sec  
MCREST 0.00000000 sec  
MCMRK 0.01500000 sec

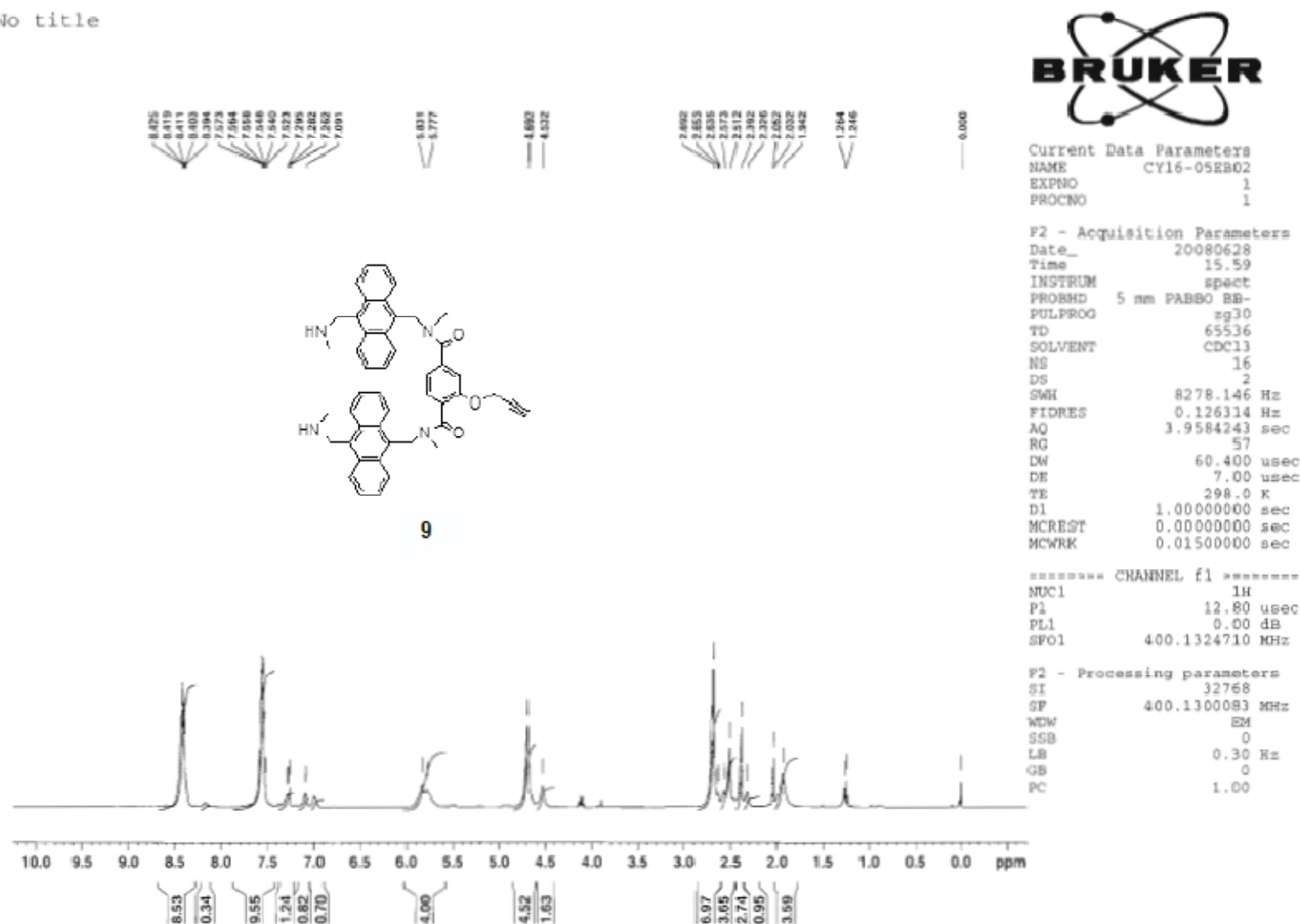
----- CHANNEL f1 -----  
NUC1 1H  
P1 12.80 usec  
PL1 0.00 dB  
SFO1 400.1324710 MHz

F2 - Processing parameters  
SI 32768  
SF 400.1300046 MHz  
NMR 894  
SSB 0  
LB 0.30 Kz  
GB 0  
PC 1.40

No title



No title



170.88  
168.59  
153.68  
131.57  
131.48  
130.46  
127.99  
126.73  
126.51  
126.32  
125.54  
125.42  
125.36  
124.96  
77.87  
77.55  
77.23  
76.77  
56.35  
48.47  
41.90  
37.65  
34.47



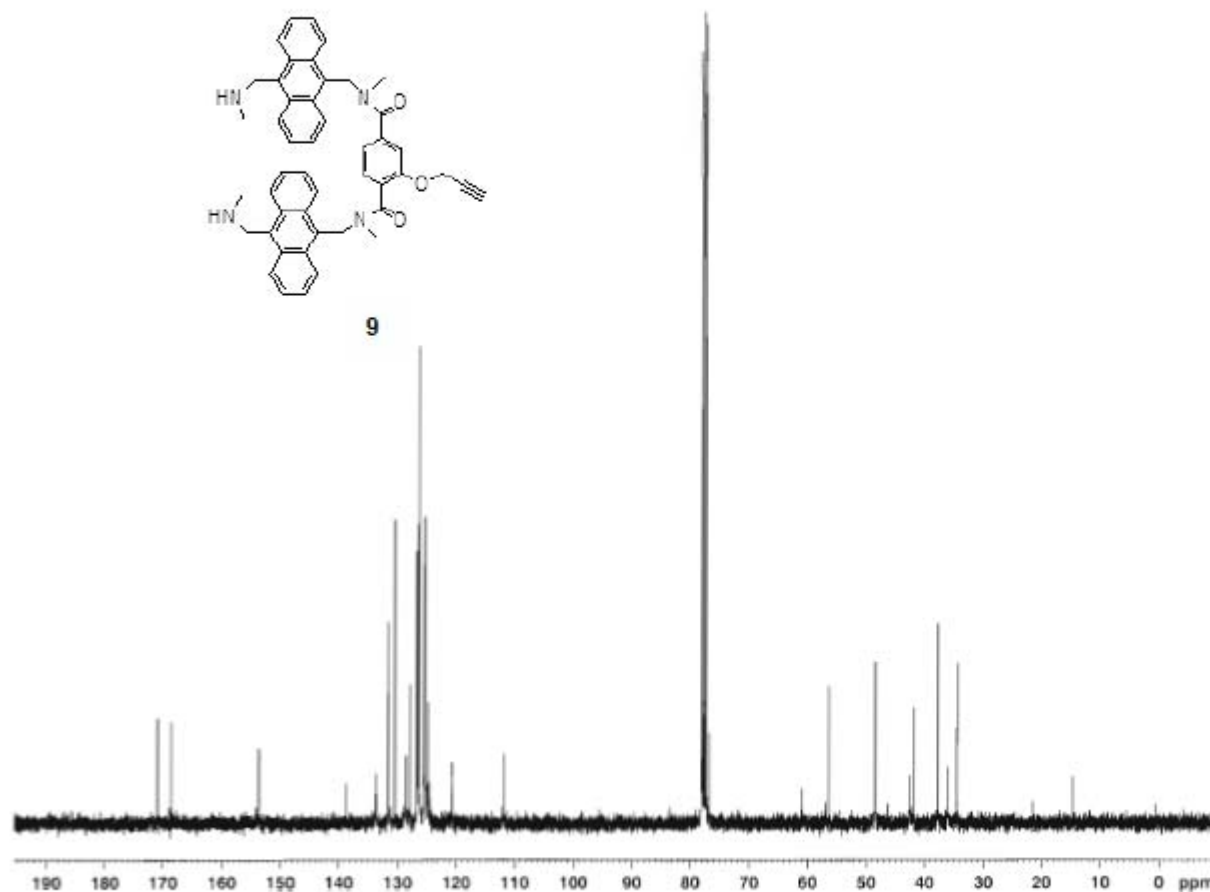
Current Data Parameters  
NAME CY16-05EH02  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20080628  
Time 16.44  
INSTRUM spect  
PROBHD 5 mm PASBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 4  
SWH 23980.814 Hz  
FIDRES 0.365918 Hz  
AQ 1.3664756 sec  
RG 32768  
DW 20.850 usec  
DE 7.00 usec  
TE 298.4 K  
D1 2.00000000 sec  
d11 0.03000000 sec  
DELTA 1.89999998 sec  
MCREST 0.00000000 sec  
MCWRX 0.01500000 sec

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.00 usec  
PL1 -3.00 dB  
SFO1 100.6228298 MHz

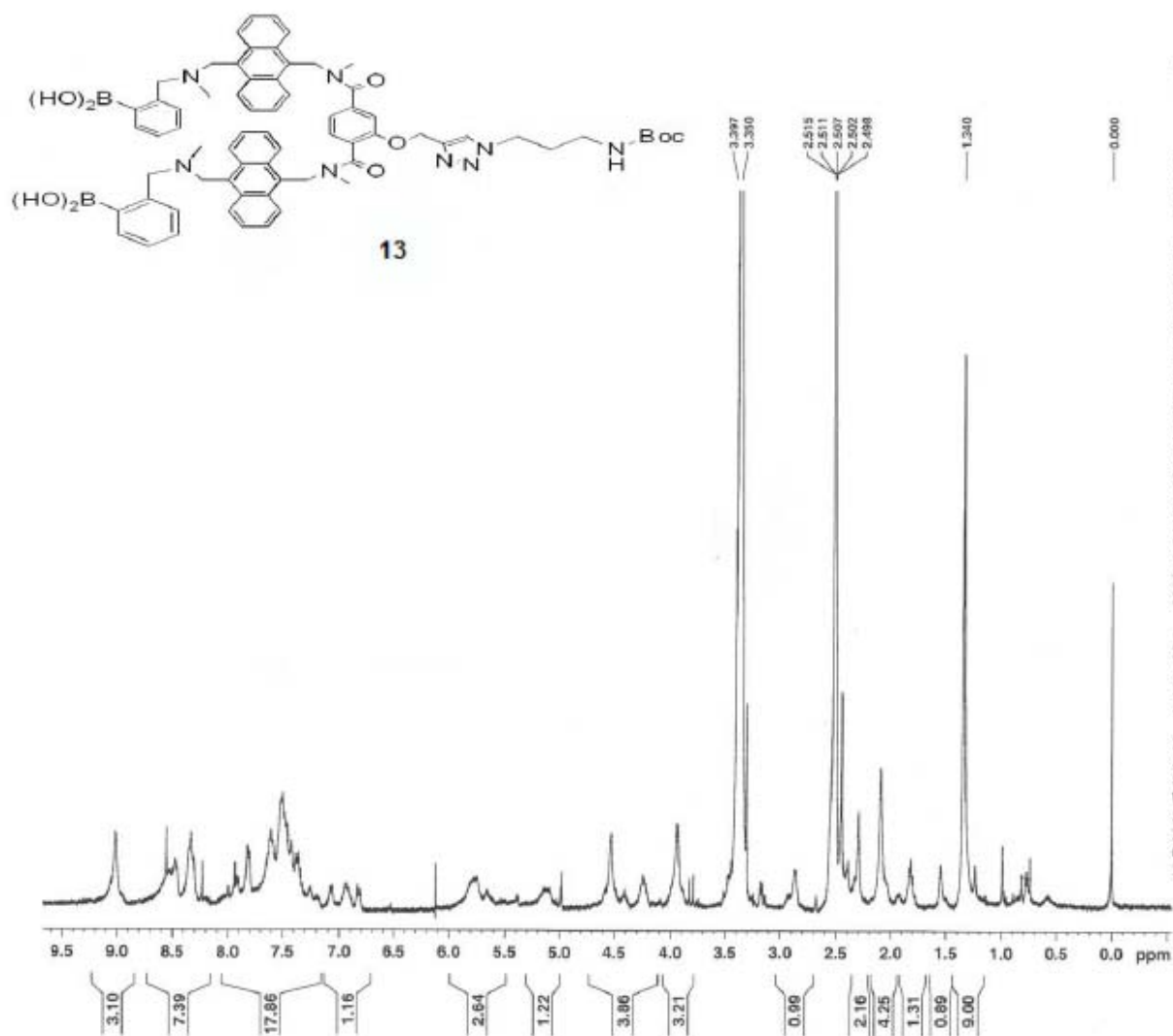
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 70.00 usec  
PL2 -1.00 dB  
PL12 14.00 dB  
PL13 14.00 dB  
SFO2 400.1316005 MHz

F2 - Processing parameters  
SI 32768  
SF 100.6127271 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40









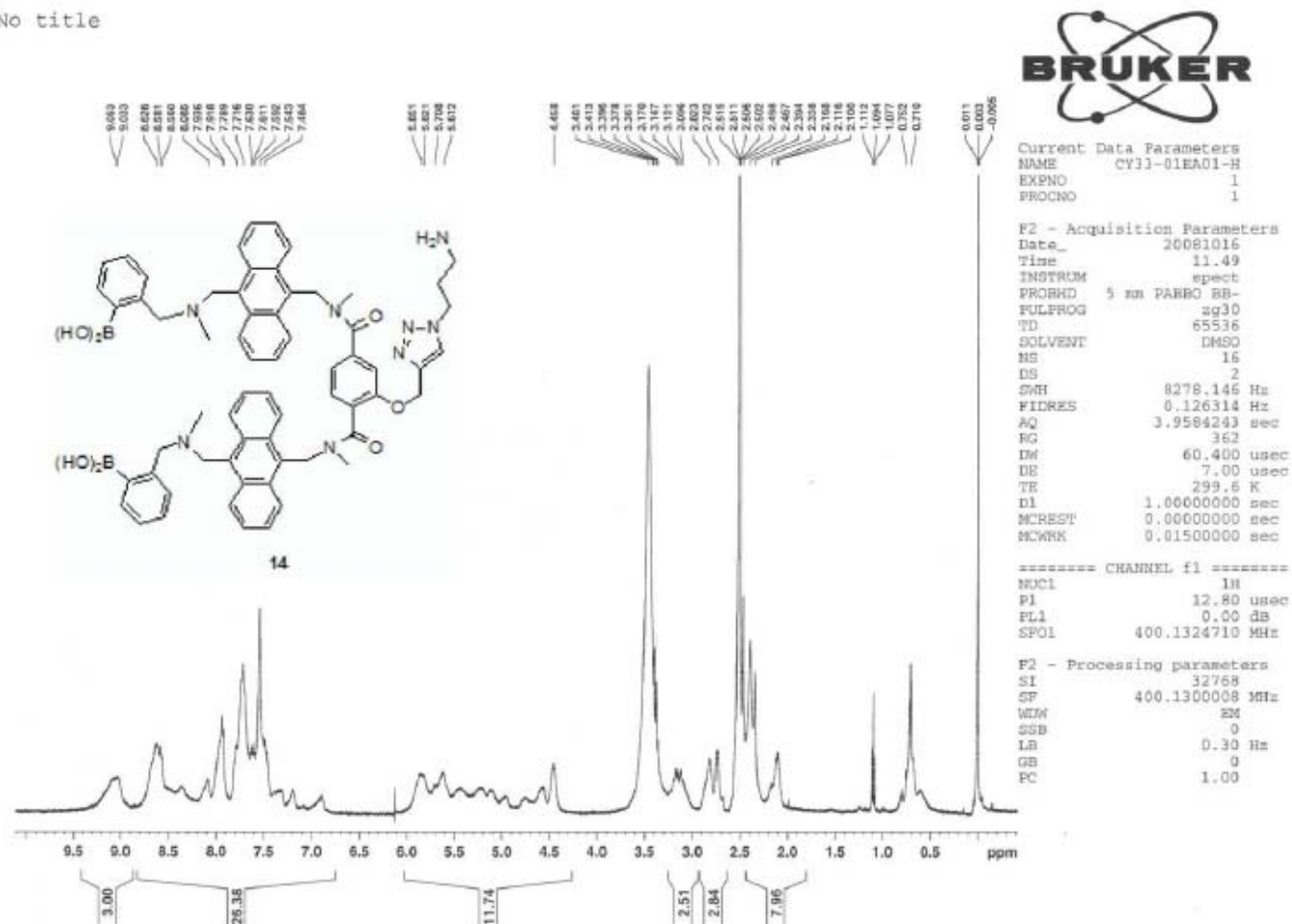
Current Data Parameters  
NAME CY32-01EA02-H2  
EXPNO 1  
PROCNO 1

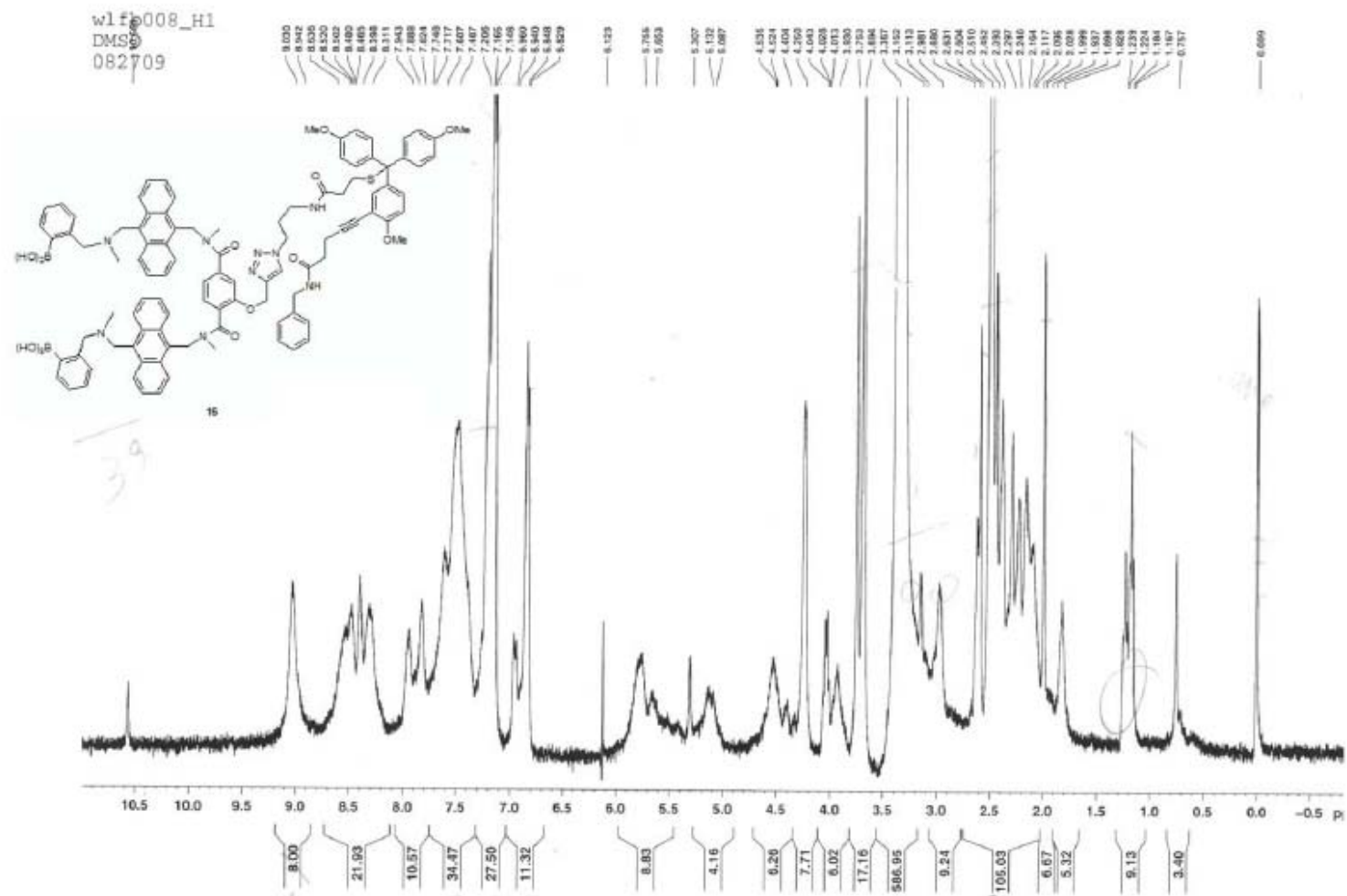
F2 - Acquisition Parameters  
Date\_ 20081016  
Time 11.35  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 16  
DS 2  
SWH 8278.146 Hz  
FIDRES 0.126314 Hz  
AQ 3.9584243 sec  
RG 406.4  
DW 60.400 usec  
DE 7.00 usec  
TE 299.8 K  
D1 1.00000000 sec  
MCREST 0.00000000 sec  
MCWRK 0.01500000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
NUC1 1H  
P1 12.80 usec  
PL1 0.00 dB  
SFO1 400.1324710 MHz

F2 - Processing parameters  
SI 32768  
SF 400.1300008 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

No title





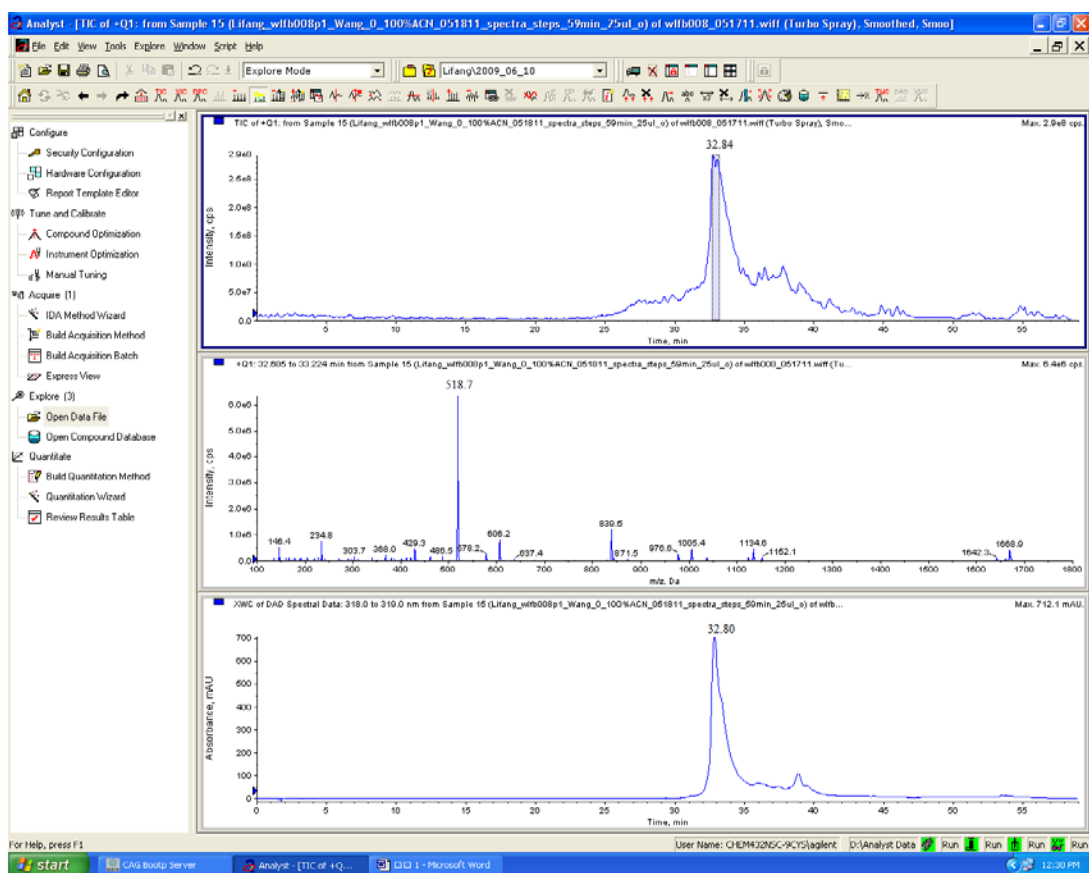


Figure S2 LCMS of compound 16

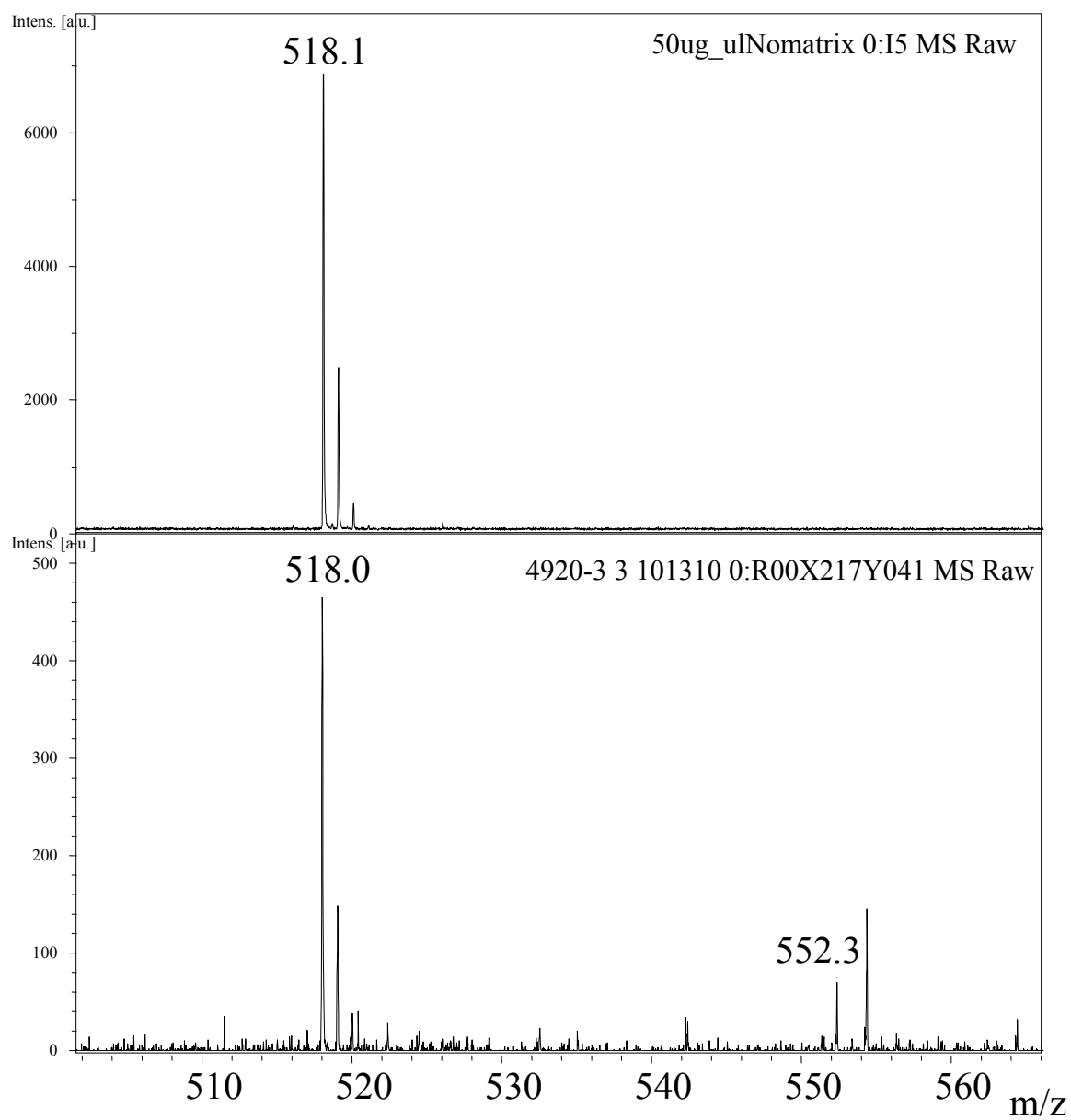


Figure S3 MALDI-IMS of compound **16** without matrix (top)  
and directly from tissue (bottom)