## Electronic Supplementary Information

# Practical manganese-catalysed highly enantioselective cis-dihydroxylation of electron-deficient alkenes and detection of cis-dioxomanganese(V) intermediate by high-resolution ESI-MS analysis 

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## NMR spectroscopy and mass spectrometry

${ }^{1} \mathrm{H}$ NMR and ${ }^{13}$ NMR spectra were recorded on a Bruker DPX-300 spectrometer, with chemaical shifts relative to tetramethylsilane. EI mass spectra were measured on a Finnigan MAT 95 mass spectrometer. ESI mass spectra were obstianed on a Waters Micromess Q-Tof Premier quadrupole time-of-flight tandem mass spectrometer.

## Preparation of $\left[\mathrm{Mn}^{\mathrm{II}}((S, S)-B Q C N) \mathrm{Cl}_{2}\right](1)$

A mixture of $(S, S)$-BQCN $(0.50 \mathrm{mmol})$ and $\mathrm{MnCl}_{2} \cdot 4 \mathrm{H}_{2} \mathrm{O}(2.50 \mathrm{mmol})$ in acetonitrile/ methanol ( $1: 2 \mathrm{v} / \mathrm{v}, 15 \mathrm{~mL}$ ) was refluxed for 18 h . The mixture was then filtered and concentrated by rotary evaporation, followed by addtion of diethyl ether for precipitation of $\mathbf{1}$. The resulted yellow crude product was washed by small amount of methanol and recrystallized from acetonitle-diethyl ether to give $\mathbf{1}$ as a yellow crystalline solid. Yield: 94\%. ESI-MS: $m / z 486.1$ ([Mn $\left.\left.{ }^{\mathrm{II}}((S, S)-\mathrm{BQCN}) \mathrm{Cl}\right]^{+}\right)$.

## General procedure for asymmetric cis-dihydroxylation of alkenes with Oxone catalysed by 1

A solution of Oxone $(1.0 \mathrm{mmol})$ and $\mathrm{NaHCO}_{3}(3.0 \mathrm{mmol})$ in water $(6 \mathrm{~mL})$ was added in two portions to a solution of alkene ( 0.5 mmol ) and $\mathbf{1}(2-5 \mathrm{~mol} \%)$ in acetonitrile ( 6 mL ) at room temperature within 5 min . The reaction mixture was stirred and monitored by TLC. Upon quenching by aqueous saturated $\mathrm{Na}_{2} \mathrm{SO}_{3}$ solution, the mixture was extracted with ethyl acetate ( 10 mL ) conatining 1,4-dichlorobenzene (GC internal standard, 0.1 mmol ) and the aliquot of the extract was analyzed by GC. Afterwards, the mixture was further extracted with ethyl acetate ( $3 \times 50 \mathrm{~mL}$ ) and the oragnic crude products were identified and quantified by ${ }^{1} \mathrm{H}$ NMR analysis. The cis-diol product was isolated after purification by column chromatography on silica gel. The enatiomeric excess of purified cis-diol product was determined by chiral HPLC (Chiralpak OD-3 and AD-3).

## Characterization of cis-diol products

The previously known cis-diol products corresponding to substrates $\mathbf{2 a}, \mathbf{4 a}, \mathbf{5 a}, \mathbf{6 b}$ and 7 were characterised on the basis of the spectral data described in literature.
(a) B. Plietker, M. Niggemann and A. Pollrich, Org.
Biomol. Chem., 2004, 2, 1116; (b) B. Plietker and M.
Niggemann, J. Org. Chem., 2005, 70, 2402.

(product for 2b)
${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}$ ) $\delta 4.54(\mathrm{~d}, J=4.46 \mathrm{~Hz}, 1 \mathrm{H})$, $5.10(\mathrm{~d}, J=4.44 \mathrm{~Hz}, 1 \mathrm{H}), 6.95(\mathrm{~d}, J=7.46 \mathrm{~Hz}, 2 \mathrm{H}), 7.22(\mathrm{t}, J$ $=7.4 \mathrm{~Hz}, 1 \mathrm{H}), 7.31-7.41(\mathrm{~m}, 5 \mathrm{H}), 7.49(\mathrm{~d}, J=7.13 \mathrm{~Hz}, 2 \mathrm{H})$. ${ }^{13} \mathrm{C}$ NMR ( $75 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}$ ) $\delta 76.38,77.19,122.56,127.01$, 128.03, 128.82, 129.24, 130.38, 142.13, 151.97, 172.55. EI-MS: $m / z 258[M]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{15} \mathrm{H}_{14} \mathrm{O}_{4}$, calcd 258.0887, found 258.0880.

(product for 2c)


(product for 3a)
${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 1.44(\mathrm{~s}, 9 \mathrm{H}), 4.26(\mathrm{~d}, J=3.61$ $\mathrm{Hz}, 1 \mathrm{H}), 4.91$ (d, $3.57 \mathrm{~Hz}, 1 \mathrm{H}$ ), 7.28-7.46 (m, 5H). ${ }^{13} \mathrm{C}$ NMR $\left(75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 27.97,74.96,74.99,83.35,126.64$, 128.03, 128.42, 140.22, 172.00. EI-MS: $m / z 238[\mathrm{M}]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{13} \mathrm{H}_{18} \mathrm{O}_{4}$, calcd 238.1200, found 238.1195 .
${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 3.56$ (br, 2H), 3.81 ( $\mathrm{s}, 3 \mathrm{H}$ ), $4.60(\mathrm{~d}, J=6.25 \mathrm{~Hz}, 2 \mathrm{H}), 5.26(\mathrm{~d}, J=12.15 \mathrm{~Hz}, 1 \mathrm{H}), 5.27(\mathrm{~d}$, $J=12.16 \mathrm{~Hz}, 1 \mathrm{H}), 7.30-7.38(\mathrm{~m}, 5 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( 75 MHz , $\left.\mathrm{CDCl}_{3}\right) \delta 53.16,68.05,72.23,72.31,128.44,128.71,128.73$, 134.93, 171.51, 172.04. EI-MS: $m / z 254$ [M] ${ }^{+}$; HRMS (EI): $\mathrm{m} / \mathrm{z}$ for $\mathrm{C}_{12} \mathrm{H}_{14} \mathrm{O}_{6}$, calcd 254.0785, found 254.0784.

(product for 3b)

(product for 3c)

${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 3.24(\mathrm{~d}, J=7.07 \mathrm{~Hz}, 2 \mathrm{H})$, $4.61(\mathrm{~d}, J=6.63 \mathrm{~Hz}, 2 \mathrm{H}), 5.26(\mathrm{~d}, J=12.18 \mathrm{~Hz}, 2 \mathrm{H})$, 5.27 (d, $J=12.17,2 \mathrm{H}), 7.33-7.42(\mathrm{~m}, 10 \mathrm{H}) .{ }^{13} \mathrm{C} \mathrm{N}$ MR ( $75 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 68.21,72.22,128.53,128.82,134.89$, 171.50. EI-MS: $m / z 239$ [M - $\left.\mathrm{PhCH}_{2}\right]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{11} \mathrm{H}_{11} \mathrm{O}_{6}$, calcd 239.0550, found 239.0551 .
${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 3.19(\mathrm{~d}, J=5.55 \mathrm{~Hz}, 2 \mathrm{H})$, $4.60(\mathrm{~d}, J=4.98 \mathrm{~Hz}, 2 \mathrm{H}), 5.04(\mathrm{~d}, J=11.94 \mathrm{~Hz}, 2 \mathrm{H})$, $5.08(\mathrm{~d}, J=11.94 \mathrm{~Hz}, 2 \mathrm{H}), 7.23-7.27(\mathrm{~m}, 5 \mathrm{H}), 7.32-7.35$ $(\mathrm{m}, 5 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $75 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta$ 68.19, 73.04, 128.74, 128.83, 128.86, 134.63, 170.97. EI-MS: m/z 239 [M - $\left.\mathrm{PhCH}_{2}\right]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{11} \mathrm{H}_{11} \mathrm{O}_{6}$, calcd 239.0550, found 239.0550 .

(product for 4b)
${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 0.94(\mathrm{t}, J=7.13 \mathrm{~Hz}, 3 \mathrm{H}), 1.05$ $(\mathrm{t}, J=7.11 \mathrm{~Hz}, 3 \mathrm{H}), 2.60-2.72(\mathrm{~m}, 1 \mathrm{H}), 2.77-2.87(\mathrm{~m}, 1 \mathrm{H})$, 3.02-3.13 (m, 1H), 3.49-3.61 (m, 1H), $4.30(\mathrm{~d}, J=6.06 \mathrm{~Hz}$, $1 \mathrm{H}), 4.74(\mathrm{~d}, J=6.05 \mathrm{~Hz}, 1 \mathrm{H}), 7.26-7.40(\mathrm{~m}, 5 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 12.64,13.88,40.52,40.91,72.82,76.11$, 126.70, 128.23, 128.42, 139.26, 171.18. EI-MS: m/z 238 [M + $\left.\mathrm{H}^{+}\right]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{13} \mathrm{H}_{20} \mathrm{O}_{3} \mathrm{~N}$, calcd 238.1438, found 238.1440.

(product for $\mathbf{5 b}$ )
${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}$ ) $\delta 4.25(\mathrm{~d}, J=2.76 \mathrm{~Hz}, 1 \mathrm{H}), 5.13$ (d, $J=2.62 \mathrm{~Hz}, 1 \mathrm{H}), 7.12$ (t, $J=7.40 \mathrm{~Hz}, 1 \mathrm{H}$ ), $7.25-7.37$ (m, $5 \mathrm{H}), 7.48(\mathrm{~d}, J=7.28 \mathrm{~Hz}, 2 \mathrm{H}), 7.58(\mathrm{~d}, J=8.37 \mathrm{~Hz}, 2 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $\left.75 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}\right) \delta 75.50,77.69,121.48,125.57$, 127.59, 128.39, 129.10, 129.80, 138.91, 143.11, 173.28. EI-MS: $m / z 257$ [M] ${ }^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{15} \mathrm{H}_{15} \mathrm{O}_{3} \mathrm{~N}$, calcd 257.1046, found 257.1040.

(product for $\mathbf{6 a}$ )
${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 3.81-3.92(\mathrm{~m}, 2 \mathrm{H}), 4.30(\mathrm{t}, J=$ $3.39 \mathrm{~Hz}, 1 \mathrm{H}), 5.21(\mathrm{~d}, J=12.23 \mathrm{~Hz}, 1 \mathrm{H}), 5.22(\mathrm{~d}, J=12.23$, $1 \mathrm{H}), 7.29-7.39(\mathrm{~m}, 5 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $75 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 64.17$, 67.69, 71.97, 128.38, 128.67, 128.76, 135.13, 173.02. EI-MS: $m / z 196[M]^{+}$; HRMS (EI): $m / z$ for $\mathrm{C}_{10} \mathrm{H}_{12} \mathrm{O}_{4}$, calcd 196.0730, found 196.0730.

## Detection of reaction intermediates by ESI-MS

A solution of complex 1 in a mixture of acetonitrile and distilled water ( $10: 1 \mathrm{v} / \mathrm{v}$ ) was treated with Oxone (2 equiv.) and $\mathrm{NaHCO}_{3}$ (6 equiv.). The reaction mixture at different reaction time was introduced into the ESI source by a syringe pump operating at a flow of $5 \mu \mathrm{~L} \mathrm{~min}$. For accurate mass measurement, sodium formate was used as an internal reference. The mass resolution was fixed at about 8000 (full width at half-height) with mass accuracy limited within 10 ppm . The TOF-MS mass spectra were acquired in the mass range 100-1200 Th with an accumulation time of 1 sec and interscan time of 0.1 sec . In the MS/MS experiments, the parent ions were separately mass-selected by the first quadrupole mass analyzer (operating at about unit mass resolution). The selected ion was transmitted into a T-wave collision cell (filled with argon gas at $7.01 \times 10^{-3} \mathrm{Torr}$, measured in the quadrupole mass-analyzer
housing) where it underwent collision-induced dissociation at a collision energy of $10-25 \mathrm{eV}$ (laboratory frame). The parent ion and fragment ions were analyzed by the TOF mass analyzer and detected by a multichannel plate (MCP) detector at 1.8 kV . The MS/MS spectrum was obtained by averaging 150-200 scans.

Table S1. Crystal data and structural refinement for complex 1.

|  | $1 \cdot \mathrm{MeCN}$ |
| :---: | :---: |
| Formula | $\mathrm{C}_{26} \mathrm{H}_{28} \mathrm{Cl}_{2} \mathrm{MnN}_{4} \cdot \mathrm{C}_{2} \mathrm{H}_{3} \mathrm{~N}$ |
| Mr | 563.42 |
| Crystal system | Monoclinic-P |
| Space group | P2 $1_{\text {(\#4) }}$ |
| $a, ~ \AA$ | 11.3578(13) |
| $b, \AA$ | 8.7479(10) |
| $c, \AA$ | 14.5673(17) |
| $\alpha$, deg | 90 |
| $\beta$, deg | 103.890(2) |
| $\gamma, \operatorname{deg}$ | 90 |
| $F(000)$ | 586 |
| $V, \AA^{3}$ | 1405.0(3) |
| Z | 2 |
| $\rho_{\text {calc, }}, \mathrm{g} \mathrm{cm}^{-3}$ | 1.332 |
| $\mu\left(\mathrm{MoK}_{\alpha}\right), \mathrm{mm}^{-1}$ | 0.685 |
| $2 \theta_{\text {max }}$, deg | 57.48 |
| Reflections collected | 9382 |
| Independent reflections | 5814 |
| Parameters | 329 |
| Final $R$ indices | $R 1=0.0287$ |
| $\left(F^{2}>2 \sigma\left(F^{2}\right)\right.$ | $R \mathrm{~W}=0.0671$ |
| Goodness-of-fit | 1.015 |
| Flack parameter | 0.031(13) |
| Largest diff. peak/hole, e $\AA^{-3}$ | 0.20/-0.19 |

Table S2 Oxidation of methyl cinnamte (2a) with Oxone at room temperature using various manganese complexes in situ generated from reaction of ligand with $\mathrm{Mn}(\mathrm{II})$ or Mn (III) salt ${ }^{a}$

$S, S$-BQCN


6-Me ${ }_{2}$-BPBP


6-Me2-BPMCN


| Entry | Ligand used | Mn salt used | Conv.$(\%)^{b}$ | Yield based on conversion (\%) |  | D/E |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | cis-Diol ${ }^{\text {b }}$ | Epoxide ${ }^{\text {b }}$ |  |
| 1 | --- | $\mathrm{Mn}(\mathrm{OAc})_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ | 5 | 0 | 56 | --- |
| 2 | $S, S$-BQCN | $\mathrm{Mn}(\mathrm{OAc})_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ | 93 | $66^{\text {c }}$ | 19 | 3.5 |
| 3 |  | $\mathrm{Mn}(\mathrm{OTf})_{2}$ | 97 | 28 | 52 | 0.54 |
| 4 | $6-\mathrm{Me}_{2}$-BPBP | $\mathrm{Mn}(\mathrm{OAc})_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ | 7 | 0 | 73 | --- |
| 5 |  | $\mathrm{Mn}(\mathrm{OTf})_{2}$ | 27 | 23 | 43 | 0.53 |
| 6 | 6-Me ${ }_{2}$-BPMCN | $\mathrm{Mn}(\mathrm{OAc})_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ | 7 | 0 | 91 | --- |
| 7 |  | $\mathrm{Mn}(\mathrm{OTf})_{2}$ | 24 | 11 | 42 | 0.26 |

${ }^{a}$ Reaction conditions: Mn salt ( $5 \mathrm{~mol} \%, 0.025 \mathrm{mmol}$ ) and ligand ( $5 \mathrm{~mol} \% 0.025 \mathrm{mmol}$ ) was stirred in MeCN ( 6 mL ) for 10 min . After addition of methyl cinnamate ( $\mathbf{2 a}$ ) ( 0.5 mmol ), Oxone ( 2 equiv.) and $\mathrm{NaHCO}_{3}$ ( 6 equiv.) in $\mathrm{H}_{2} \mathrm{O}(6 \mathrm{~mL})$ were added in 2 portions within 5 min , R.T., $2 \mathrm{~h} .{ }^{b}$ Determined by ${ }^{1} \mathrm{H}$ NMR. ${ }^{c}$ Isolated yield.

## Chiral HPLC chromatographic analysis of cis-diol of 2a

- Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane (7:93) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 2a.

(II): cis-Diol from asymmetric cis-dihydroxylation of 2a with Oxone catalysed by $\mathbf{1}$.


| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | RetTirne <br> [min] | Type | Width <br> [min] | $\begin{gathered} \mathrm{Area} \\ {\left[\mathrm{mu}^{+} \mathrm{T} \mathrm{~J}^{+}\right]} \end{gathered}$ | Height [ mLU ] | $\begin{gathered} \text { Area } \\ 8 \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 21.407 | MF | 0. 6646 | 1.08976e4 | 273.27890 | 96.3402 |
| 2 | 24.920 | FM | 0.7951 | 413.98526 | 8. 67786 | 3.6598 |

(III) cis-Diol from asymmetric cis-dihydroxylation of 2a with Oxone catalysed by the manganese complex in situ generated from reaction of $\mathrm{Mn}(\mathrm{OAc})_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ with $S, S$-BQCN.


| Peak \# | $\begin{gathered} \text { RetTime } \\ \text { [min] } \end{gathered}$ | Type | Width <br> [min] | $\begin{gathered} \text { Area } \\ {\left[\mathrm{mu}^{+} \mathrm{J}\right]} \end{gathered}$ | Heicht <br> [muT] | Area $8$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 21.420 |  | 0.6816 | 1.81322 E 4 | 443.35577 | 90.75 .54 |
| 2 | 24.931 |  | 0.7750 | 1847.00012 | 39.71991 | 9.2446 |

(IV) cis-Diol from gram-scale asymmetric cis-dihydroxylation of 2a with Oxone catalysed by 1 .


Fig. S1 Chiral HPLC chromatographic analysis of cis-diol of 2a. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of $\mathbf{2 a}$ catalysed by 1. (III) Asymmetric cis-dihydroxylation of 2a catalysed by in situ generated manganese complex. (IV) Gram-scale asymmetric cis-dihydroxylation of 2a catalysed by $\mathbf{1}$.

## Chiral HPLC chromatographic analysis of cis-diol of 2b

* Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
a Condition: isopronanol $/ n$-hexane (7:93) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 2b

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{2 b}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S2 Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{2 b}$. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 2b catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of 2c

* Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane (1:99) at $0.5 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 2c

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{2 c}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S3 Chiral HPLC chromatographic analysis of cis-diol of 2c. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 2c catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of 3a

* Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane (10:90) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 3a

(II): cis-Diol from asymmetric cis-dihydroxylation of 3a with Oxone catalysed by $\mathbf{1}$.


Fig. S4 Chiral HPLC chromatographic analysis of cis-diol of 3a. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 3a catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of 3b

* Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane ( $10: 90$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 3b

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{3 b}$ with Oxone catalysed by $\mathbf{1}$.

(II): cis-Diol from gram-scale asymmetric cis-dihydroxylation of 3b with Oxone catalysed by 1 .


Fig. S5 Chiral HPLC chromatographic analysis of cis-diol of 3b. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 3b catalysed by 1. (III) Gram-scale asymmetric cis-dihydroxylation of 3b catalysed by $\mathbf{1}$.

## Chiral HPLC chromatographic analysis of cis-diol of 4a

* Column: Analytical Chiralpak AD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
a Condition: isopronanol $/ n$-hexane ( $10: 90$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of $\mathbf{4 a}$.

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{4 a}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S6 Chiral HPLC chromatographic analysis of cis-diol of 4a. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 4a catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{4 b}$

* Column: Analytical Chiralpak AD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
a Condition: isopronanol $/ n$-hexane ( $10: 90$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of 4b.


| Peak \# | $\begin{gathered} \text { RetTime } \\ {[\text { minin] }} \end{gathered}$ | Type | $\begin{gathered} \text { Width } \\ {[\mathrm{min}]} \end{gathered}$ | $\begin{gathered} \text { Area } \\ {\left[\mathrm{ma}^{+} \mathrm{B}\right]} \end{gathered}$ | Heicfht $\text { [ } \mathrm{mBU}]$ | $\begin{gathered} \text { Area } \\ \frac{8}{8} \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 21.959 | WINI | 0.4884 | 1.078.57e4 | 368.08688 | 50.1169 |
| 2 | 25.843 | IWIM | 0.5871 | 1.07354@4 | 304.73276 | 49.88 |

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{4 b}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S7 Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{4 b}$. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 4b catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of 5a

* Column: Analytical Chiralpak AD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
a Condition: isopronanol $/ n$-hexane ( $10: 90$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of $\mathbf{5 a}$.

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{5 a}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S8 Chiral HPLC chromatographic analysis of cis-diol of 5a. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 5a catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{5 b}$

* Column: Analytical Chiralpak AD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane ( $10: 90$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of $c i s$-diol of $\mathbf{5 b}$.

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{5 b}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S9 Chiral HPLC chromatographic analysis of cis-diol of 5b. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of 5b catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{6 a}$

* Column: Analytical Chiralpak AD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
* Condition: isopronanol $/ n$-hexane ( $7: 93$ ) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of $\mathbf{6 a}$.

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{6 a}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S10 Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{6 a}$. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of $\mathbf{6 a}$ catalysed by 1.

## Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{6 b}$

* Column: Analytical Chiralpak OD-3 ( $25 \mathrm{~cm} \times 0.46 \mathrm{~cm} \times 3 \mu \mathrm{~m}$ ).
a Condition: isopronanol $/ n$-hexane (7:93) at $1.0 \mathrm{~mL} / \mathrm{min}$, enantiomeric excess determined at 210 nm .
(I): Racemic standard of cis-diol of $\mathbf{6 b}$.

(II): cis-Diol from asymmetric cis-dihydroxylation of $\mathbf{6 b}$ with Oxone catalysed by $\mathbf{1}$.


Fig. S11 Chiral HPLC chromatographic analysis of cis-diol of $\mathbf{6 b}$. (I): Racemic cis-diol standard. (II): Asymmetric cis-dihydroxylation of $\mathbf{6 b}$ catalysed by $\mathbf{1}$.

A: $[\mathrm{BQCN}-\mathrm{Me}]^{+}$
B: $\left[\mathrm{BQCN}+\mathrm{H}^{+}\right]^{+}$
$\mathrm{C}:[\mathrm{Mn}(\mathrm{BQCN})]^{+}$
D: $\left\{\left[\mathrm{Mn} 2(\mathrm{BQCN})_{2}(\mathrm{O})_{3}\right]-2 \mathrm{H}^{+}\right\}^{2+}$
$\mathrm{E}:\left[\mathrm{Mn}(\mathrm{BQCN})(\mathrm{O})_{2}\right]^{+}$


Fig. S12 Electrospray ionization mass spectrometric measurements of the reaction mixture of $\mathbf{1}$ with Oxone ( 2 equiv.) in $\mathrm{MeCN} / \mathrm{H}_{2} \mathrm{O}(10: 1 \mathrm{v} / \mathrm{v})$ at different reaction time $(t): 0 \mathrm{sec}$ (i.e. before addition of Oxone), 30 sec and 15 min .


Fig. S13 Electrospray ionization mass spectrometric measurement of $[\mathrm{Mn}(\mathrm{BQCN}) \mathrm{Cl}]^{+}$for $\mathbf{1}$ in MeCN : (Upper) Simulated isotope pattern of $\left[\mathrm{MnC}_{26} \mathrm{H}_{28} \mathrm{~N}_{4} \mathrm{Cl}\right]^{+}$and (Lower) Experimental mass measurement.


Fig. S14 Electrospray ionization mass spectrometric measurement of $[\mathrm{Mn}(\mathrm{BQCN}) \mathrm{O}]^{2+}$ in the reaction of $\mathbf{1}$ with Oxone (2 equiv.): (Upper) Simulated isotope pattern of $\left[\mathrm{MnC}_{26} \mathrm{H}_{28} \mathrm{~N}_{4} \mathrm{O}\right]^{2+}$ and (Lower) Experimental mass measurement.


Fig. S15 Electrospray ionization mass spectrometric measurement of $\left[\mathrm{BQCN}+\mathrm{H}^{+}\right]^{+}$ in the reaction of $\mathbf{1}$ with Oxone ( 2 equiv.): (Upper) Simulated isotope pattern of $\left[\mathrm{C}_{25} \mathrm{H}_{25} \mathrm{~N}_{4}\right]^{+}$and (Lower) Experimental mass measurement.


Fig. S16 Electrospray ionization mass spectrometric measurement of [BQCN-Me] ${ }^{+}$in the reaction of 1 with Oxone (2 equiv.): (Upper) Simulated isotopic pattern of $\left[\mathrm{C}_{26} \mathrm{H}_{29} \mathrm{~N}_{4}\right]^{+}$and (Lower) Experimental mass measurement.


Fig. S17 Electrospray ionization mass spectrometric measurement of $[\mathrm{Mn}(\mathrm{BQCN})]^{+}$ in the reaction of $\mathbf{1}$ with Oxone (2 equiv.): (Upper) Simulated isotope pattern of $\left[\mathrm{MnC}_{26} \mathrm{H}_{28} \mathrm{~N}_{4}\right]^{+}$and Lower) Experimental mass measurement.


Fig. S18 Electrospray ionization mass spectrometric measurement of $\left\{\left[\mathrm{Mn}_{2}(\mathrm{BQCN})_{2}(\mathrm{O})_{3}\right]-2 \mathrm{H}^{+}\right\}^{2+}$ in the reaction of $\mathbf{1}$ with Oxone (2 equiv.): (Upper) Simulated isotope pattern of $\left[\mathrm{Mn}_{2} \mathrm{C}_{52} \mathrm{H}_{54} \mathrm{~N}_{8} \mathrm{O}_{3}\right]^{2+}$ and (Lower) Experimental mass measurement.


Fig. S19 Collision-induced dissociation of $[\mathrm{Mn}(\mathrm{BQCN}) \mathrm{Cl}]^{+}$for $\mathbf{1}$ in MeCN recorded at the collision energy of 25 eV .


Fig. S20 Collision-induced dissociation of $\left\{\left[\mathrm{Mn}_{2}(\mathrm{BQCN})_{2}(\mathrm{O})_{3}\right]-2 \mathrm{H}^{+}\right\}^{2+}$ for the reaction of $\mathbf{1}$ with Oxone (2 equiv.) recorded at the collision energy of 12 eV .


$\mathrm{C}_{17} \mathrm{H}_{2} \mathrm{~N}_{3}$
$\mathrm{MW}: 268$

M.W.: 129:

$\underset{435.1399}{\left[\mathbf{L}+\mathbf{K}^{+}\right]^{+}}$
TOF MSMS 482.99ES+


Fig. S21 Collision-induced dissociation of $\left[\mathrm{Mn}(\mathrm{BQCN})(\mathrm{O})_{2}\right]^{+}$for the reaction of $\mathbf{1}$ with Oxone (2 equiv.) recorded at the collision energy of $10 \mathrm{eV} . \mathrm{L}=\mathrm{BQCN}=$ $\mathrm{C}_{26} \mathrm{H}_{28} \mathrm{~N}_{4}$.

