Supporting Information

Cis-glycofused benzopyran compounds as new amyloid β peptide ligands

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Synthesis

General Remarks

All solvents were dried with molecular sieves for at least 24 h prior to use. Thin layer chromatography (TLC) was performed on silica gel 60 F254 plates (Merck) with detection using UV light when possible, or by charring with a solution of concd. H₂SO₄/EtOH/H₂O (5:45:45) or a solution of (NH₄)₆Mo₇O₂₄ (21 g), Ce(SO₄)₂ (1 g), concd. H₂SO₄ (31 mL) in water (500 mL). Flash column chromatography was performed on silica gel 230-400 mesh (Merck). ¹H and ¹³C NMR spectra were recorded at 25 °C unless otherwise stated, with a Varian Mercury 400 MHz instrument. Chemical shift assignments, reported in ppm, are referenced to the corresponding solvent peaks. HRMS were recorded on a QSTAR elite LC/MS/MS system with a nanospray ion source. Optical rotations were measured at room temperature using an Atago Polax-2L polarimeter and are reported in units of $10^{-1} \text{ deg} \cdot \text{cm}2 \cdot \text{g}^{-1}$.

General synthetic strategy for the synthesis of protected compounds **10-19**: (J. S. Yadav, B. V. S. Reddy, L. Chandraiah, B. Jagannadh, S. Kiran Kumar and Ajit C. Kunwar *Tetrahedron Letters*, 2002, 43, 4527).

A mixture containing the appropriate O-hydroxybenzaldehyde (1-7) (2.5 equiv.), trimethylorthoformiate (2.5 equiv.) and scandium triflate (3% mol) in CH_2Cl_2 is stirred at r.t. for 20 min. The mixture is then cooled at 0°C and the appropriate tri-*O*-benzyl glycal (8-9) is added. The reaction is then left stirring at r.t. for 30 min. The reaction is then diluted with CH_2Cl_2 , washed with water, dried over Na_2SO_4 , filtrated and the solvent is removed under reduced pressure. The crude is purified by flash chromatography (eluent Toluene/AcOEt 9.75/0.25) to afford pure compounds 10-19.



(2R,3R,4R,4aS,5R/S,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-2,3,4,4a,5,10ahexa hydropyrano[2,3-b]chromene (10): yield 59%, C5 R/S 92/8

(5R)



¹H.NMR (400 MHz, CDCl₃) δ 7.55 (d, *J* = 7.6 Hz, 1H, H6), 7.37 – 7.22 (m, 15H, Ar), 7.23 – 7.15 (m, 1H, H8), 6.99 (t, *J* = 7.5 Hz, 1H, H7), 6.83 (d, *J* = 8.1 Hz, 1H, H9), 5.66 (d, *J* = 2.9 Hz, 1H, H10a), 4.96 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.77 (d, *J* = 4.3 Hz, 1H, H5), 4.62 – 4.31 (m, 5H, OCH₂Ph), 4.18 (t, *J* = 6.4 Hz, 1H, H2), 3.79 (s, 1H, H3), 3.68 – 3.63 (m, 1H, H4), 3.63 – 3.59 (m, 2H, CH₂O), 3.58 (s, 3H, OMe), 3.36 – 3.26 (m, 1H, H4a); ¹³C NMR (101 MHz, CDCl₃) δ 152.27, 139.03, 138.89, 138.15, 129.12, 128.62, 128.44, 128.41, 128.38, 128.13, 128.01, 127.95, 127.77, 127.55, 126.26, 122.43, 121.38, 115.59, 97.74, 76.22, 75.62, 75.16, 73.97, 73.71, 72.78, 71.71, 69.13, 57.10, 34.72. [α]_D²⁰= +5,3 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 575.2, [M + K]⁺ = 591.2; found [M + Na]⁺ = 575.3, [M + K]⁺ = 591.3.

(5S)



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.24 (m, 13H, Ar), 7.24 – 7.20 (m, 1H, H6), 7.18 (dd, *J* = 7.0, 2.2 Hz, 2H, Ar), 7.07 (d, *J* = 7.4 Hz, 1H, H8), 6.88 (m, 2H, H7, H9), 5.71 (d, *J* = 3.2 Hz, 1H, H10a), 4.92 (d, *J* = 11.5 Hz, 1H, OCH₂Ph), 4.54 (ddd, *J* = 34.5, 23.4, 11.7 Hz, 5H, OCH₂Ph), 4.40 (d, *J* = 2.1 Hz, 1H, H5), 4.20 (M, 1H, H2), 3.98 (s, 1H, H3), 3.72 – 3.64 (m, 2H, CH₂O), 3.37 (s, 3H, OMe), 3.32 (dd, *J* = 11.8, 2.4 Hz, 1H, H4), 3.06 – 2.98 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 153.89, 138.79, 138.09, 137.76, 131.20, 130.35, 128.67, 128.64, 128.50, 128.25, 128.21, 128.03, 128.01, 127.85, 127.83, 120.81, 118.54, 116.96, 95.00, 75.17, 74.79, 74.77, 73.79, 71.62, 71.53, 71.52, 68.94, 56.38, 37.70, 29.93. [α]_D²⁰= +8,7 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 575.2, [M + K]⁺ = 591.2; found [M + Na]⁺ = 575.3, [M + K]⁺ = 591.3.

(2R,3R,4R,4aR,5R,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-7-nitro-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (11): yield 40%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 8.44 – 8.41 (m, 1H, H6), 8.08 (dd, *J* = 9.0, 2.8 Hz, 1H, H8), 7.43 – 7.17 (m, 15H, Ar), 6.87 (d, *J* = 9.0 Hz, 1H, H9), 5.73 (d, *J* = 2.9 Hz, 1H, H10a), 4.96 (d, *J* = 11.3 Hz, 1H, OCH₂Ph), 4.73 (t, *J* = 7.4 Hz, 1H, H5), 4.64 – 4.36 (m, 5H, OCH₂Ph), 4.11 (t, *J* = 6.4 Hz, 1H, H2), 3.86 (s, 1H, H3), 3.64 (dd, *J* = 9.2, 5.8 Hz, 2H, CH₂O), 3.59 (d, *J* = 7.4 Hz, 3H, OMe), 3.50 (dd, *J* = 11.1, 2.5 Hz, 1H, H4), 3.41 – 3.32 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 157.58, 142.33, 138.72, 138.01, 137.95, 128.68, 128.53, 128.42, 128.39, 128.23, 128.18, 128.08, 127.94, 127.81, 125.33, 123.31, 123.21, 116.20, 98.87, 75.28, 75.23, 74.68, 73.83, 73.02, 72.39, 72.13, 68.95, 57.10, 33.96.[α]_D²⁰= -4,5 (c=1, CHCl₃). MS: m/z calcd for [M + H]⁺ = 598.2, [M + Na]⁺ = 620.2, [M + K]⁺ = 636.2; found [M + H]⁺ = 598.3, [M + Na]⁺ = 620.4, [M + K]⁺ = 636.4.

(2R,3R,4R,4aR,5R,10aR)-3,4,7-tris(benzyloxy)-2-benzyloxymethyl-5-methoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene (12): yield 35%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 7.52 – 7.25 (m, 20H, Ar), 7.23 (d, *J* = 2.2 Hz, 1H, H6), 6.86 (dd, *J* = 8.8, 3.0 Hz, 1H, H8), 6.81 – 6.74 (m, 1H, H9), 5.64 (d, *J* = 2.9 Hz, 1H, H10a), 5.10 – 4.95 (m, 3H, OCH₂Ph), 4.76 (d, *J* = 4.4 Hz, 1H, H5), 4.64-4.34 (m, 5H, OCH₂Ph), 4.20 (t, *J* = 6.5 Hz, 1H, H2), 3.83 (s, 1H, H3), 3.67 (dd, *J* = 11.1, 2.6 Hz, 1H, H4), 3.65 – 3.61 (m, 2H, CH₂O), 3.58 (s, 3H, OMe), 3.36 – 3.25 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 149.98, 139.05, 138.93, 138.16, 130.59, 129.64, 128.61, 128.43, 128.40, 128.36, 128.13, 127.97, 127.94, 127.75, 127.52, 126.52, 121.97, 115.37, 97.62, 76.33, 75.66, 75.16, 73.93, 73.71, 72.69, 71.65, 69.15, 57.11, 34.78, 20.99. [α]_D²⁰= - 5,5 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 681.3, [M + K]⁺ = 697.3; found [M + Na]⁺ = 681.5, [M + K]⁺ = 697.4.

(2R,3R,4R,4aR,5R/S,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5,7-dimethoxy-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (13): yield 73%, C5 R/S 85/15

(5R)



¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.21 (m, 15H, Ar), 7.11 (d, *J* = 0.9 Hz, 1H, H6), 6.76 (d, 2H,H8 and H9), 5.61 (d, *J* = 2.9 Hz, 1H, H10a), 4.96 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.74 (d, *J* = 4.4 Hz, 1H, H5), 4.60-4.36 (m, 5H, OCH₂Ph), 4.18 (t, *J* = 6.4 Hz, 1H, H2), 3.80 (s, 4H, ArOMe and H3), 3.66 (dd, *J* = 11.1, 2.6 Hz, 1H, H4), 3.61 (dt, *J* = 7.0, 3.4 Hz, 2H, CH₂O), 3.57 (s, 3H, OMe), 3.33 – 3.24 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 154.39, 146.10, 139.05, 138.92, 138.16, 128.62, 128.44, 128.41, 128.38, 128.14, 127.97, 127.95, 127.76, 127.54, 123.12, 116.33, 115.18, 110.81, 97.61, 76.27, 75.76, 75.16, 73.92, 73.72, 72.77, 71.65, 69.14, 57.07, 56.06, 34.72. [α]_D²⁰= +7,1 (c=1, CHCl₃); MS: m/z calcd for [M + K]⁺ = 621.2; found [M + K]⁺ = 621.5. (5S)



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.15 (m, 15H, Ar), 6.84 – 6.76 (m, 2H, H8 and H9), 6.58 (s, 1H, H6), 5.64 (d, *J* = 3.1 Hz, 1H, H10a), 4.91 (d, *J* = 11.5 Hz, 1H, OCH₂Ph), 4.66 – 4.40 (m, 5H, OCH₂Ph), 4.35 (d, *J* = 2.0 Hz, 1H, H5), 4.26 – 4.17 (m, 1H, H2), 4.00 (s, 1H, H3), 3.76 (s, 3H, ArOMe), 3.70 – 3.63 (m, 2H, CH₂O), 3.40 (s, 3H, OMe), 3.34 (dd, *J* = 11.8, 2.3 Hz, 1H, H4), 3.03 – 2.94 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 153.63, 147.66, 138.79, 138.10, 137.74, 128.63, 128.50, 128.46, 128.42, 128.25, 128.21, 128.03, 128.03, 127.97, 127.89, 127.83, 127.72, 127.50, 119.01, 117.67, 116.63, 115.27, 99.18, 94.88, 74.97, 74.80, 73.79, 71.60, 71.49, 71.42, 70.65, 70.02, 69.88, 68.96, 56.57, 55.96, 37.60. [α]_D²⁰= +6,6 (c=1, CHCl₃); MS: m/z calcd for [M + K]⁺ = 621.2; found [M + K]⁺ = 621.6.

(2R,3R,4R,4aR,5R/S,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-7-methyl-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (14): yield 91%, C5 R/S 95/5

(5R)



¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, *J* = 7.1 Hz, 1H, H6), 7.31 – 7.21 (m, 15H, Ar), 6.99 (d, *J* = 7.3 Hz, 1H, H8), 6.72 (d, *J* = 8.2 Hz, 1H, H9), 5.61 (d, *J* = 2.9 Hz, 1H, H10a), 4.96 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.74 (d, *J* = 4.5 Hz, 1H, H5), 4.62 – 4.33 (m, 5H, OCH₂Ph), 4.18 (t, *J* = 6.4 Hz, 1H, H2), 3.80 (s, 1H, H3), 3.66 (dd, *J* = 11.2, 2.6 Hz, 1H, H4), 3.61 (dd, *J* = 6.3, 4.1 Hz, 2H, CH₂O), 3.57 (s, 3H,OMe), 3.33 – 3.23 (m, 1H, H4a), 2.31 (s, 3H, Me). ¹³C NMR (101 MHz, CDCl₃) δ 149.98, 139.05, 138.93, 138.16, 130.59, 129.64, 128.61, 128.43, 128.40, 128.36, 128.13, 127.97, 127.75, 127.52, 126.52, 121.97, 115.37, 97.63, 76.92, 76.33, 75.66, 75.16, 73.93, 73.71, 72.69, 71.65, 69.15, 57.1, 34.78, 20.99. [α]_D²⁰= -5,2 (c=1, CHCl₃); MS: m/z calcd for [M + H]⁺ = 567.3, [M + Na]⁺ = 589.3, [M + K]⁺ = 605.2; found [M + H]⁺ = 567.6, [M + Na]⁺ = 589.5, [M + K]⁺ = 605.6.

(5S)



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.13 (m, 15H, Ar), 7.01 (dd, J = 8.3, 1.7 Hz, 1H, H8), 6.82 (s, 1H, H6), 6.75 (d, J = 8.3 Hz, 1H, H9), 5.66 (d, J = 3.2 Hz, 1H, H10a), 4.92 (d, J = 11.5 Hz, 1H, OCH₂Ph), 4.65 – 4.42 (m, 5H, OCH₂Ph), 4.37 – 4.30 (d, J = 2.0 Hz 1H, H5), 4.26 – 4.17 (m, 1H, H2), 3.98 (s, 1H, H3), 3.72 – 3.63 (m, 2H, CH₂O), 3.38 (s, 3H, ArOMe), 3.36 – 3.29 (dd, J = 2.38, 11.78 Hz, 1H, H4), 3.06 – 2.93 (m, 1H, H4a), 2.26 (s, 3H, Me). ¹³C NMR (101 MHz, CDCl₃) δ 151.53, 138.81, 138.10, 137.77, 131.35, 131.03, 129.91, 128.64, 128.60, 128.54, 128.49, 128.26, 128.21, 128.01, 127.93, 127.82, 118.18, 116.64, 94.89, 75.00, 74.84, 74.80, 73.79, 71.55, 71.47, 71.35, 68.96, 56.42, 37.60, 29.93, 20.77. [α]_D²⁰= -2,1 (c=1, CHCl₃); MS: m/z calcd for [M + H]⁺ = 567.3, [M + Na]⁺ = 589.3, [M + K]⁺ = 605.2; found [M + H]⁺ = 567.6, [M + Na]⁺ = 589.5, [M + K]⁺ = 605.6.

(2R,3R,4R,4aR,5R/S,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5,8-dimethoxy-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (15): yield 64%, C5 R/S 53/47

(5R)



¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 8.6 Hz, 1H, H6), 7.37 – 7.21 (m, 15H, Ar),6.56 (dd, *J* = 8.5, 2.4 Hz, 1H, H7), 6.39 (d, *J* = 2.4 Hz, 1H, H8), 5.62 (d, *J* = 2.8 Hz, 1H, H10a), 4.96 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.71 (d, *J* = 4.4 Hz, 1H, H5), 4.60-4.35 (m, 5H, OCH₂Ph), 4.19 (t, *J* = 6.4 Hz, 1H, H2), 3.80 (s, 1H, H3), 3.78 (s, 3H, ArOMe), 3.66 (dd, *J* = 11.1, 2.4 Hz, 1H, H4), 3.60 (t, *J* = 6.3 Hz, 2H, CH₂O), 3.55 (s, 3H, OMe), 3.31 – 3.24 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 160.56, 153.13, 139.03, 138.97, 138.15, 128.61, 128.43, 128.40, 128.12, 127.96, 127.75, 127.52, 127.14, 114.73, 107.86, 100.74, 97.96, 76.04, 75.67, 75.16, 74.05, 73.72, 72.86, 71.82, 69.18, 56.99, 55.55, 34.91, 29.92. [α]_D²⁰= -2,2 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 605.3, [M + K]⁺ = 621.2; found [M + Na]⁺ = 605.6, [M + K]⁺ = 621.5.

(5S)



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.16 (m, 15H, Ar), 6.97 (d, *J* = 8.4 Hz, 1H, H6), 6.47 (dd, *J* = 8.3, 2.5 Hz, 1H, H7), 6.43 (d, *J* = 2.3 Hz, 1H, H8), 5.71 (d, *J* = 3.2 Hz, 1H, H10a), 4.97 – 4.87 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.65 – 4.43 (m, 5H, OCH₂Ph), 4.37 (d, *J* = 2.1 Hz, 1H, H5), 4.19 (d, *J* = 5.5 Hz, 1H, H2), 3.98 (s, 1H, H3), 3.77 (s, 3H, ArOMe), 3.70 – 3.63 (m, 2H,CH₂O), 3.35 (s, 3H, OMe), 3.32 (d, *J* = 2.4 Hz, 1H, H4), 3.00 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 161.44, 155.01, 138.80, 138.10, 137.84, 131.97, 128.67, 128.65, 128.50, 128.25, 128.20, 128.03, 128.02, 127.84, 111.11, 108.00, 101.38, 95.17, 75.34, 74.78, 74.35, 73.79, 71.70, 71.60, 71.57, 69.00, 56.13, 55.54, 37.84, 29.93. [α]_D²⁰= -4,6 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 605.3, [M + K]⁺ = 621.2; found [M + Na]⁺ = 605.7, [M + K]⁺ = 621.6.

(2R,3R,4R,4aR,5R,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-8-methyl-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (16): yield 45%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.8 Hz, 1H, H6), 7.37 – 7.20 (m, 15H, Ar), 6.81 (d, *J* = 7.8 Hz, 1H, H7), 6.66 (s, 1H, H9), 5.63 (d, *J* = 2.8 Hz, 1H, H10a), 4.97 (d, *J* = 11.4 Hz, 1H, OCH₂Ph), 4.74 (d, *J* = 4.2 Hz, 1H, H5), 4.62 – 4.34 (m, 5H, OCH₂Ph), 4.19 (t, *J* = 6.3 Hz, 1H, H2), 3.81 (s, 1H, H3), 3.66 (dd, *J* = 11.1, 2.6 Hz, 1H, H4), 3.62 (dd, *J* = 6.2, 4.6 Hz, 2H, CH₂O), 3.56 (s, 3H, OMe), 3.34 – 3.21 (m, 1H, H4a), 2.31 (s, 3H, Me). ¹³C NMR (101 MHz, CDCl₃) δ 152.11, 139.20, 139.07, 139.00, 138.19, 128.61, 128.44, 128.41, 128.37, 128.12, 127.97, 127.94, 127.75, 127.51, 126.10, 122.28, 119.48, 116.04, 97.74, 76.22, 75.74, 75.16, 74.03, 73.70, 72.84, 71.69, 69.16, 57.06, 34.89, 21.41. [α]_D²⁰= -6,2 (c=1, CHCl₃). MS: m/z calcd for [M + K]⁺ = 605.2; found [M + K]⁺ = 605.2.

(2R,3S,4R,4aR,5R,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-7-methyl-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (17): yield 66%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 7.43 – 7.18 (m, 14H, Ar and H6), 7.09 (dd, J = 6.7, 2.7 Hz, 2H, Ar), 6.99 (d, J = 6.3 Hz, 1H, H8), 6.72 (d, J = 8.2 Hz, 1H, H9), 5.60 (d, J = 3.0 Hz, 1H, H10a), 4.82 (d, J = 10.7 Hz, 1H, OCH₂Ph), 4.74 (d, J = 4.4 Hz, 1H, H5), 4.71 – 4.44 (m, 5H, OCH₂Ph), 4.07 (d, J = 9.9 Hz, 1H, H2), 3.86 – 3.79 (m, 2H, CH₂O), 3.74 (dd, J = 10.8, 1.9 Hz, 1H, H3), 3.72 – 3.65 (m, 1H, H4), 3.55 (s, 3H, OMe), 2.83 (ddd, J = 10.5, 4.4, 3.2 Hz, 1H, H4a), 2.30 (s, 3H, Me). ¹³C NMR (101 MHz, CDCl₃) δ 149.83, 139.14, 138.40, 138.19, 130.78, 129.78, 128.61, 128.58, 128.44, 128.17, 128.01, 127.91, 127.89, 127.55, 126.48, 121.83, 115.50, 97.17, 78.56, 78.54, 76.43, 75.60, 74.92, 73.77, 72.55, 68.67, 57.39, 40.09, 20.98. [α]_D²⁰= +3,7 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 589.3, [M + K]⁺ = 605.2; found [M + Na]⁺ = 589.4, [M + K]⁺ = 605.3. (2R,3S,4R,4aR,5R,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5,8-dimethoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene (18): yield 37%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 8.6 Hz, 1H, H6), 7.38 – 7.19 (m, 10H, Ar), 7.15 – 7.05 (m, 2H, Ar), 6.56 (dd, *J* = 8.5, 2.3 Hz, 1H, H7), 6.40 (t, *J* = 5.9 Hz, 1H, H9), 5.61 (d, *J* = 2.8 Hz, 1H, H10a), 4.83 (d, *J* = 10.6 Hz, 1H, H4a), 4.72 (d, *J* = 4.4 Hz, 1H, OCH₂Ph), 4.70-4.44 (m, 5H, OCH₂Ph), 4.08 (d, *J* = 10.0 Hz, 1H, H2), 3.88 – 3.73 (m, 3H, H3 and CH₂O), 3.79 (s, 3H, OMe), 3.73 – 3.65 (m, 1H, H4), 3.52 (s, 3H, OMe), 2.89 – 2.75 (m, 1H, H4a). ¹³C NMR (101 MHz, CDCl₃) δ 160.66, 152.96, 139.13, 138.37, 138.18, 128.62, 128.60, 128.44, 128.18, 128.15, 128.02, 127.93, 127.91, 127.56, 127.12, 114.56, 108.09, 100.84, 97.50, 78.51, 78.45, 76.18, 75.63, 74.97, 73.78, 72.70, 68.67, 57.29, 55.57, 40.20. [α]_D²⁰ = +11,6 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 605.3, [M + K]⁺ = 621.2; found [M + Na]⁺ = 605.6, [M + K]⁺ = 621.4.

(2R,3S,4R,4aR,5R,10aR)-3,4-bis(benzyloxy)-2-benzyloxymethyl-5-methoxy-8-methyl-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene (19): yield 21%, C5 R/S 100/0



¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, J = 7.9 Hz, 1H, H6), 7.39 – 7.18 (m, 13H, Ar), 7.14 – 7.03 (m, 2H, Ar), 6.80 (d, J = 7.7 Hz, 1H, H7), 6.65 (s, 1H, H9), 5.62 (d, J = 2.7 Hz, 1H, H10a), 4.82 (d, J = 10.6 Hz, 1H, OCH₂Ph), 4.74 (d, J = 3.8 Hz, 1H, H5), 4.72 – 4.39 (m, 5H, OCH₂Ph), 4.07 (d, J = 10.0 Hz, 1H, H2), 3.89 – 3.72 (m, 3H, H3 and CH₂O), 3.68 (t, J = 9.7 Hz, 1H, H4), 3.53 (s, 3H, OMe), 2.87 – 2.78 (m, 1H, H4a), 2.30 (s, 3H, Me). ¹³C NMR (101 MHz, CDCl₃) δ 151.94, 139.37, 139.14, 138.39, 138.20, 128.62, 128.59, 128.44, 128.17, 128.02, 127.92, 127.90, 127.55, 126.05, 122.43, 119.33, 116.17, 97.28, 78.52, 76.93, 76.34, 75.61, 74.96, 73.77, 72.58, 68.68, 57.33, 40.17, 21.38. [α]_D²⁰= +8,3 (c=1, CHCl₃). MS: m/z calcd for [M + Na]⁺ = 589.3, [M + K]⁺ = 605.2; found [M + Na]⁺ = 589.5, [M + K]⁺ = 605.2.

General synthetic strategy for the deprotection of compounds **10-19** (C7 R only) to afford compounds **20-29**. To a 6 mM solution of the protected compound in AcOEt/MeOH 1:1, previously degassed, $Pd(OH)_2$ 5% mol is added and the reaction mixture is stirred under H₂ atmosphere for 45min.-1.5 h. Then the catalist is removed by filtration and the solvent evaporated under reduced pressure to afford pure compounds **20-29**.



(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5-methoxy-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene-3,4-diol (20): yield 97%



¹H NMR (400 MHz, CD₃OD) δ 7.45 (d, *J* = 7.7 Hz, 1H, H6), 7.18 (t, *J* = 7.7 Hz, 1H, H8), 6.94 (dd, *J* = 17.4, 9.9 Hz, 1H, H7), 6.78 (d, *J* = 8.2 Hz, 1H, H9), 5.62 (d, *J* = 3.1 Hz, 1H, H10a), 4.91 (d, *J* = 2.4 Hz, 1H, H5), 3.99 (t, *J* = 6.0 Hz, 1H, H2), 3.81 (s, 2H, H3 and H4), 3.80 – 3.74 (m, 2H, CH₂O), 3.69 (s, 3H, OMe), 3.00 – 2.92 (m, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 156.35, 132.98, 130.04, 125.32, 124.83, 119.22, 100.69, 81.51, 76.68, 71.54, 71.23, 65.57, 61.65, 38.46. [α]_D²⁰= - 7,6 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 305.1, [M + K]⁺ = 321.1; found [M + Na]⁺ = 305.3, [M + K]⁺ = 321.2.

(2R,3R,4R,4aS,5R,10aR)-7-amino-2-hydroxymethyl-5-methoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (21): yield 94%



¹H NMR (400 MHz, CD₃OD) δ 6.89 (s, 1H, H6), 6.65-6.55 (m, 2H, H8 and H9), 5.52 (d, *J* = 3.0 Hz, 1H, H10a), 4.83 (d, *J* = 4.9 Hz, 1H, H5), 3.98 (t, *J* = 5.8 Hz, 1H, H2), 3.87 – 3.78 (m, 2H, H3 and H4), 3.79 – 3.73 (m, 1H, CH₂O), 3.67 (s, 3H, OMe), 2.96 – 2.85 (m, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 149.10, 144.77, 125.69, 121.06, 119.69, 118.60, 117.30, 114.55, 100.39, 81.79, 80.99, 76.53, 71.56, 71.43, 65.59, 61.71, 38.72. [α]_D²⁰= +13,3 (c=1, CHCl₃). MS: m/z calcd for [M + H]⁺ = 298.1 [M + Na]⁺ = 320.1, [M + K]⁺ = 336.1; found [M + H]⁺ = 298.3, [M + Na]⁺ = 320.3, [M + K]⁺ = 336.3.

(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5-methoxy-2,3,4,4a,5,10a-hexahydropyrano[2,3-b]chromene-3,4,7-triol (22): yield 100%



¹H NMR (400 MHz, CD₃OD) δ 6.88 (s, 1H, H6), 6.62 (s, 2H, H8 and H9), 5.54 (d, J = 2.8 Hz, 1H, H10a), 4.84 (d, J = 4.8 Hz, 1H, H5), 3.98 (t, J = 5.9 Hz, 1H, H2), 3.84 (d, J = 2.7 Hz, 1H, H4), 3.81 (s, 1H, H3), 3.76 (dd, J = 5.8, 2.3 Hz, 2H, CH₂O), 3.68 (s, 3H, OMe), 2.98 – 2.84 (m, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 155.33, 149.19, 125.97, 119.88, 119.77, 116.18, 116.06, 100.47, 81.66, 76.55, 71.57, 71.35, 65.59, 61.67, 38.61. [α]_D²⁰= +11,1 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 321.1; found [M + Na]⁺ = 321.2.

(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5,7-dimethoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (23): yield 96%



¹H NMR (400 MHz, CD₃OD) δ 7.00 (d, J = 2.1 Hz, 1H, H6), 6.77 (dd, J = 8.9, 2.3 Hz, 1H, H8), 6.71 (d, J = 8.8 Hz, 1H, H9), 5.57 (d, J = 3.0 Hz, 1H, H10a), 4.87 (s, 1H, H5), 3.98 (t, J = 6.0 Hz, 1H, H2), 3.83 – 3.79 (m, 2H, H3 and H4), 3.76 (dd, J = 6.1, 3.0 Hz, 2H, CH₂O), 3.74 (s, 3H, OMe),

3.68 (s, 3H, OMe), 2.98 - 2.89 (m, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 158.33, 150.09, 125.97, 119.98, 118.90, 114.61, 100.59, 81.54, 76.60, 71.60, 71.26, 65.57, 61.52, 58.83, 38.47. [α]_D²⁰= +8,9 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 335.1; found [M + Na]⁺ = 335.3.

(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5-methoxy-7-methyl-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (24): yield 95%



¹H NMR (400 MHz, CD₃OD) δ 7.24 (s, 1H, H6), 6.98 (d, *J* = 8.2 Hz, 1H, H8), 6.66 (d, *J* = 8.3 Hz, 1H, H9), 5.57 (d, *J* = 3.0 Hz, 1H, H10a), 3.98 (t, *J* = 6.0 Hz, 1H, H2), 3.83 – 3.78 (m, 2H, H3 and H4), 3.77 (dd, *J* = 6.0, 2.9 Hz, 2H, CH₂O), 3.68 (s, 3H, OMe), 2.96 – 2.89 (m, 1H, H4a), 2.26 (s, 3H, Me). ¹³C NMR (101 MHz, CD₃OD) δ 150.14, 130.22, 129.54, 126.29, 120.97, 115.09, 96.65, 77.67, 72.66, 67.62, 67.37, 61.64, 57.73, 34.59, 19.57. [α]_D²⁰= +13,3 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 319.1, [M + K]⁺ = 335.1; found [M + Na]⁺ = 319.4, [M + K]⁺ = 335.4.

(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5,8-dimethoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (25): yield 97%



¹H NMR (400 MHz, CD₃OD) δ 7.32 (d, *J* = 8.6 Hz, 1H, H6), 6.54 (dd, *J* = 8.6, 2.4 Hz, 1H, H7), 6.35 (d, *J* = 2.4 Hz, 1H, H9), 5.58 (d, *J* = 3.0 Hz, 1H, H10a), 4.84 (d, *J* = 4.9 Hz, 1H, H5), 4.00 (t, *J* = 6.0 Hz, 1H, H2), 3.86 – 3.79 (m, 2H, H3 and H4), 3.77 (dd, *J* = 6.0, 3.3 Hz, 2H, CH₂O), 3.74 (s, 3H, OMe), 3.67 (s, 3H, OMe), 2.96-2.86 (m, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 160.95, 153.28, 126.97, 113.53, 107.31, 100.40, 96.94, 77.53, 72.80, 67.62, 67.29, 61.64, 57.61, 54.54, 34.63. [α]_D²⁰= -7,6 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 335.1, [M + K]⁺ = 351.1; found [M + Na]⁺ = 335.4, [M + K]⁺ = 351.3.

(2R,3R,4R,4aS,5R,10aR)-2-hydroxymethyl-5-methoxy-8-methyl-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (26): yield 97%



¹H NMR (400 MHz, CD₃OD) δ 7.29 (d,*J* = 7.8 Hz, 1H, H6), 6.76 (d, *J* = 7.8 Hz, 1H, H7), 6.60 (s, 1H, H9), 5.57 (d, *J* = 3.0 Hz, 1H, H10a), 4.85 (d, *J* = 4.7 Hz, 1H, H5), 3.98 (t, *J* = 5.9 Hz, 1H, H3), 3.83 – 3.73 (m, 4H, CH₂O, H2, H4), 3.67 (s, 3H, OMe), 2.96 – 2.85 (m, 1H, H4a), 2.25 (s, 3H, Me). ¹³C NMR (101 MHz, CD₃OD) δ 156.14, 143.22, 133.19, 129.62, 125.66, 122.29, 119.72, 100.65, 81.54, 76.64, 71.24, 65.63, 61.61, 38.51, 23.91. [α]_D²⁰= +8,3 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺= 319.1; found [M + Na]⁺= 319.3.

(2R,3S,4R,4aS,5R,10aR)-2-(hydroxymethyl)-5-methoxy-7-methyl-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (27): yield 98%



¹H NMR (400 MHz, CD₃OD) δ 7.12 (s, 1H, H6), 6.85 (d, *J* = 8.3 Hz, 1H, H8), 6.54 (d, *J* = 8.3 Hz, 1H, H9), 5.38 (d, *J* = 2.9 Hz, 1H, H10a), 3.75 – 3.59 (m, 3H, CH₂O and H2), 3.53 (s, 4H, H3 and OMe), 3.34 (t, *J* = 8.9 Hz, 1H, H4), 2.50 – 2.40 (m, 1H, H4a), 2.13 (s, 3H, Me). ¹³C NMR (101 MHz, CD₃OD) δ 150.01, 130.29, 129.54, 126.43, 121.13, 115.15, 96.39, 77.56, 73.82, 70.37, 70.10, 61.29, 57.52, 40.03, 19.56. [α]_D²⁰= +13,3 (c=1, CHCl₃); MS: m/z calcd for [M + K]⁺ = 335.1; found [M + K]⁺ = 335.3.

(2R,3S,4R,4aS,5R,10aR)-2-hydroxymethyl-5,8-dimethoxy-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (28): yield 97%



¹H NMR (400 MHz, CD₃OD) δ 7.33 (d, *J* = 8.6 Hz, 1H, H6), 6.54 (dd, *J* = 8.6, 2.4 Hz, 1H, H7), 6.36 (d, *J* = 2.4 Hz, 1H, H9), 5.51 (t, *J* = 6.6 Hz, 1H, H10a), 4.83 (d, *J* = 4.8 Hz, 1H, H5), 3.87 – 3.66 (m, 6H, CH₂O and H4 and OMe), 3.66 (d, *J* = 6.3 Hz, 3H, OMe), 3.48 (t, *J* = 9.2 Hz, 1H, H3),

2.57 (ddd, J = 10.3, 4.9, 3.1 Hz, 1H, H4a). ¹³C NMR (101 MHz, CD₃OD) δ 160.95, 153.13, 127.12, 113.68, 107.43, 100.38, 96.66, 77.40, 73.93, 70.26, 70.02, 61.24, 57.39, 54.54, 40.05. $[\alpha]_D^{20} = +11.6$ (c=1, CHCl₃); MS: m/z calcd for $[M + Na]^+ = 335.1$, $[M + K]^+ = 351.1$; found $[M + Na]^+ = 335.5$, $[M + K]^+ = 351.5$.

(2R,3S,4R,4aS,5R,10aR)-2-hydroxymethyl-5-methoxy-8-methyl-2,3,4,4a,5,10ahexahydropyrano[2,3-b]chromene-3,4-diol (29): yield 98%



¹H NMR (400 MHz, CD₃OD) δ 7.31 (d, *J* = 7.8 Hz, 1H, H6), 6.78 (d, *J* = 7.7 Hz, 1H, H7), 6.62 (s, 1H, H9), 5.52 (d, *J* = 2.9 Hz, 1H, H10a), 4.85 (d, *J* = 4.7 Hz, 1H, H5), 3.85 (dd, *J* = 13.7, 4.5 Hz, 1H, CH₂O), 3.76 (q, *J* = 4.2 Hz, 2H, CH₂O and H2), 3.68 – 3.62 (m, 4H, H4 and OMe), 3.48 (t, *J* = 9.0 Hz, 1H, H3), 2.58 (ddd, *J* = 10.4, 4.9, 3.1 Hz, 1H, H4a), 2.27 (s, 3H, Me). ¹³C NMR (101 MHz, CD₃OD) δ 152.07, 139.31, 126.11, 121.84, 118.54, 115.68, 96.45, 77.50, 73.84, 70.30, 70.04, 61.25, 57.47, 40.04, 19.99. [α]_D²⁰= -7,8 (c=1, CHCl₃); MS: m/z calcd for [M + Na]⁺ = 319.1, [M + K]⁺ = 335.1; found [M + Na]⁺ = 319.4, [M + K]⁺ = 335.3.
































































































Peptide synthesis and purification

Procedure

Aβ1-42 was prepared by solid-phase peptide synthesis on a 433A synthesizer (Applied Biosystems) using Fmoc-protected L-amino acid derivatives, NOVASYN-TGA resin on a 0.1 mM scale.¹ Peptide was cleaved from the resin as previously described and purified by reverse phase HPLC on a semi-preparative C4 column (Waters) using water:acetonitrile gradient elution. Peptide identity was confirmed by MALDI-TOF analysis (model Reflex III, Bruker). Peptide purity was always above 95%.

NMR spectroscopy binding studies

Material and Methods

NMR experiments were recorded on a Varian 400-MHz Mercury. A batch of A β 1-42 was selected that contained pre-amyloidogenic seeds highly toxic to N2a cells.² Immediately before use, lyophilized A β 1-42 was dissolved in 10 mM NaOD in D₂O at a concentration of 160 μ M, then diluted 1:1 with 10 mM phosphate buffer saline, pH 7.4 containing 100 mM NaCl (PBS) and one of the tested compounds. Compounds **20-29** were dissolved in PBS, pH 7.4, and added to the peptide solution. The pH of each sample was verified with a Microelectrode (Mettler Toledo) for 5 mm NMR tubes and adjusted with NaOD or DCl. All pH values were corrected for isotope effect. Basic sequences were employed for 2D-TOCSY, 2D-NOESY and STD experiments. For STD, a train of Gaussian-shaped pulses each of 50 ms was employed to saturate selectively the protein envelope; the total saturation time of the protein envelope was adjusted by the number of shaped pulses and was varied between 5 s and 0.5 s.

Supporting Figures



Figure 1S. ¹H spectra (A, C, E, G, I, M, O, Q, S and U) and 1D-STD spectra (B, D, F, H, L, N, P, R, T and V) of mixtures dissolved in deuterated PBS at 25°C containing A β 1-42 (80 uM) and a test molecule (1.6 mM) (A and B, compound 13; C and D, compound 14; E and F, compound 15; G and H, compound 16; I and L compound 17; M and N, compound 18; O and P, compound 19; Q and R, compound 20; S and T, compound 21 e U and V, compound 22). ¹H spectra were acquired with 64 scans, 1D-STD spectra with 512 scans and 2 s of saturation time.



Figure 28. 2D-NOESY spectra of compounds 21 (A) e 24 (C) dissolved in deuterated PBS, pH 7.5, 25°C, mixing time 0.9 s. trNOESY of mixture containing A β 1-42 (80 uM) and compound 21 (B) or compound 24 (D) dissolved in deuterated PBS, pH 7.5, 25°C, mixing time 0.3 s. Positive cross-peak are blue, negative ones red.



Figure 3S. A) ¹H NMR spectrum of the mixture containing A β 1-42 (80 uM) and compound **24** (1.6 mM) in PBS, pH=7.5, 25°C; **B-F** STD-NMR spectra of the same mixture acquired with different saturation times. (**B**, 0.5 s; **C**, 1,2 s; **D**, 2,0 s; **E** 3,0 s; **F**, 5,0 s).

Molecular Mechanics (MM) and Molecular Dynamics (MD) calculations

Material and Methods

Molecular mechanics and dynamics studies were conducted with MacroModel 9.8.207³ as implemented in version 9.1.207 of the Maestro suite,⁴ using MM3* force field.⁵ The starting coordinates for dynamics calculations were those obtained after energy minimization of the structures, followed by conformational search. In particular, a systematic variation of the torsional degrees of freedom of the molecules permitted different starting structures to be constructed that were further minimized to provide the corresponding local minima. For each compound the conformer with the lowest energy was considered. Simulations were carried out over 5 ns at 298 K with a 0.25 fs time step and a 20 ps equilibration step; 100 structures were sampled and minimized for further analysis. The continuum GB/SA solvent model⁶ was employed and the general PRCG (Polak–Ribiere Conjugate Gradient) method for energy minimization was used. An extended cut-off was applied and the SHAKE procedure for bonds was not selected.

The values of the key proton-proton distances H2-H3, H2-H4, H3-H4, H4a-H10a, H4a-H5 and H5-H10a as well as the values of the dihedral angle H2-C2-C3-H3, H3-C3-C4-H4, H4a-C4a-C10a-H10a and H4a-C4a-C5-H5 were monitored during the MD and are reported in supporting information. All compounds showed the same values of H2-H4, H4a-H5, H4a-H10a, and H5-H10a distances (Fig. 4S), and of the dihedral angles H4a-C4a-C10a-H10a (average value θ = 62°) and H4a-C4a-C5-H5 (average value θ = -53°), which are the diagnostic parameters to identify molecule conformation. On the other hand, on the basis of the distances H2-H3 and H3-H4 (Fig. 6S) and the dihedral angles H2-C2-C3-H3 and H3-C3-C4-H4, molecules **20-29** could be clustered into two groups (**20-26**, with average $\theta_{H2-C2-C3-H3}$ = -60°, $\theta_{H3-C3-C4-H4}$ = 60° and **27-29**, with average $\theta_{H2-C2-C3-H3}$ = -180°, average $\theta_{H3-C3-C4-H4}$ =180°), depending on C3 stereochemistry (Fig.s 7S-16S).

Supporting Figures



Figure 4S. Superimposition of the 30 structures with the lowest energy calculated through MD simulations in water, 298K; A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Fig. 5S. Superimposition of the 30 structures with the lowest energy calculated through MD simulations in water, 298K; A, compound 21; B, compound 24



Fig. 6S. Average values for H2-H3, H2-H4, H3-H4, H4a-H10a and H5-H10a interproton distances. Concerning H2-H3 and H3-H4, compounds **20-29** can be clustered into two groups (bottom).



Figure 7S. H2-H3 distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 8S. H2-H4 distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figura 9S. H3-H4 distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.


Figure 10S. H4a-H10a distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 11S. H4a-H5 distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 128. H10a-H5 distance (Å). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 13S. H2-C2-C3-H3 dihedral angle (°). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 14S. H3-C3-C4-H4 dihedral angle (°). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.



Figure 15S. H4a-C4a-C10a-H10a dihedral angle (°). A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29..



Figure 16S. H4a-C4a-C5-H5 dihedral angle (°).A, compound 20; B, compound 21; C, compound 22; D, compound 23; E, compound 24; F, compound 25; G, compound 26; H, compound 27; I, compound 28; L, compound 29.

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