

Stereoselective Synthesis of Tetracyclic Indolines via Gold-Catalyzed Cascade Cyclization Reactions

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General Methods. ¹H-NMR spectra were recorded on Varian 200 (200 MHz), Varian 400 (400 MHz) spectrometers. Chemical shifts are reported in ppm from TMS with the solvent resonance as the internal standard (deuteriochloroform: δ 7.27 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = duplet, pd = pseudo duplet, t = triplet, pt = pseudo triplet, q = quartet, pq = pseudo quartet, br = broad, bs = broad singlet, m = multiplet), coupling constants (Hz). ¹³C-NMR spectra were recorded on a Varian 200 (50 MHz), Varian 400 (100 MHz) spectrometers with complete proton decoupling. Chemical shifts are reported in ppm from TMS with the solvent as the internal standard (deuteriochloroform: δ 77.0 ppm). GC-MS spectra were taken by EI ionization at 70 eV on a Hewlett-Packard 5971 with GC injection. They are reported as: *m/z* (rel. intense). LC-electrospray ionization mass spectra were obtained with Agilent Technologies MSD1100 single-quadrupole mass spectrometer. Chromatographic purification was done with 240-400 mesh silica gel. Elemental analyses were carried out by using a EACE 1110 CHNOS analyzer. Supplementary crystallographic data (CCDC 814206-814207) for **2c** and **5a** can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: (+44) 1223-336-033; or e-mail: deposit@ccdc.cam.ac.uk).

Materials. All reactions were carried out under inert gas and under anhydrous conditions, if not further specified. Anhydrous solvents were supplied by Fluka in Sureseal® bottles and used without any further purification. Targeted tosyl-tryptamines were synthesized in variable yields (50-86%), from the commercially available tryptamines, following conventional protocols. tryptamines/TsCl/TEA (1/1.2/2), CH₂Cl₂, rt, 4 h.¹ 2-(1*H*-indol-3-ylmethyl)-malonic acid dialkyl esters were synthesized as previously described.² Propargylic bromide **A** was synthesized from monoprotected butyn-1,4-diol following a conventional protocol. In a three-necked round bottomed flask, equipped with a dropping funnel, under nitrogen atmosphere, 10 mmol of monoprotected butyn-1,4-diol and 11 mmol of NBS were dissolved in 25 ml of DCM. At the same time, 11 mmol of PPh₃ were dissolved in 15 ml of DCM in the adding funnel and then the solution dropped at 0 °C. The suspension was allowed to warm up to r.t. and stirred for 2 hs. DCM was then removed under vacuum. The crude was then treated with 30 ml of pentane and stirred for 1 h. Afterwards the organic layer was filtered with a Gooch funnel and pentane removed under vacuum to afford propargylic bromide **A** with yields greater than 90% and NMR purity > 95%.

Synthesis of indolyl propargylic silyl ethers **1a'-j'**

To a solution of 2-(2,5-disubstituted-1*H*-indole-3-ylmethyl) malonic acid dialkyl ester (1eq) in THF, NaH (60% suspension in mineral oil, 2 eq.) was added at 0 °C. The suspension was stirred for 30 min at 0 °C, then **2** (1.2 eq) was added drop-wise. The reaction was warmed up to room temperature and stirred for 3 h. After extractive work-up (H₂O/EtOAc) the organic layer was separated, dried with Na₂SO₄, filtrated and concentrated under vacuum. The residue

was purified by flash chromatography. Compounds **1f'**, **1g'** and **1i'** were not isolated and deprotected as reaction crudes.

1a': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 77%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.15 (s, 6H); 0.92 (s, 9H); 1.23 (t, J = 7.2 Hz, 6H); 2.83 (s, 2H); 3.57 (s, 2H); 4.12-4.21 (m, 4H); 4.46 (s, 2H); 7.07-7.18 (m, 3H); 7.33 (d, J = 8.4 Hz, 1H); 7.67 (d, J = 7.6 Hz, 1H); 8.08 (s, 1H). LC-MS: 472 (M+1), 494 (M+Na).

1b': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 48%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.02 (s, 6H); 0.96 (s, 9H); 2.86 (s, 2H); 3.66 (s, 2H); 3.67 (s, 6H); 4.43 (t, J = 2.0 Hz, 2H); 6.99 (d, J = 2.0 Hz, 1H); 7.10 (ddd, J₁ = 18.0 Hz, J₂ = 8.0 Hz, J₃ = 1.2 Hz, 2H); 7.28 (d, J = 8.0 Hz, 1H); 7.59 (d, J = 8.0 Hz, 1H); 8.31 (s, 1H). LC-MS: 444 (M+1), 466

(M+Na).

1c': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 56%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.10 (s, 6H); 0.87 (s, 9H); 1.43 (s, 9H); 2.75 (s, 2H); 3.47 (s, 2H); 4.36 (s, 2H); 7.05 (s, 1H); 7.09 (pt, J = 6.8 Hz, 1H); 7.16 (pt, J = 7.2 Hz, 1H); 7.32 (d, J = 8.0 Hz, 1H); 7.79 (d, J = 8.0 Hz, 1H); 8.10 (s, 1H). LC-MS: 550 (M+Na).

1d': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 58%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.13-0.15 (m, 6H); 0.92-0.94 (m, 9H); 1.23-1.27 (m, 6H); 2.83 (t, J = 2.0 Hz, 2H); 3.55 (s, 2H); 4.12-4.21 (m, 4H); 4.38 (s, 2H); 6.99 (dd, J₁ = 8.4 Hz, J₂ = 1.6 Hz, 1H); 7.03 (d, J = 2.4 Hz, 1H); 7.16 (d, J = 8.4 Hz, 1H); 7.43 (d, J = 8.4 Hz, 1H); 7.97 (s, 1H).

LC-MS 486 (M); 509 (M+ Na).

1e': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 60%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.13 (s, 6H); 0.90 (s, 9H); 1.26 (t, J = 7.2 Hz, 6H); 2.85 (s, 2H); 3.54 (s, 2H); 3.86 (s, 3H); 4.17 (q, J = 7.2 Hz, 4H); 4.35 (s, 2H); 6.83 (d, J = 8.8 Hz, 1H); 7.01 (d, J = 2.8 Hz, 1H); 7.19-7.27 (m, 2H); 7.95 (s, 1H). LC-MS: 502 (M+1).

1h': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 46%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.14 (s, 6H); 0.91 (s, 9H); 1.27 (t, J = 7.2 Hz, 6H); 2.81 (s, 2H); 3.53 (s, 2H); 4.17 (q, J = 7.2 Hz, 4H); 4.39 (s, 2H); 7.10 (d, J = 2.4 Hz, 1H); 7.23-7.27 (m, 2H); 7.75 (d, J = 2.0 Hz, 1H); 8.13 (s, 1H). LC-MS: 552 (M+1), 573 (M+Na).

1j': Viscous yellow oil. Flash chromatography (cHex:AcOEt = 8:2). Yield = 30%. ¹H-NMR (400 MHz, CDCl₃) δ: 0.15 (s, 6H); 0.92 (s, 9H); 1.22 (t, J = 7.2 Hz, 6H); 2.40 (s, 3H); 2.81 (s, 3H); 3.51 (s, 2H); 4.15 (q, J = 7.2 Hz, 4H); 4.37 (t, J = 2.0 Hz, 2H); 7.01-7.10 (m, 2H); 7.22 (d, J = 8.4 Hz, 1H); 7.60 (d, J = 7.6 Hz, 1H); 7.80 (s, 1H). LC-MS: 486 (M+1).

Synthesis of indolyl propargylic alcohols 1a-j.

In a one-necked round bottom flask, the desired silyloxy compound **1'** was dissolved in 10 ml of reagent grade THF and then TBAF (1 eq.) was added at the reaction mixture. After stirring for 2-3 h, the reaction was monitored by TLC analysis and judged complete. After extractive work-up (H₂O/AcOEt), the collected organic layers were dried over anhydrous Na₂SO₄ and concentrated under vacuum. The crude was purified by flash-chromatography.

1a. Yellow solid. Flash chromatography (CH₂Cl₂:MeOH = 95:5). Yield = 80%. Mp: 80- 82 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.23 (t, J = 7.2 Hz, 6H); 2.82 (s, 2H); 3.57 (s, 2H); 4.10-4.29 (m, 4H); 4.30 (s, 2H); 7.03 (d, J = 2.0 Hz, 1H); 7.11 (ddd, J₁ = 18.6 Hz, J₂ = 8.0 Hz, J₃ = 1.2 Hz, 2H); 7.32 (d, J = 1.2 Hz, 1H); 7.64 (d, J = 8.0 Hz, 1H); 8.28 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9(2C), 23.1, 27.3, 50.9, 58.2, 61.6(2C), 81.4, 82.0, 109.4, 111.1, 118.7, 119.3, 121.9, 123.6, 128.0, 135.8, 170.3(2C). LC-MS: 340 (M-H₂O); 358 (M+1); 380 (M+Na). Anal. calcd for (C₂₀H₂₃NO₅: 357.40): C, 67.21; H, 6.49; N: 3.92. Found: C, 67.15; H, 6.50; N: 3.88.

1b. Viscous yellow oil. Flash chromatography (DCM:MeOH = 95:5). Yield = 90%. ¹H-NMR (400 MHz, CDCl₃) δ: 2.86 (s, 2H); 3.56 (s, 2H); 3.67 (s, 6H); 4.27 (t, J = 2.0 Hz, 2H); 7.00 (d, J = 2.0 Hz, 1H); 7.10 (ddd, J₁ = 18.0 Hz, J₂ = 8.0 Hz, J₃ = 1.2 Hz, 2H); 7.28 (d, J = 8.0 Hz, 1H); 7.59 (d, J = 8.0 Hz, 1H); 8.31 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 23.1, 27.5, 51.0, 52.7, 52.8(2C), 81.1, 82.1, 109.1, 111.1, 118.6, 119.4, 121.9, 123.6, 127.9, 135.8, 170.7(2C). LC-MS: 330 (M+1); 352 (M+Na). Anal. calcd for (C₁₈H₁₉NO₅: 329.35): C, 65.64; H, 5.81; N: 4.25. Found: C, 65.55; H, 5.70; N: 4.10.

1c. White solid. Flash chromatography (DCM:MeOH = 95:5). Yield = 72%. Mp: 117-119 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.44 (s, 18H); 2.76 (s, 2H); 3.48 (s, 2H); 4.28 (s, 2H); 7.04 (s, 2H); 7.13 (ddd, J₁ = 17.0 Hz, J₂ = 8.0 Hz, J₃ = 1.2 Hz, 2H); 7.31 (dd, J = 8.0 Hz, J₂ = 1.2 Hz, 1H); 7.77 (d, J = 8.0 Hz, 1H); 8.13 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 23.2, 26.7, 27.8(6C), 51.3, 52.8, 81.6(2C), 81.8, 82.2, 110.1, 110.9, 119.3, 121.9(2C), 123.2, 128.4, 135.7, 169.5(2C). LC-MS: 302 (M-CO₂tBu); 436 (M+Na). Anal. calcd for (C₂₄H₃₁NO₅: 413.51): C, 69.71; H, 7.56; N: 3.39. Found: C, 69.68; H, 7.43; N: 3.25.

1d. Yellow amorphous solid. Flash chromatography (DCM:MeOH = 95:5). Yield = 91%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.26 (t, J = 7.2 Hz, 6H); 2.44 (s, 3H); 2.83 (t, J = 2.0 Hz, 2H); 3.55 (s, 2H); 4.12-4.24 (m, 4H); 4.31 (s, 2H); 6.99-7.01 (m, 2H); 7.22 (d, J = 8.4 Hz, 1H); 7.45 (s,

1H); 7.96 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 14.1(2C), 21.6, 23.2, 27.4, 51.3, 58.2, 61.8 (2C), 81.6, 82.2, 108.9, 110.9, 118.6, 123.6, 123.8, 128.4, 128.6, 134.3, 170.5(2C). LC-MS: 372 (M+1), 394 (M+Na). Anal. calcd for (C₂₁H₂₅NO₅; 371.43): C, 67.91; H, 6.78; N: 3.77. Found: C, 67.80; H, 6.66; N: 3.85.

1e. Viscous orange oil. Flash chromatography (DCM:MeOH = 95:5). Yield = 45%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.23 (t, J = 7.2 Hz, 6H); 2.8 (s, 2H); 3.53 (s, 2H); 3.86 (s, 3H); 4.16 (q, J = 7.2 Hz, 4H); 4.30 (s, 2H); 6.83 (dd, J₁ = 8.4 Hz, J₂ = 2.4 Hz, 1H); 6.97 (d, J = 2.4 Hz, 1H); 7.20 (dd, J₁ = 5.8 Hz, J₂ = 2.8 Hz 2H); 8.10 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.1, 30.9, 51.1, 55.8, 58.5, 60.4, 61.6(2C), 81.7, 82.0, 100.9, 109.4, 111.7, 112.2, 124.1, 128.7, 131.0, 154.0, 170.2(2C). LC-MS: 388 (M+1), 410 (M+Na). Anal. calcd for (C₂₁H₂₅NO₆; 387.43): C, 65.10; H, 6.50; N: 3.62. Found: C, 65.01; H, 6.45; N: 3.70.

1f. Viscous yellow oil. Flash chromatography (DCM:MeOH = 95:5). Yield = 51%. ¹H-NMR (400 MHz, CDCl₃) δ: 2.80 (t, J = 2.0 Hz, 2H); 3.54 (s, 2H); 3.74 (s, 6H); 4.33 (t, J = 2.4 Hz, 2H); 7.07 (d, J = 2.4 Hz, 1H); 7.13 (d, J = 2.0 Hz, 1H); 7.26 (d, J = 2.0 Hz, 1H); 7.62 (d, J = 2.0 Hz, 1H); 8.10 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 23.1, 27.3, 51.3, 52.8, 58.0(2C), 81.3, 82.3, 109.4, 112.1, 118.5, 122.4, 124.9, 125.4, 134.2, 159.0, 170.4(2C). LC-MS: 364 (M+1), 386 (M+Na). Anal. calcd for (C₁₈H₁₈ClNO₅; 363.79): C, 59.43; H, 4.99; N: 3.85. Found: C, 59.27; H, 5.06; N: 3.66.

1g. White solid. Flash chromatography (DCM:MeOH = 95:5). Yield = 60%. Mp = 107-109°C ¹H-NMR (400 MHz, CDCl₃) δ: 1.26 (t, J = 7.2 Hz, 6H); 2.79 (t, J = 2.0 Hz, 2H); 4.12 (s, 2H); 4.12-4.24 (m, 4H); 4.33 (dt, J₁ = 4.0 Hz, J₂ = 2.0 Hz, 2H), 7.08 (d, J = 2.4 Hz, 1H), 7.12 (dd, J₁ = 8.8 Hz, J₂ = 2.0 Hz, 1H); 7.14 (s, 1H); 7.26 (d, J = 2.8 Hz, 1H); 7.65 (d, J = 2.0 Hz, 1H); 8.12 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 14.0(2C), 23.0, 27.1, 51.3, 57.9, 61.8(2C), 81.5, 82.2, 112.1, 118.5, 122.3, 124.8, 125.3, 129.2, 132.0, 134.1, 170.1(2C). LC-MS: 392 (M+1), 414 (M+Na). Anal. calcd for (C₂₀H₂₂ClNO₅; 391.85): C, 61.30; H, 5.66; N: 3.57. Found: C, 61.19; H, 5.70; N: 3.40.

1h. Yellow solid. Flash chromatography (DCM:MeOH = 95:5). Yield = 74%. Mp = 124-126 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.27 (t, J = 7.2 Hz, 6H); 2.79 (s, 2H); 3.53 (s, 2H); 4.12-4.22 (m, 4H); 4.33 (s, 2H); 7.05 (d, J = 2.4 Hz, 1H); 7.22 (dd, J₁ = 20.8 Hz, J₂ = 8.0 Hz, 2H); 7.79 (s, 1H); 8.24 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9(2C), 22.9, 27.0, 51.2, 57.8, 61.8(2C), 81.3, 82.2, 109.2, 112.5, 112.7, 121.6, 124.7, 124.7, 129.8, 134.4, 170.1(2C). LC-MS: 436 (M), 437(M+1), 438 (M+2). Anal. calcd for (C₂₀H₂₂BrNO₅; 436.30): C, 55.06; H, 5.08; N: 3.21. Found: C, 55.11; H, 5.12; N: 3.10.

1i. Viscous white oil. Flash chromatography (DCM:MeOH = 95:5). Yield = 60%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.24 (t, J = 7.2 Hz, 6H); 2.81 (t, J = 1.6 Hz, 2H), 3.53 (s, 2H); 4.12-4.24

(m, 4H); 4.31 (s, 2H); 7.04 (d, $J = 2.0$ Hz, 1H); 7.07 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H); 7.33 (d, $J = 1.6$ Hz, 1H); 7.56 (d, $J = 8.0$ Hz, 1H); 8.12 (s, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 14.0(2C), 23.1, 27.2, 51.3, 58.1, 61.7(2C), 81.4, 82.1, 109.9, 111.0, 119.8, 120.3, 124.1, 126.7, 128.0, 136.2, 170.1(2C). LC-MS: 414 (M+Na). Anal. calcd for ($\text{C}_{20}\text{H}_{22}\text{ClNO}_5$; 391.85): C, 61.30; H, 5.66; N, 3.57. Found: C, 61.33; H, 5.75; N, 3.51.

1j. Viscous orange oil. Flash chromatography (DCM:MeOH = 95:5). Yield = 74%. $^1\text{H-NMR}$ (400 MHz CDCl_3) δ : 1.15 (t, $J = 7.2$ Hz, 6H); 2.27 (s, 3H); 2.73 (s, 3H); 3.44 (s, 2H); 4.12 (q, $J = 7.2$ Hz, 4H); 4.22 (s, 2H); 6.96-7.00 (m, 2H); 7.13 (d, $J = 8.0$ Hz, 1H); 7.50 (d, $J = 8.0$ Hz, 1H); 7.85 (s, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 12.0, 13.9(2C), 23.1, 26.8, 51.2, 58.6, 61.6(2C), 81.9, 82.0, 105.5, 110.0, 118.3, 119.1, 121.1, 129.3, 133.6, 135.2, 170.5(2C). LC-MS: 371 (M+1); 394 (M+Na). Anal. calcd for ($\text{C}_{21}\text{H}_{25}\text{NO}_5$; 371.43): C, 67.91; H, 6.78; N, 3.77. Found: C, 67.95; H, 6.70; N, 3.67.

Synthesis of indolyl alcohols 4.

In a Schlenk tube equipped with a magnetic stirring bar, (4-bromo-but-2-ynyloxy)-*tert*-butyl-dimethyl-silane **A** (1.2 eq.), was added to the corresponding *N*-tosyl tryptamine (1 eq.), K_2CO_3 (2 eq.) dissolved in 5 ml of acetone.³ The reaction mixture was stirred at 60 °C for 12 h, and then quenched with water. The aqueous layer was extracted with ethyl acetate (3× 15 ml). The combined organic layers were dried with Na_2SO_4 and the solvents evaporated under vacuum. The residue was purified by flash-chromatography.

4a'. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 80:20). Yield = 50%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 0.05 (s, 6H); 0.87 (s, 9H); 2.40 (s, 3H); 3.08 (t, $J = 7.6$ Hz, 2H); 3.51 (t, $J = 7.6$ Hz, 2H); 4.08 (s, 2H); 4.20 (s, 2H); 7.10-7.24 (m, 5H); 7.37 (d, $J = 8.1$ Hz, 1H); 7.62 (d, $J = 7.6$ Hz, 1H); 7.73 (d, $J = 8.0$ Hz, 2H); 8.01 (s, 1H). LC-MS: 497 (M+1).

4b'. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 80:20). Yield = 35%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 0.05 (s, 6H); 0.87 (s, 9H); 2.41 (s, 3H); 3.02 (t, $J = 7.2$ Hz, 2H); 3.48 (t, $J = 7.2$ Hz, 2H); 4.11 (t, $J = 2.0$ Hz, 2H); 4.19 (t, $J = 2.0$ Hz, 2H); 7.15 (pt, $J = 2.0$ Hz, 2H); 7.26-7.28 (m, 4H); 7.52 (d, $J = 2.0$ Hz, 1H); 7.33 (d, $J = 8.0$ Hz, 1H); 8.07 (s, 1H). LC-MS: 533 (M+1).

4c'. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 90:10). Yield = 37%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 0.048 (s, 6H); 0.88 (s, 9H); 2.40 (s, 3H); 3.31 (t, $J = 8.4$ Hz, 2H); 3.49 (t, $J = 8.4$ Hz, 2H); 3.88 (s, 3H); 3.96 (t, $J = 2.0$

Hz, 2H); 4.18 (t, $J = 2.0$ Hz, 2H); 6.87 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz, 1H); 7.07 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz, 1H); 7.24-7.28 (m, 3H); 7.72 (d, $J = 7.2$ Hz, 2H); 7.92 (br, 1H). LC-MS: 527 (M+1), 549 (M+Na).

4d'. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 91:9). Yield = 44%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 0.048 (s, 6H); 0.88 (s, 9H); 2.40 (s, 3H); 3.06 (t, $J = 7.2$ Hz, 2H); 3.49 (t, $J = 7.2$ Hz, 2H); 3.75 (s, 3H); 4.08 (t, $J = 2.0$ Hz, 2H); 4.21 (t, $J = 2.0$ Hz, 2H); 6.94 (s, 1H); 7.09-7.13 (m, 1H); 7.22-7.31 (m, 5H); 7.60 (d, $J = 8.0$ Hz, 2H); 7.71 (d, $J = 8.0$ Hz, 2H). LC-MS: 533 (M+Na).

The cleavage of the silyl protecting group was performed in analogy to the previously described method for **1'**.

4a. White solid. Flash-chromatography (DCM:MeOH = 95:5). Yield: 70%. Mp = 109-111 °C. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 2.32 (s, 3H); 2.99 (t, $J = 7.6$ Hz, 2H); 3.43 (t, $J = 7.6$ Hz, 2H); 3.90 (s, 2H); 4.08 (s, 2H); 7.00-7.13 (m, 5H); 7.28 (d, $J = 7.2$ Hz, 1H); 7.52 (d, $J = 7.2$ Hz, 1H); 7.64 (dd, $J_1 = 6.6$ Hz, $J_2 = 2.0$ Hz, 2H); 8.01 (s, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 21.5, 24.4, 37.1, 47.1, 50.7, 78.8, 83.7, 111.2, 112.3, 118.6, 119.4, 122.1, 122.3, 127.2, 127.8(2C), 129.4(2C), 135.9, 136.2, 143.5. LC-MS: 383 (M+1), 405 (M+Na). Anal. calcd for ($\text{C}_{21}\text{H}_{22}\text{N}_2\text{O}_3\text{S}$: 382.48): C, 65.95; H, 5.80; N, 7.32. Found: C, 65.88; H, 5.89; N: 7.22.

4b. Pale yellow wax. Flash-chromatography (DCM:MeOH = 95:5). Yield: 60%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 2.41 (s, 3H); 2.99 (t, $J = 8.0$ Hz, 2H); 3.48 (t, $J = 8.0$ Hz, 2H); 4.05 (s, 2H); 4.16 (s, 2H); 7.11 (s, 1H); 7.13 (d, $J = 2.0$ Hz, 1H); 7.26-7.28 (m, 3H); 7.51 (d, $J = 1.6$ Hz, 1H); 7.71 (d, $J = 8.0$ Hz, 2H); 8.27 (s, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 21.6, 24.2, 37.1, 46.9, 50.9, 78.8, 84.1, 112.2, 112.4, 118.2, 122.5, 123.9, 125.3, 127.9 (2C), 128.5, 129.6 (2C), 134.6, 135.9, 143.7. LC-MS: 439 (M+Na). Anal. calcd for ($\text{C}_{21}\text{H}_{21}\text{ClN}_2\text{O}_3\text{S}$: 416.92): C, 60.50; H, 5.08; N, 6.72. Found: C, 60.41; H, 5.00; N: 6.85.

4c. Yellow viscous oil. Flash-chromatography (*c*Hex:EtOAc = 60:40). Yield: 71%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 2.40 (s, 3H); 3.04 (t, $J = 8.0$ Hz, 2H); 3.50 (t, $J = 8.0$ Hz, 2H); 3.87 (s, 3H); 4.00 (s, 2H); 4.19 (s, 2H); 6.87 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz, 1H); 7.06 (d, $J = 2.4$ Hz, 1H); 7.14 (d, $J = 2.4$ Hz, 1H); 7.25-7.27 (m, 3H); 7.73 (d, $J = 8.0$ Hz, 2H); 8.00 (br, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 21.4, 24.4, 37.1, 46.8, 50.6, 56.1, 78.6, 84.0, 101.4, 111.6, 111.9, 111.9, 123.2, 127.7(2C), 129.4(2C), 131.5, 135.9, 143.5, 153.8. LC-MS: 413 (M+1), 435 (M+Na). Anal. calcd for ($\text{C}_{22}\text{H}_{24}\text{N}_2\text{O}_3\text{S}$: 412.50): C, 64.06; H, 5.86; N, 6.79. Found: C, 64.15; H, 5.79; N: 6.71.

4d. White wax. Flash-chromatography (DCM:MeOH = 95:5). Yield: 43%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 2.41 (s, 3H); 3.07 (t, $J = 7.2$ Hz, 2H); 3.50 (t, $J = 7.2$ Hz, 2H); 3.76 (s, 3H); 3.99 (s, 2H); 4.19 (t, $J = 1.6$ Hz, 2H); 6.96 (s, 1H); 7.11 (ddd, $J_1 = 8.0$ Hz, $J_2 = 7.2$ Hz, $J_3 = 1.2$ Hz, 1H); 7.23-7.51 (m, 4H); 7.60 (d, $J = 8.0$ Hz, 1H); 7.73 (dd, $J_1 = 6.8$ Hz, $J_2 = 2.0$ Hz, 1H). $^{13}\text{C-NMR}$

(100 MHz, CDCl₃) δ: 24.3, 29.7, 37.1, 47.2, 50.8, 78.9, 83.7, 109.3, 110.7, 118.7, 118.9, 121.7, 127.0, 127.7, 127.9, 129.3(2C); 136.0, 137.0, 143.5. LC-MS: 397 (M+1). Anal. calcd for (C₂₂H₂₄N₂O₃S: 396.50): C, 66.64; H, 6.10; N, 7.07. Found: C, 66.55; H, 6.01; N, 7.15.

General procedure for the gold-catalyzed diastereoselective domino cyclization.

In a 10 ml two-necked round bottomed flask, substrate **1/4** (0.2 mmol) was dissolved 1.0 ml of dry, followed by the addition of catalyst **3b** (5 mol%). The mixture was stirred overnight at rt, then the crude directly charged into a plug of silica for flash-chromatography purification.

2a. White wax. Flash chromatography (cHex:AcOEt = 7:3). Yield = 74%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.18-1.21 (m, 6H); 2.37 (d, J = 14.4 Hz, 1H); 2.89 (d, J = 14.0 Hz, 1H); 3.03 (d, J = 15.2 Hz, 1H); 3.62 (dd, J₁ = 15.3 Hz, J₂ = 3.6 Hz, 1H); 4.08 (s, 2H); 4.18-4.37 (m, 4H); 4.97 (s, 1H); 5.71 (t, J = 3.6 Hz, 1H); 6.68 (d, J = 8.0 Hz, 1H); 6.73 (pt, J = 8.0 Hz, 1H); 7.06 (dd, J₁ = 14.4 Hz, J₂ = 7.5 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 41.0, 45.1, 52.3, 57.8, 61.1, 61.8, 61.9, 93.4, 109.6, 119.3, 119.6, 123.2, 127.9, 135.0, 137.5, 148.4, 171.5, 171.7. LC-MS: 358 (M+1); 380 (M+23). Anal. calcd for (C₂₀H₂₃NO₅: 357.40): C, 67.21; H, 6.49; N, 3.92. Found: C, 67.40; H, 6.31; N, 3.92.

2b. Pale yellow oil. Flash chromatography (cHex:AcOEt = 6:4). Yield = 75%. ¹H-NMR (400 MHz, CDCl₃) δ: 2.41 (d, J = 14.4 Hz, 1H); 2.87 (d, J = 14.0 Hz, 1H); 3.05 (d, J = 15.6 Hz, 1H); 3.63 (dd, J₁ = 15.6 Hz, J₂ = 3.2 Hz, 1H); 3.71 (s, 3H); 3.76 (s, 3H); 4.08 (dd, J₁ = 5.2 Hz, J₂ = 2.8 Hz, 2H); 4.96 (s, 1H); 5.72 (d, J = 2.8 Hz, 1H); 6.66 (d, J = 0.8 Hz, 1H); 6.74 (t, J = 6.4 Hz, 1H); 7.03-7.07 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 41.0, 45.1, 52.3, 53.0, 53.1, 61.1, 93.3, 109.6, 119.4, 119.6, 123.1, 127.9, 134.8, 137.3, 148.4, 156.3, 171.9, 172.2. LC-MS: 330 (M+1), 352 (M+Na). Anal. calcd for (C₁₈H₁₉NO₅: 329.36): C, 65.64; H, 5.81; N, 4.25. Found: C, 65.60; H, 5.96; N, 4.19.

2c. White solid. Flash chromatography (cHex:AcOEt = 85:15). Yield = 81%.⁴ Mp = 155-157 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.28 (s, 9H); 1.41 (s, 9H); 2.25 (d, J = 14.0 Hz, 1H); 2.82 (d, J = 14.0 Hz, 1H); 2.91 (d, J = 15.2 Hz, 1H); 3.55 (dd, J₁ = 15.2 Hz, J₂ = 2.4 Hz, 1H); 4.08 (s, 2H); 4.95 (s, 1H); 5.67 (t, J = 2.4 Hz, 1H); 6.71 (dt, J₁ = 15.2 Hz, J₂ = 8.0 Hz, 2H); 7.01-7.10 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 27.7(3C), 27.8(3C), 40.8, 44.9, 52.3, 59.1, 61.2, 81.6, 93.4, 109.6, 118.8, 119.6, 123.3, 127.7, 135.2, 137., 170.5, 170.9. LC-MS: 414 (M+1); 436 (M+Na). Anal. calcd for (C₂₄H₃₁NO₅: 412.51): C, 69.71; H, 7.56; N, 3.39. Found: C, 69.75; H, 7.44; N, 3.21.

2d. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 7:3). Yield = 70%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.20 (t, J = 7.2 Hz, 6H); 2.31 (d, J = 14.0 Hz, 1H); 2.89 (d, J = 14.0 Hz, 1H); 3.02 (d, J = 15.6 Hz, 1H); 3.61 (dd, J₁ = 15.6 Hz, J₂ = 2.0 Hz, 1H); 3.73 (s, 3H); 4.13-4.93 (m, 4H); 4.24 (s, 2H); 4.90 (s, 1H); 5.70 (t, J = 2.0 Hz, 1H); 6.61 (d, J = 1.6 Hz, 2H); 6.73 (pt, J = 1.6 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 20.9, 41.1, 44.9, 52.3, 57.9, 61.3, 61.7, 61.8, 93.6, 109.4, 119.3, 123.9, 128.2, 128.9, 135.3, 137.6, 146.0, 171.4, 171.7. LC-MS: 372 (M+1), 394 (M+Na). Anal. calcd for (C₂₁H₂₅NO₅: 371.43): C, 67.91; H, 6.78; N: 3.77. Found: C, 67.83; H, 6.69; N: 3.60.

2e. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 7:3). Yield = 45%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.20 (t, J = 7.2 Hz, 6H); 2.31 (d, J = 14.0 Hz, 1H); 2.89 (d, J = 14.0 Hz, 1H); 3.02 (d, J = 15.6 Hz, 1H); 3.61 (dd, J₁ = 15.6 Hz, J₂ = 2.0 Hz, 1H); 3.73 (s, 3H); 4.12-4.22 (m, 4H); 4.24 (s, 2H); 4.90 (s, 1H); 5.70 (t, J = 2.0 Hz, 1H); 6.61 (d, J = 1.6 Hz, 2H); 6.73 (pt, J = 1.6 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 40.9, 44.6, 52.8, 55.9, 57.7, 61.5, 61.8, 61.9, 93.9, 110.1, 110.4, 112.7, 119.3, 136.3, 137.1, 142.0, 153.9, 171.4, 171.7. LC-MS: 388 (M+1). Anal. calcd for (C₂₁H₂₅NO₆: 387.43): C, 65.10; H, 6.50; N: 3.62. Found: C, 65.23; H, 6.41; N: 3.78.

2f. White wax. Flash chromatography (*c*Hex:AcOEt = 6:4). Yield = 75%. ¹H-NMR (400 MHz, CDCl₃) δ: 2.38 (d, J = 14.0 Hz, 1H); 2.85 (d, J = 14.0 Hz, 1H); 3.08 (d, J = 15.6 Hz, 1H); 3.57 (dt, J₁ = 16.0 Hz, J₂ = 2.8 Hz, 1H); 3.76 (s, 6H); 4.08 (t, J = 2.8 Hz, 2H); 4.93 (s, 1H); 5.74 (t, J = 2.8 Hz, 1H); 6.59 (d, J = 8.0 Hz, 1H); 7.01 (d, J = 10.4 Hz, 1H); 7.27 (d, J = 3.2 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 30.9, 40.9, 44.9, 52.5, 53.1, 61.4, 65.8, 93.5, 110.4, 119.9, 123.6, 124.1, 127.7(2C), 136.7, 147.9, 171.7, 172.2. LC-MS: 364 (M+1); 386 (M+Na). Anal. calcd for (C₁₈H₁₈ClNO₅: 363.79): C, 59.43; H, 4.88; N: 3.85. Found: C, 59.33; H, 4.97; N: 3.73.

2g. Viscous yellow oil. Flash chromatography (*c*Hex:AcOEt = 7:3). Yield = 75%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.27 (t, J = 7.2 Hz, 6H); 2.36 (d, J = 14.4 Hz, 1H); 2.87 (d, J = 14.0 Hz, 1H); 3.06 (d, J = 15.6 Hz, 1H); 3.56 (dd, J₁ = 15.6 Hz, J₂ = 2.8 Hz, 1H); 4.07-4.09 (m, 2H); 4.17-4.25 (m, 4H); 4.93 (s, 1H); 5.73 (dd, J₁ = 5.4 Hz, J₂ = 2.4 Hz, 1H); 6.58-6.60 (m, 1H); 6.99-7.02 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 40.9, 44.8, 52.5, 57.8, 61.4, 61.9, 62.0, 93.5, 110.4(2C), 119.8, 123.6, 124.1, 127.7, 136.9, 147.1, 171.2, 171.6. LC-MS: 414 (M +Na). Anal. calcd for (C₂₀H₂₂ClNO₅: 391.85): C, 61.30; H, 5.66; N: 3.57. Found: C, 61.21; H, 5.54; N: 3.45.

2h. White solid. Flash chromatography (*c*Hex:AcOEt = 7:3). Yield = 75%. Mp = 155-157 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.24 (t, J = 7.2 Hz, 6H); 2.36 (d, J = 14.4 Hz, 1H); 2.86 (d, J = 14.4 Hz, 1H); 3.06 (d, J = 15.6 Hz, 1H); 3.55 (dd, J₁ = 15.6 Hz, J₂ = 2.4 Hz, 1H); 4.08 (s, 2H); 4.17-4.26 (m, 4H); 4.93 (s, 1H); 5.73 (s, 1H); 6.55 (dd, J₁ = 6.4 Hz, J₂ = 2.0 Hz, 1H); 7.15 (dd, J₁ = 7.2 Hz, J₂ = 2.0 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 40.9, 42.7, 44.8, 52.5, 57.8, 61.3, 61.9, 93.4, 111.0, 119.8,

126.4, 130.5, 136.9, 137.2, 147.6, 169.2, 171.2, 171.6. LC-MS: 436 (M); 437 (M+1); 438 (M+2). Anal. calcd for (C₂₀H₂₂BrNO₅: 436.30): C, 55.06; H, 5.08; N: 3.21. Found: C, 55.17; H, 4.99; N: 3.12.

2i. Viscous yellow oil. Flash chromatography (cHex:AcOEt = 6:4). Yield = 78%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.27 (t, J = 7.2 Hz, 6H); 2.33 (d, J = 14.0 Hz, 1H); 2.87 (d, J = 14.8 Hz, 1H); 3.02 (d, J = 15.6 Hz, 1H); 3.58 (dd, J₁ = 15.8 Hz, J₂ = 2.8 Hz, 1H); 4.06 (s, 2H); 4.12-4.22 (m, 4H); 4.97 (s, 1H); 5.72 (d, J = 2.8 Hz, 1H); 6.65 (s, 1H); 6.68 (d, J = 8.0 Hz, 1H); 6.96 (d, J = 8.0 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 14.0, 14.1, 40.8, 45.0, 51.8, 57.7, 61.1(2C), 61.9, 93.5, 109.8, 119.3, 119.5, 124.0, 133.6, 135.6, 137.2, 149.7, 171.4, 171.6. LC-MS: 414 (M+Na). Anal. calcd for (C₂₀H₂₂ClNO₅: 391.85): C, 61.30; H, 5.66; N: 3.57. Found: C, 61.35; H, 5.73; N: 3.46.

2j. Viscous yellow oil. Flash chromatography (cHex:AcOEt = 7:3). Yield = 58%. ¹H-NMR (400 MHz, CDCl₃) δ: 1.20 (t, J = 7.2 Hz, 6H); 1.53 (s, 3H); 2.36 (d, J = 14.0 Hz, 1H); 2.94 (d, J = 14.0 Hz, 1H); 3.02 (d, J = 15.6 Hz, 1H); 3.62 (d, J = 16.0 Hz, 1H); 4.03 (d, J = 16.0 Hz, 1H); 4.13-4.26 (m, 5H); 5.62 (pt, J = 3.8 Hz, 1H); 6.64 (d, J = 8.0 Hz, 1H); 6.70 (pt, J = 7.2 Hz, 1H); 7.01-7.08 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 13.9, 14.0, 18.8, 40.7, 42.8, 54.6, 57.1, 60.9, 61.8(2C), 95.2, 109.7, 117.5, 119.5, 123.2, 127.6, 136.8, 137.4, 147.4, 171.7, 172.0. LC-MS: 371 (M+1); 394 (M+Na). Anal. calcd for (C₂₁H₂₅NO₅: 371.43): C, 67.91; H, 6.78; N: 3.77. Found: C, 67.85; H, 6.59; N: 3.71.

5a. White solid. Flash-chromatography (cHex:AcOEt = 7:3). Yield: 76%. Mp = 109-111 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.85 (dt, J₁ = 12.6 Hz, J₂ = 3.6 Hz, 1H); 2.00 (td, J₁ = 12.0 Hz, J₂ = 4.0 Hz, 1H); 2.34 (s, 3H); 3.27 (dt, J₁ = 12.6 Hz, J₂ = 3.6 Hz, 1H); 3.53 (td, J₁ = 12.0 Hz, J₂ = 4.0 Hz, 1H); 3.77 (dd, J₁ = 16.4 Hz, J₂ = 4.0 Hz, 1H); 4.10 (dd, J₁ = 16.4 Hz, J₂ = 5.6 Hz, 1H); 4.21 (s, 2H); 5.05 (s, 1H); 5.68 (t, J = 5.6 Hz, 1H); 6.61 (d, J = 7.6 Hz, 1H); 6.74 (pt, J = 7.6 Hz, 1H); 7.06 (pt, 8.0 Hz, 1H); 7.19-7.28 (m, 2H); 7.41 (d, J = 7.6 Hz, 1H); 7.63(d, J = 8.0 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 21.5, 29.7, 32.6, 45.5, 59.1, 72.2, 100.9, 109.7, 117.5, 119.6, 126.6, 127.3(2C), 128.9, 129.4, 129.8(2C), 136.1, 143.5, 146.5, 148.5. LC-MS: 383 (M+1), 405 (M+Na). Anal. calcd for (C₂₁H₂₂N₂O₃S: 382.48): C, 65.95; H, 5.80; N: 7.32. Found: C, 65.88; H, 5.69; N: 7.22.

5b. White solid. Flash-chromatography (cHex:AcOEt = 7:3). Yield: 60.^[4] Mp = 100-103 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.94 (dt, J₁ = 14.0 Hz, J₂ = 3.6 Hz, 1H); 2.07 (dd, J₁ = 14.0 Hz, J₂ = 4.8 Hz, 1H); 2.46 (s, 3H); 3.38 (dt, J₁ = 9.6 Hz, J₂ = 4.0 Hz, 1H); 3.58 (dt, J₁ = 12.8 Hz, J₂ = 2.8 Hz, 1H); 3.88 (dd, J₁ = 15.2 Hz, J₂ = 4.8 Hz, 1H); 4.14-4.19 (m, 1H); 4.29-3.30 (m, 2H); 4.73 (s, 1H); 5.15 (s, 1H); 5.77-5.80 (m, 1H); 6.60 (d, J = 8.0 Hz, 1H); 7.09 (dd, J₁ = 8.0 Hz, J₂ = 1.6 Hz, 1H); 7.35 (d, J = 7.6 Hz, 2H); 7.46 (d, J = 2.0 Hz, 1H); 7.72 (d, J = 7.6 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 21.5, 23.3, 32.6, 45.4, 45.6, 55.1, 59.1, 72.2, 73.5, 79.4, 86.7, 101.2, 110.5, 118.3, 126.7, 127.3(2C), 128.8, 129.8(2C), 131.4, 143.6, 147.1. LC-MS: 417 (M+1), 439 (M+Na). Anal. calcd for (C₂₁H₂₁ClN₂O₃S: 416.92): C, 60.50; H, 5.08; N: 6.72. Found: C, 60.41; H, 5.19; N: 6.62.

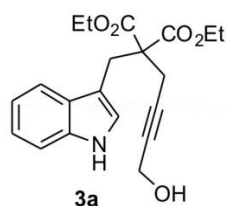
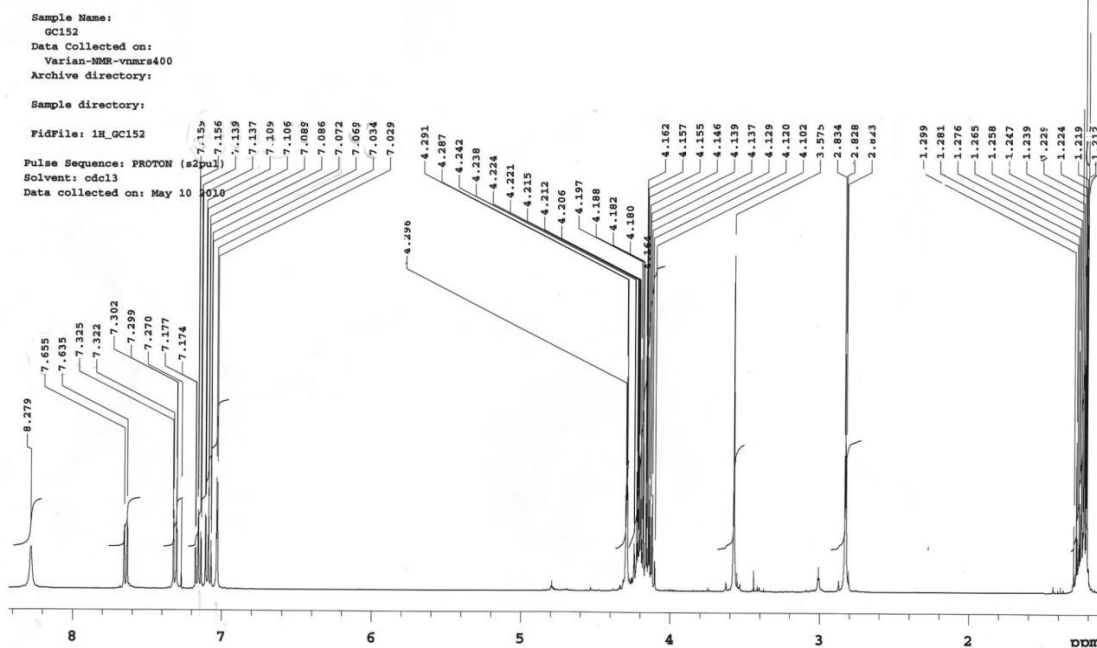
5c. White solid. Flash-chromatography (cHex:AcOEt = 65:35). Yield: 52%.^[4] Mp = 72-74 °C (mixture of isomers). ¹H-NMR (400 MHz, CDCl₃) δ: 1.99 (dq, J₁ = 14.0 Hz, J₂ = 3.2 Hz, 1H); 2.01 (dd, J₁ = 7.2 Hz, J₂ = 5.2 Hz, 1H); 2.46 (s, 3H); 3.05-3.15 (m, 1H); 3.19 (dp, J₁ = 14.0 Hz, J₂ = 7.2 Hz, 1H); 3.51 (dd, J₁ = 8.0 Hz, J₂ = 5.2 Hz, 1H); 3.68-3.72 (m, 1H); 3.82 (s, 3H); 4.21 (m, 2H); 5.23 (s, 1H); 5.67-5.69 (m, 1H); 6.61 (d, J = 8.8 Hz, 1H); 6.67 (dd, J₁ = 8.8 Hz, J₂ = 6.4 Hz, 1H); 6.85 (s, 1H); 7.35 (d, J = 7.6 Hz, 2H); 7.70 (d, J = 7.6 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 21.5, 26.9, 32.1, 45.3, 45.4, 56.1, 72.2, 101.6, 110.6, 112.3, 115.2, 117.9, 127.4(2C), 129.6, 129.8(2C), 142.3, 143.6, 146.1, 154.1. LC-MS: 413 (M+1), 435 (M+Na). Anal. calcd for (C₂₂H₂₄N₂O₃S: 412.50): C, 64.06; H, 5.86; N, 6.79. Found: C, 64.00; H, 5.85; N, 6.85.

5d. White solid. Flash-chromatography (cHex:AcOEt = 8:2). Yield of **5d**: 53. Mp = 160-162 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 1.85 (dt, J₁ = 14.0 Hz, J₂ = 3.6 Hz, 1H); 1.98 (td, J₁ = 11.6 Hz, J₂ = 3.6 Hz, 1H); 2.38 (s, 3H); 2.88 (s, 3H); 3.26-3.29 (m, 1H); 3.54 (td, J₁ = 12.4 Hz, J₂ = 4.0 Hz, 1H); 3.78 (dd, J₁ = 16.4 Hz, J₂ = 4.0 Hz, 1H); 4.09-4.13 (m, 2H); 4.17-4.21 (m, 1H); 4.92 (s, 1H); 5.66 (t, J = 4.8 Hz, 1H); 6.41 (d, J = 7.6 Hz, 1H); 6.67 (pt, J = 7.6 Hz, 1H); 7.10 (t, J = 7.6 Hz, 1H); 7.27 (d, J = 7.6 Hz, 1H); 7.35 (d, J = 6.4 Hz, 1H); 7.63 (d, J = 8.0 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 24.3, 26.7, 37.1, 47.2, 50.8, 54.1, 78.9, 83.7, 109.3, 110.7, 118.6, 118.9, 121.7, 127.0, 127.6, 127.9(2C), 129.3(2C), 136.7, 140.0, 143.4. LC-MS: 397 (M+1). Anal. calcd for (C₂₂H₂₄N₂O₃S: 396.50): C, 66.64; H, 6.10; N, 7.07. Found: C, 66.41; H, 6.00; N, 7.01.

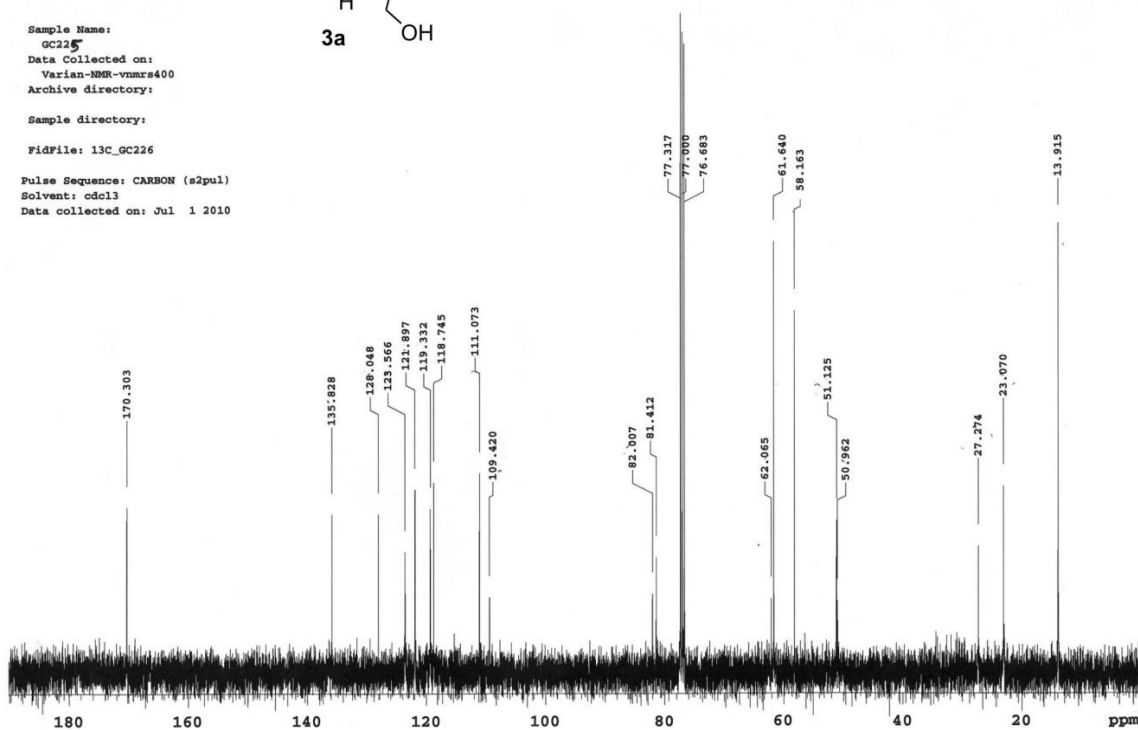
X-ray data.

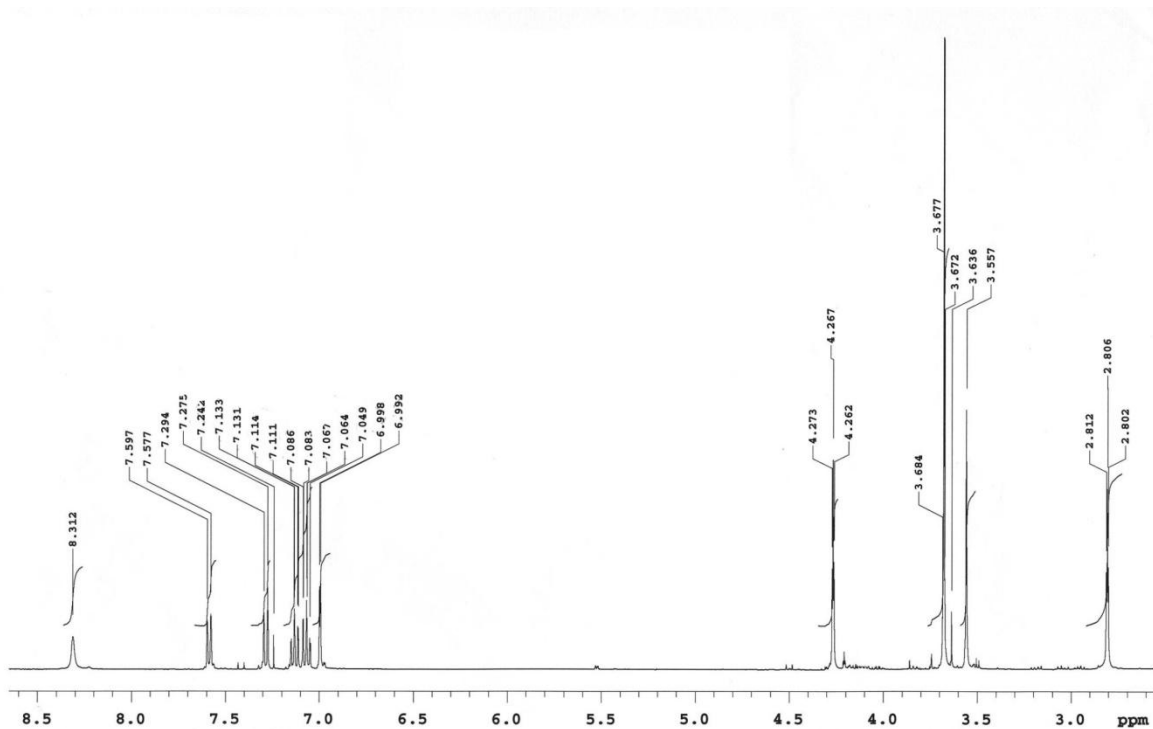
Crystal data for **2c**: C₂₄H₃₁NO₅, *M* = 413.50, colorless plate, 0.30 x 0.20 x 0.15 mm, monoclinic, space group *C2* (No. 5), *a* = 17.796(6), *b* = 6.252(2), *c* = 20.307(6) Å, β = 90.601(4)°, *V* = 2259(1) Å³, *Z* = 4, *D_c* = 1.216 g/cm³, *F*(000) = 888, CCD area detector Smart ApexII, MoK_α radiation, λ = 0.71073 Å, *T* = 293(2)K, 2θ_{max} = 50.8°, 10746 reflections collected, 4162 unique (*R*_{int} = 0.0321). The structure was solved and refined using the programs SIR-97⁵ and SHELXTL,⁶ respectively. Final *Goof* = 1.029, *RI* = 0.0381, *wR2* = 0.0897, *R* indices based on 3442 reflections with *I* > 2σ(*I*) (refinement on *F*²), 278 parameters, 1 restraint. Lp and absorption corrections applied, μ = 0.085 mm⁻¹.

Crystal data for **5a**: C₂₁H₂₂NO₅S, *M* = 382.47, colorless plate, 0.25 x 0.15 x 0.07 mm, monoclinic, space group *C2/c* (No. 15), *a* = 16.779(4), *b* = 13.615(4), *c* = 16.919(4) Å, β = 100.050(5)°, *V* = 3806(2) Å³, *Z* = 8, *D_c* = 1.335 g/cm³, *F*(000) = 1616, CCD area detector Smart ApexII, MoK_α radiation, λ = 0.71073 Å, *T* = 293(2)K, 2θ_{max} = 49.76°, 16998 reflections collected, 3298 unique (*R*_{int} = 0.1056). The structure was solved and refined using the programs SIR-97⁵ and SHELXTL,⁶ respectively. Final *Goof* = 1.087, *RI* = 0.0949, *wR2* = 0.1530, *R* indices based on 1363 reflections with *I* > 2σ(*I*) (refinement on *F*²), 245 parameters, 1 restraint. Lp and absorption corrections applied, μ = 0.194 mm⁻¹.



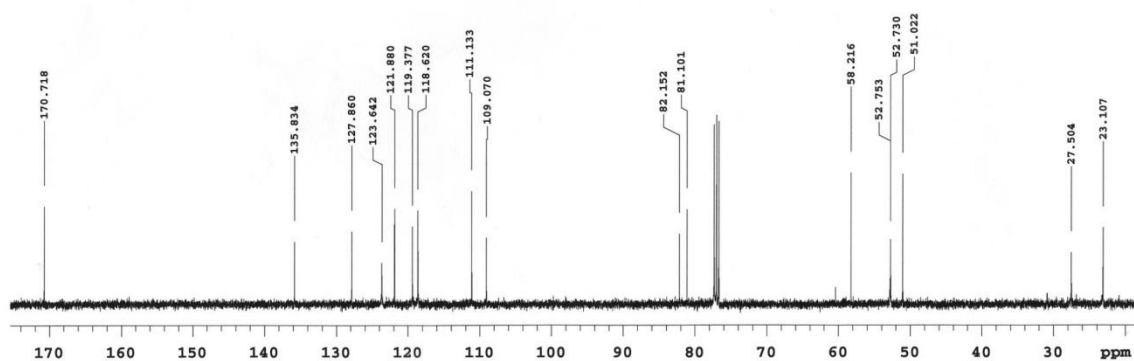
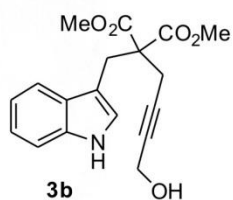
Sample Name: GC225
Data Collected on: Varian-NMR-vnmr400
Archive directory:
Sample directory:
FidFile: 13C_GC225
Pulse Sequence: CARBON (s2pul)
Solvent: cdcl3
Data collected on: Jul 1 2010

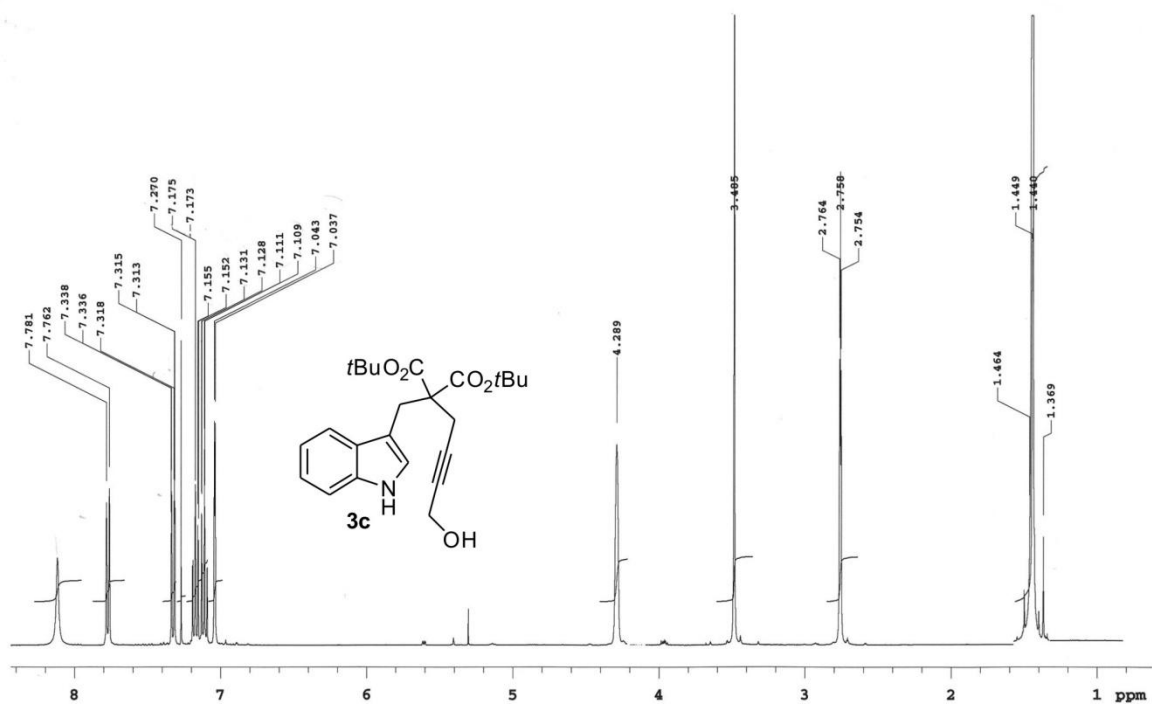




STANDARD CARBON PARAMETERS

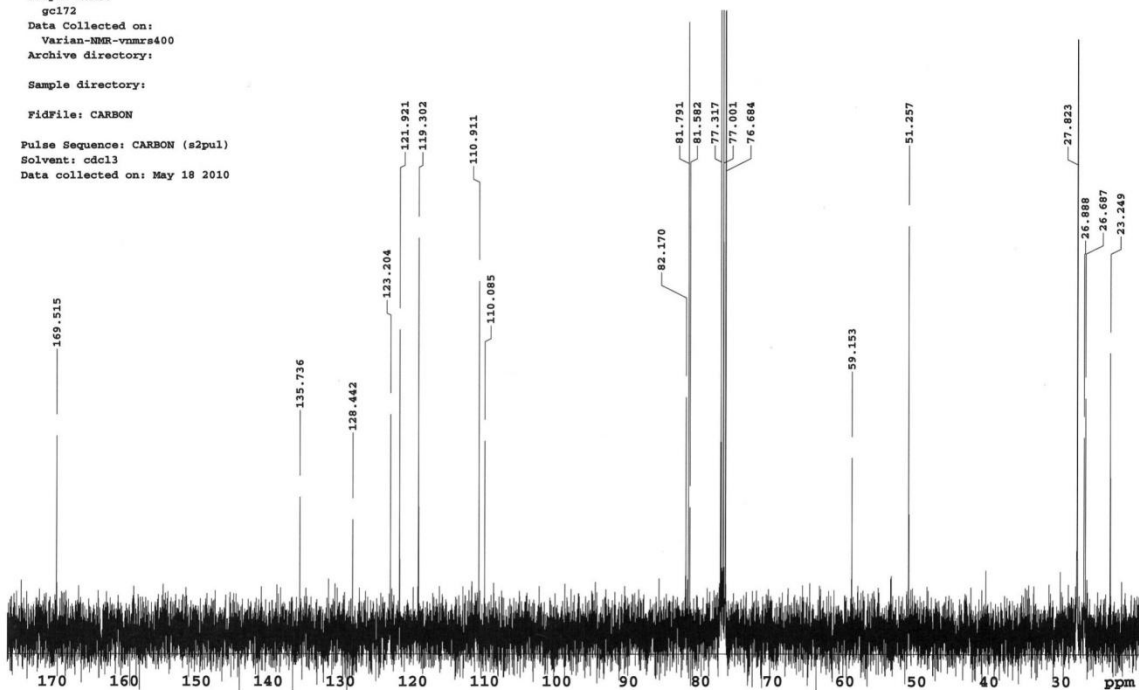
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Pulse Sequence: CARBON (s2pul)
Solvent: cdcl3
Data collected on: Jun 7 2010

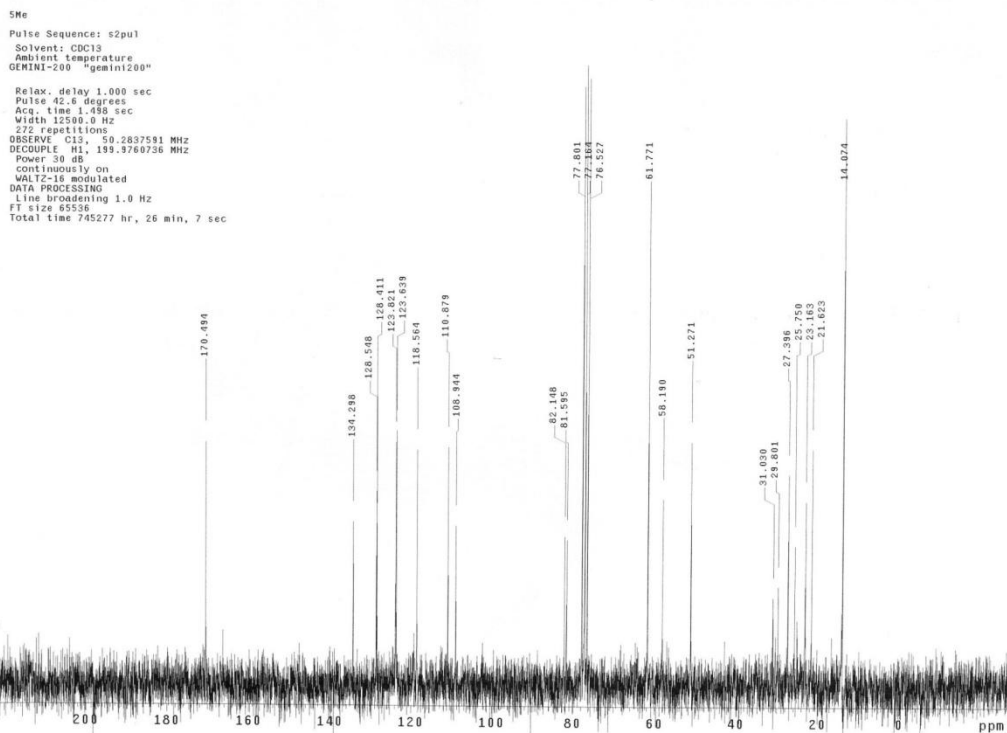
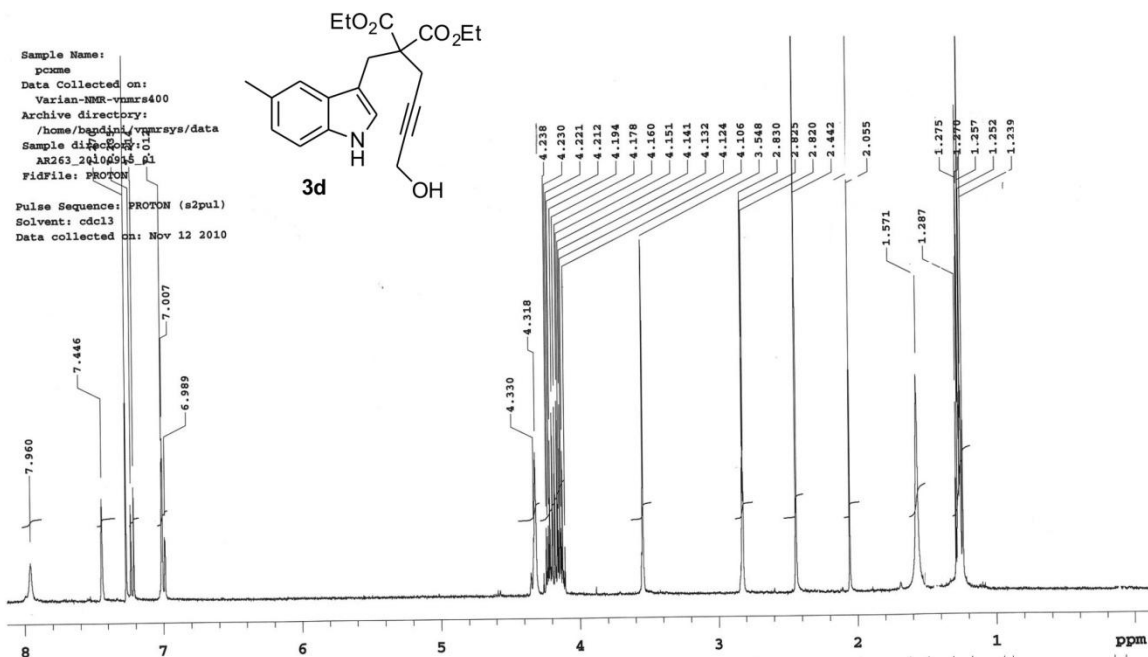


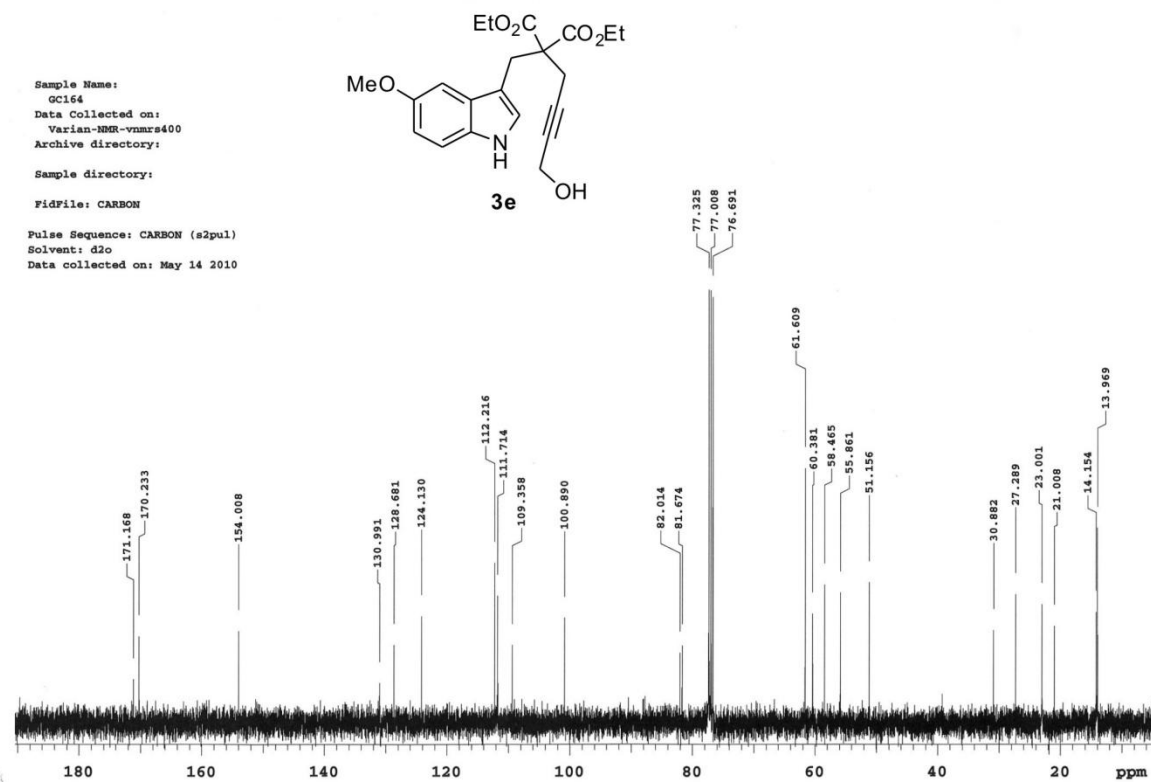
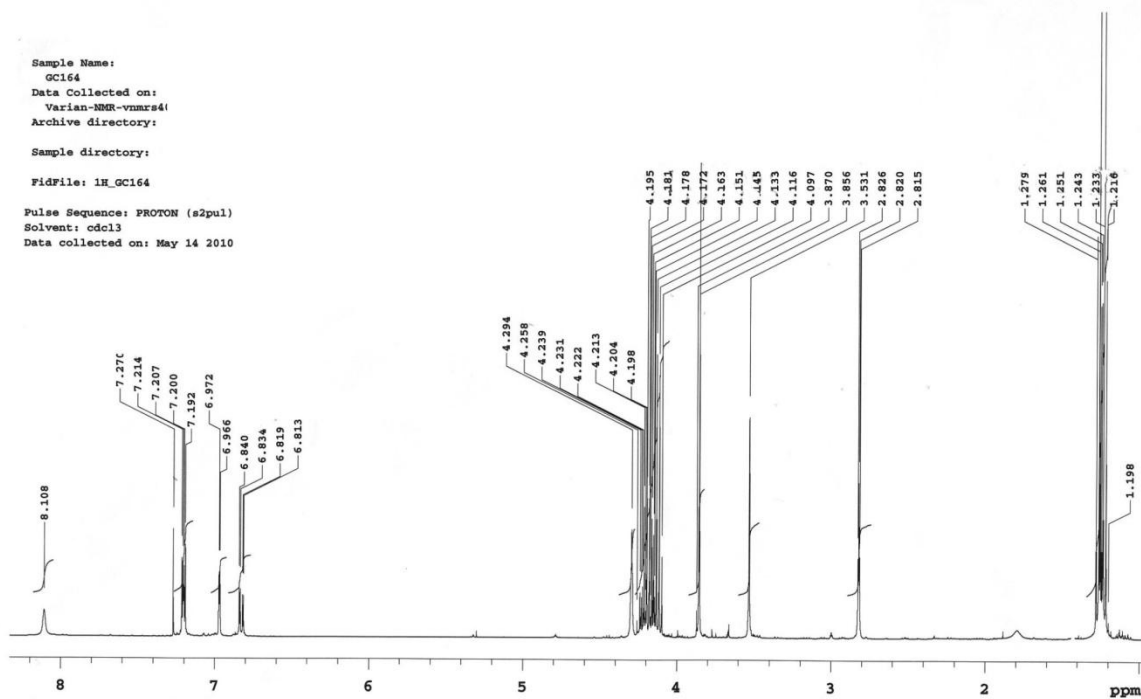


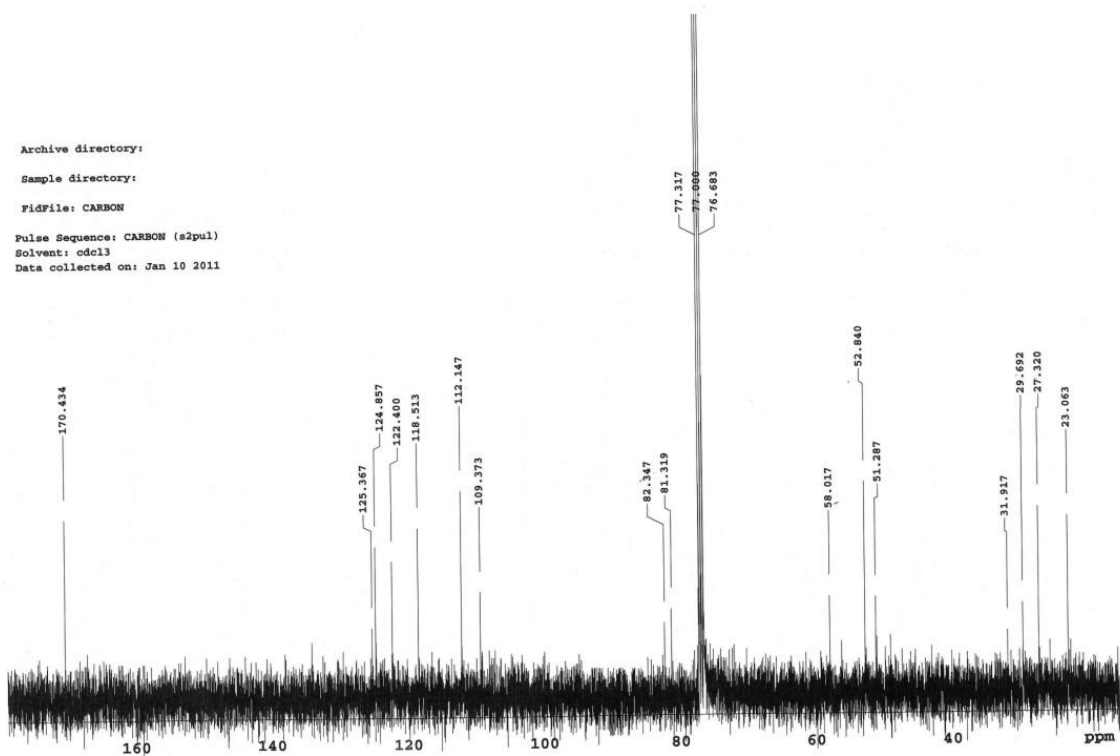
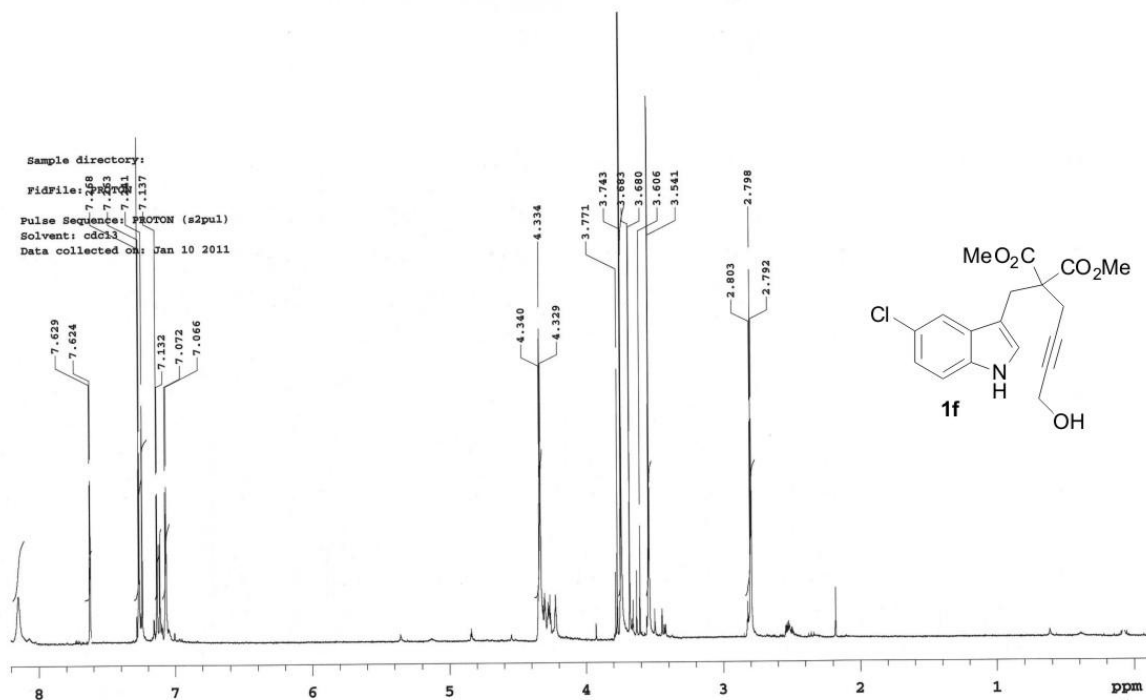
PC9

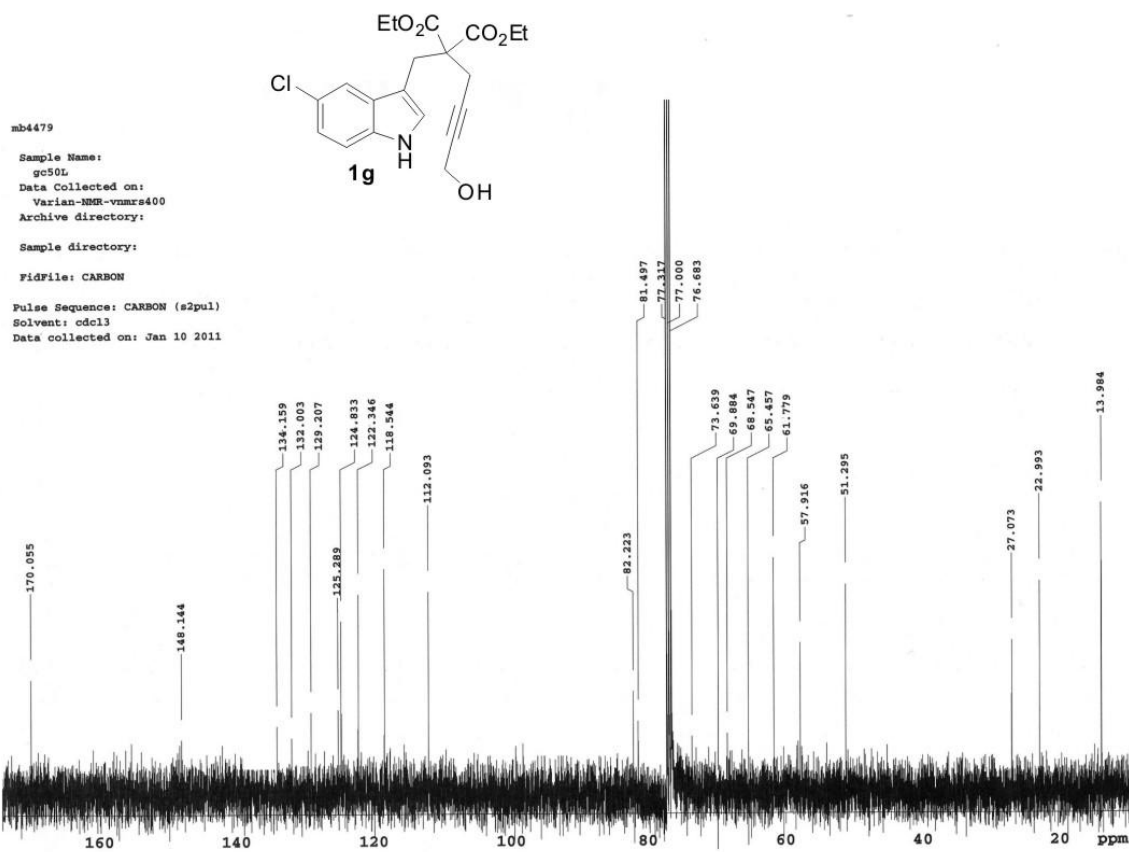
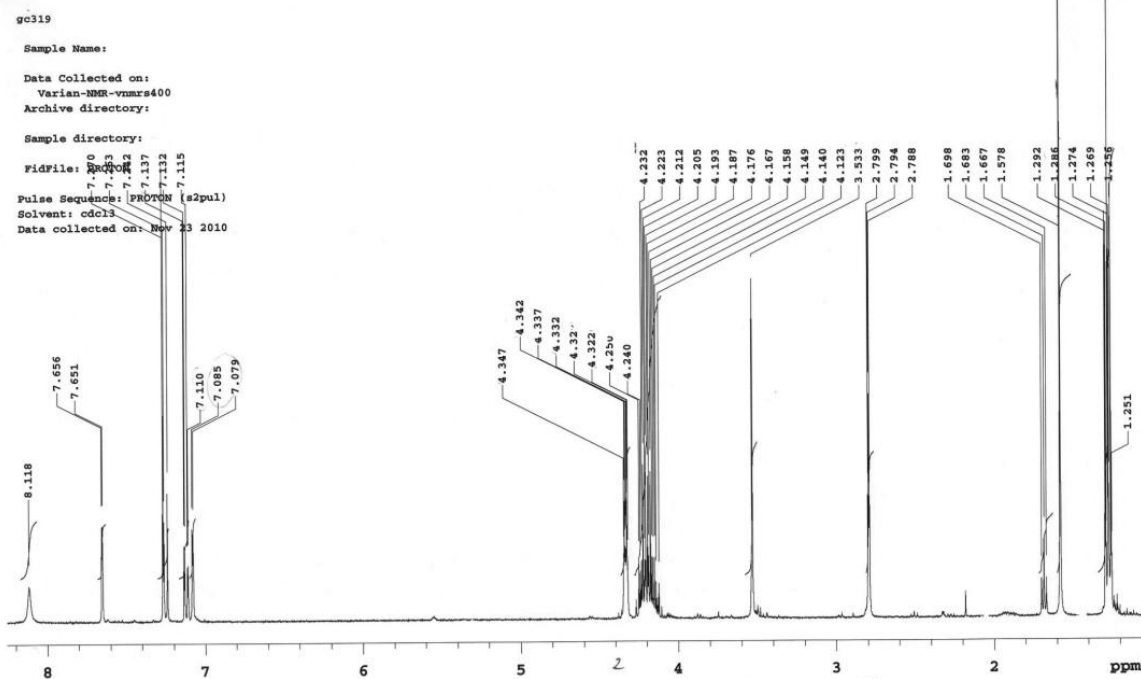
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gc172
Data Collected on:
Varian-NMR-vnmrs400
Archive directory:
Sample directory:
Fidfile: CARBON
Pulse Sequence: CARBON (s2pul)
Solvent: cdcl3
Data collected on: May 18 2010

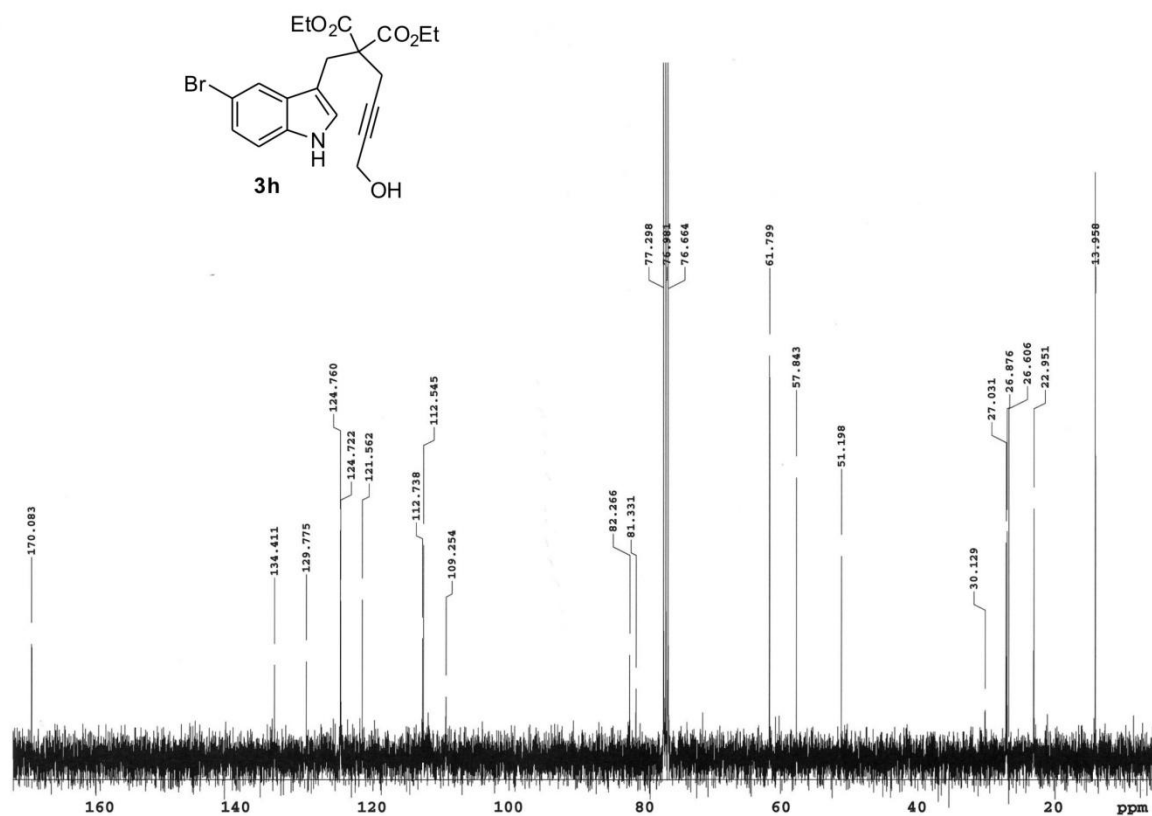
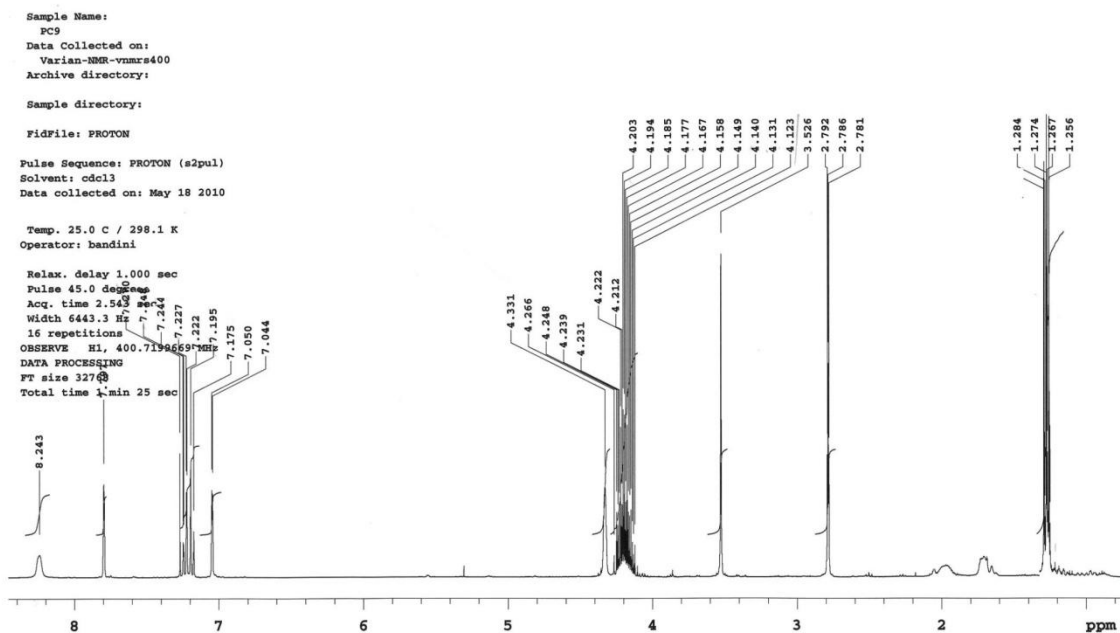


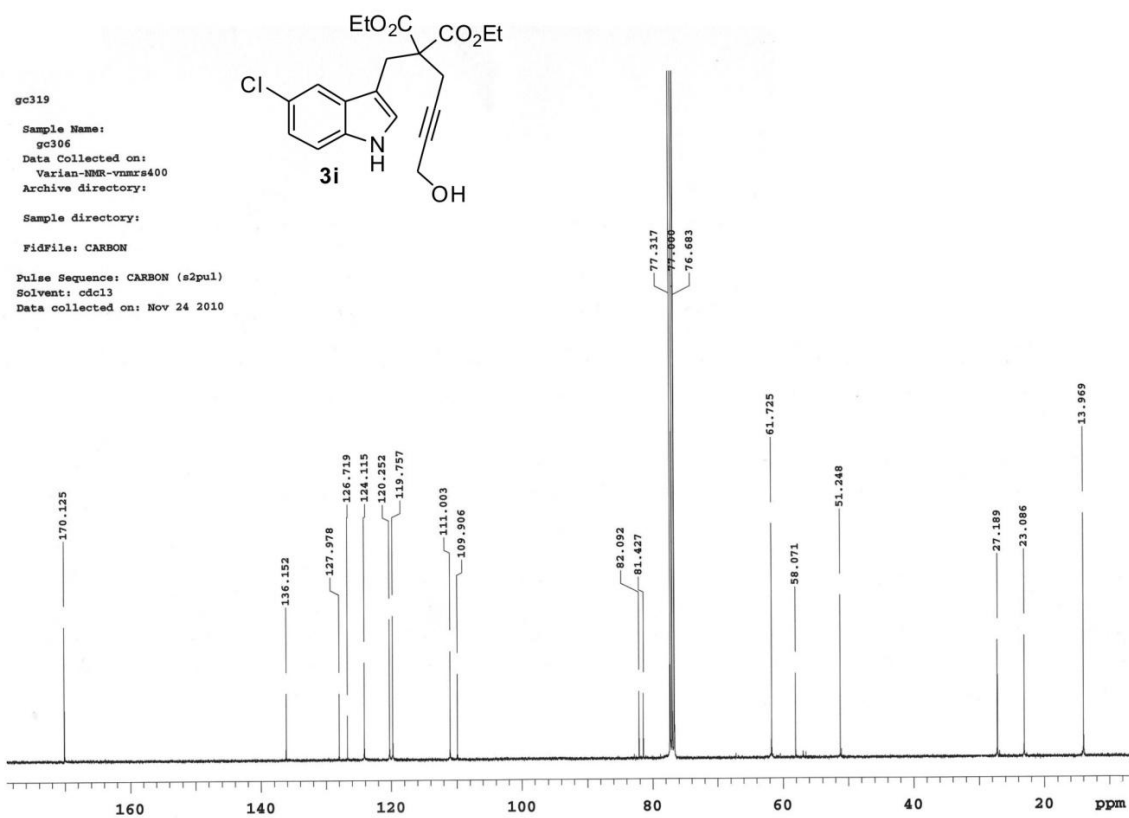
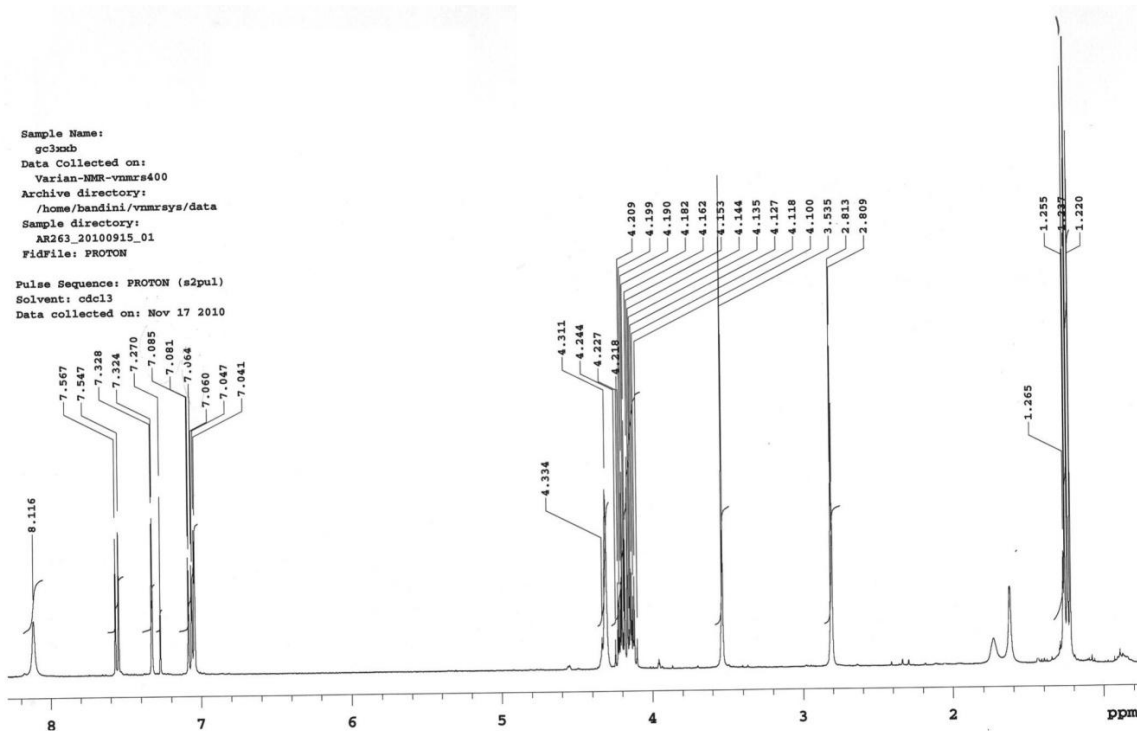


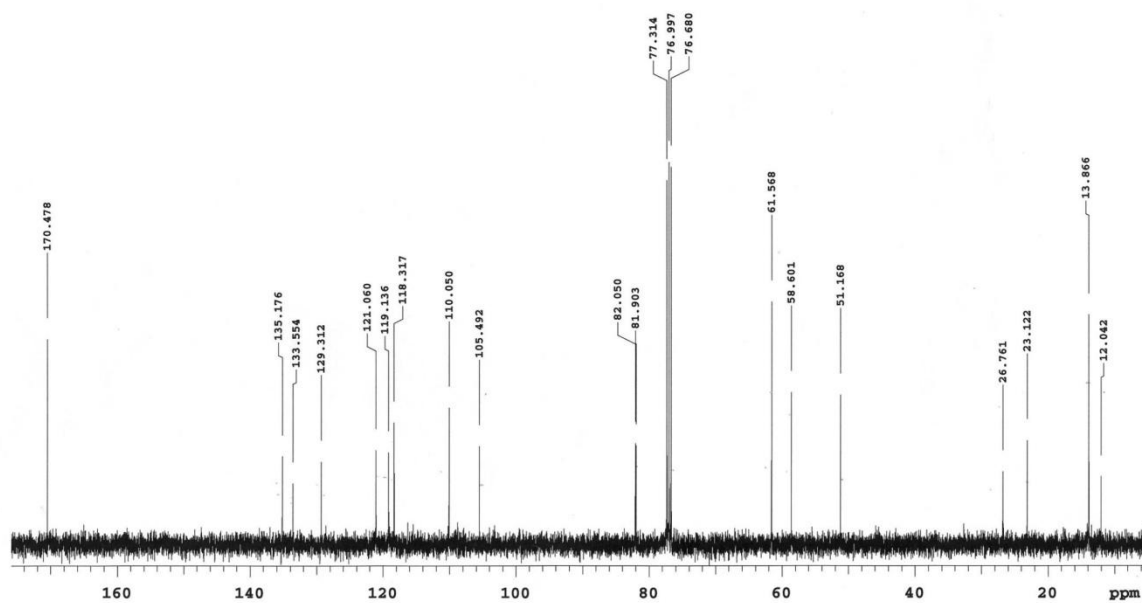
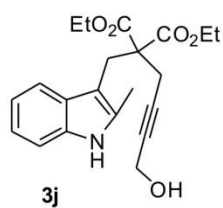
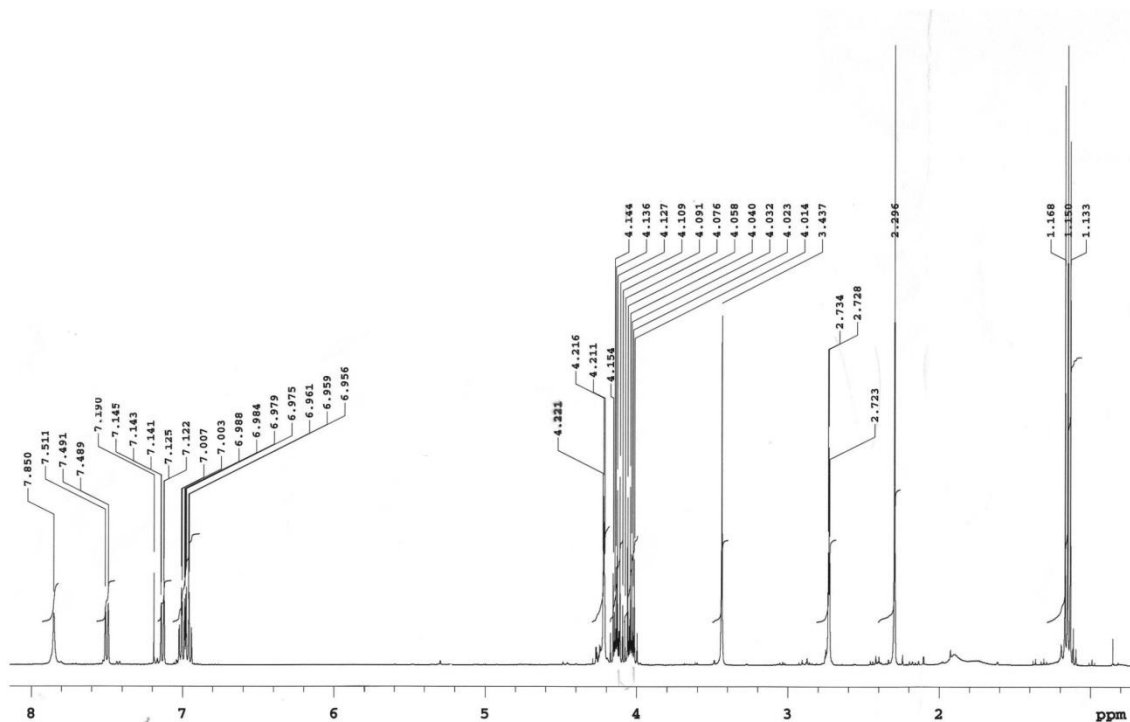


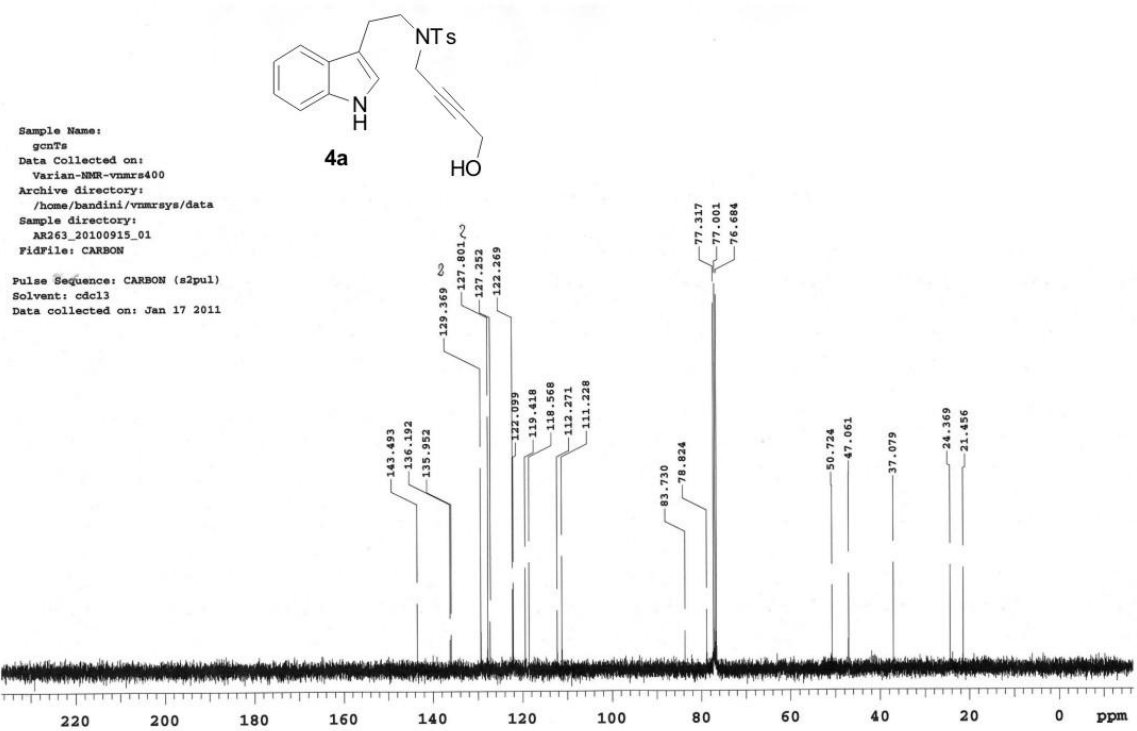
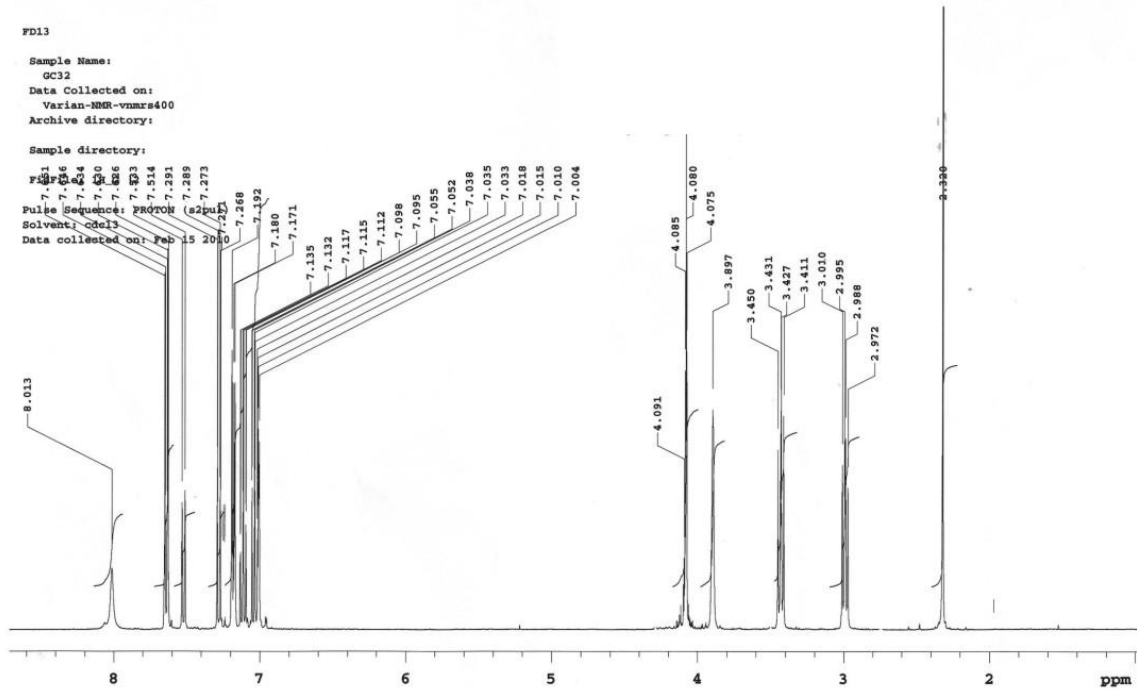


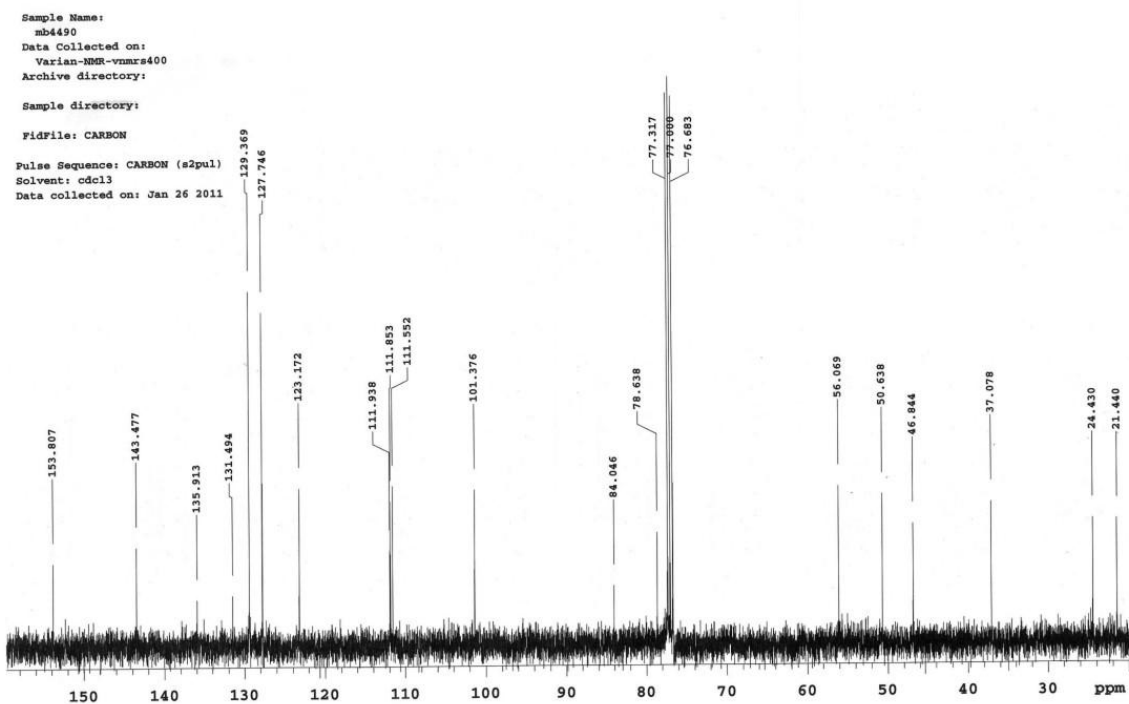
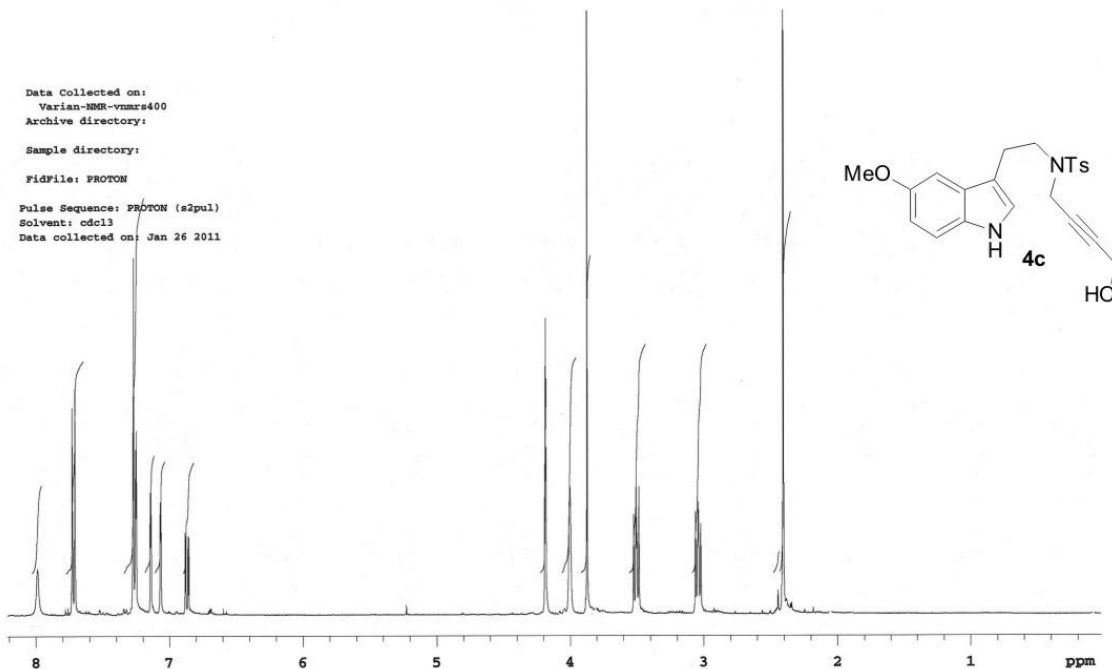


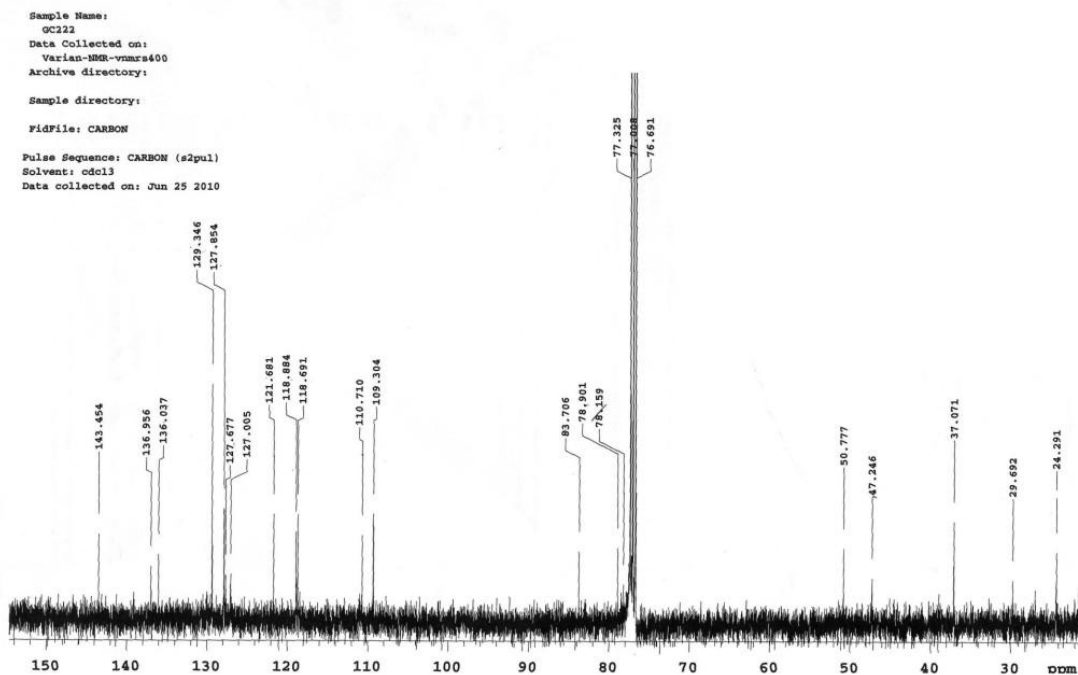
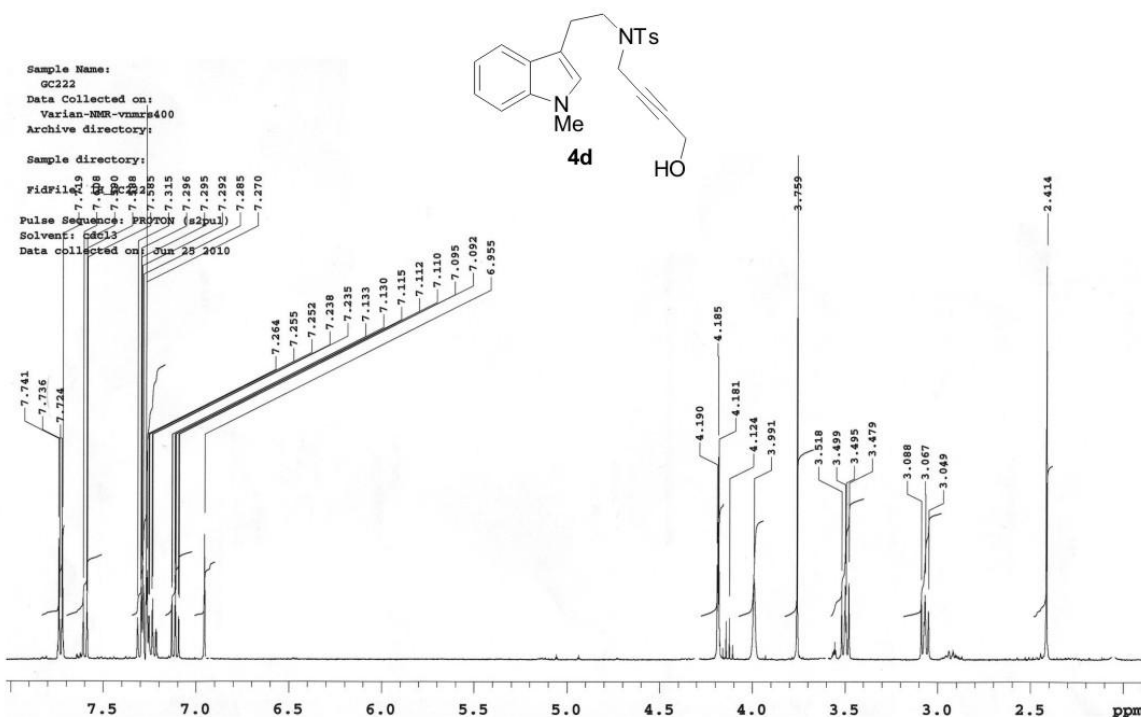


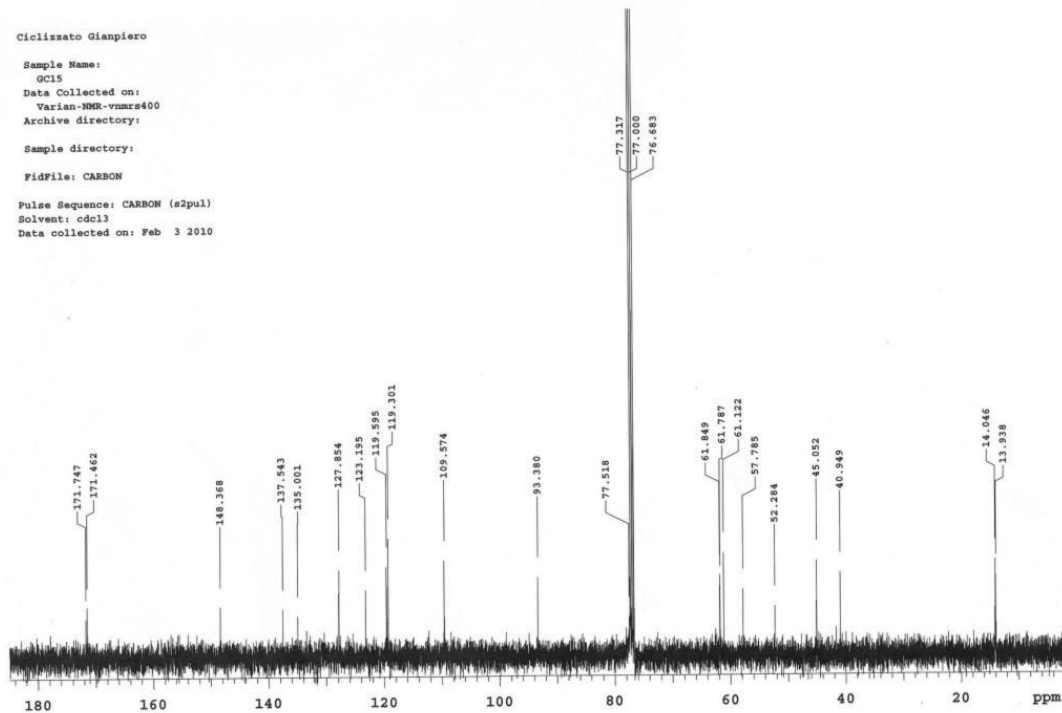
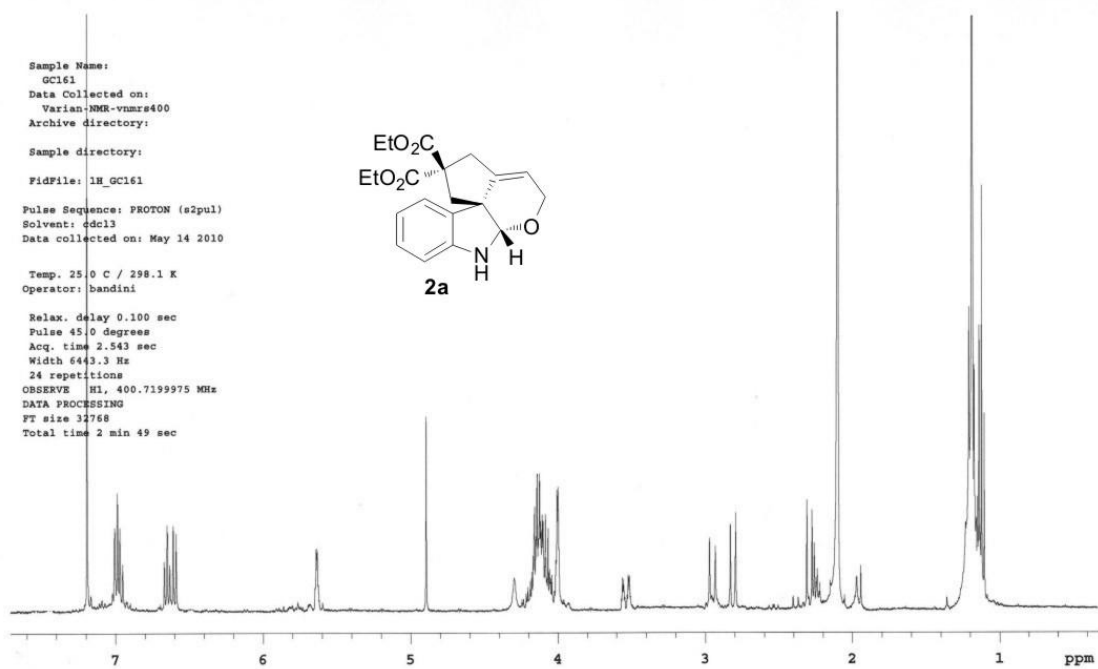




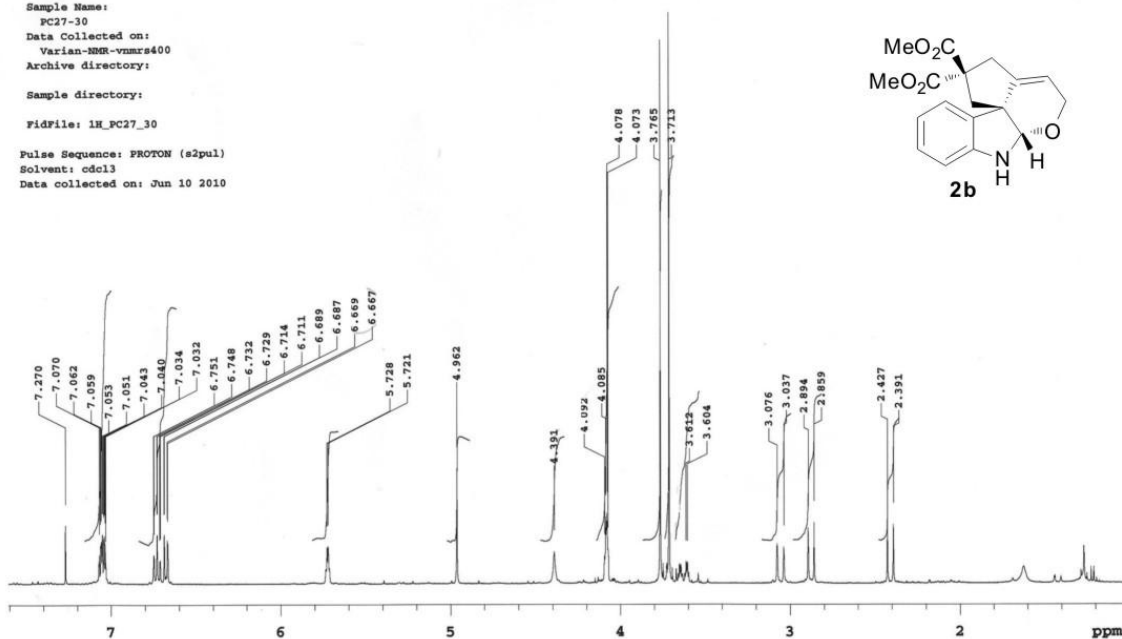
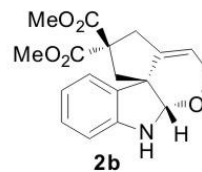




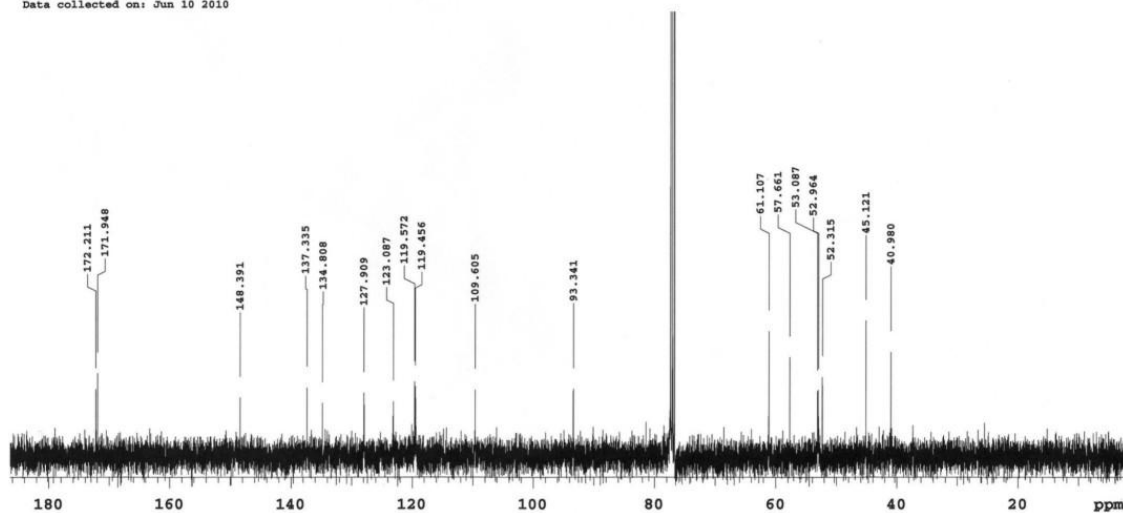


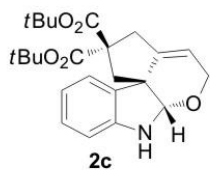
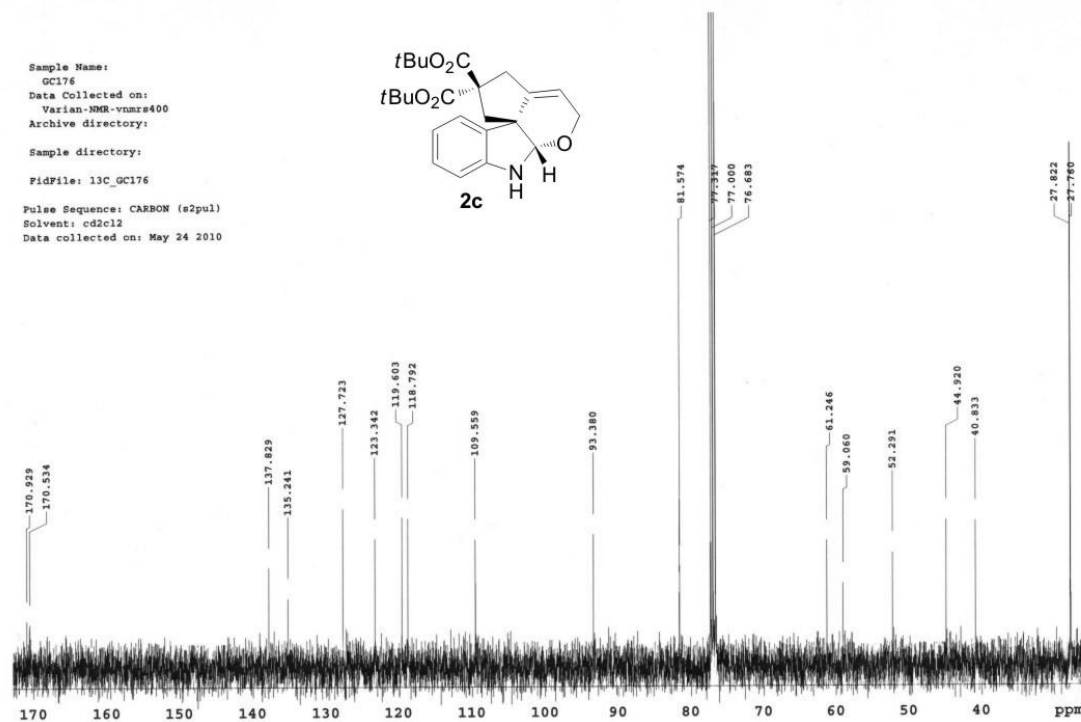
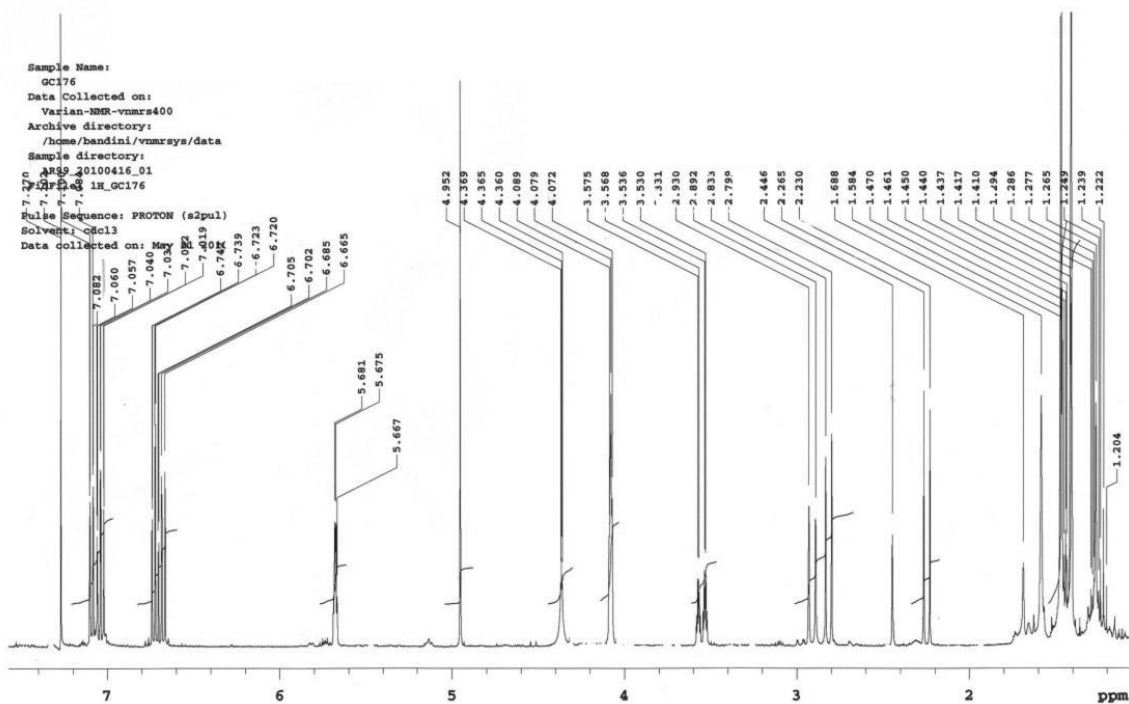


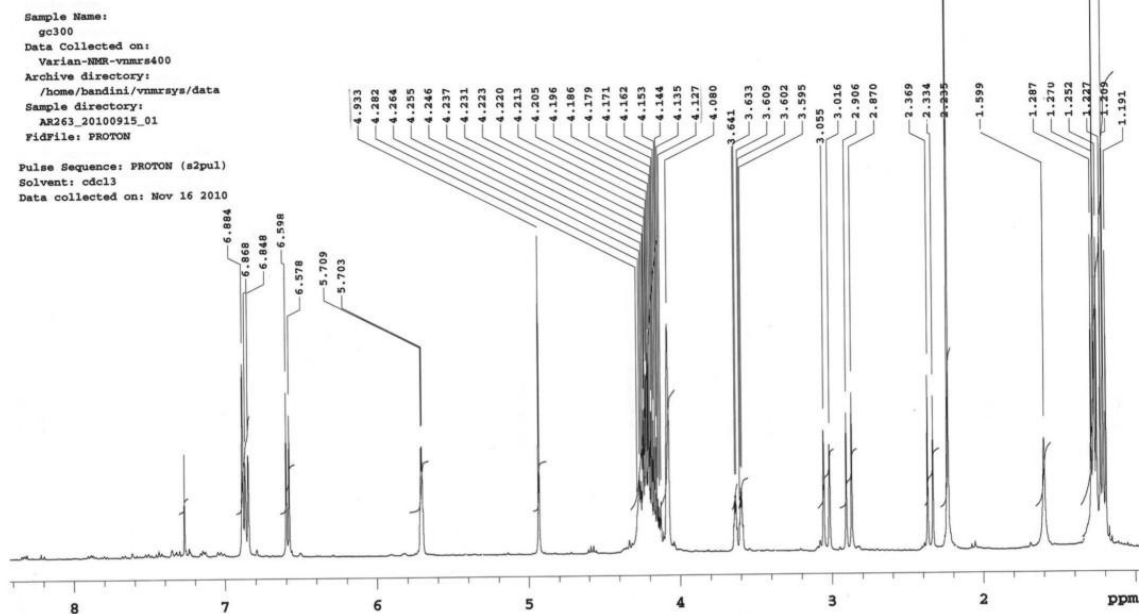
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PC27-30
Data Collected on:
Varian-NMR-vnmrs400
Archive directory:
Sample directory:
FidFile: 1H_PC27_30
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Jun 10 2010



Sample Name:
PC27-30
Data Collected on:
Varian-NMR-vnmrs400
Archive directory:
Sample directory:
FidFile: CARBON
Pulse Sequence: CARBON (s2pul)
Solvent: cd2cl2
Data collected on: Jun 10 2010

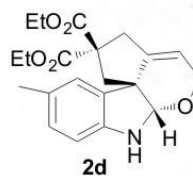
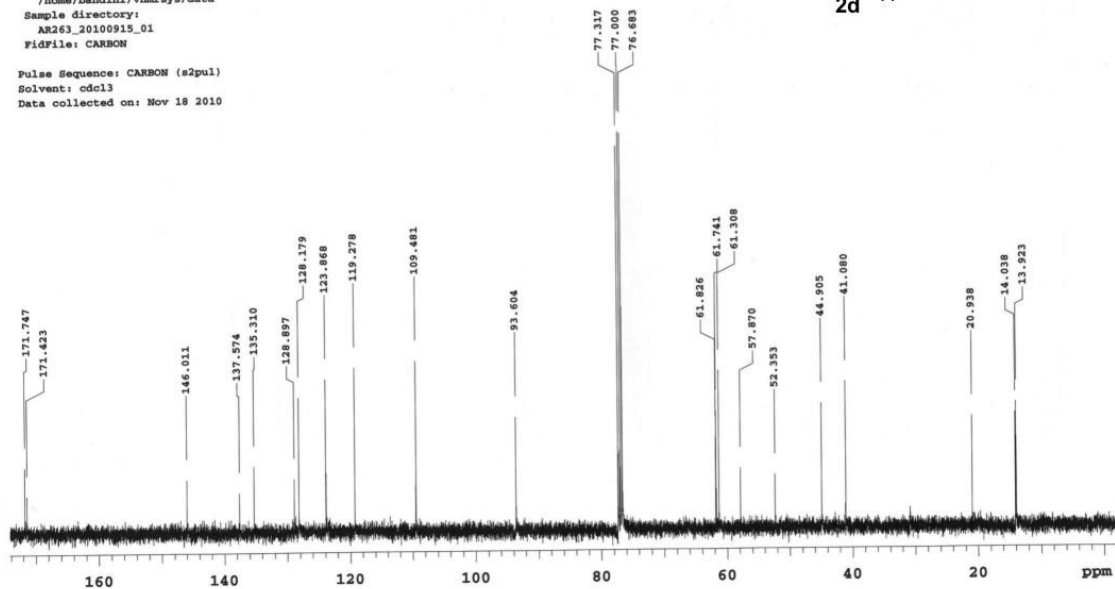


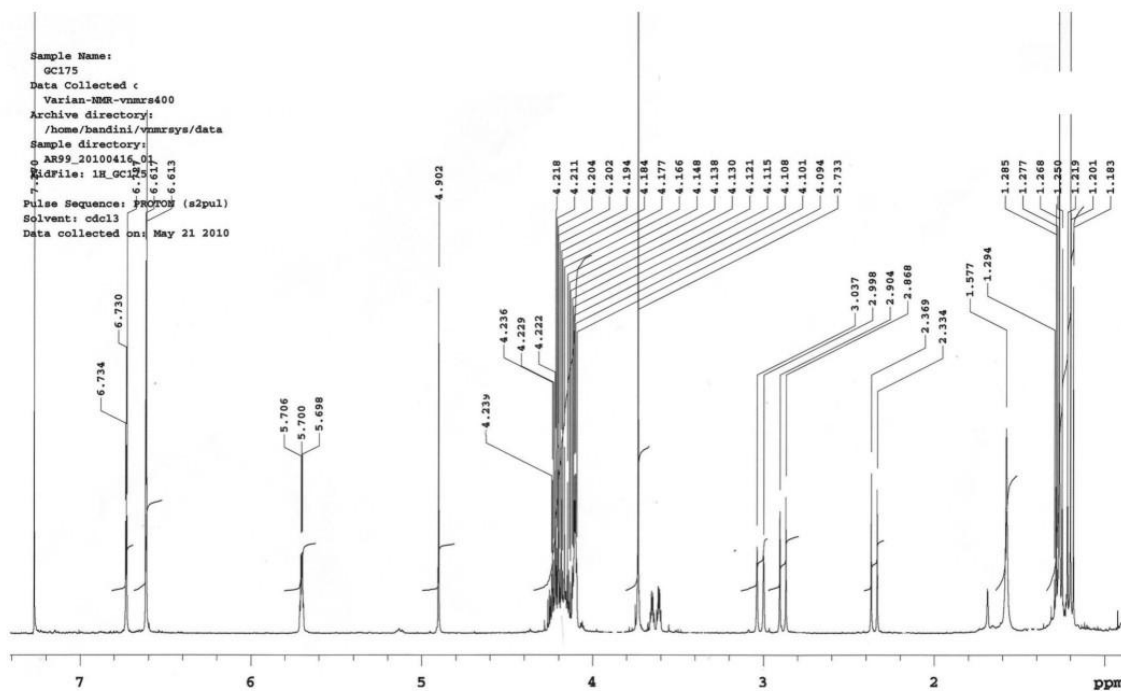




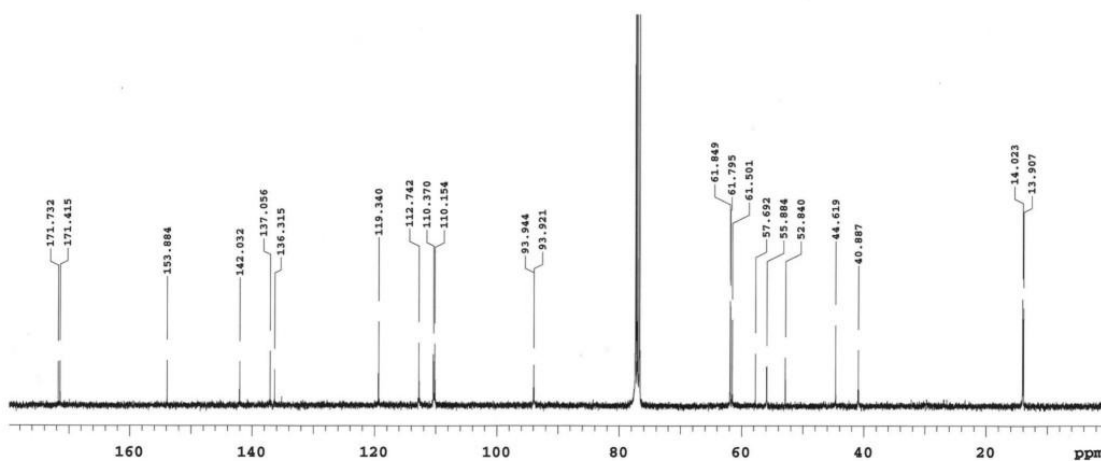
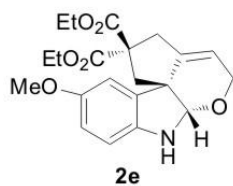
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Data Collected on: Varian-NMR-vnmrs400
Archive directory: /home/bandini/vnmrsys/data
Sample directory: AR263_20100915_01
Fidfile: CARBON

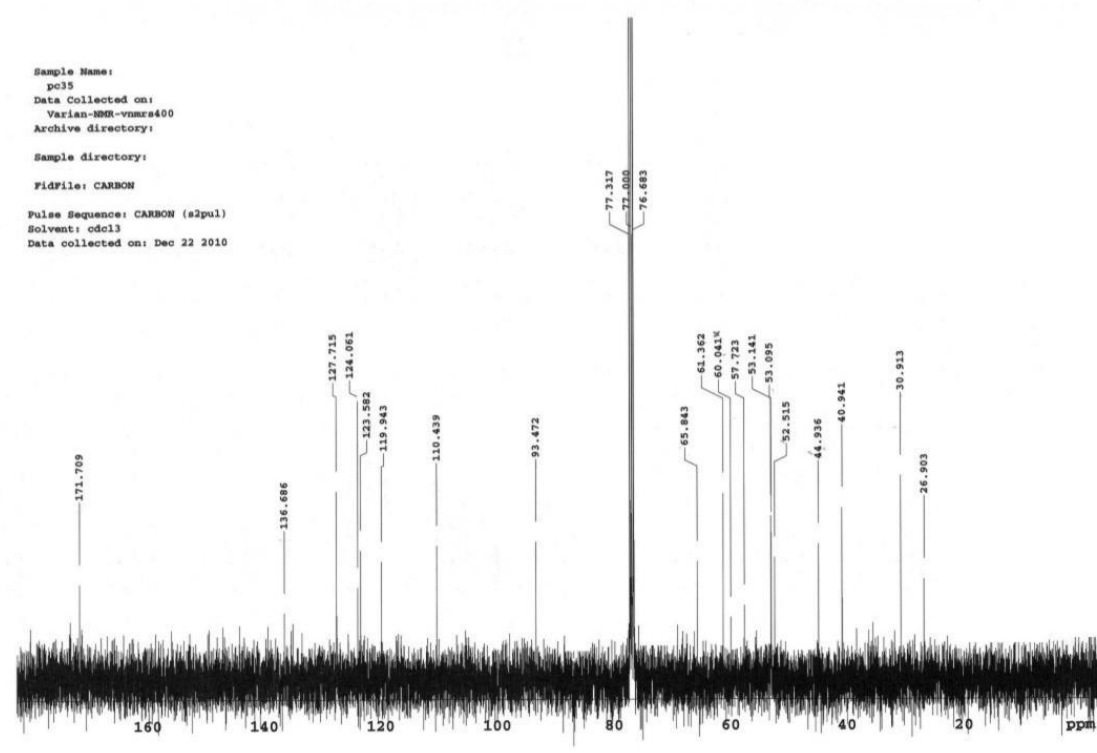
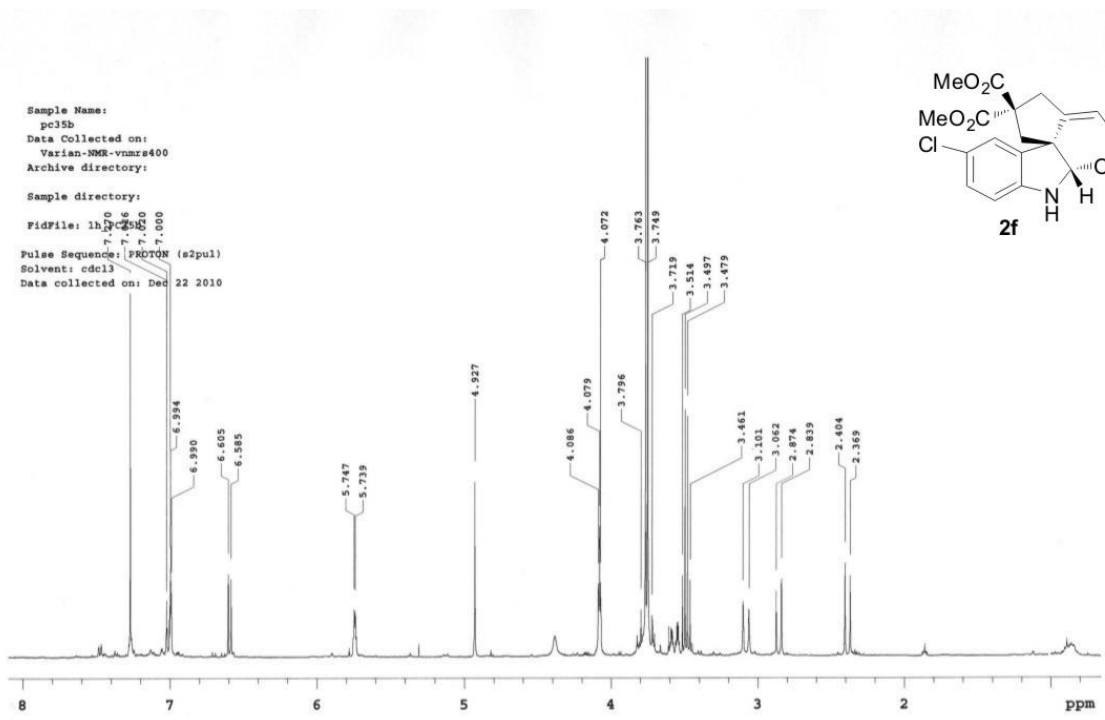
Pulse Sequence: CARBON (s2pul)
Solvent: cdcl3
Data collected on: Nov 18 2010

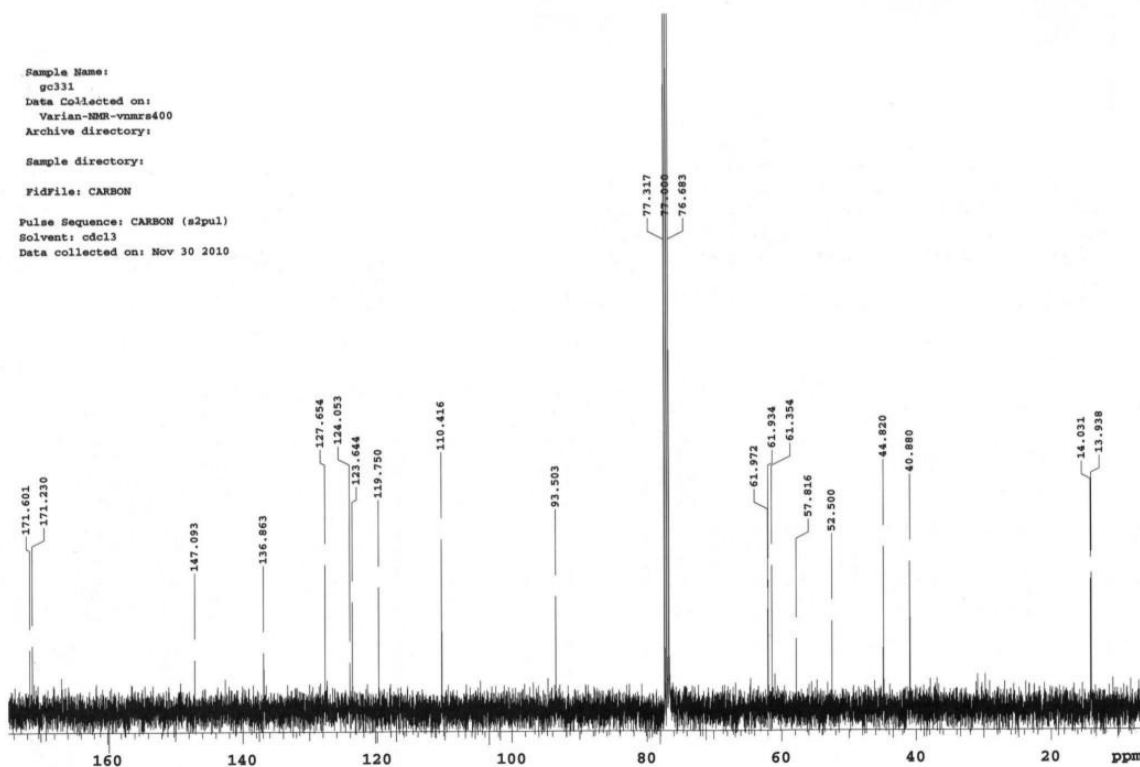
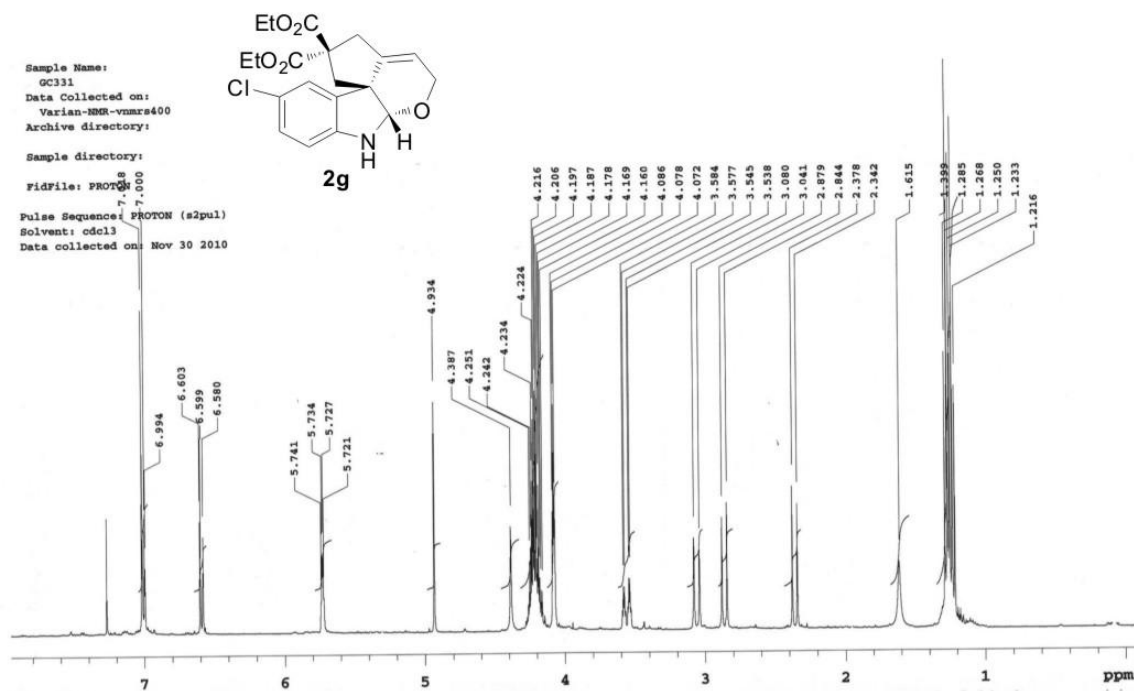


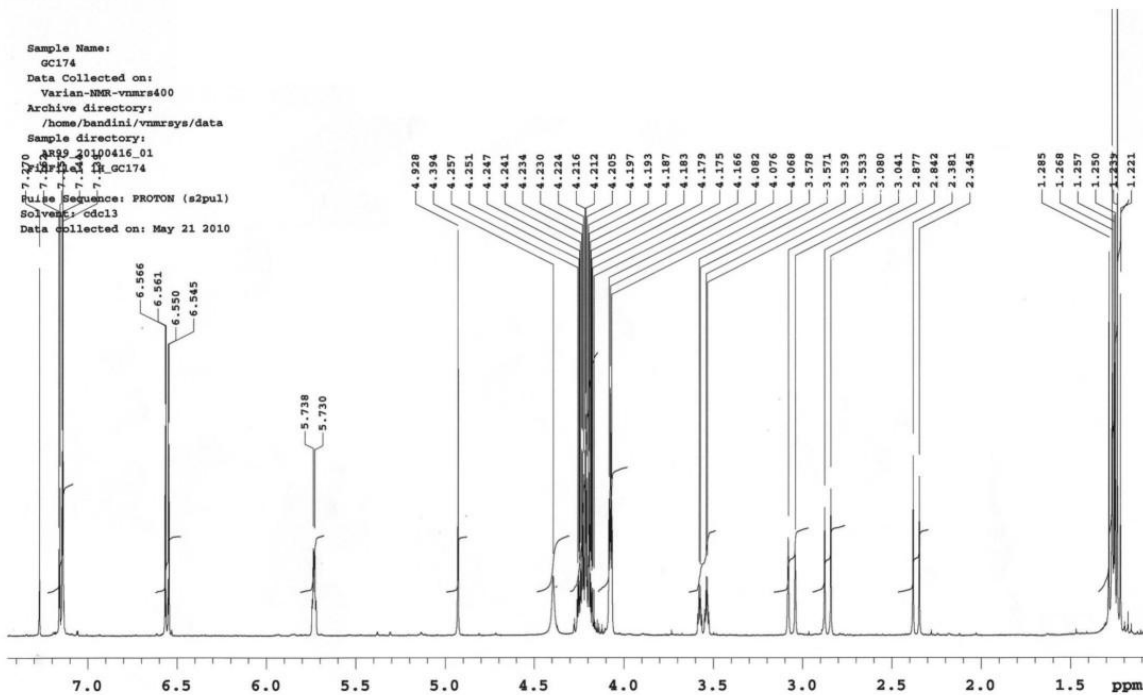


Sample Name: GC175
 Data Collected on: Varian-NMR-vnmrs400
 Archive directory:
 Sample directory:
 Fidfile: CARBON
 Pulse Sequence: CARBON (s2pul)
 Solvent: cdcl3
 Data collected on: May 28 2010



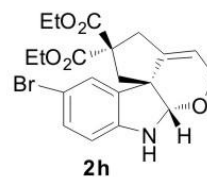
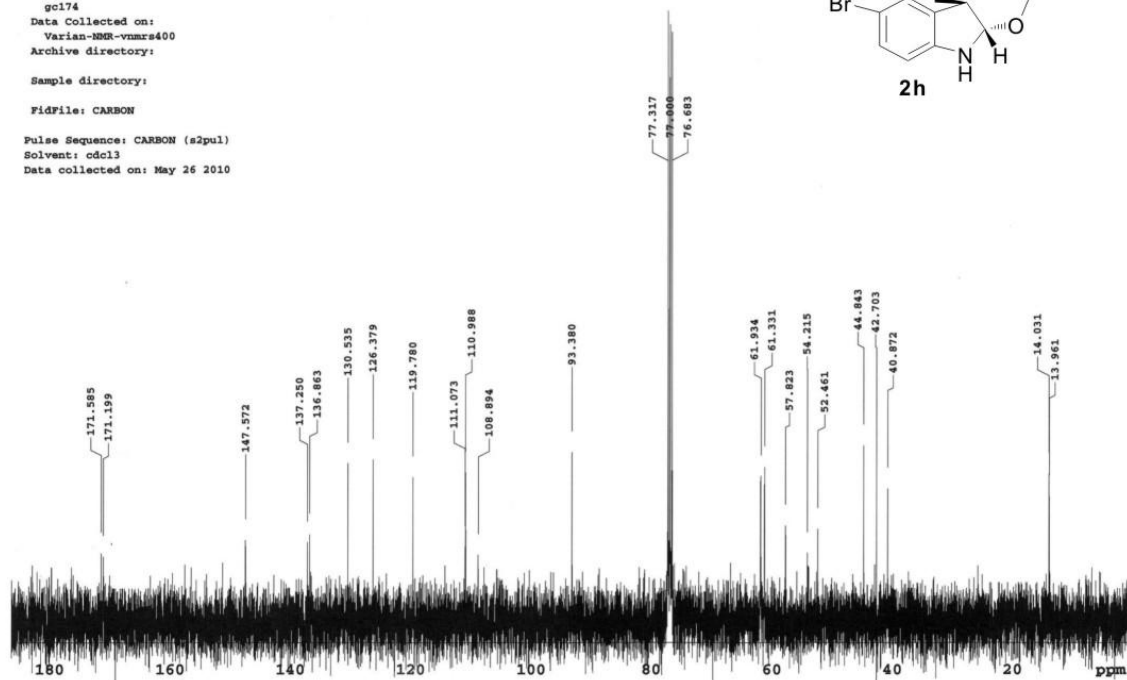


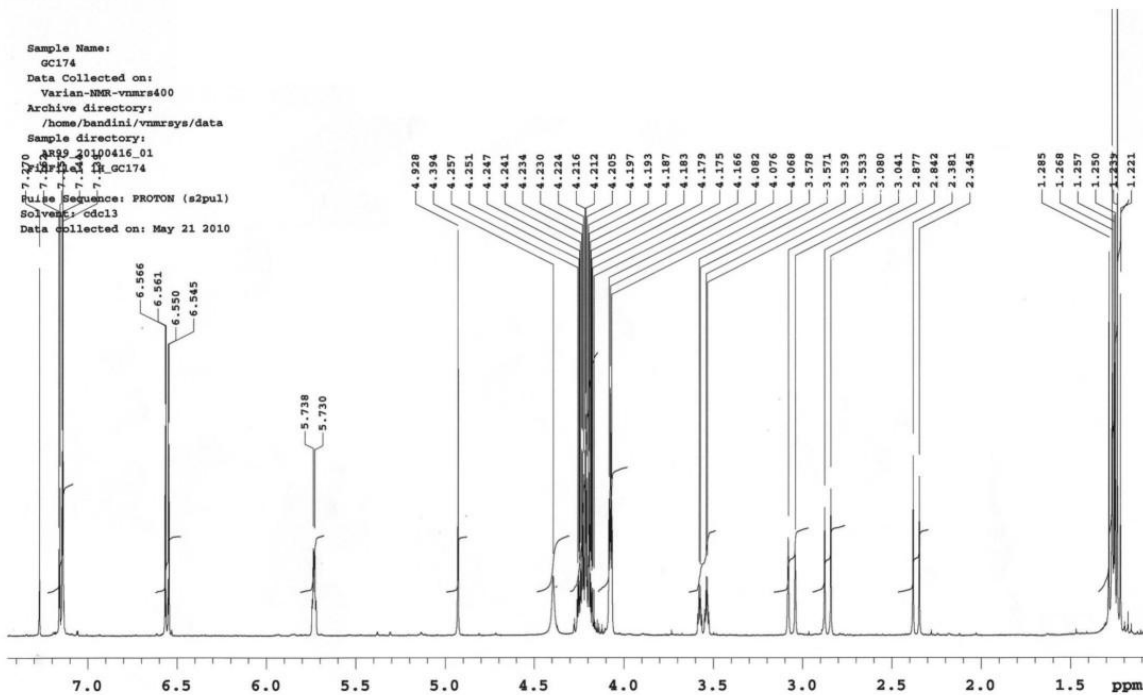




STANDARD CARBON PARAMETERS

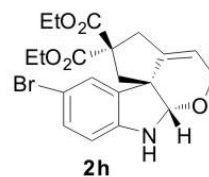
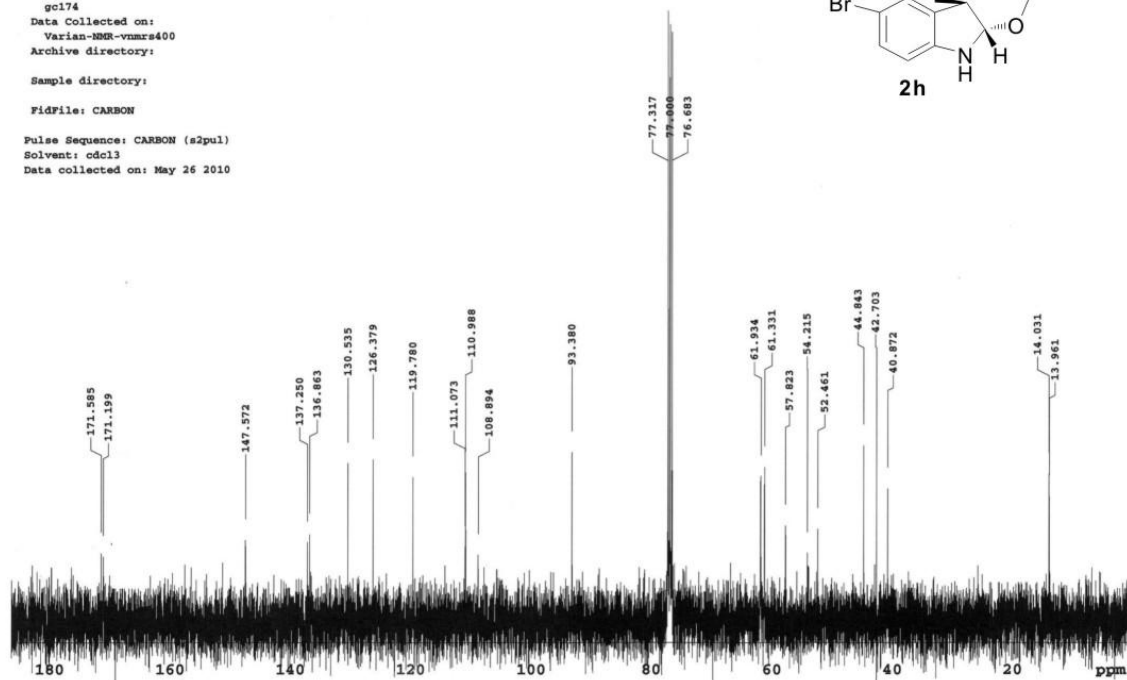
Sample Name: gc174
 Data Collected on: Varian-NMR-vnmrs400
 Archive directory:
 Sample directory:
 FIDfile: CARBON
 Pulse Sequence: CARBON (s2pul)
 Solvent: cdcl3
 Data collected on: May 26 2010

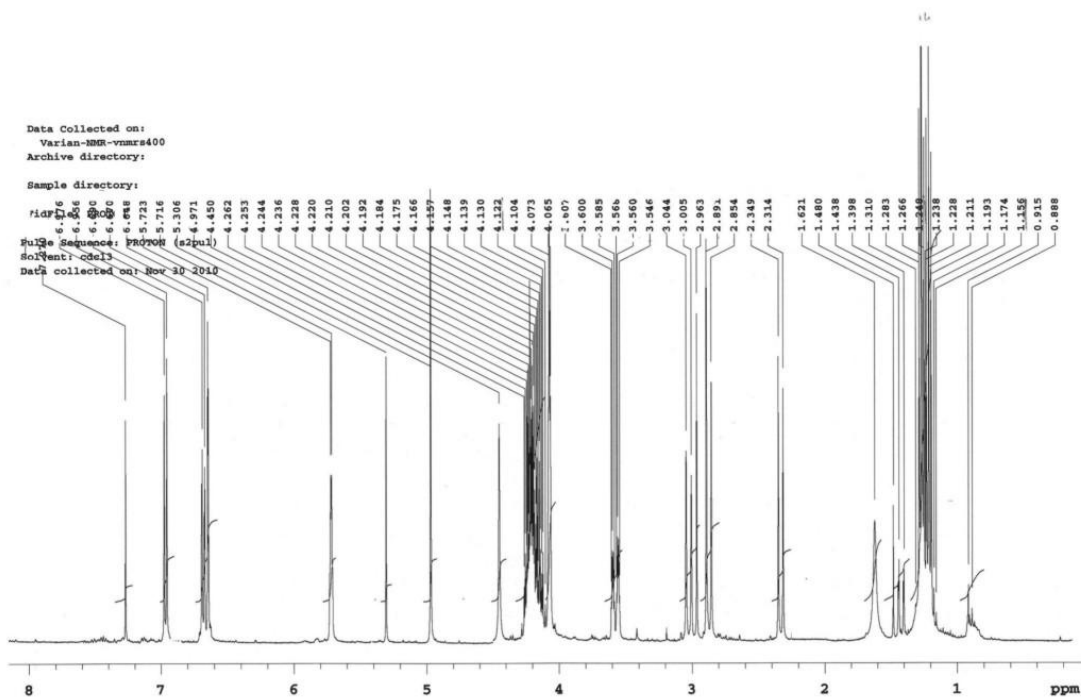




STANDARD CARBON PARAMETERS

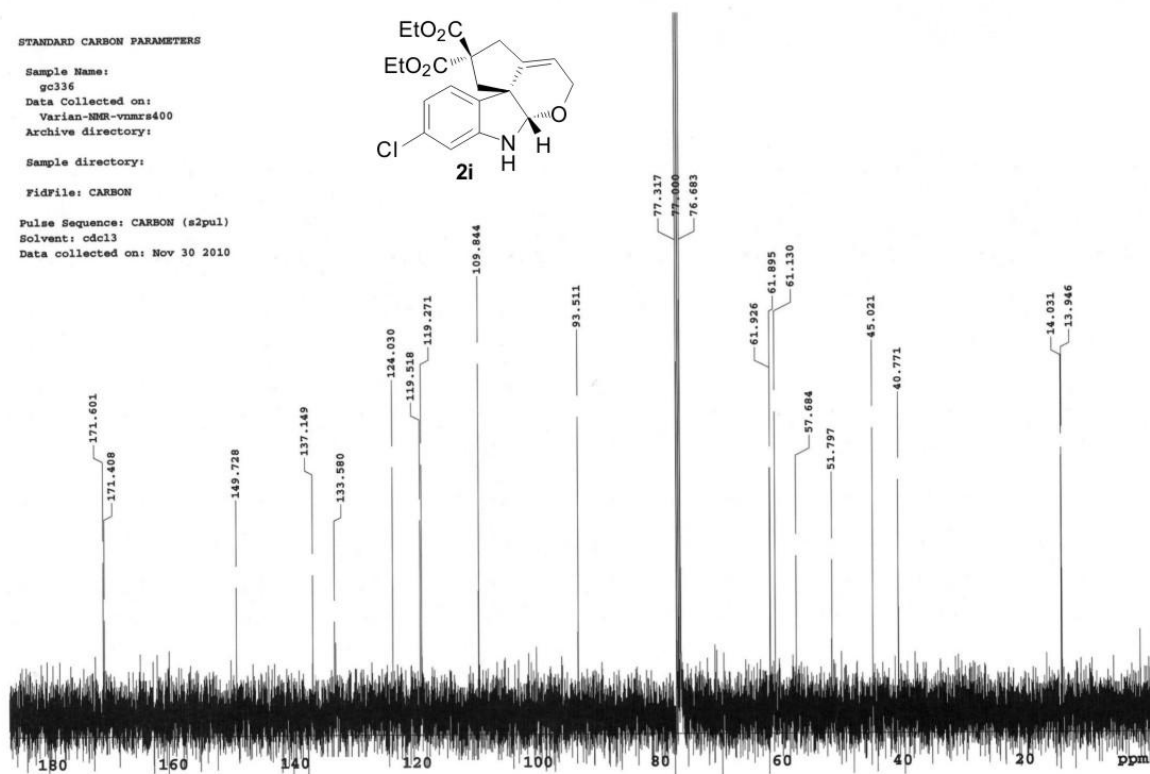
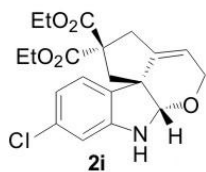
Sample Name: gc174
 Data Collected on: Varian-NMR-vnmrs400
 Archive directory:
 Sample directory:
 FIDfile: CARBON
 Pulse Sequence: CARBON (s2pul)
 Solvent: cdcl3
 Data collected on: May 26 2010

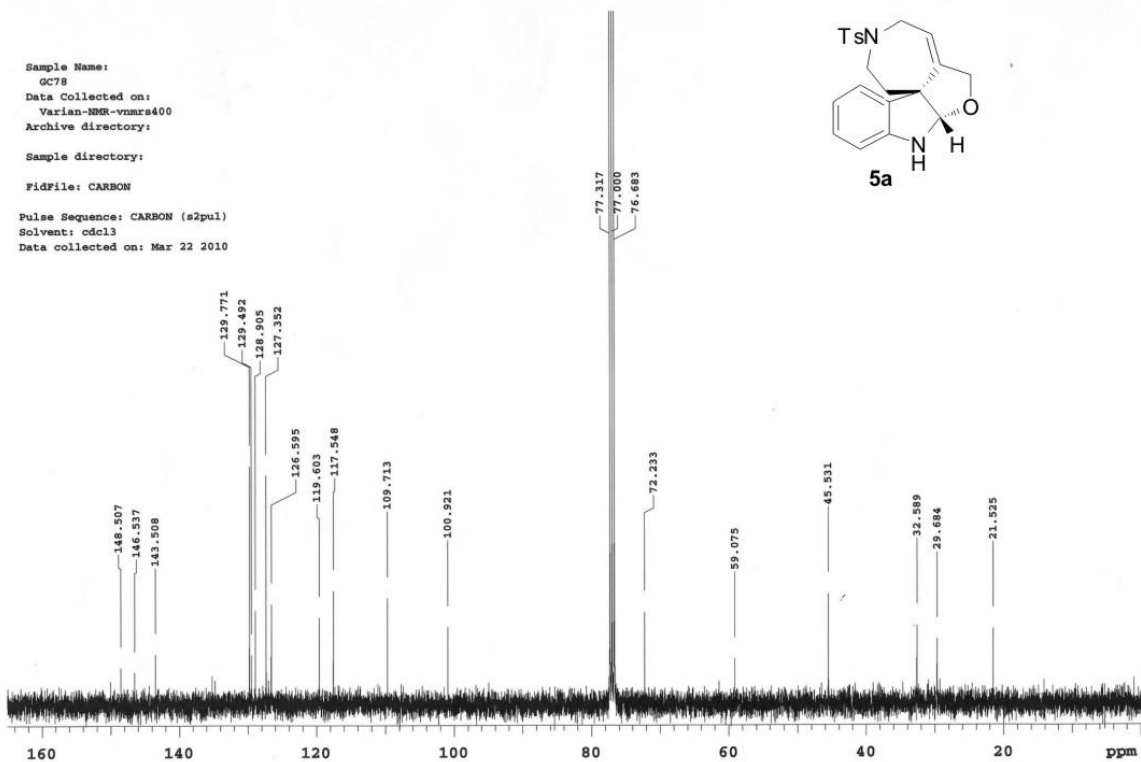
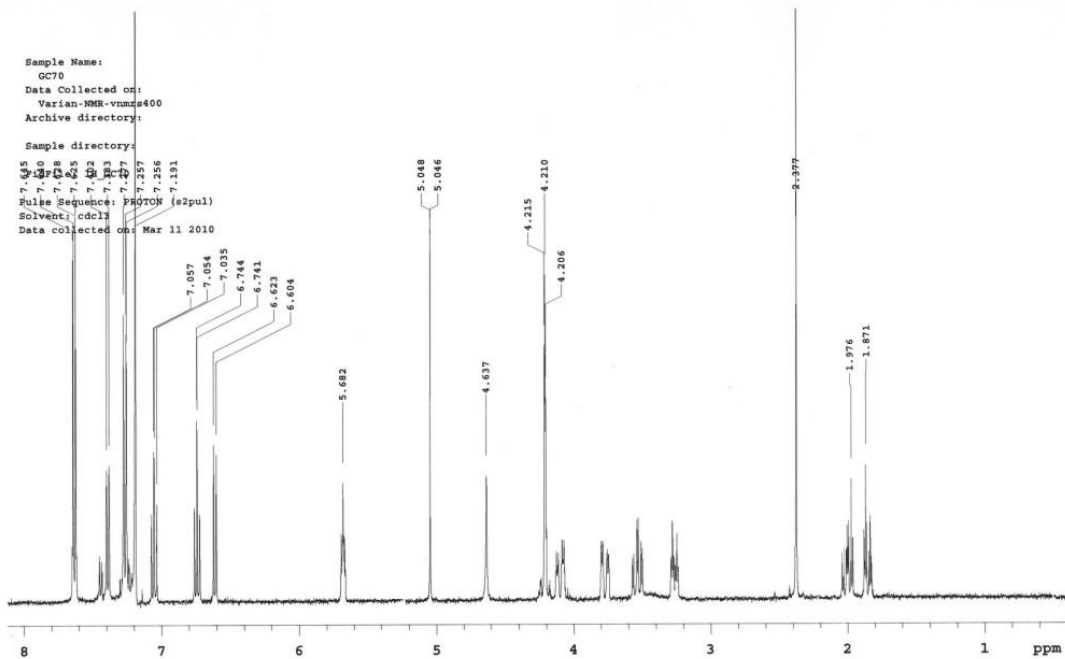


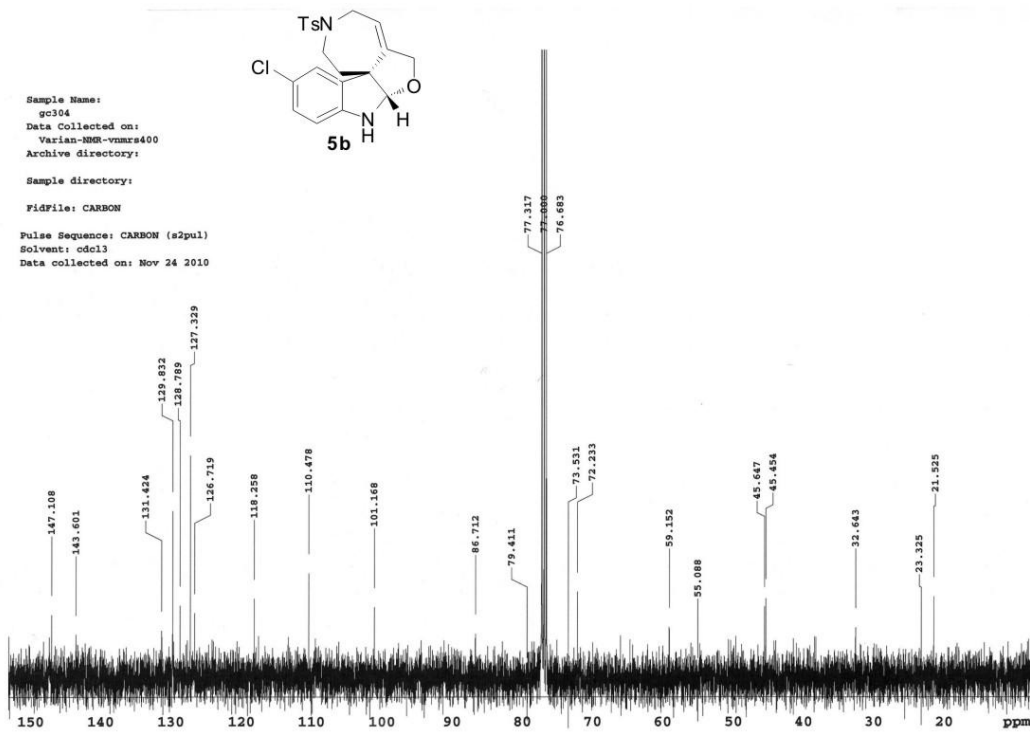
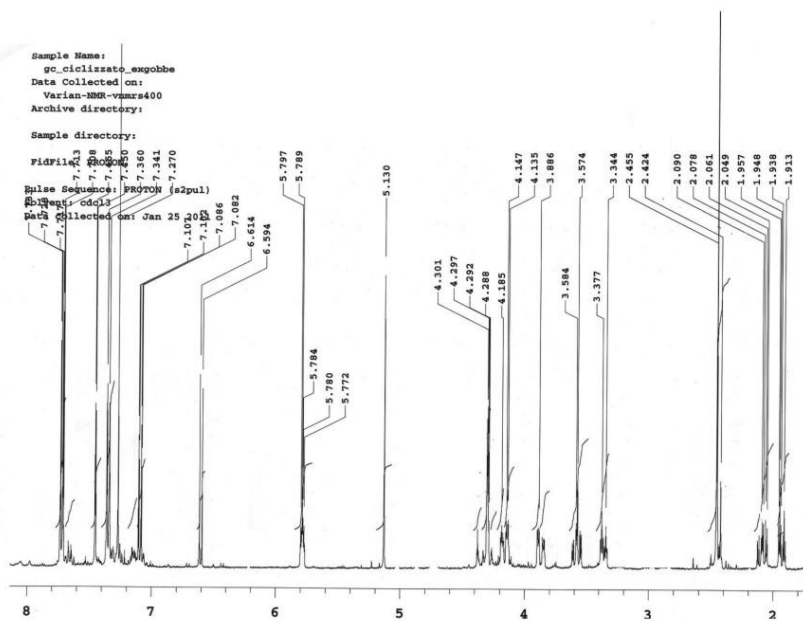


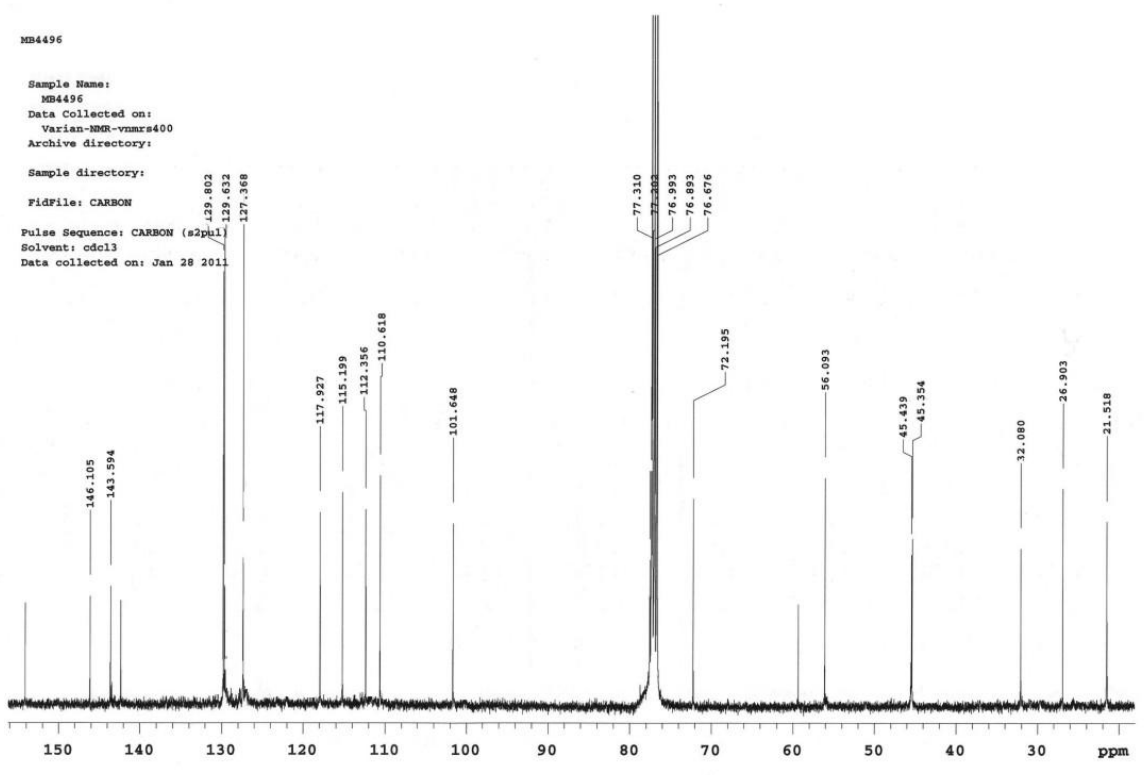
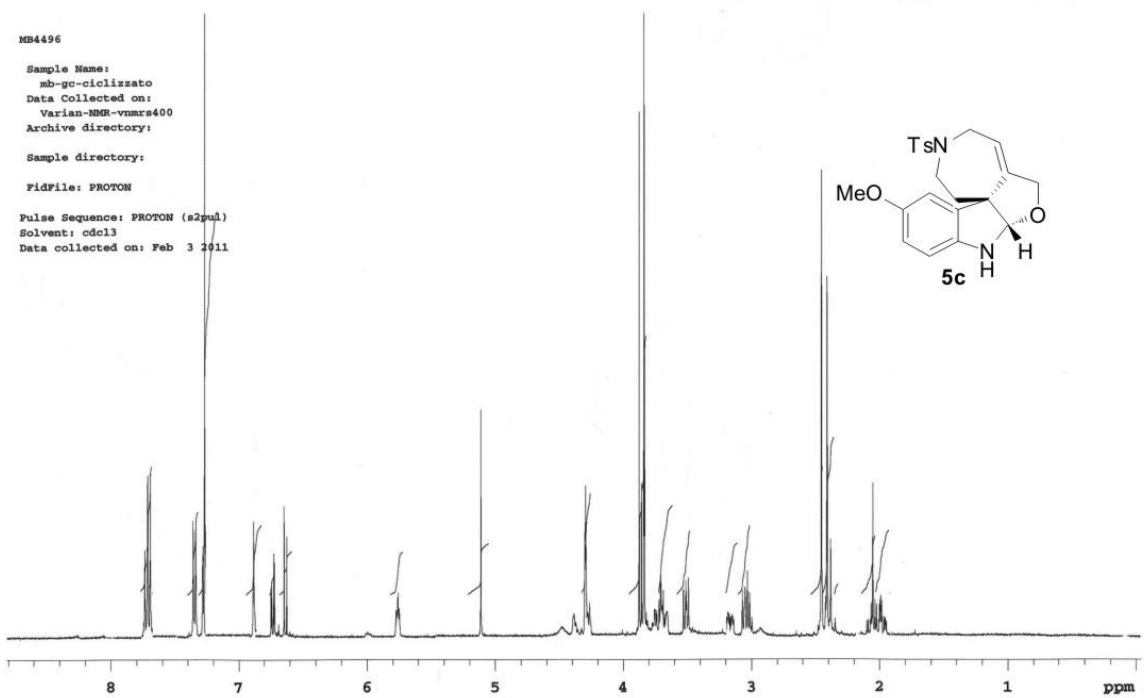
STANDARD CARBON PARAMETERS

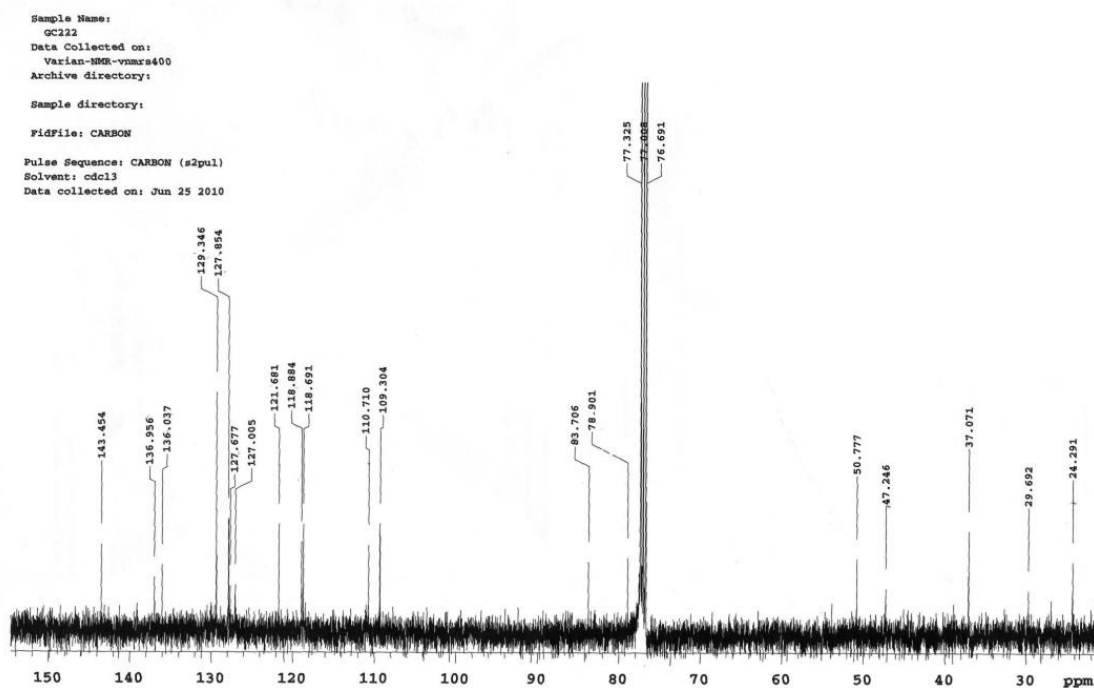
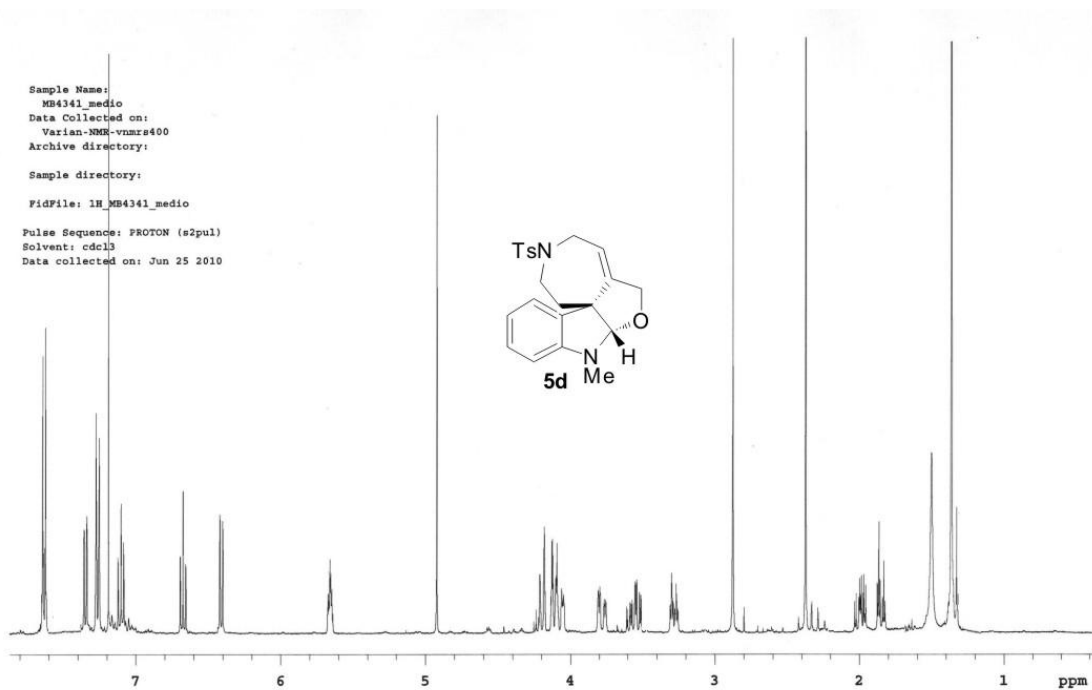
Sample Name:
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 Varian-NMR-vnmrs400
 Archive directory:
 Sample directory:
 F1dfile: CARBON
 Pulse Sequence: CARBON (s2pul)
 Solvent: cdcl3
 Data collected on: Nov 30 2010











References

- ¹ S. S. Kinderman, M. M. T. Wekking, J. H. van Maarseveen, H. E. Schoemaker, H. Hiemstra, F. P. J. T. Rutjes, *J.Org.Chem.* **2005**, **70**, 5519.
- ² D. T. Jones, G. D. Artman, R. M. Williams and M. Robert, *Tetrahedron Lett.* 2007, **48**, 1291.
- ³ S. S. Kinderman, M. M. T. Wekking, J. H. van Maarseveen, H. E. Schoemaker, H. Hiemstra, F. P. J. T. Rutjes, *J. Org. Chem.* **2005**, **70**, 5519.
- ⁴ With 10 mol% of catalyst.
- ⁵ A. Altomare, M. C. Burla, M. Camalli, G. L. Cascarano, C. Giacovazzo, A. Guagliardi, A. G. G. Moliterni, G. Polidori and R. Spagna, *J. Appl. Cryst.*, 1999, **32**, 115.
- ⁶ G. M. Sheldrick, *SHELXTLplus (Windows NT Version) Structure Determination Package; Version 5.1*. Bruker Analytical X-ray Instruments Inc.: Madison, WI, USA, 1998.