

Electronic Supplementary Information

A Platinum Chugaev Carbene.Complex as Potent-anti Cancer Agent

Georges Alves,^{a,b,e} Laurent Morel ,^{a,b,e} Malika El-Ghozzi,^{d,e} Daniel Avignant,^{d,e} Bertrand Legeret,^{c,e} Lionel Nauton,^{c,e} Federico Cisnetti, ^{c,e} and Arnaud Gautier,*^{c,e}

a CNRS, UMR 6247, laboratoire GReD, 24 Ave des Landais, F-63177, Aubière, France.

b INSERM, U 931, laboratoire GReD, 24 Ave des Landais, F-63177 Aubière, France.

c CNRS, UMR 650, laboratoire SEESIB, 24 Ave des Landais, F-63177 Aubière, France. Tel : +33 (0)4 73 40 76 46 ; Fax : +33 (0)4 73 40 77 17 ; E-mail : arnaud.gautier@univ-bpclermont.fr.

d CNRS, UMR 6002, laboratoire LMI, 24 Ave des Landais, F-63177 Aubière, France.

e Clermont Université, Université Blaise Pascal, BP 10448, F-63000 Clermont-Ferrand, France.

Table of content.

S1. Material and methods.

S2. Synthesis of complex 1 and sample preparation.

S3. ^1H NMR spectrum of complex 1.2dmso.

S4. ^{13}C NMR spectrum of complex 1.2dmso.

S5. ^{195}Pt NMR spectrum of complex 1.2dmso.

S6. Crystal data and structure refinement of complex 1.2dmso.

S7. UV spectra of the products 1.2dmso, 2 and 3.

S8. Chromatogram of the product 2.

S9. Expansion of the mass spectrum of the product 2.

S10. ^{195}Pt NMR spectrum of the product 2

S11. Chromatogram of the product 3.

**S12. Expansion of the mass spectrum of the product 3 at $m/z= 699$
(100%abundance)**

**S13. Full mass spectrum of the product 3 (5 and 100%
abundance).**

S14. ^{195}Pt NMR spectrum of the product 3.

S15. Reaction of 1 with guanosine.

S1. Material and methods.

Solution NMR spectra were recorded in Fourier Transform mode at 25 °C on a Bruker AVANCE 400 (^1H at 400 MHz, ^{13}C at 100 MHz) or a Bruker AV 500 (^{195}Pt at 107 MHz) spectrometer. Data are reported as chemical shifts (δ) in parts per million (ppm) with respect to the standard reference compounds tetramethylsilane (^1H , ^{13}C) or Na_2PtCl_6 (^{195}Pt). Spin multiplicity is described by the following abbreviations: s = singlet, d = doublet. Coupling constants (J) are reported in hertz (Hz). IR spectrum was obtained as solid KBr pellets on a Shimadzu Fourier Transform Infrared Spectrophotometer FTIR-8400S and is reported in wave numbers (cm^{-1}). Electrospray (positive mode) high-resolution mass spectra were recorded on a Q-TOF micro spectrometer (Waters), using an internal lock mass (H_3PO_4) and an external lock mass (leucine-enkephalin $[\text{M}+\text{H}]^+$: $m/z = 556.2766$). UV spectra were recorded with a Perkin-Elmer lambda 25 spectrophotometer. IC₅₀ are realized at the Gif-sur-Yvette facility using the MTT method. X-ray data were recorded with a Bruker APEX-II CCD diffractometer. HPLC were realized with the aid of a Dionex ASI-100 automated sample injector equipped with P580 pump and using a UVD340 detector. Reverse phase C18 column (Dionex, 5 μm , 120 $\text{\AA}$, 4.6X250mm) was used with the following conditions: A: water, 0.1% TFA; B: Methanol; flow: 0.8mL/ min. From 0 to 3 min : isocratic 10% A; from 3 to 20 min : gradient up to 90% B; from 20 to 40 minutes: isocratic 90% B.

Dichloromethane was distilled over CaH_2 and methanol over Mg/I_2 . They were *degassed by bubbling argon during a few minutes* prior to be used.

Cell proliferation assay. IC₅₀ values were determined at the CNRS, ICSN ‘ciblotheque cellulaire’ facility using the following automatized protocol: Cells were plated in 96-well tissue culture plates in 200 μL medium and treated 24 h later with compounds dissolved in DMSO; compound concentrations ranged 0.05 μM to 100 μM and were prepared by use of a Biomek 2000 (Beckman). Control cells received the same volume of DMSO (1% final volume). After 72 h exposure to the drug, MTS reagent (Promega) was added and incubated for 3 h at 37 °C: the absorbance was monitored at 490 nm and results are expressed as the inhibition of cell proliferation calculated as the ratio $[(\text{OD}_{490} \text{ treated}/\text{OD}_{490} \text{ control})\times 100]$. For IC₅₀ determinations (50% inhibition of cell proliferation) experiments were performed in duplicate.

DNA gel electrophoresis. A solution (3.10^{-4}M) of **1** was prepared in DMF. 1 μL of this solution was added to 1.4 μg of plasmid pcDNA 4T0 in the presence of NaClO_4 (10^{-2}M) in 30

μL and incubated 24h at 37°C. Samples were then analysis by agarose (1%) electrophoresis and DNA was revealed with BET under UV light.

S2. Synthesis of complex 1 and sample preparation.

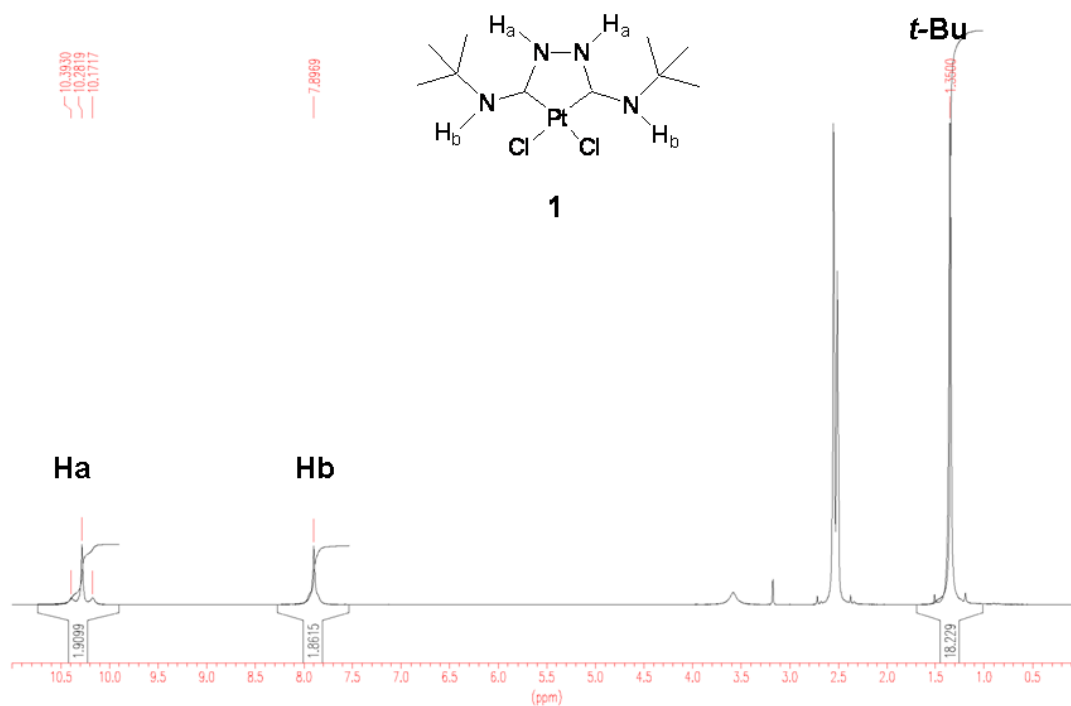
To a solution of Pt(COD)Cl₂ (300 mg, 0.802 mmol, 1.0 eq.) in 8 mL of degassed dichloromethane is added *tert*-butyl isocyanide (190 µL, 1.68 mmol, 2.1 eq.). The solution is stirred 1^h at room temperature and evaporated. The resulting white solid is dissolved in 8 mL of degassed ethanol and hydrazine monohydrate is added (90 µL, 1.68 mmol, 2.1 eq.) to furnish a pale yellow solution that is stirred overnight. The solvent is removed under reduced pressure to yielding a yellow solid that is heated at 75°C in 10 mL of aqueous HCl (4 M) until a white precipitate forms. The solution is cooled to room temperature, filtered and dried on air resulting in a white solid (248 mg, 0.53 mmol, 67%). This compound is insoluble in water, poorly soluble in DMSO but highly in DMF. Suitable crystals for X-ray diffraction of **1.2dmsO** were grown from by layering diethyl ether onto a saturated DMSO solution of **1**.ⁱ Mass spectra and ¹⁹⁵Pt NMR for cysteine and glutathione were realized using these crystals. ¹H NMR (400MHz, DMSO-d₆): 10.18 (d, ³J_{H-Pt} = 88.5Hz, 1H, Ha), 7.90 (d, ³J_{H-Pt} = 31.4Hz, 1H, H_b), 1.35 (s, 18H, H^{*tert*-bu}). ¹³C NMR (100MHz, DMF-d₇): 163.6 (C-Pt, ¹J_{C-Pt} = 1413 Hz), 54.5 (C-(CH₃)₃), ³J_{C-Pt} = 46.6 Hz), 28.3 (C-(CH₃)₃). ¹⁹⁵Pt NMR (107MHz, DMF + 10%D₂O): -3498. IR (KBr): ν(cm⁻¹): 3300, 2962, 1570, 1520, 1473, 1464, 1412, 1375, 1233, 1207, 1020, 590. Elemental analysis. Calcd for C₁₀H₂₂Cl₂N₄Pt: C: 25.87%, H: 4.78%, N: 12.07%. Found C: 26.04%, H: 4.92%, N: 12.15%.

Sample preparation:

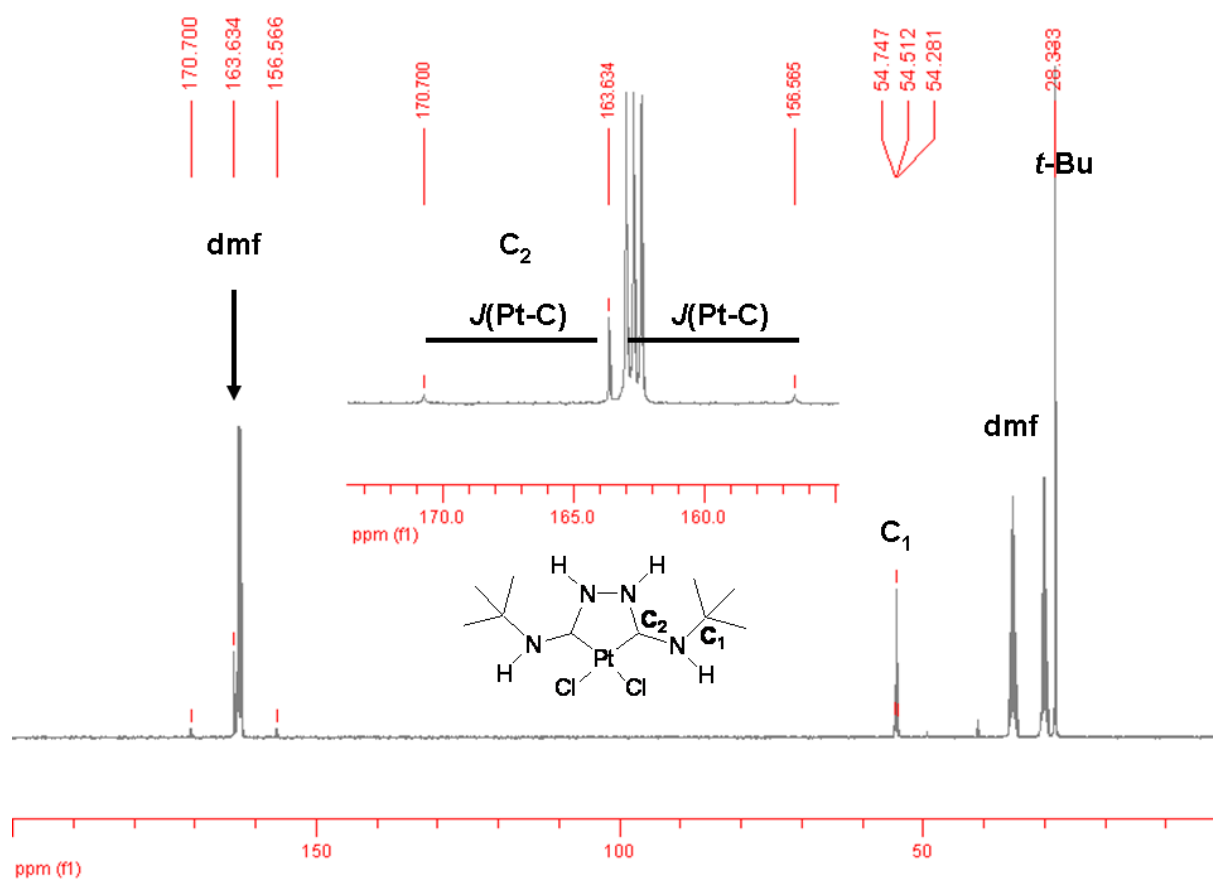
¹⁹⁵Pt NMR and mass spectrometry: solutions of 0.10 M of cysteine or glutathione were prepared in 0.5 M phosphate buffer (pH = 7.45 for ¹⁹⁵Pt NMR) or in 0.5 M triethyl ammonium carbonate (pH = 8.6 for mass spectra). To 1.0 mL of these solutions was added **1.2dmsO** (0.025mmol, 11.7 mg) dissolved in 100µL of DMF under vigorous stirring.

¹H NMR (interaction with guanine): **1** (3.0 mg, 6.5·10⁻³mmol) is dissolved in 0.5 mL of dmf-d₇ and 0.2 mL of D₂O containing a small amount of 3-(trimethylsilyl)-1-propane sulfonic acid sodium salt as internal standard. This solution is added to solid guanosine monohydrate (5.5mg, 0.019 mmol, 3.0 equivalents), stirred 2h and monitored by ¹H NMR.

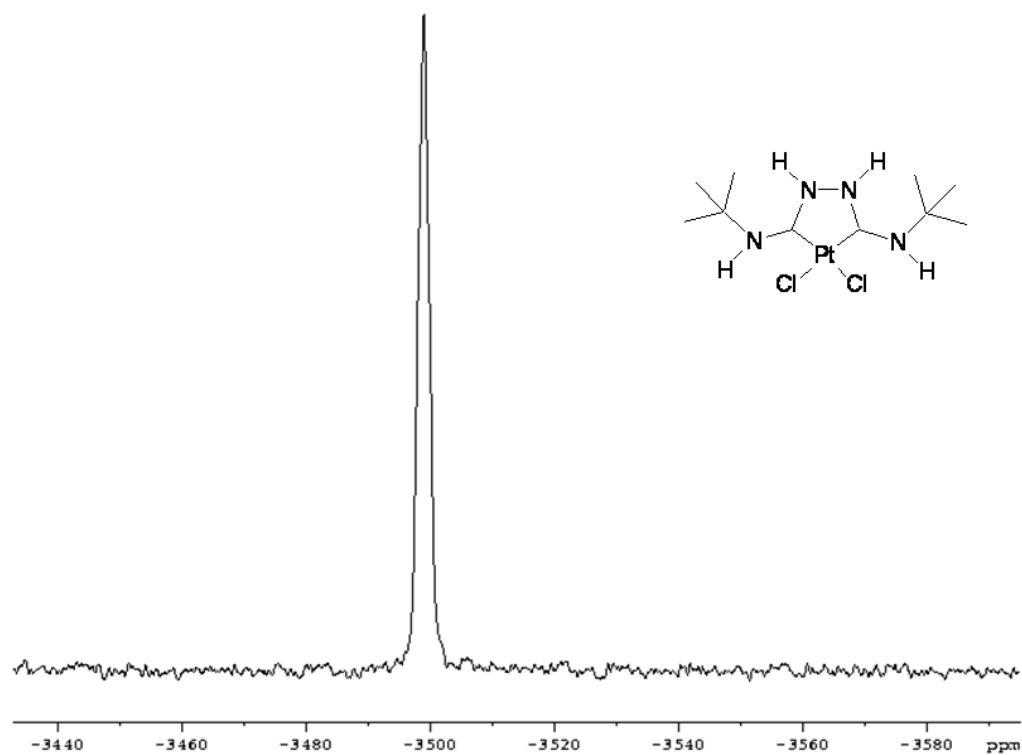
S3. ^1H of Chugaev complex 1.2dmso.



S4. ^{13}C of Chugaev complex 1.



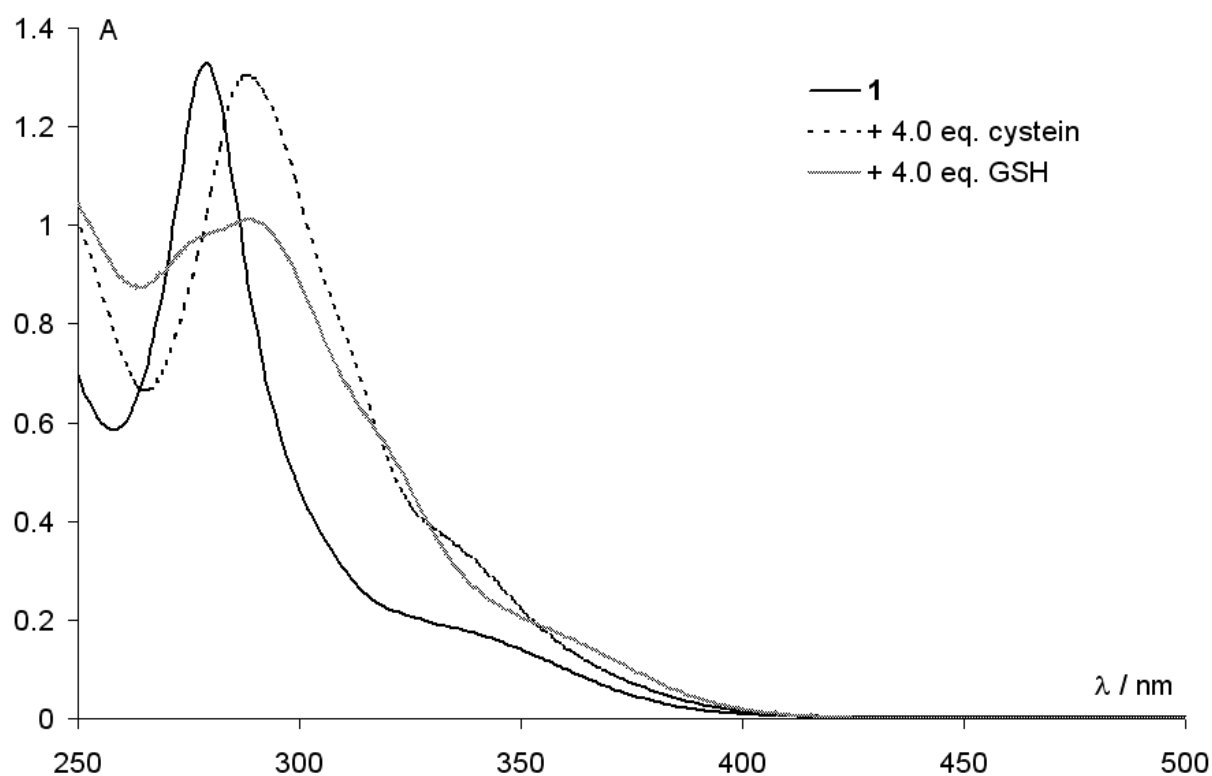
S5. ¹⁹⁵Pt of complex 1.



S6. Crystal data and structure refinement of complex 1.2dmso.

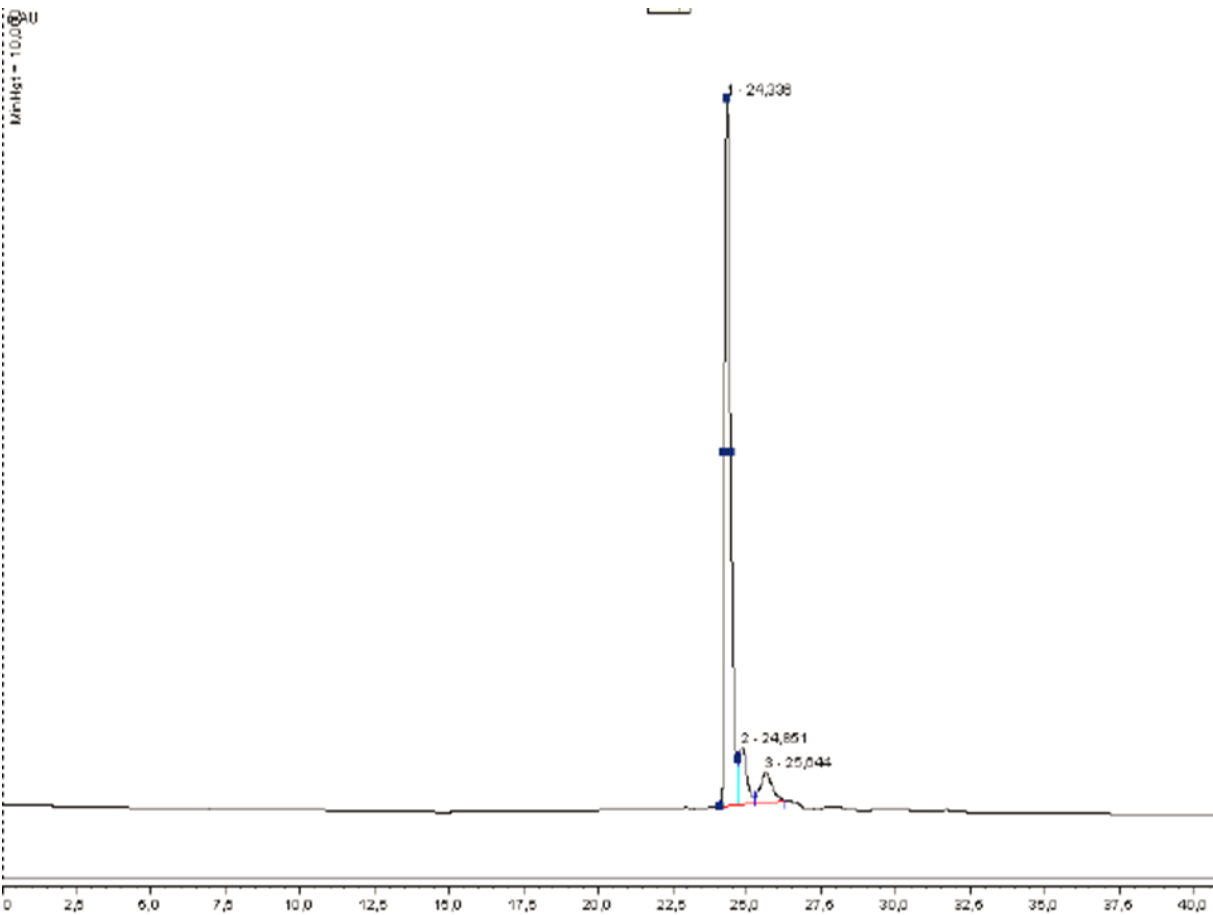
Chemical formula	C _{6.50} H _{15.50} Cl ₁ N ₂ O ₁ Pt _{0.50} S ₁
M_w	302.77
Crystal system / Space group	Monoclinic / C2/c
Unit cell parameters	
a [Å]	18.1852(6)
b [Å]	11.5308(4)
c [Å]	12.0790(4)
β [°]	100.628(2)
V [Å ³]	2489.39(15)
Z	4
$\rho_{\text{calcd.}}$ [g·cm ⁻³]	1.616
F(000)	1188
μ [mm ⁻¹]	6.03
Data collection	
θ range [°]	2 – 35
Measured reflections	19827
Independent reflections	5420
Observed reflections	4207 with $I > 2\sigma(I)$
Absorption correction	Multi-scan
Tmin./ Tmax.	0.3227 / 0.7469
Number of refined parameters	265
R_1 / ($F > 2\sigma(F)$)	0.03
wR_2 (F)	0.027
s	1.0756
$\Delta\rho$ max / $\Delta\rho$ min (e. Å ⁻³)	1.08/ -1.21

S7. UV spectra of the products **1**, **2** and **3**.

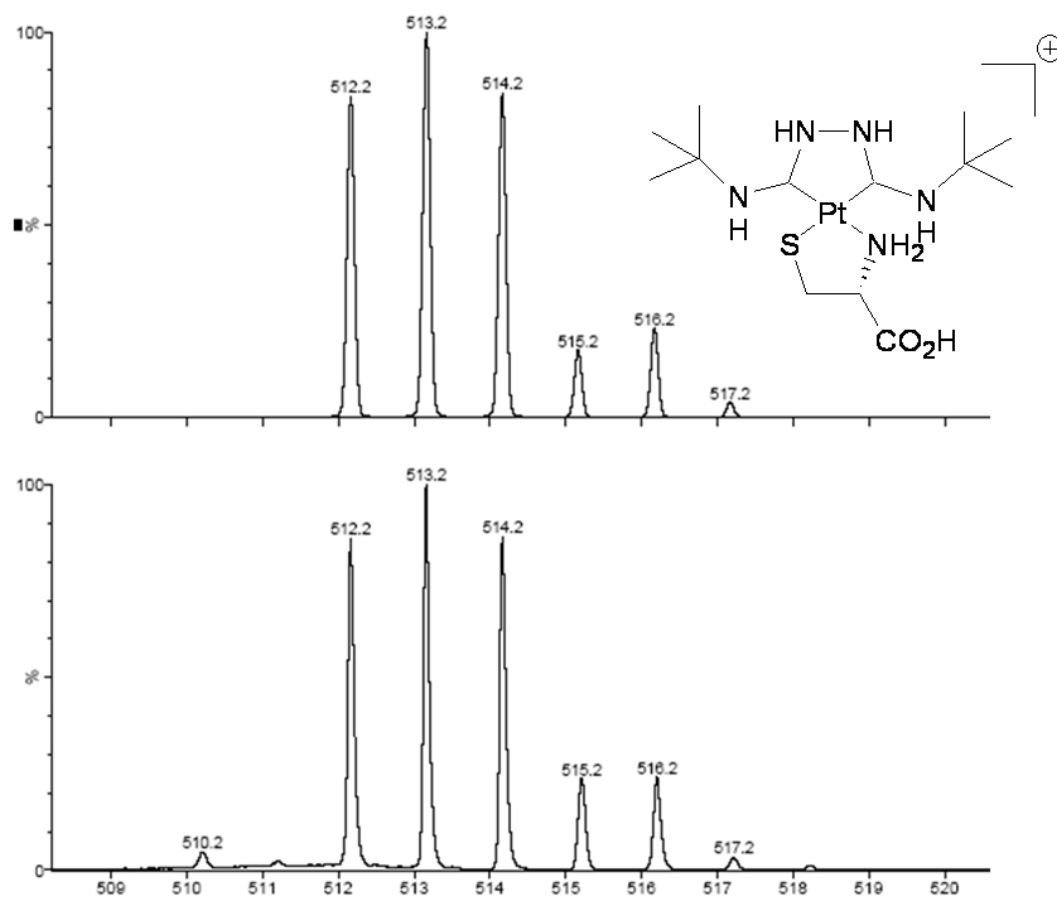


UV spectra of **1** and adducts with cysteine and glutathione. Concentration = 1.0×10^{-4} M, 0.1 M pH = 7.45 phosphate buffer. The aqueous solution of **1** was prepared by dilution of a 0.05 M stock solution in DMF.

S8. Chromatogram of the product 2.

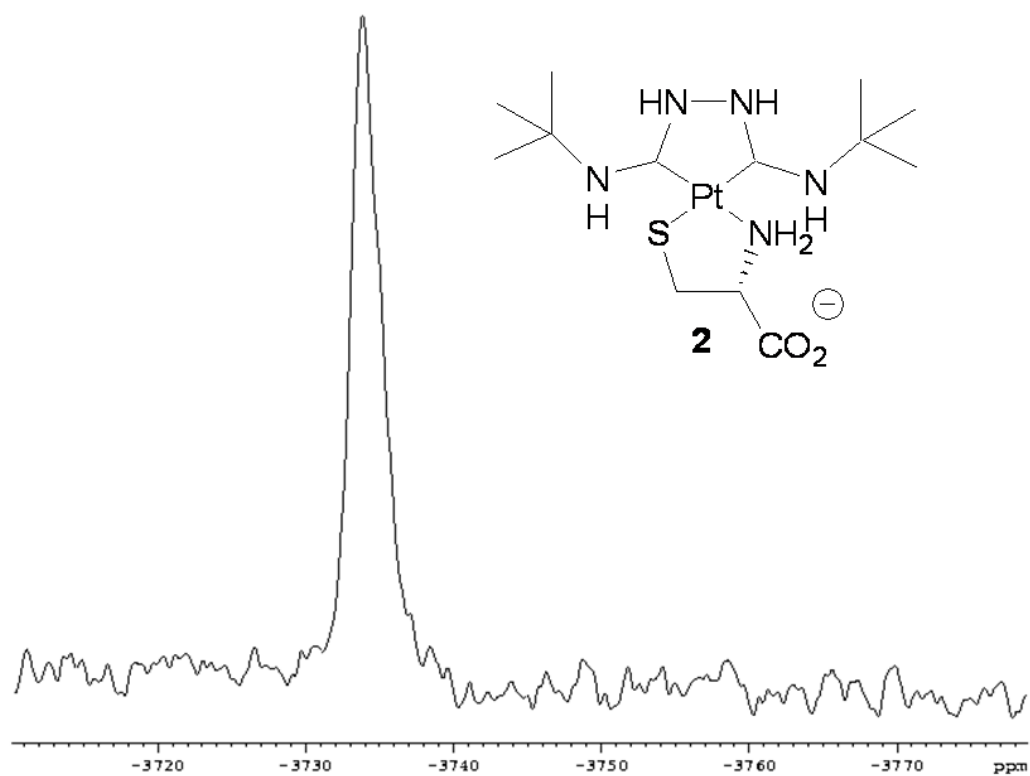


S9. Expansion of the mass spectrum of the product 2.

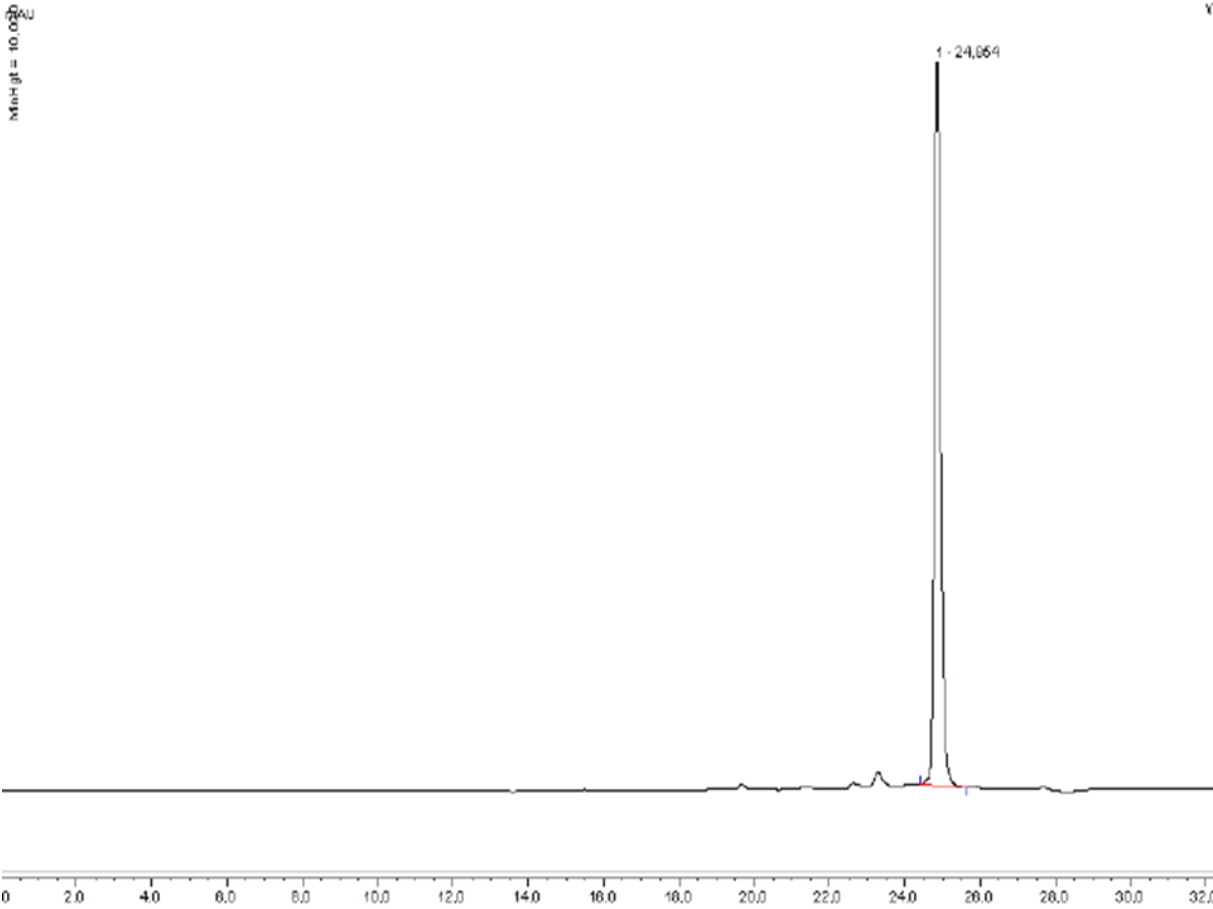


Simulated (up) and observed (down) mass spectrum of 2.

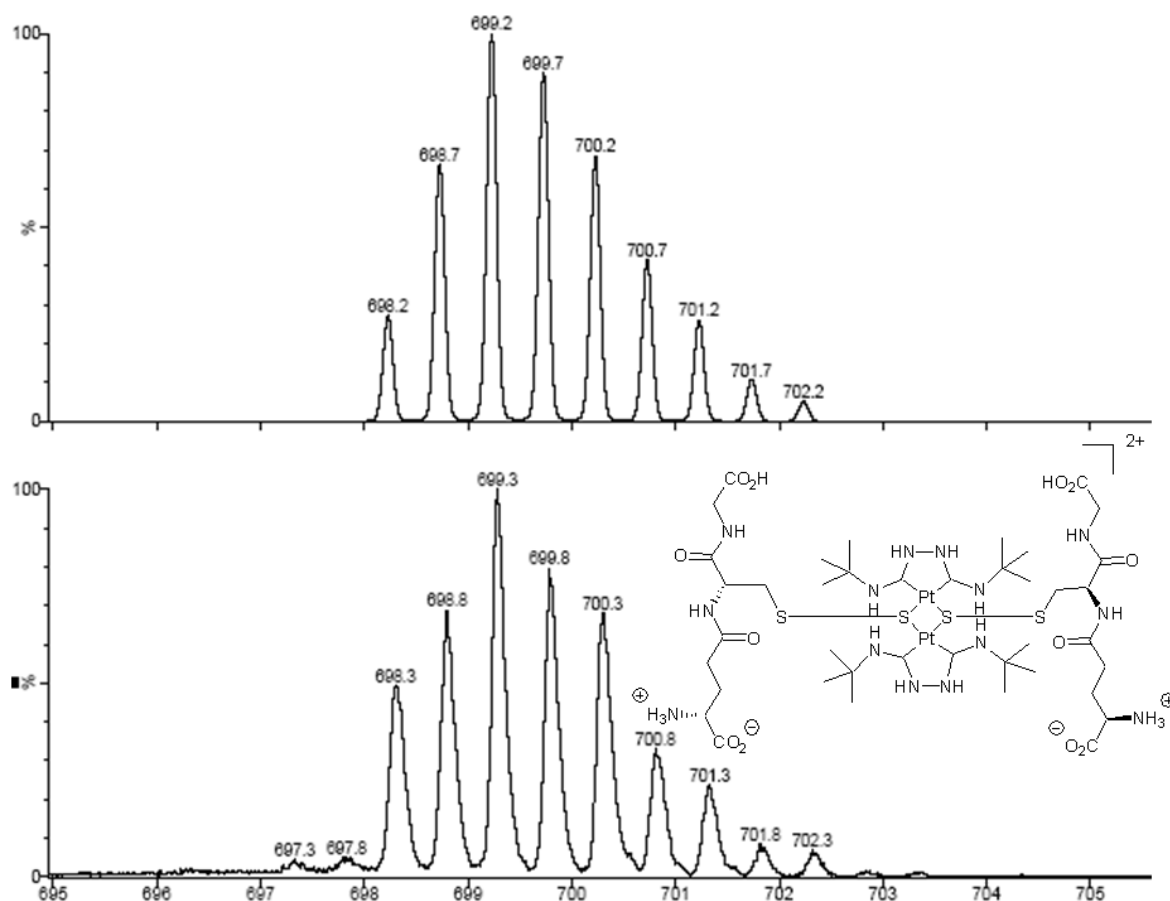
S10. ^{195}Pt of the product 2.



S11. Chromatogram of the product 3.

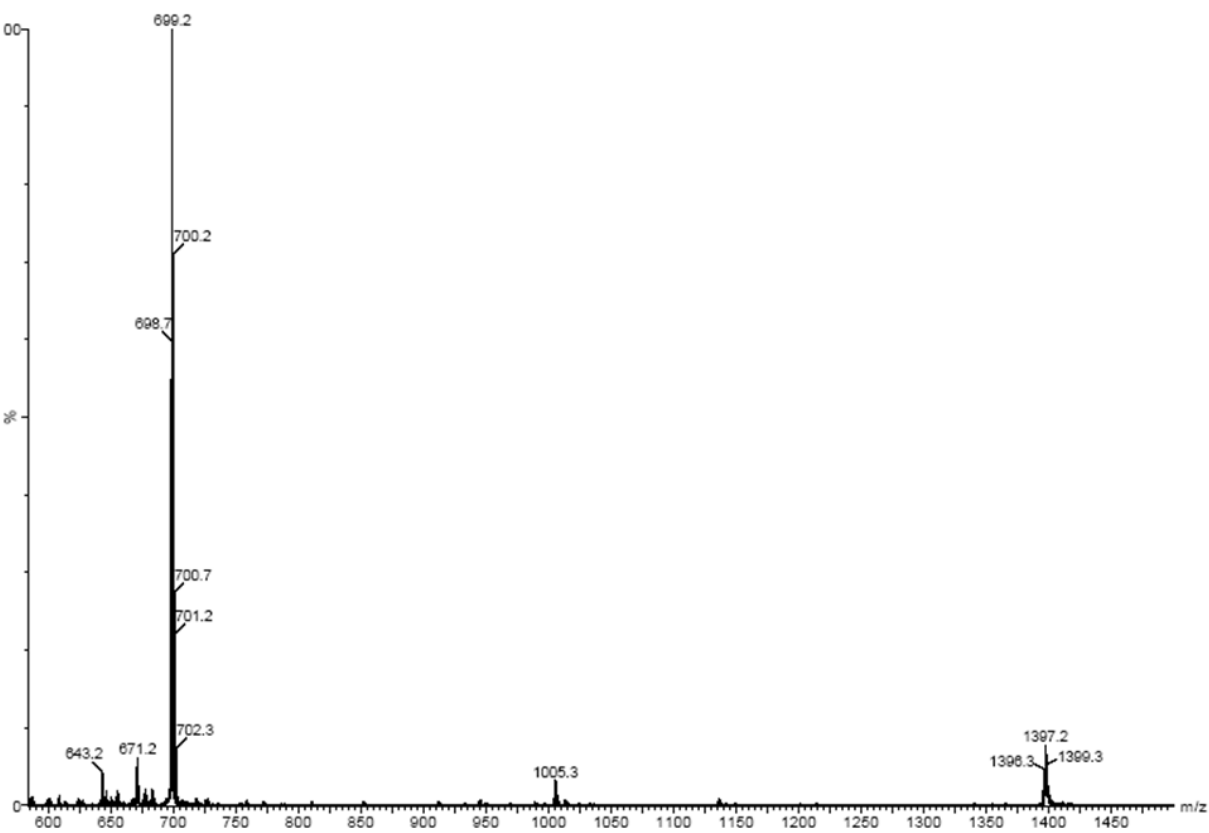


S12. Expansion of the mass spectrum of the product 3 at $m/z = 699$ (100% abundance).

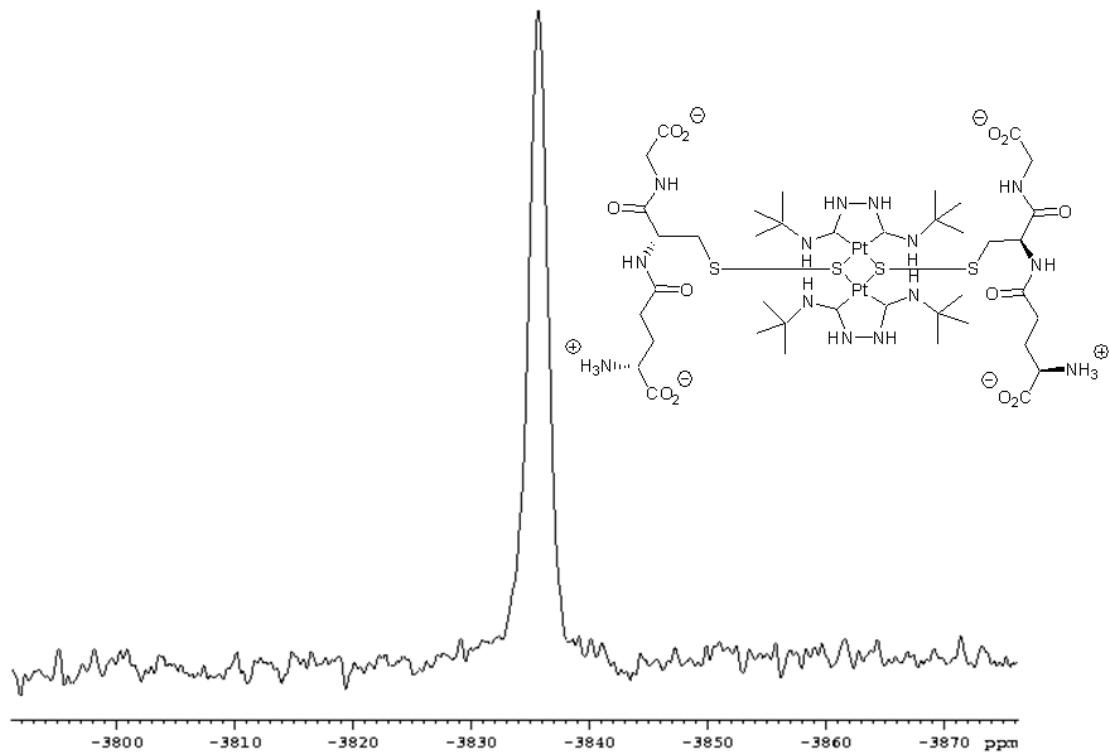


Simulated (up) and observed (down) mass spectrum of 3.

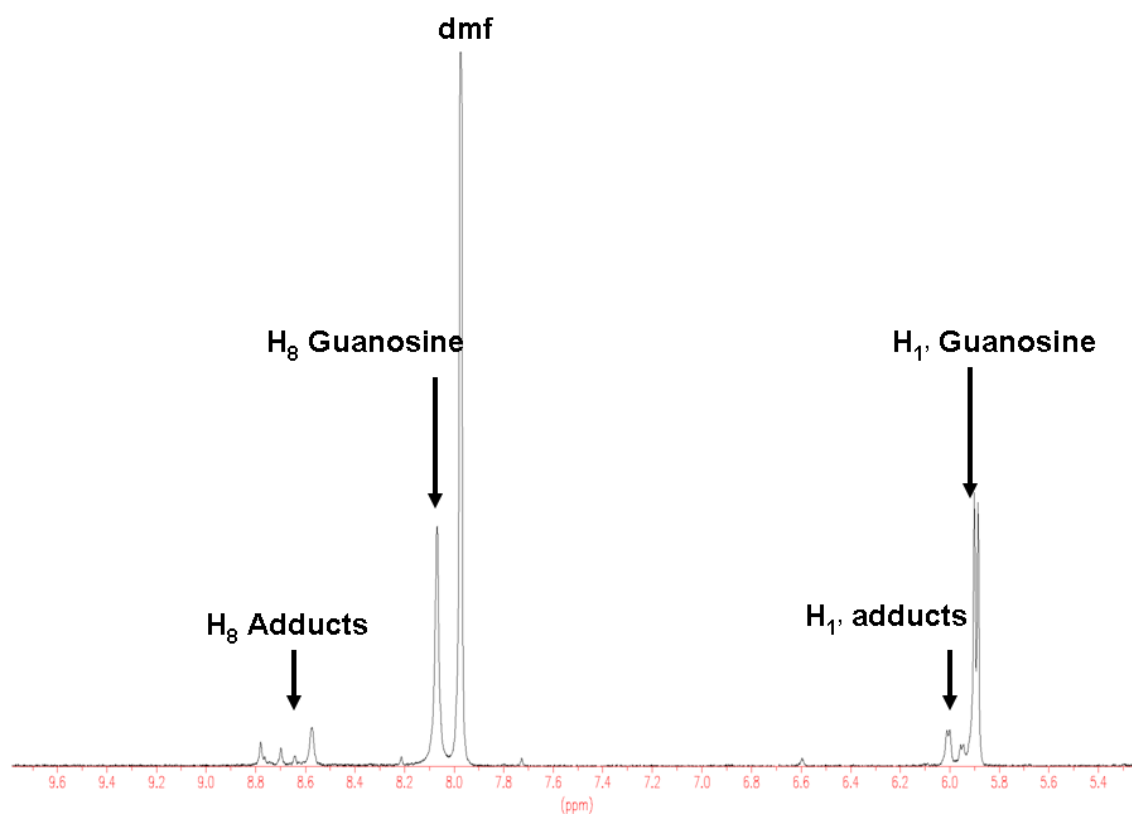
S13. Full mass spectrum of the product 3 (5 and 100% abundance).



S14. ¹⁹⁵Pt of the product 3



S15. Reaction of 1 with guanosine.



ⁱ (a) A. Yoshitha, Wanniarachchi, L. M. Slaughter, *Chem Commun*, 2007, 3294-3296.