

1 Supplementary Material for Chemical Communications
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4 **Supplementary Data**

6 **Label-Free Fluorescent Detection of Cu(II) Ions Based on the** 7 **DNA Cleavage-Dependent Graphene-Quenched DNazymes**

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10 **Reagents and Instruments**

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12 Cu(II)-dependent DNazymes (Cu-Sub: 5'-T₁₀AGCTTCTTTCTAATACGGCTTACC-3' and Cu-Enz:
13 5'-GGTAAGCCTGGGCCTTTCTTTTAAGAAAGAAC-3') were purchased from Takara Biotechnology Co.
14 (Dalian, China) purified by high-performance liquid chromatography (HPLC). GelRed (10000×) in water was
15 purchased from Biotium. NaCl, CuCl₂, BaCl₂, CaCl₂, CdCl₂, FeCl₃, MnCl₂, Pb(NO₃)₂ and ZnCl₂ were of analytical
16 reagent grade and purchased from the Kemiou Agent Co., Tianjin (China). Other reagents such as HCl, HNO₃, and
17 H₂O₂ (30%) and anhydrous ethanol were used as received without any treatment. Sodium ascorbate was obtained from
18 Sinopharm Chemical Reagent Co., Ltd (China). Ultrapure water obtained from a Millipore water purification system
19 (resistivity > 18.0 MΩ cm⁻¹, Laikie Instrument Co., Ltd, Shanghai, China) was used throughout the experiments.
20 Phosphate buffer solution (PBS, 20 mM) with pH 7.4 was prepared by mixing the stock solution of Na₂HPO₄ and
21 NaH₂PO₄. FL measurements were performed using a Hitachi F-4500 spectrofluorimeter with a scan rate at 2400
22 nm/min. The excitation wavelength was at 530 nm. The slits for excitation and emission were set at 5 nm/10 nm.

24 **Preparation of Graphene Oxide and Graphene**

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26 Graphene oxide was prepared by our reported method.¹ An environment-friendly hydrothermal route was employed to
27 convert graphene oxide to graphene. The resultant solution of graphene (0.1 mg/mL) was stored at room temperature
28 and employed in the following experiments.

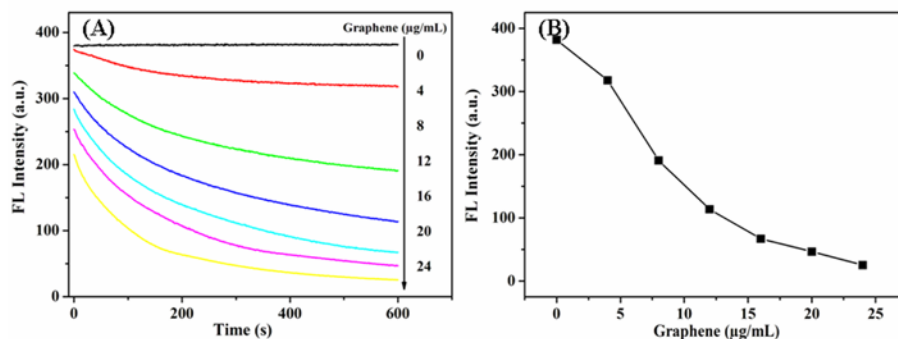
30 **Preparation of DNazymes**

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32 In brief, 1.8 μM Cu-Sub and 1.8 μM Cu-Enz were firstly mixed in a buffer (pH 7.4) containing 20 mM PBS, 0.5 M
33 NaCl. Then the mixture was warmed to 95 °C for 2 min in a water bath and subsequently allowed to cool naturally to
34 room temperature (25 °C). The obtained DNazymes were stored at 4 °C.

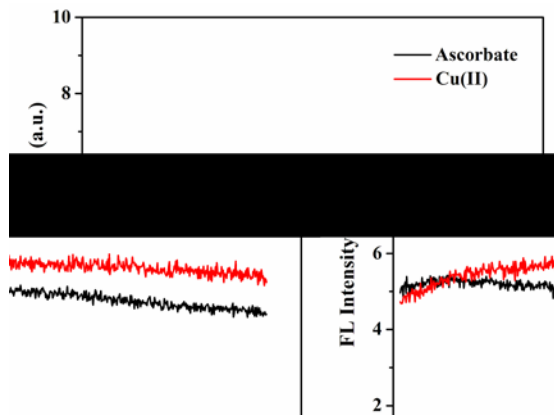
36 **Fluorescent Assay for Cu(II)**

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38 In a typical experiment, 200 μL of GelRed (100×) and 800 μL of DNazymes were incubated for 20 min at 25 °C. The
39 final concentration of Cu(II)-dependent DNazymes was 1.45 μM. Subsequently, 340 μL of PBS buffer (20 mM, 50
40 mM NaCl, pH 7.4), 20 μL of DNazymes-GelRed, 120 μL of graphene solution (0.1 mg/mL) were sequentially added
41 into a microcentrifuge tube. After 15 min reaction, the solution was transferred to a quartz cell. Finally, 10 μL of
42 ascorbate (10 mM) and 10 μL of different concentrations of Cu(II) were added for time-dependent FL measurement at
43 $\lambda_{\text{ex}}/\lambda_{\text{em}} = 530/613$ nm. The final volume of the solution was fixed at 500 μL. The control experiment was carried out
44 under the same condition without the addition of Cu(II).

1 **Figures**



16 Fig. S1 (A) Kinetics study for the FL changes of the DNAzymes-GelRed complex with different concentrations of graphene.
17 (B) FL changes of DNAzymes-GelRed complex as a function of graphene with a series of concentrations. Experimental
18 conditions: 20 mM PBS, 50 mM NaCl, pH 7.4, 58 nM DNAzymes, GelRed (20 \times), reaction time: 10 min (25 $^{\circ}$ C), excitation
19 wavelength: 530 nm, emission wavelength: 613 nm.
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37 Fig. S2 Kinetics study for FL changes of the label-free graphene-quenched DNAzymes in the absence of Cu(II) or ascorbate.
38 Experimental conditions: 20 mM PBS, 50 mM NaCl, pH 7.4, 58 nM DNAzymes, 24 μ g/mL graphene, GelRed (20 \times), 0.2 mM
39 ascorbate or 0.5 mM Cu(II), reaction time: 10 min (25 $^{\circ}$ C), excitation wavelength: 530 nm, emission wavelength: 613 nm.
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52 **Reference**

53 1 M. Liu, H. M. Zhao, X. Quan, S. Chen and X. F. Fan, *Chem. Commun.*, 2010, **46**, 7909-7911.
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