

Iron (III) Chloride as an Efficient Catalyst for Stereoselective Synthesis of Glycosyl Azides and a Cocatalyst with Cu(0) for the Subsequent Click Chemistry

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SUPPORTING INFORMATION

Detailed procedures for FeCl₃ catalyzed azido glycosylation and analytical data of azido glycosides **1-10**; Detailed procedures for FeCl₃:Cu catalyzed dipolar cycloaddition and analytical data for glycosyl 1,2,3-triazole conjugates **11-16**; procedure for one pot synthesis of **17**; ¹H and ¹³C NMR spectra for **1-17** (63 pages).

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General. ¹H NMR and ¹³C NMR spectra were recorded on 400 MHz ¹H (100 MHz ¹³C) spectrometers in deuteriochloroform with chloroform as an internal reference unless otherwise stated. Chemical shifts are reported in ppm (δ). Coupling constants, *J*, are reported in Hz. Electrospray ionization (ESI) mass spectra were recorded with data reported in the form *m/e* (intensity relative to base peak). Analytical TLC was performed on silica gel glass plates. Visualization was accomplished with UV light or with phosphomolybdic acid (PMA) and KMnO₄ staining agents. Column (flash) chromatography was performed using 32-63 μm silica gels. THF and toluene were dried over Na with benzophenone-ketyl intermediate under nitrogen atmosphere and distilled before use. DMF, CH₂Cl₂ and CHCl₃ were dried over CaH and distilled before use. MeOH was dried over magnesium turnings under nitrogen atmosphere and distilled before use. Oxygen was purged in CHCl₃ before use. All reaction products were isolated as chromatographically pure materials. All reagents and catalysts used for azido glycosylation reaction were purchased from Across Organics. The acetate

derivatives of glucose¹, galactose², mannose³, ribose, xylose⁴, maltose, lactose⁵, maltotriose⁶ were synthesized by using 2 equiv of acetic anhydride (for each hydroxyl group) in the presence of 10 mol% of Sodium acetate under reflux condition. *N*-protected glucosamine derived substrates and propargyl-*O*-linked glucose peracetate⁷ were synthesized by using literature methods.

Caution: Organic azides (e.g. TMSN₃) are potentially explosive and corrosive compounds and should be handled with great care,⁸ despite we did not sense any adverse effects during our studies at small scale reactions.

Attention: It has been documented in the literatures that, ionic azides react with CH₂Cl₂ to form explosive azidochloromethane and/or diazidomethane.⁹ It has been noted that, if these azides were formed, they would be converted to stable triazole derivatives in the subsequent azide-alkyne cycloaddition reactions. However, we did not observe these side products under our catalytic conditions.

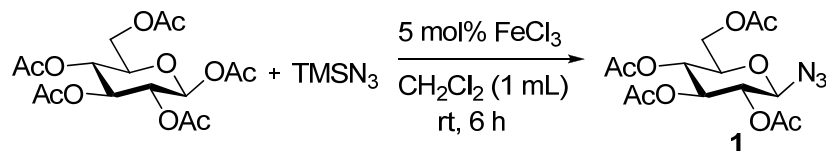
Table S1. Effect of solvents on the azido glycosylation of glucose β -peracetate.^a

Entry	Solvent	Time (h)	Conversion (%) ^b	Yield (%) ^c
1	CH ₃ CN	36	5	n.d.
2	CH ₃ COCH ₃	36	0	--
3	CH ₃ NO ₂	36	decomposed	--
4	THF	36	0	--
5	TBME	40	55	47
6	toluene	40	95	86
7 ^d	toluene	7	quantitative	91
8	CH ₂ Cl ₂	6	quantitative	96

^aCarried out by using TMSN₃ (0.38 mmol, 1.5 equiv), 5 mol% FeCl₃ catalyst in 1 mL of solvent. ^bConversion of β -D-glucose peracetate was determined by ¹H NMR analysis of the crude reaction mixture. ^cIsolated, purified yield. ^dReaction was conducted at 70 °C.

(A) Detailed procedures for FeCl₃ catalyzed azido glycosylation and analytical data of azido glycosides 1-10.

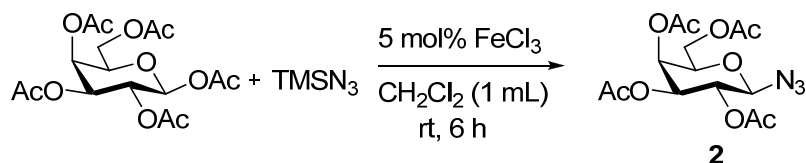
2,3,4,6-tetra-O-acetyl-6-deoxy- β -D-glucopyranosyl azide (1).¹⁰



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*R*,4*S*,5*R*,6*R*)-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2,3,4,5-tetrayl tetraacetate (97.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been

stirred for 5 min at ambient temperature, a solution of trimethyl silyl azide i.e. TMSN₃ (50 µL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 6 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 1/2) on silica gel to afford 89 mg (96%) of 2,3,4,6-tetra-*O*-acetyl-6-deoxy-β-D-glucopyranosyl azide **1** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 5.19 (t, *J* = 9.2, 1H), 5.07 (t, *J* = 10, 1H), 4.92 (t, *J* = 9.2, 1H), 4.63 (d, *J* = 8.8, 1H), 4.25 (dd, *J* = 12.4, 4.8, 1H), 4.14 (dd, *J* = 12.4, 2, 1H), 3.80-3.76 (m, 1H), 2.07 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃), 1.98 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 170.0, 169.2, 169.1, 87.8, 74.0, 72.5, 70.6, 67.9, 61.6, 20.6, 20.4; MS ESI (C₁₄H₁₉O₉N₃, 373): 396 (M+Na⁺, 100); TLC R_f 0.34 (EtOAc/hexanes, 1/2); m.p. 124-125 °C (lit.¹¹ m.p. 127 °C); [α]_D²⁵ -30.2 (c 1.0, CHCl₃) (lit.¹⁰ [α]_D²³ -31 (c 1.0, CHCl₃)).

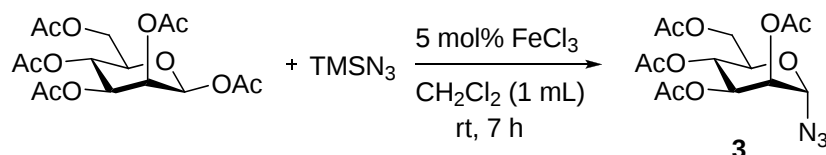
2,3,4,6-tetra-*O*-acetyl-6-deoxy-β-D-galactopyranosyl azide (2**).¹⁰**



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*S*,4*S*,5*R*,6*R*)-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2,3,4,5-tetraol tetraacetate (97.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 6 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 1/2) on silica gel to afford 89 mg (96%) of 2,3,4,6-tetra-*O*-acetyl-6-deoxy-β-D-galactopyranosyl azide **2** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 5.41 (d, *J* = 2.4, 1H), 5.15 (dd, *J* = 10, 8.8, 1H), 5.02 (dd, *J* = 10.4, 3.6, 1H), 4.59 (d, *J* = 8.8, 1H), 4.18-4.14 (m, 2H), 4.00 (td, *J* = 6.8, 0.6, 1H),

2.16 (s, 3H, COCH₃), 2.08 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 1.98 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 170.0, 169.9, 169.2, 88.1, 72.7, 70.6, 67.95, 66.8, 61.2, 20.5, 20.4, 20.35; MS ESI (C₁₄H₁₉O₉N₃, 373): 396 (M+Na⁺, 100); TLC R_f0.35 (EtOAc/hexanes, 1/2); m.p. 95-97 °C (lit.¹¹ m.p. 94-95 °C); [α]_D²⁵ -15.4 (c 1.0, CHCl₃) (lit.¹⁰ [α]_D²³ -14 (c 1.0, CHCl₃)).

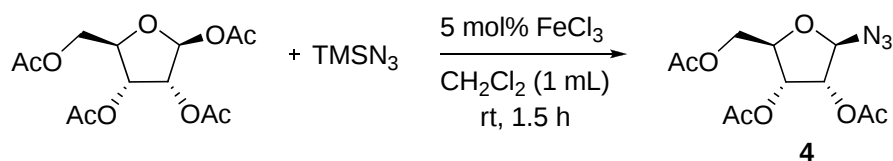
2,3,4,6-tetra-*O*-acetyl-6-deoxy-α-*D*-mannopyranosyl azide (3).¹²



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (3*S*,4*S*,5*S*,6*R*)-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2,3,4,5-tetraol tetraacetate (97.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 7 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL).

The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 1/2) on silica gel to afford 83 mg (89%) of 2,3,4,6-tetra-*O*-acetyl-6-deoxy- α -D-mannopyranosyl azide **3** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 5.37 (d, *J* = 1.5, 1H), 5.28-5.20 (m, 2H), 5.13 (t, *J* = 2.3, 1H), 4.28 (dd, *J* = 12.4, 5.5, 1H), 4.16-4.11 (m, 2H), 2.18 (s, 3H, COCH₃), 2.14 (s, 3H, COCH₃), 2.1 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 169.8, 169.7, 169.6, 87.4, 70.6, 69.1, 68.2, 65.6, 62.1, 20.7, 20.61, 20.6, 20.5; MS ESI (C₁₄H₁₉O₉N₃, 373): 396 (M+Na⁺, 100); TLC R_f 0.32 (EtOAc/hexanes, 1/2). m.p. 103-105 °C (lit.¹³ m.p. 104.1-104.9 °C); [α]_D²⁵ +181.1 (c 1.0, CHCl₃) (lit.¹³ [α]_D²³ +184 (c 1.0, CHCl₃)).

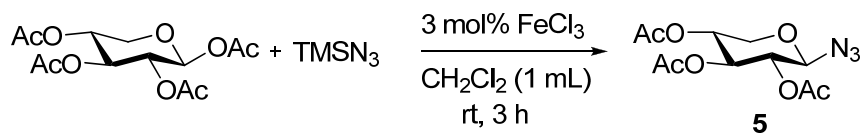
2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl azide (4**).¹⁴**



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (1.2 mg, 0.0075 mmol, 0.03 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*R*,4*R*,5*R*)-5-(acetoxymethyl)tetrahydrofuran-2,3,4-triyl triacetate (79.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at

ambient temperature, a solution of TMSN₃ (50 μL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 1.5 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 1/2) on silica gel to afford 72 mg (96%) of 2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl azide **4** as a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 5.33 (d, *J* = 1.4, 1H), 5.30 (dd, *J* = 6.6, 5.0, 1H), 5.10 (dd, *J* = 4.8, 1.6, 1H), 4.38 (dd, *J* = 12, 3.2, 1H), 4.34-4.30 (m, 1H), 4.12 (dd, *J* = 12.2, 4.2, 1H), 2.092 (s, 3H, COCH₃), 2.09 (s, 3H, COCH₃), 2.04 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 169.5, 169.3, 92.6, 79.3, 74.4, 70.4, 62.9, 20.6, 20.4, 20.3; MS ESI (C₁₁H₁₅O₇N₃, 301): 324 (M+Na⁺, 100); TLC R_f 0.34 (EtOAc/hexanes, 1/2); [α]_D²⁵ +111.3 (c 1.0, CHCl₃) (lit.¹⁴ [α]_D²³ +116 (c 1.0, CHCl₃)).

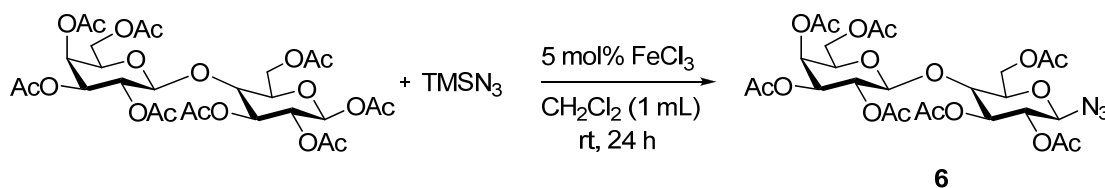
2,3,5-tri-*O*-acetyl- β -D-xylopyranosyl azide (5**).¹⁰**



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (1.2 mg, 0.0075 mmol, 0.03 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*S*,4*S*)-tetrahydro-2*H*-pyran-2,3,4,5-tetrayl tetraacetate (79.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μ L, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 3 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 $\times$ 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 1/2) on silica gel to afford 69 mg (92%) of 2,3,5-tri-*O*-acetyl- β -D-xylopyranosyl azide **5** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 5.18 (t, *J* = 8.8, 1H), 5.00-4.95 (m, 1H), 4.87 (t, *J* = 8.4, 1H), 4.63 (d, *J* = 8.0, 1H), 4.21 (dd, *J* = 12, 5.6, 1H), 3.44 (dd, *J* = 11.6, 9.2, 1H), 2.07 (s, 3H, COCH₃), 2.042 (s, 3H, COCH₃), 2.04 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 169.9, 169.7, 169.3, 88.3, 71.5,

70.4, 68.4, 64.3, 20.6, 20.6; MS ESI ($C_{11}H_{15}O_7N_3$, 301): 324 ($M+Na^+$, 100); TLC R_f 0.34 (EtOAc/hexanes, 1/2); m.p. 84-85 °C (lit.¹⁵ m.p. 87 °C); $[\alpha]_D^{25}$ +88.4 (c 1.0, $CHCl_3$) (lit.¹⁵ $[\alpha]_D^{25}$ +86 (c 1.0, $CHCl_3$)).

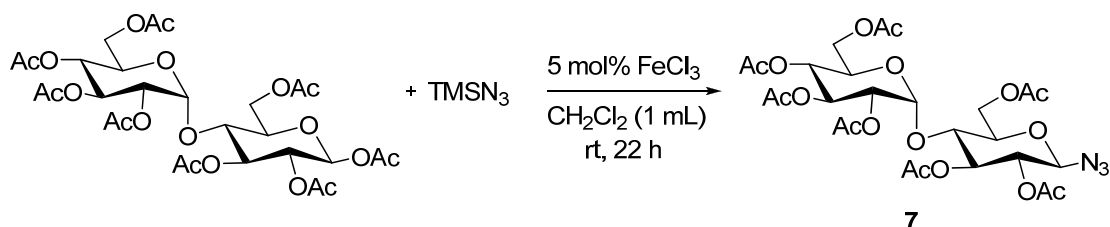
2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- α -D-galactopyranosyl)- β -D-glucopyranosylazide (6).¹⁰



To a 10 mL, round bottomed flask was placed 5 mol% of $FeCl_3$ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH_2Cl_2 (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*S*,4*S*,6*R*)-6-(acetoxymethyl)-5-((2*S*,3*S*,4*S*,6*R*)-3,4,5-triacetoxy-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2-yloxy)tetrahydro-2*H*-pyran-2,3,4-triyl triacetate (169.5 mg, 0.25 mmol, 1 equiv) in CH_2Cl_2 (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of $TMSN_3$ (50 μ L, 44 mg, 0.38 mmol, 1.5 equiv) in CH_2Cl_2 (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 24 h, the reaction mixture was quenched with saturated aqueous $NaHCO_3$ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH_2Cl_2 (2×3 mL). The combined organic layers were washed with brine (5 mL), dried ($MgSO_4$), filtered and

evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 149 mg (90%) of 2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- α -D-galactopyranosyl)- β -D-glucopyranosyl azide **6** as a amorphous solid. ^1H NMR (400 MHz, CDCl_3) δ 5.34 (d, J = 3.6, 1H), 5.20 (t, J = 9.4, 1H), 5.09 (dd, J = 10.4, 8, 1H), 4.94 (dd, J = 10.4, 3.6, 1H), 4.85 (t, J = 9.6, 1H), 4.62 (d, J = 8.8, 1H), 4.52-4.46 (m, 2H), 4.14-4.05 (m, 3H), 3.87 (t, J = 6.8, 1H), 3.81 (t, J = 9.6, 1H), 3.70-3.71 (m, 1H), 2.14 (s, 3H, COCH_3), 2.13 (s, 3H, COCH_3), 2.08 (s, 3H, COCH_3), 2.06 (s, 3H, COCH_3), 2.05 (s, 3H, COCH_3), 2.04 (s, 3H, COCH_3), 1.95 (s, 3H, COCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ 170.3, 170.1, 170.0, 169.6, 169.5, 169.1, 101.1, 87.7, 75.7, 74.8, 72.5, 71.0, 70.9, 70.7, 69.0, 66.6, 61.7, 60.8, 20.8, 20.7, 20.6, 20.5; MS ESI ($\text{C}_{26}\text{H}_{35}\text{O}_{17}\text{N}_3$, 661): 684 ($\text{M}+\text{Na}^+$, 100); TLC R_f 0.20 (EtOAc/hexanes, 1/1); $[\alpha]_{\text{D}}^{25}$ -21.2 (c 1.0, CHCl_3) (lit.¹⁰ $[\alpha]_{\text{D}}^{23}$ -20.4 (c 1.0, CHCl_3)).

2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosylazide (7**).¹⁰**

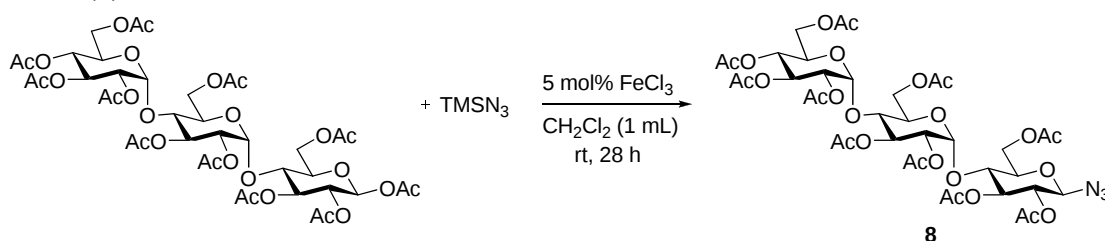


To a 10 mL, round bottomed flask was placed 5 mol% of FeCl_3 (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH_2Cl_2 (0.5 mL) under nitrogen atmosphere. A solution of

(2*S*,3*S*,4*S*,5*R*,6*R*)-6-(acetoxymethyl)-5-((3*S*,4*S*,5*S*,6*R*)-3,4,5-triacetoxy-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2-yl)oxytetrahydro-2*H*-pyran-2,3,4-triyl triacetate (169.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 µL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 22 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 152 mg (92%) of 2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl-β-*D*-glucopyranosyl)-β-*D*-glucopyranosyl azide **7** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 5.40 (d, *J* = 4, 1H), 5.35 (t, *J* = 10, 1H), 5.26 (t, *J* = 9.0, 1H), 5.05 (t, *J* = 9.8, 1H), 4.85 (dd, *J* = 10.8, 4.0, 1H), 4.78 (t, *J* = 8.8, 1H), 4.70 (d, *J* = 8.8, 1H), 4.51 (dd, *J* = 12, 2.4, 1H), 4.26-4.24 (m, 12.4, 2H), 4.07-4.03 (m, 2H), 4.01-3.99 (m, 1H), 3.79-3.70 (m, 1H), 2.15 (s, 3H, COCH₃), 2.10 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 2.04 (s, 3H, COCH₃), 2.02 (s, 3H, COCH₃), 2.01 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃); ¹³C NMR (100 MHz,

CDCl₃) δ 170.4, 170.3, 169.9, 169.8, 169.3, 169.3, 95.6, 77.3, 77.0, 76.7, 74.9, 72.3, 71.4, 69.9, 69.1, 68.5, 67.9, 62.4, 61.4, 20.7, 20.6, 20.5, 20.4; MS ESI (C₂₆H₃₅O₁₇N₃, 661): 684 (M+Na⁺, 100); TLC R_f 0.15 (EtOAc/hexanes, 1/1); m.p. 181-183 °C (lit.^{16a} m.p. 180-182 °C); [α]_D²⁵ -32.1 (c 1.0, CHCl₃) (lit.^{16b} [α]_D²⁴ -31 (c 1.0, CHCl₃)).

2,3,4,6-tetra-O-acetyl-6-deoxy- α -D-glucopyranosyl-(1-4)-2,3,6-tri-O-acetyl-6-deoxy- α -D-glucopyranosyl-(1-4)-2,3,6-tri-O-acetyl-6-deoxy- β -D-glucopyranosyl azide (8).¹⁷



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*R*,4*S*,5*R*,6*R*)-6-(acetoxymethyl)-5-((2*R*,3*R*,4*S*,5*R*,6*R*)-3,4-diacetoxy-6-(acetoxymethyl)-5-((2*R*,3*R*,4*S*,5*R*,6*R*)-3,4,5-triacetoxy-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2-yloxy)tetrahydro-2*H*-pyran-2-yloxy)tetrahydro-2*H*-pyran-2,3,4-triyl triacetate (241.5 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μ L, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 28 h, the reaction mixture was

quenched with saturated aqueous NaHCO_3 solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH_2Cl_2 (2×3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO_4), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 206 mg (87%) of 2,3,4,6-tetra-*O*-acetyl-6-deoxy- α -D-glucopyranosyl-(1-4)-2,3,6-tri-*O*-acetyl-6-deoxy- α -D-glucopyranosyl-(1-4)-2,3,6-tri-*O*-acetyl-6-deoxy- β -D-glucopyranosyl azide **8** as a white solid. ^1H NMR (400 MHz, CDCl_3) δ 5.37-5.32 (m, 2H), 5.32 (d, $J = 6.4$, 1H), 5.25-5.21 (m, 2H), 5.03 (t, $J = 9.8$, 1H), 4.82 (dd, $J = 10.4$, 4.0, 1H), 4.76-4.67 (m, 3H), 4.49-4.41 (m, 2H), 4.29 (dd, $J = 12.2$, 4.4, 1H), 4.21 (dd, $J = 12.4$, 3.6, 1H), 4.15 (dd, $J = 12.4$, 3.2, 1H), 4.02 (dd, $J = 12.4$, 2.0, 1H), 3.96 (d, $J = 9.2$, 1H), 3.93-3.87 (m, 3H), 3.81-3.79 (m, 1H), 2.15 (s, 3H, COCH_3), 2.12 (s, 3H, COCH_3), 2.06 (s, 3H, COCH_3), 2.02 (s, 3H, COCH_3), 2.0 (s, 6H, COCH_3), 1.98 (s, 3H, COCH_3), 1.97 (s, 6H, $2 \times \text{COCH}_3$), 1.96 (s, 3H, COCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ 170.5, 170.4, 170.3, 170.4, 170.2, 169.7, 169.6, 169.4, 169.3, 95.8, 95.6, 87.3, 74.8, 74.0, 73.4, 72.4, 71.6, 71.4, 70.3, 70.0, 69.2, 69.0, 68.4, 67.8, 62.6, 62.2, 61.3, 20.8, 20.7, 20.5, 20.5, 20.4; MS ESI ($\text{C}_{38}\text{H}_{51}\text{O}_{25}\text{N}_3$, 949): 972 ($\text{M}+\text{Na}^+$, 100); TLC R_f 0.10 (EtOAc/hexanes, 1/1); m.p. 95-96 °C; $[\alpha]_{\text{D}}^{25} +69.2$ (c 1.0, CHCl_3) (lit.^{17b} $[\alpha]_{\text{D}}^{25} +65$ (c 1.2, CHCl_3)).

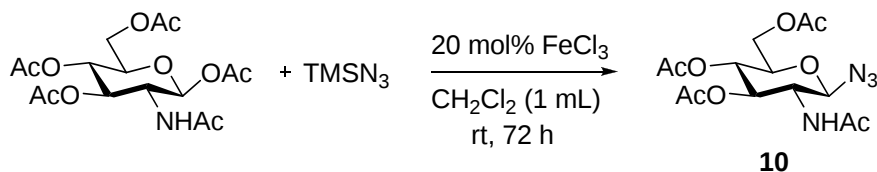
3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl azide (9).¹⁸



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*R*,4*R*,5*S*,6*R*)-6-(acetoxymethyl)-3-(1,3-dioxoisindolin-2-yl)tetrahydro-2*H*-pyran-2,4,5-triyl triacetate (119 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μL, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 22 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 105 mg (91%) of 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl azide **9** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.87 (dd, *J* = 5.4, 3.0, 2H), 7.76 (dd, *J* =

5.5, 3.0, 2H), 5.80 (dd, $J = 10.5, 9.2$, 1H), 5.65 (d, $J = 9.4$, 1H), 5.19 (t, $J = 9.7$, 1H), 4.35 (dd, $J = 4.7, 12.4$, 1H), 4.24 (dd, $J = 12.5, 2.3$, 1H), 4.24 (dd, $J = 10.6, 9.2$, 1H), 3.99-3.96 (m, 1H), 2.12 (s, 3H, COCH₃), 2.03 (s, 3H, COCH₃), 1.86 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 169.9, 169.3, 134.5, 131.2, 123.7, 73.9, 85.5, 70.4, 68.4, 61.7, 53.1, 20.6, 20.5, 20.3; MS ESI (C₂₀H₂₀O₉N₄, 460): 483 (M+Na⁺, 100); TLC R_f 0.28 (EtOAc/hexanes, 1/1); m.p. 135-137 °C (lit.^{18a} m.p. 135-138 °C); $[\alpha]_D^{25} +39.6$ (c 1.0, CHCl₃) (lit.^{18b} $[\alpha]_D^{25} +38$ (c 1.0, CHCl₃)).

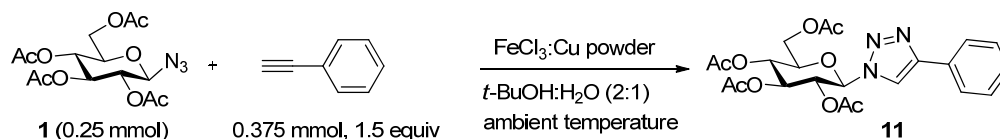
3,4,6-tri-*O*-acetyl-2-*N*-acetyl-2-deoxy- β -D-glucopyranosyl azide (10).¹⁰



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl₃ (8 mg, 0.05 mmol, 0.2 equiv) in anhydrous CH₂Cl₂ (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*R*,4*R*,5*S*,6*R*)-3-acetamido-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2,4,5-triyl triacetate (97 mg, 0.25 mmol, 1 equiv) in CH₂Cl₂ (0.25 mL) was added. After having been stirred for 5 min at ambient temperature, a solution of TMSN₃ (50 μ L, 44 mg, 0.38 mmol, 1.5 equiv) in CH₂Cl₂ (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 72 h, the

reaction mixture was quenched with saturated aqueous NaHCO₃ solution (5 mL). Two layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (5 mL), dried (MgSO₄), filtered and evaporated under reduced pressure. The crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 86 mg (93%) of 3,4,6-tri-*O*-acetyl-2-*N*-acetyl-2-deoxy-β-D-glucopyranosyl azide **10** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 6.09 (d, *J* = 8.8, 1H, NH), 5.25 (t, *J* = 10, 1H), 5.08 (t, *J* = 9.6, 1H), 4.78 (d, *J* = 9.2, 1H), 4.25 (dd, *J* = 4.8, 12.4, 1H), 4.16-4.13 (m, 1H), 3.91 (dd, *J* = 19.4, 9.3, 1H), 3.81 (m, 1H), 2.09 (s, 3H, COCH₃), 2.024 (s, 3H, COCH₃), 2.02 (s, 3H, COCH₃), 1.97 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.9, 170.7, 170.6, 169.3, 88.3, 76.7, 73.9, 72.1, 68.1, 61.9, 54.0, 23.1, 20.7, 20.6, 20.5; MS ESI (C₁₄H₂₀O₈N₄, 372): 395 (M+Na⁺, 100); TLC R_f 0.20 (EtOAc/hexanes, 1/1); m.p. 161-162 °C (lit.¹⁹ m.p. 158-161 °C); [α]_D²⁵ -45.9 (c 1.0, CHCl₃) (lit.¹⁰ [α]_D²³ -47.7 (c 1.0, CHCl₃)).

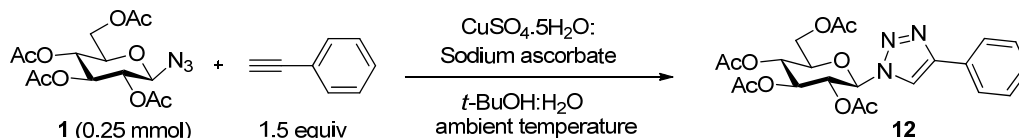
Table S2: Optimization of FeCl₃/Cu(0) ratio for cycloaddition reactions (i. e. click chemistry) between β-azido glucose and 2-phenylethyne.^a



Entry	FeCl ₃ mol%	Cu powder mol%	FeCl ₃ :Cu ratio	Time, h	Yield (%)
1	20	0	1:0	48	No reaction
2	0	100	0:1	48	No reaction
3	20	20	1:1	48	No reaction
4	20	30	1:1.5	48	Sluggish reaction
5	20	40	1:2	40	92
6	20	100	1:5	10	94

^aReaction details: β-azido glucose (0.25 mmol, 1 equiv), 2-phenylethyne (0.375 mmol, 1.5 equiv) in *t*-BuOH:H₂O (2 mL:1 mL). ^bIsolated yield after purification by silica gel column chromatography.

Table S3: Control experiments for cycloaddition reactions between β-azido glucose and 2-phenylethyne^a by using Sharpless and Fokin's reagents.²⁰



Entry	CuSO ₄ (mol%)	sodium ascorbate (mol%)	CuSO ₄ : sodium ascorbate ratio	Time (h)	Conversion (%) ^b	Yield (%) ^c
1	3	30	1:10	48	8	n.d.
2	20	40	1:2	20	7	n.d.
3	20	40	1:2	70	30	24 ^d
4	20	100	1:5	20	15	n.d.
5	20	100	1:5	70	45	40

^aReaction details: β-azido glucose (0.25 mmol, 1 equiv), 2-phenylethyne (0.375 mmol, 1.5 equiv) in *t*-BuOH:H₂O (2 mL:1 mL). ^bConversion of β-azido glucose **1** was determined by ¹H NMR analysis of the crude reaction mixture. ^cIsolated yield after purification by silica gel column chromatography. n.d. = not determined. ^dRemaining starting material was recovered.

Table S4: Control experiments for cyclo-addition reaction between β -azido glucose and 2-phenylethyne^a by using bimetallic catalyst Cu(OTf)₂:Cu system.²¹

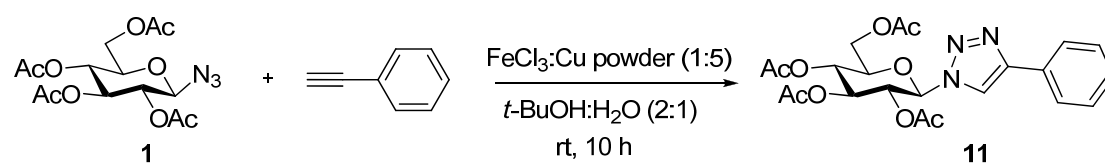
1 (0.25 mmol) + 1.5 equiv $\xrightarrow[\text{Solvent, ambient temperature}]{\text{Cu(OTf)}_2: \text{Cu Powder}}$ 12

Entry	Cu(OTf) ₂ (mol%)	Cu Powder (mol%)	Cu(OTf) ₂ : Cu(0) ratio	Solvent	Time (h)	Conv. (%) ^b	Yield (%) ^c
1	10	10	1:1	CH ₃ CN	40	21	16 ^d
2	10	50	1:5	CH ₃ CN	12	quant.	92
3	10	10	1:1	<i>t</i> -BuOH:H ₂ O	35	0	n.d.
4	10	50	1:5	<i>t</i> -BuOH:H ₂ O	35	9	n.d.

^aReaction details: β -azido glucose (0.25 mmol, 1 equiv), 2-phenylethyne (0.375 mmol, 1.5 equiv) in CH₃CN (6 mL) or *t*-BuOH:H₂O (2 mL:1 mL). ^bConversion of β -azido glucose **1** was determined by ¹H NMR analysis of the crude reaction mixture. ^cIsolated yield after purification by silica gel column chromatography. n.d. = not determined. ^dRemaining starting material was recovered.

(B) Detailed procedures for FeCl₃:Cu catalyzed 1,3-dipolar cycloaddition and analytical data for glycosyl 1,2,3-triazole conjugates 11-16.

Phenyl 2,3,4,6-tetra-*O*-acetyl-6-deoxy- β -D-glucopyranosyl 1,2,3-triazole (11).²²

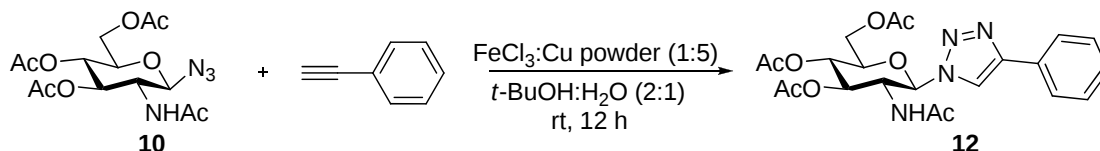


In a 10 mL, round bottomed flask were placed 20 mol% of FeCl₃ (8.1 mg, 0.05 mmol, 0.2 equiv) and Cu_(s) powder (16 mg, 0.25 mmol, 1.0 equiv) in 1:1 mixture of *t*-BuOH:H₂O (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 2,3,4,6-tetra-*O*-acetyl-6-deoxy- β -D-glucopyranosyl azide

1 (93 mg, 0.25 mmol, 1 equiv) in *t*-BuOH (1 mL) was added followed by addition of ethynylbenzene (41 μ L, 38 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 10 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 $\times$ 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1) on silica gel to afford 109 mg (92%) of phenyl 2,3,4,6-tetra-*O*-acetyl-6-deoxy- β -D-glucopyranosyl 1,2,3-triazole **11** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 8.0 (s, 1H), 7.83 (d, *J* = 7.2, 2H), 7.43 (t, *J* = 7.2, 2H), 7.36 (d, *J* = 7.6, 1H), 5.93 (d, *J* = 9.2, 1H), 5.52 (t, *J* = 9.4, 1H), 5.44 (t, *J* = 9.4, 1H), 5.27 (t, *J* = 9.6, 1H), 4.33 (dd, *J* = 12.6, 5.0, 1H), 4.16 (dd, *J* = 12.6, 1.8, 1H), 4.05-4.02 (m, 1H), 2.084 (s, 3H, COCH₃), 2.08 (s, 3H, COCH₃), 2.04 (s, 3H, COCH₃), 1.88 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 169.9, 169.4, 169.0, 148.5, 129.9, 128.9, 128.6, 125.9, 117.7, 85.8, 75.3, 72.7, 70.2, 67.7, 61.6, 20.7, 20.6, 20.5, 20.2; MS ESI (C₂₂H₂₅O₉N₃, 475): 498 (M+Na⁺, 100); TLC R_f 0.42 (EtOAc/hexanes, 1/1); m.p. 211-213 °C (lit.^{22b} m.p.

213-215 °C); $[\alpha]_{\text{D}}^{25}$ -54.5 (*c* 1.0, CHCl₃) (lit.^{22c} $[\alpha]_{\text{D}}^{20}$ -56.2 (*c* 1.0, CHCl₃)).

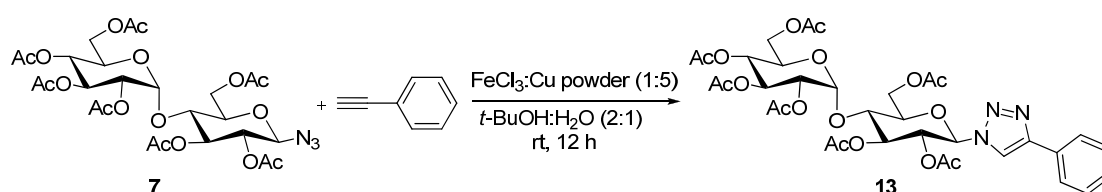
Phenyl 3,4,6-tri-*O*-acetyl-2-*N*-acetyl-6-deoxy-β-*D*-glucopyranosyl 1,2,3-triazole (12).



In a 10 mL, round bottomed flask were placed 20 mol% of FeCl₃ (8.1 mg, 0.05 mmol, 0.2 equiv) and Cu_(s) powder (16 mg, 0.25 mmol, 1.0 equiv) in 1:1 mixture of *t*-BuOH:H₂O (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 3,4,6-tri-*O*-acetyl-2-*N*-acetyl-2-deoxy-β-*D*-glucopyranosyl azide **10** (93 mg, 0.25 mmol, 1 equiv) in *t*-BuOH (1 mL) was added followed by addition of ethynylbenzene (41 μL, 38 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 12 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1)

on silica gel to afford 105 mg (89%) of phenyl 3,4,6-tri-*O*-acetyl-2-*N*-acetyl-6-deoxy- β -D-glucopyranosyl 1,2,3-triazole **13** as a off white solid. ^1H NMR (400 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$) δ 8.04 (s, 1H), 7.52 (d, $J = 7.3$, 2H), 7.13 (t, $J = 7.4$, 2H), 7.05 (d, $J = 7.2$, 1H), 5.81 (d, $J = 10$, 1H), 5.19 (t, $J = 10$, 1H), 4.99 (t, $J = 9.6$, 1H), 4.36 (t, $J = 10.2$, 1H), 4.05 (dd, $J = 12.4$, 4.8, 1H), 3.89-3.82 (m, 2H), 1.79 (s, 3H), 1.787 (s, 3H, COCH_3), 1.76 (s, 3H, COCH_3), 1.46 (s, 3H, COCH_3); ^{13}C NMR (100 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$) δ 171.5, 170.6, 170.1, 169.4, 147.6, 129.2, 128.3, 128.0, 125.2, 118.7, 85.4, 74.2, 72.0, 67.8, 61.4, 52.6, 21.4, 19.7, 19.7, 19.6; MS ESI ($\text{C}_{22}\text{H}_{26}\text{O}_8\text{N}_4$, 474): 497 ($\text{M}+\text{Na}^+$, 100); TLC R_f 0.40 (EtOAc/hexanes, 1/1); m.p. 228-229 °C; $[\alpha]_{\text{D}}^{25}$ -62.4 (c 1.0, CHCl_3).

Phenyl 2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosyl 1,2,3-triazole (13**).²²**

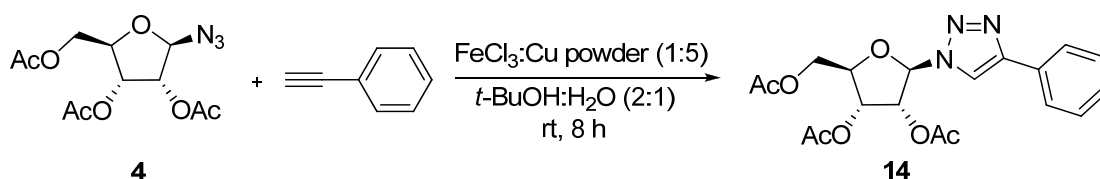


In a 10 mL, round bottomed flask were placed 20 mol% of FeCl_3 (8.1 mg, 0.05 mmol, 0.2 equiv) and $\text{Cu}_{(\text{s})}$ powder (16 mg, 0.25 mmol, 1.0 equiv) in 1:1 mixture of t -BuOH: H_2O (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosylazide **7** (165 mg, 0.25 mmol, 1 equiv)

in *t*-BuOH (1 mL) was added followed by addition of ethynylbenzene (41 μ L, 38 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 12 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 $\times$ 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 3/2) on silica gel to afford 166 mg (87%) of phenyl 2,3,6-tri-*O*-acetyl-4-*O*-(2',3',4',6'-tetra-*O*-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosyl 1,2,3-triazole **13** as a off white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.81 (d, *J* = 7.6, 2H), 7.42 (t, *J* = 7.4, 2H), 7.34 (d, *J* = 7.4, 1H), 5.94 (d, *J* = 9.2, 1H), 5.51-5.36 (m, 4H), 5.07 (t, *J* = 9.8, 1H), 4.88 (dd, *J* = 10.8, 4, 1H), 4.49 (dd, *J* = 12.4, 2.0, 1H), 4.29-4.23 (m, 2H), 4.17 (t, *J* = 8.8, 1H), 4.08 (dd, *J* = 12.6, 1.8, 1H), 4.02-3.99 (m, 2H), 2.12 (s, 3H, COCH₃), 2.10 (s, 3H, COCH₃), 2.07 (s, 3H, COCH₃), 2.03 (s, 3H, COCH₃), 2.01 (s, 3H, COCH₃), 1.85 (s, 3H, COCH₃); 1.72 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 170.5, 170.3, 169.9, 169.4, 169.3, 148.4, 129.9, 128.9, 128.6, 125.9, 95.9, 85.3, 75.4, 75.2, 72.5,

70.8, 70.0, 69.2, 68.8, 67.9, 62.5, 61.5, 20.8, 20.7, 20.6, 20.6, 20.2; MS ESI (C₃₄H₄₁O₁₇N₃, 763): 786 (M+Na⁺, 100); TLC R_f 0.21 (EtOAc/hexanes, 1/1); m.p. 170-171 °C; [α]_D²⁵ -38.5 (c 1.0, CHCl₃).

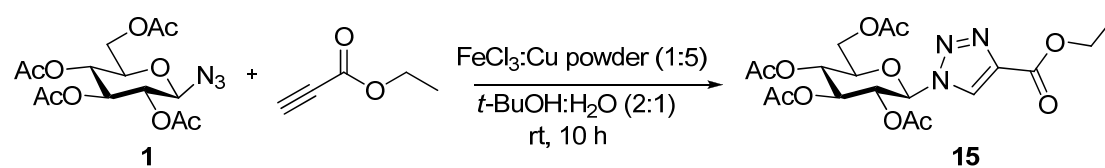
Phenyl 2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl 1,2,3-triazole (14).²²



In a 10 mL, round bottomed flask were placed 20 mol% of FeCl₃ (8.1 mg, 0.05 mmol, 0.2 equiv) and Cu_(s) powder (16 mg, 0.25 mmol, 1.0 equiv) in 1:1 mixture of *t*-BuOH:H₂O (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl azide **4** (75 mg, 0.25 mmol, 1 equiv) in *t*-BuOH (1 mL) was added followed by addition of ethynylbenzene (41 μL, 38 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 8 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove

inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1) on silica gel to afford 97 mg (96%) of phenyl 2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl 1,2,3-triazole **14** as a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ 7.96 (s, 1H), 7.80 (d, $J = 7.2$, 2H), 7.40 (t, $J = 7.6$, 2H), 7.32 (d, $J = 7.2$, 1H), 6.18 (d, $J = 3.6$, 1H), 5.9 (dd, $J = 4.8$, 4.0, 1H), 5.63 (t, $J = 5.4$, 1H), 4.48 (dd, 8.2, 4.2, 1H), 4.40 (dd, $J = 12.4$, 2.8, 1H), 4.23 (dd, $J = 12.4$, 4, 1H), 2.10 (s, 6H, $2 \times \text{COCH}_3$), 2.02 (s, 3H, COCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ 170.2, 169.3, 169.2, 148.0, 129.9, 128.8, 125.7, 118.8, 89.9, 80.0, 74.1, 70.6, 62.7, 20.5, 20.3, 20.2; MS ESI ($\text{C}_{19}\text{H}_{21}\text{O}_7\text{N}_3$, 403): 426 ($\text{M}+\text{Na}^+$, 100); TLC R_f 0.43 (EtOAc/hexanes, 1/1); $[\alpha]_{\text{D}}^{25} +87.2$ (c 1.0, CHCl_3).

2,3,4,6-tetra-*O*-acetyl-(4'-ethylcarboxylate)- β -D-glucopyranosyl 1,2,3-triazole (15).²²

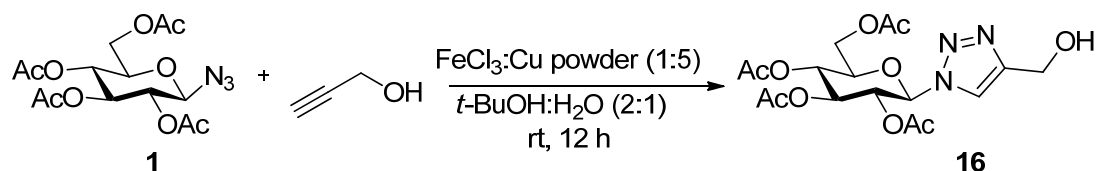


In a 10 mL, round bottomed flask, 20 mol% of FeCl_3 (8.1 mg, 0.05 mmol, 0.2 equiv) and $\text{Cu}_{(s)}$ powder (16 mg, 0.25 mmol, 1.0 equiv) were placed in 1:1 mixture of $t\text{-BuOH}:\text{H}_2\text{O}$ (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 2,3,4,6-tetra-*O*-acetyl-6-deoxy- β -D-glucopyranosyl azide **1** (93 mg, 0.25 mmol, 1 equiv) in $t\text{-BuOH}$ (1 mL) was added followed by addition of

ethyl propiolate (38 μ L, 37 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 10 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 $\times$ 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1) on silica gel to afford 112 mg (95%) of 2,3,4,6-tetra-*O*-acetyl-(4'-ethyl carboxylate)- β -D-glucopyranosyl 1,2,3-triazole **15** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 8.33 (s, 1H), 5.92 (d, *J* = 8.8, 1H), 5.46-5.37 (m, 2H), 5.24 (t, *J* = 9.6, 1H), 4.42 (q, *J* = 7.1, 2H), 4.31 (dd, *J* = 12.6, 5.0, 1H), 4.15 (dd, *J* = 12.8, 1.4, 1H), 4.03-4.0 (m, 1H), 2.08 (s, 3H, COCH₃), 2.06 (s, 3H, COCH₃), 2.02 (s, 3H, COCH₃), 1.89 (s, 3H, COCH₃), 1.41 (t, *J* = 7, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 169.8, 169.3, 168.8, 160.1, 140.9, 126.0, 85.8, 75.3, 72.3, 70.4, 67.5, 61.5, 61.4, 20.6, 20.4, 20.0, 14.2; MS ESI (C₁₉H₂₅O₁₁N₃, 494): 517 (M+Na⁺, 100); TLC R_f 0.39 (EtOAc/hexanes, 1/1); m.p. 170-171 °C (lit.^{22d} m.p. 172-174 °C); $[\alpha]_D^{25}$ -46.3 (c 1.0, CHCl₃) (lit.^{22d} $[\alpha]_D^{25}$ -62 (c 0.2, CHCl₃)).

2,3,4,6-tetra-*O*-acetyl-(4'-hydroxymethyl)-6-deoxy-β-D-glucopyranosyl

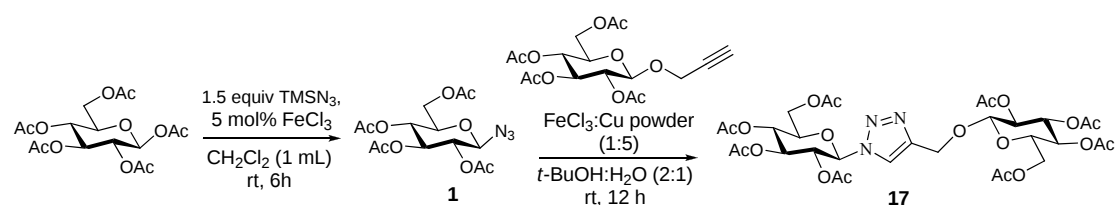
1,2,3-triazole (16).²²



In a 10 mL, round bottomed flask were placed 20 mol% of FeCl₃ (8.1 mg, 0.05 mmol, 0.2 equiv) and Cu_(s) powder (16 mg, 0.25 mmol, 1.0 equiv) in 1:1 mixture of *t*-BuOH:H₂O (1 mL : 1 mL). The resulting mixture was stirred at ambient temperature for 5 min and a solution of 2,3,4,6-tetra-*O*-acetyl-6-deoxy-β-D-gluco-pyranosyl azide **1** (93 mg, 0.25 mmol, 1 equiv) in *t*-BuOH (1 mL) was added followed by addition of prop-2-yn-1-ol (22 μL, 21 mg, 0.375 mmol, 1.5 equiv). The resulting reaction mixture was stirred at ambient temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 12 h, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered, passed through Celite bed to remove inorganic salts and concentrated under reduced pressure. The resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1) on silica

gel to afford 100 mg (93%) of 2,3,4,6-tetra-*O*-acetyl-(4'-hydroxymethyl) -6-deoxy- β -D-glucopyranosyl 1,2,3-triazole **16** as a off white solid. ^1H NMR (400 MHz, CDCl_3) δ 7.84 (s, 1H), 5.89 (d, $J = 8.4$, 1H), 5.46-5.37 (m, 2H), 5.23 (t, $J = 9.4$, 1H), 4.74 (brs, 2H), 4.25 (dd, $J = 12.6$, 5, 1H), 4.10 (d, $J = 12.4$, 1H), 4.01-3.99 (m, 1H), 3.42 (brs, 1H), 2.021 (s, 3H, COCH_3), 2.02 (s, 3H, COCH_3), 1.98 (s, 3H, COCH_3), 1.82 (s, 3H, COCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ 170.5, 169.9, 169.4, 169.0, 148.5, 120.2, 85.6, 75.0, 72.6, 70.3, 67.7, 61.5, 56.3, 20.6, 20.5, 20.4, 20.1; MS ESI ($\text{C}_{17}\text{H}_{23}\text{O}_{10}\text{N}_3$, 429): 452 ($\text{M}+\text{Na}^+$, 100); TLC R_f 0.40 (EtOAc/hexanes, 1/1); m.p. 149-151 °C (lit.^{22d} m.p. 152-154 °C); $[\alpha]_{\text{D}}^{25}$ -9.1 (c 1.0, CHCl_3) (lit.^{22c} $[\alpha]_{\text{D}}^{20}$ -6 (c 1.0, CHCl_3)).

(C) Procedure for one pot synthesis of 2,3,4,6-tetra-*O*-acetyl-4'-*O*-(2,3,4,6-tetra-*O*-acetyl-6-deoxy-glucopyranose)-6-deoxy- β -D-glucopyranosyl triazole (17**).²²**



To a 10 mL, round bottomed flask was placed 5 mol% of FeCl_3 (2 mg, 0.0125 mmol, 0.05 equiv) in anhydrous CH_2Cl_2 (0.5 mL) under nitrogen atmosphere. A solution of (2*S*,3*S*,4*S*,5*R*)-6-(acetoxymethyl)tetrahydro-2*H*-pyran-2,3,4,5-tetraol tetraacetate (98 mg, 0.25 mmol, 1 equiv) in CH_2Cl_2 (0.25 mL) was added. After having been stirred for 5 min, a solution of TMSN_3 (50 μL , 44 mg, 0.38 mmol, 1.5 equiv) in CH_2Cl_2 (0.25 mL) was added slowly. The resulting reaction mixture was stirred at ambient

temperature and the progress of the reaction was monitored by TLC. After complete consumption of starting material in 6 h as judged by TLC, the reaction mixture was concentrated under reduced pressure and the resulting residue was dried in *vacuo*. The residue was then suspended in a mixture of *t*-BuOH:H₂O (1 mL: 1 mL). After having been stirred for 5 min, a solution of 15 mol% of FeCl₃ (6.0 mg, 0.0375 mmol, 0.15 equiv) and (3*R*,4*S*,5*S*,6*R*)-2-(acetoxymethyl)-6-(prop-2-ynyloxy)tetrahydro-2*H*-pyran-3,4,5-triyl triacetate (145 mg, 0.375 mmol, 1.5 equiv) in *t*-BuOH (1 mL) was added followed by addition of Cu_(s) powder (16 mg, 0.25 mmol, 1.0 equiv). After having been stirred for 12 h at ambient temperature, the organic volatiles were removed under reduced pressure and the resulting mixture was partitioned between H₂O and CH₂Cl₂ (5 mL each). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic layers were washed with brine (10 mL), dried (MgSO₄), filtered and passed through Celite bed to remove inorganic salts. The organic layer was then concentrated under reduced pressure and the resulting crude residue was purified by column chromatography (EtOAc/hexanes, 1/1) on silica gel to afford 171 mg (90%) of **17** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.80 (s, 1H), 5.86 (d, *J* = 8.8, 1H), 5.42-5.39 (m, 2H), 5.23 (t, *J* = 9.6, 1H), 5.16 (t, *J* = 9.4, 1H), 5.08 (t, *J* = 9.6, 1H), 4.97 (td, *J* = 8.2, 1.2, 1H), 4.90 (d, *J* = 12.8, 1H), 4.79 (d, *J* = 12.8, 1H), 4.53 (d, *J* = 8.0, 1H), 4.31-4.25 (m, 2H), 4.19-4.10 (m,

1H), 4.02-3.98 (m, 1H), 3.76-3.72 (m, 1H), 2.10 (s, 3H, COCH₃), 2.06 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 2.02 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃), 1.96 (s, 3H, COCH₃), 1.94 (s, 3H, COCH₃), 1.88 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 170.4, 170.2, 169.8, 169.4, 169.3, 169.0, 144.2, 121.9, 98.7, 75.2, 72.6, 72.4, 71.7, 71.0, 70.4, 68.3, 67.6, 61.8, 61.7, 61.4, 20.7, 20.6, 20.5, 20.5, 20.0; MS ESI (C₃₀H₃₉O₁₉N₃, 745): 768 (M+Na⁺, 100); TLC R_f0.33 (EtOAc/hexanes, 1/1); m.p. 198-200 °C (lit.²³ m.p. 199-199.5 °C); [α]_D²⁵ -41.6 (c 1.0, CHCl₃).

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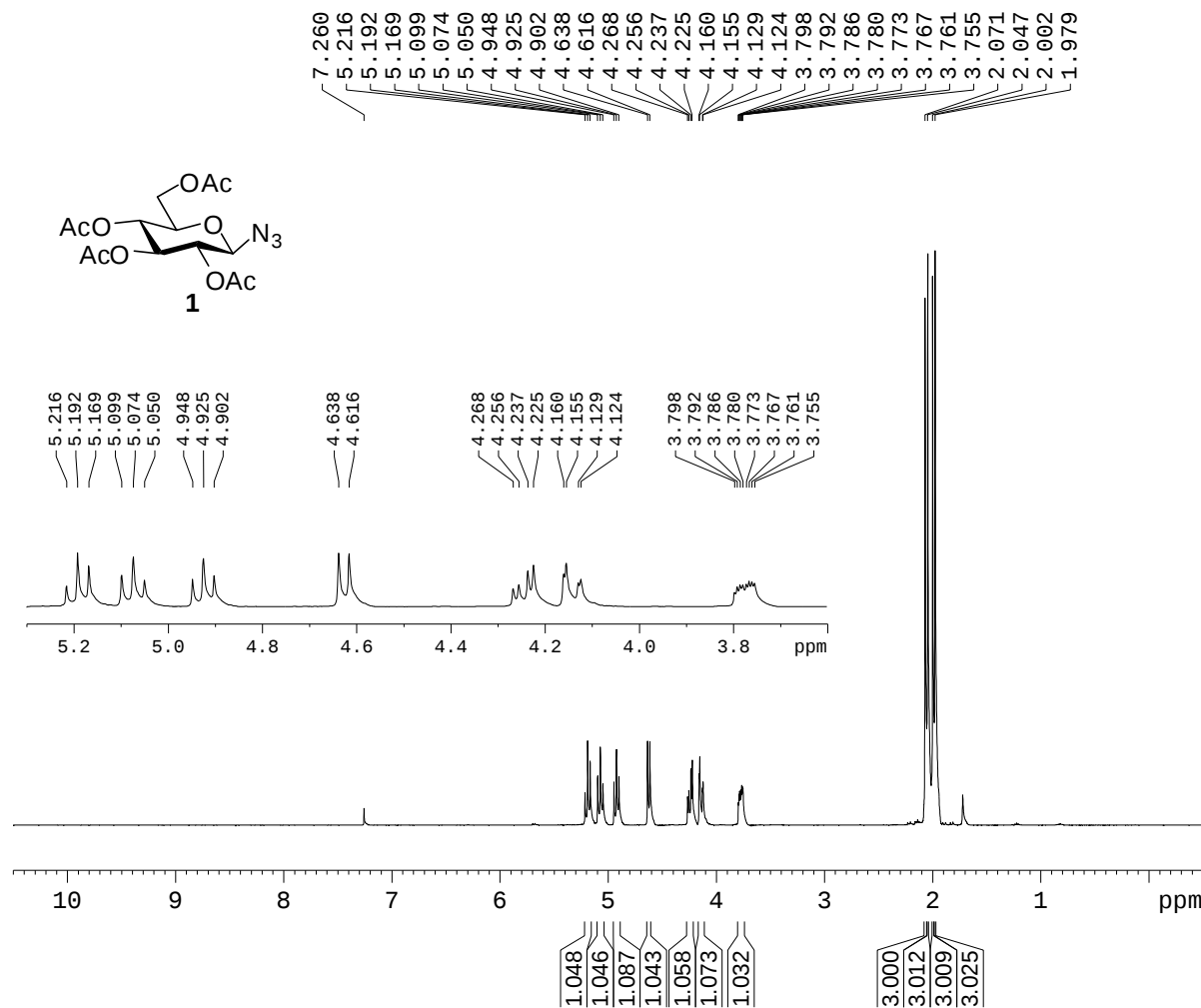
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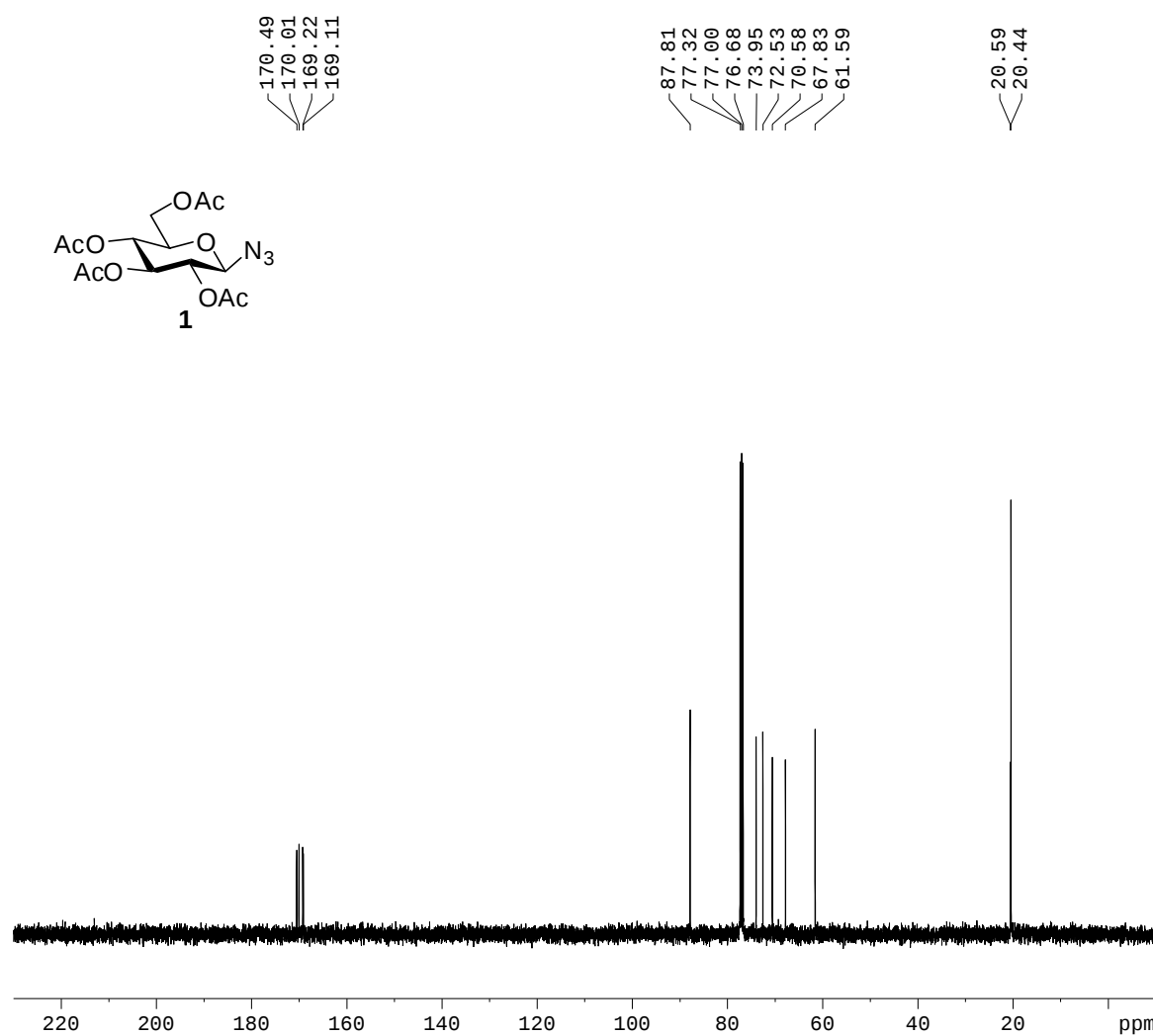
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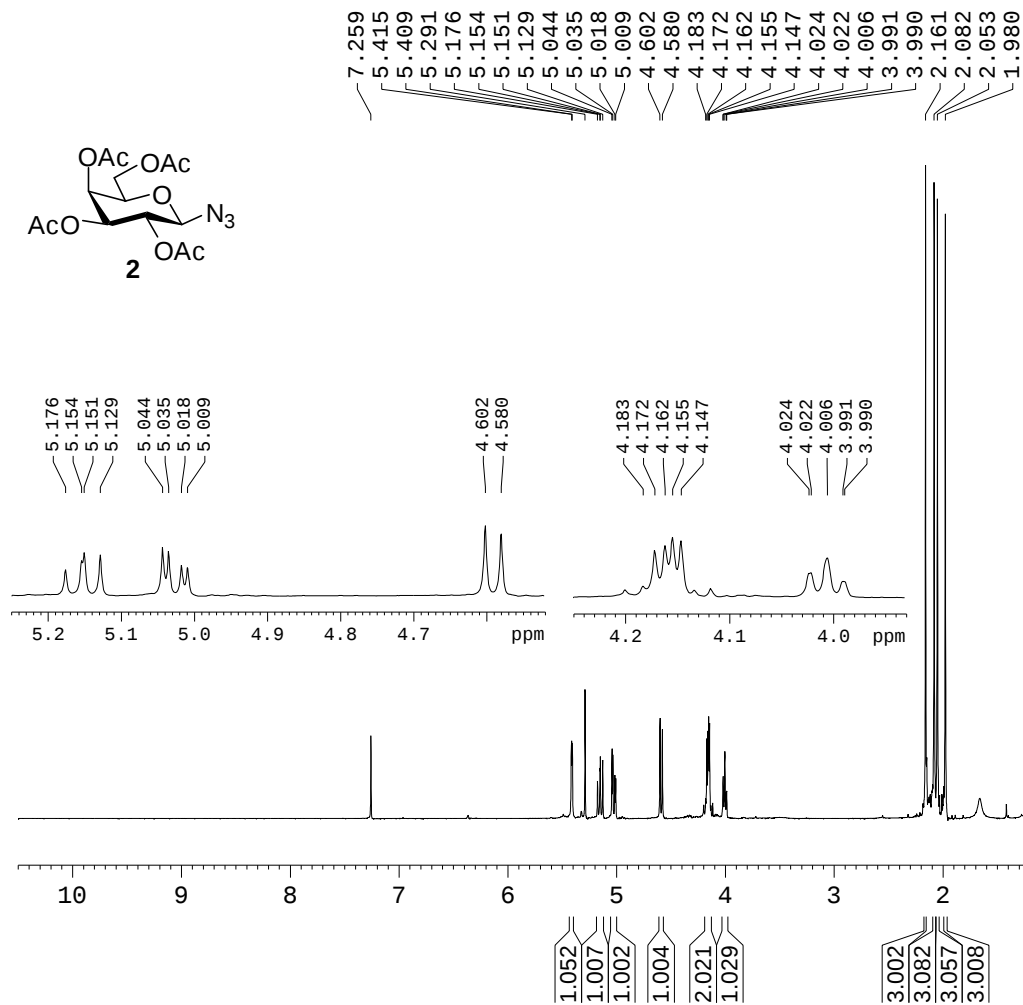
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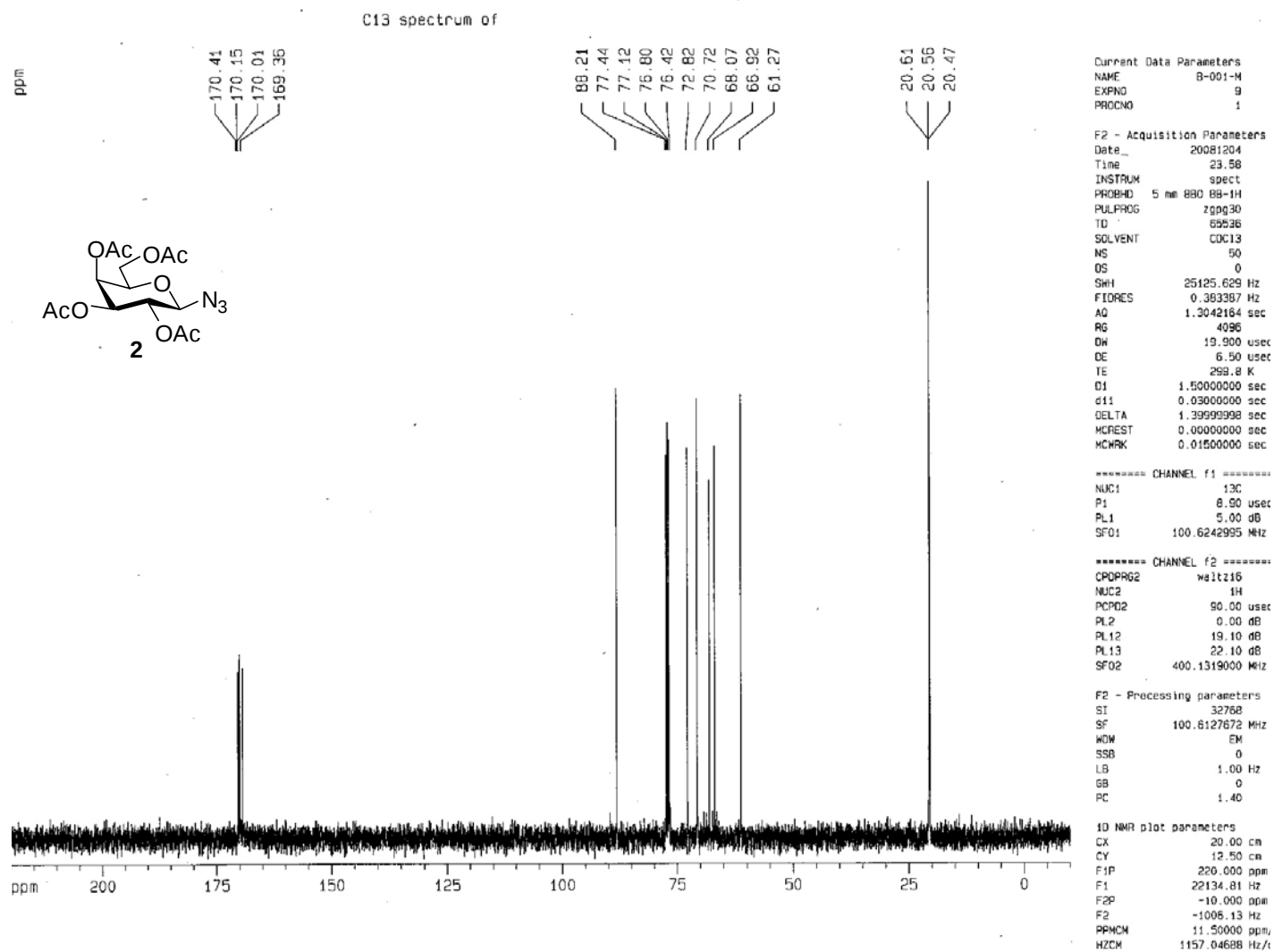
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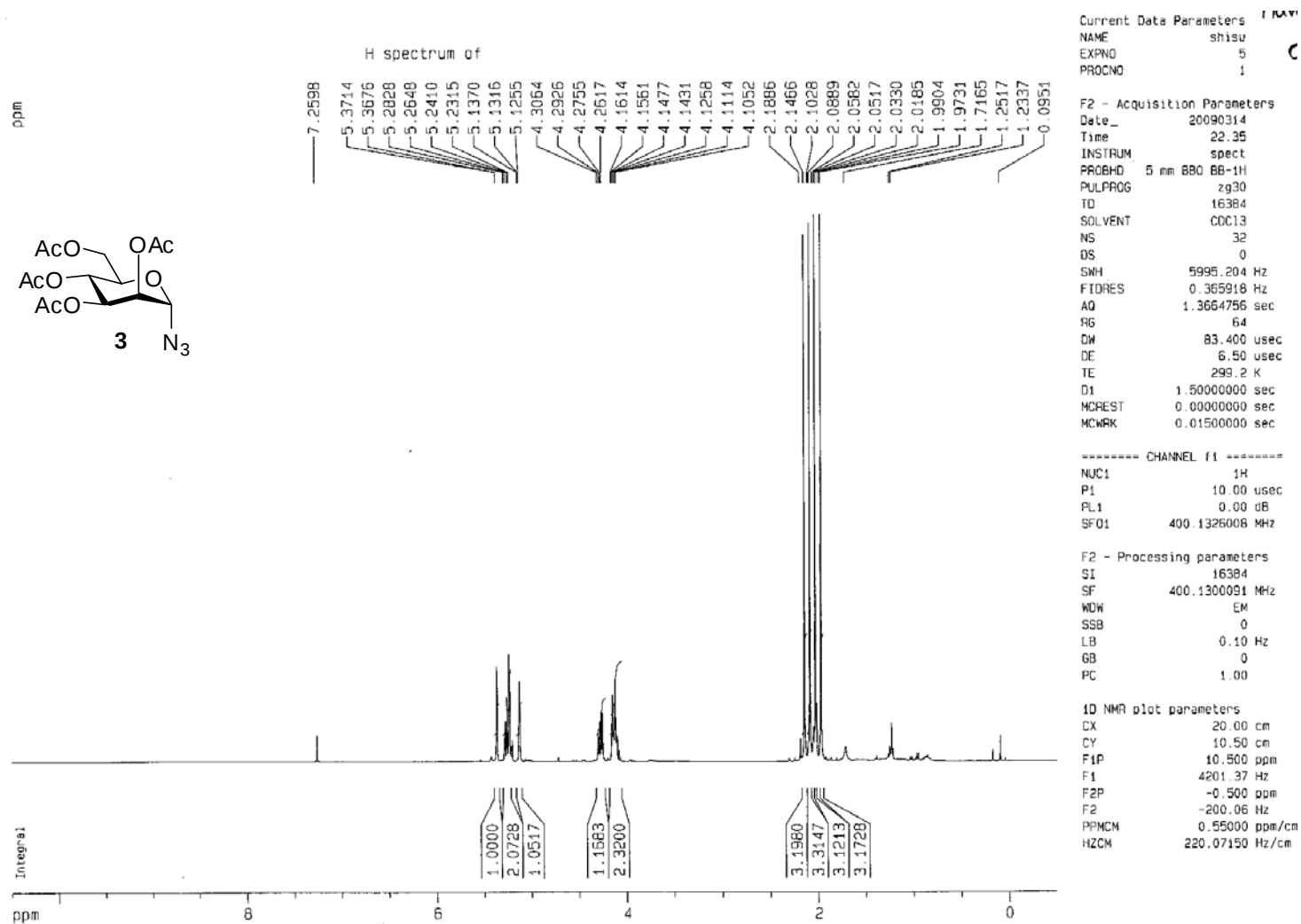
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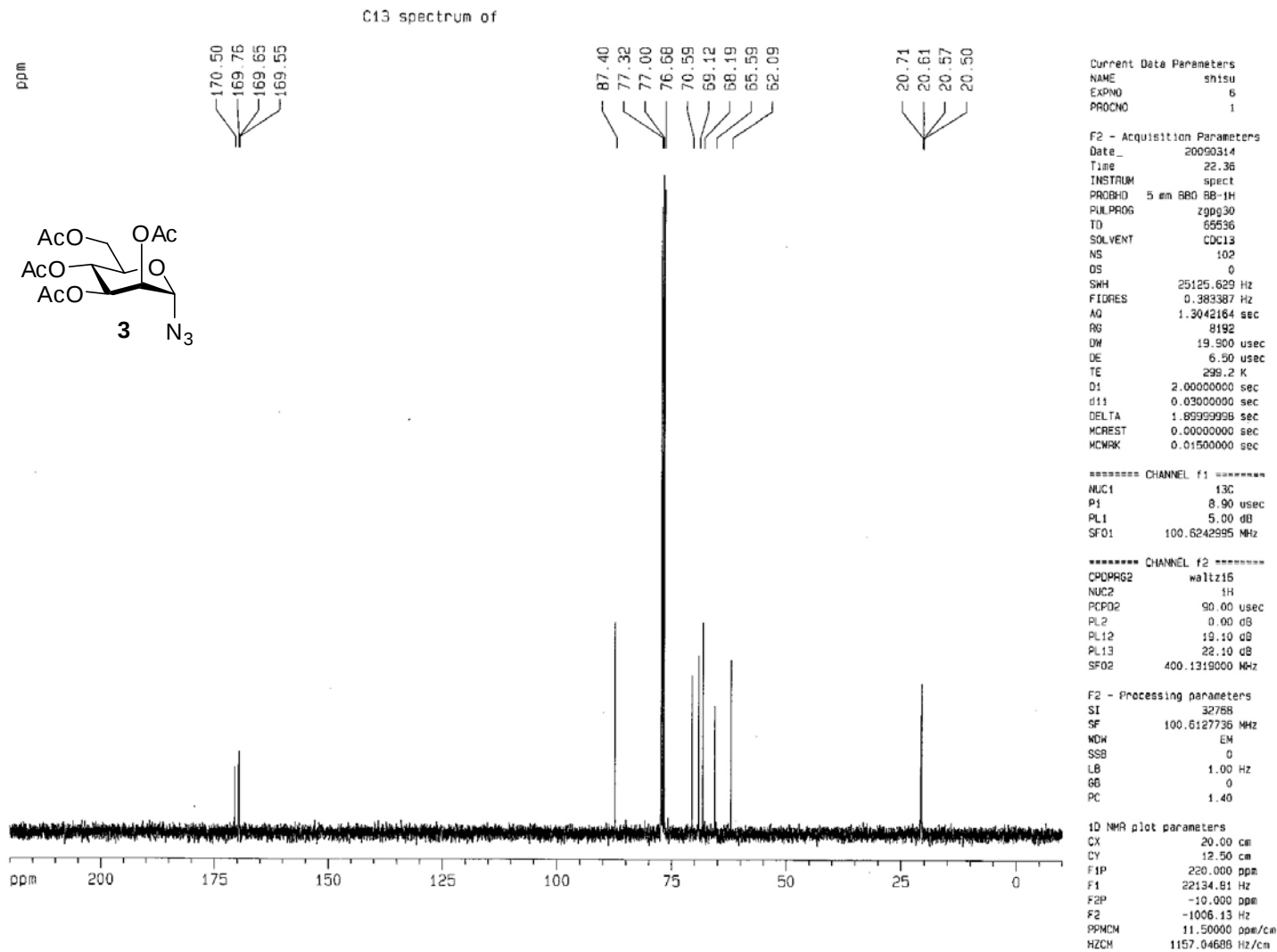
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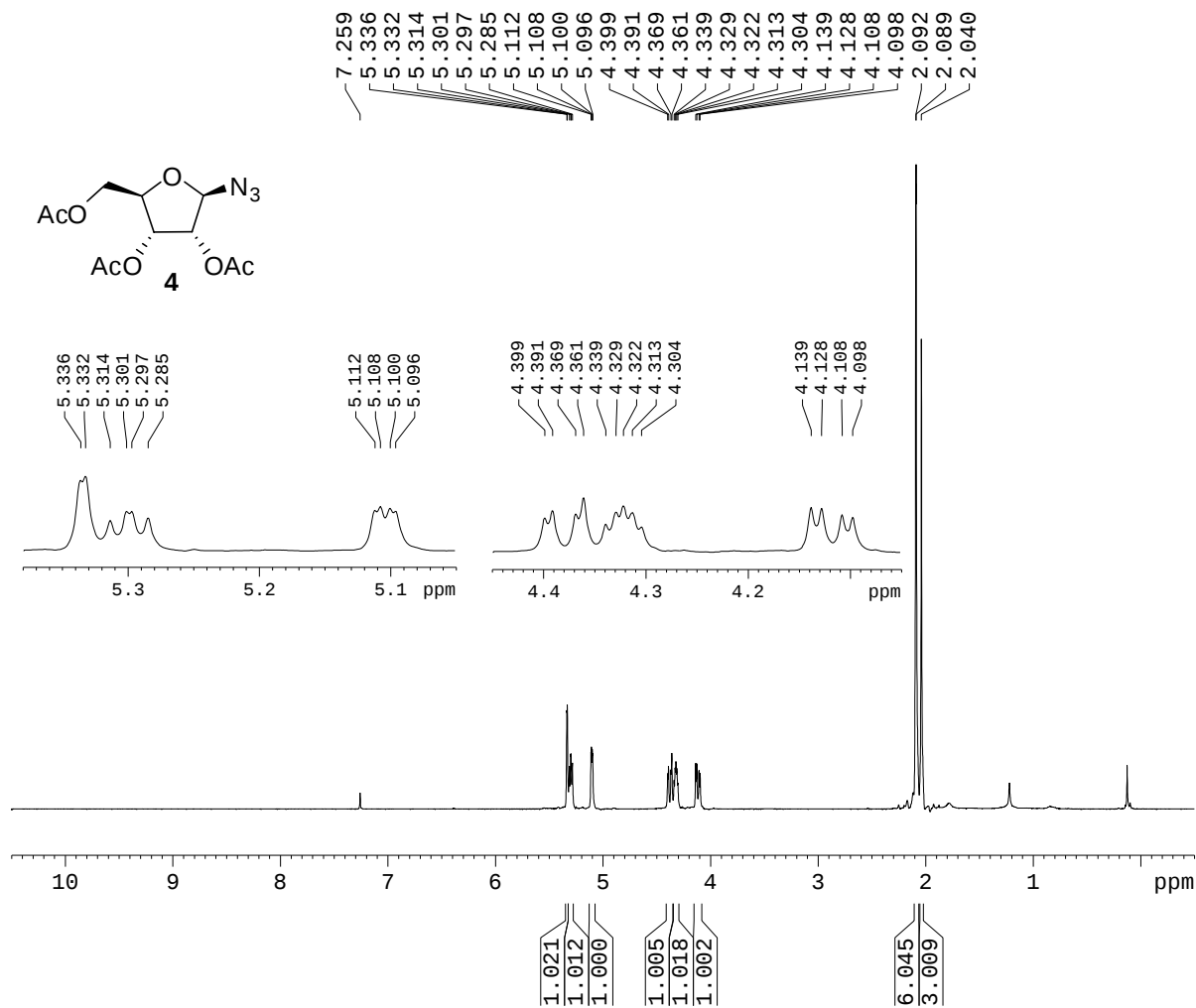
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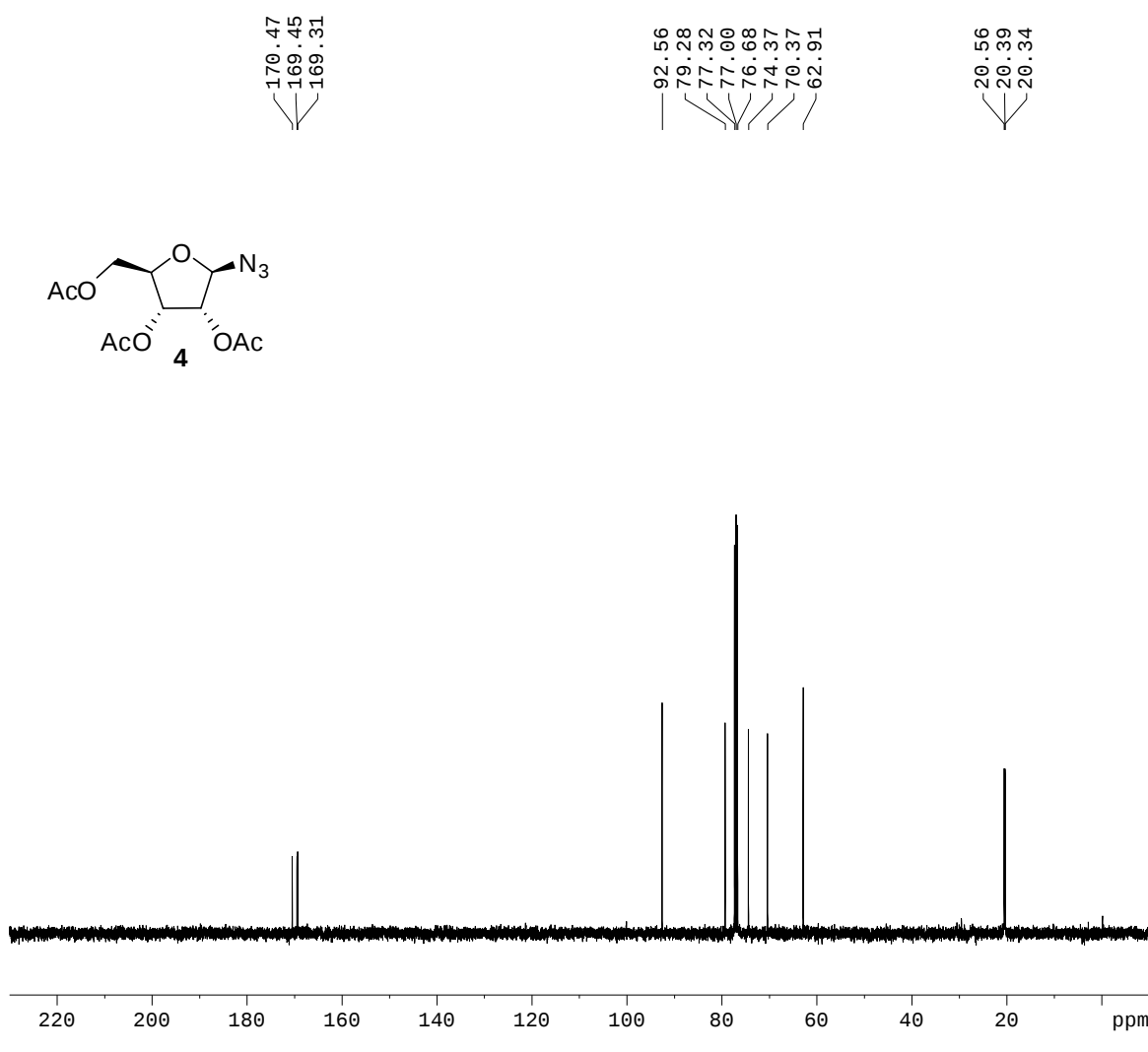
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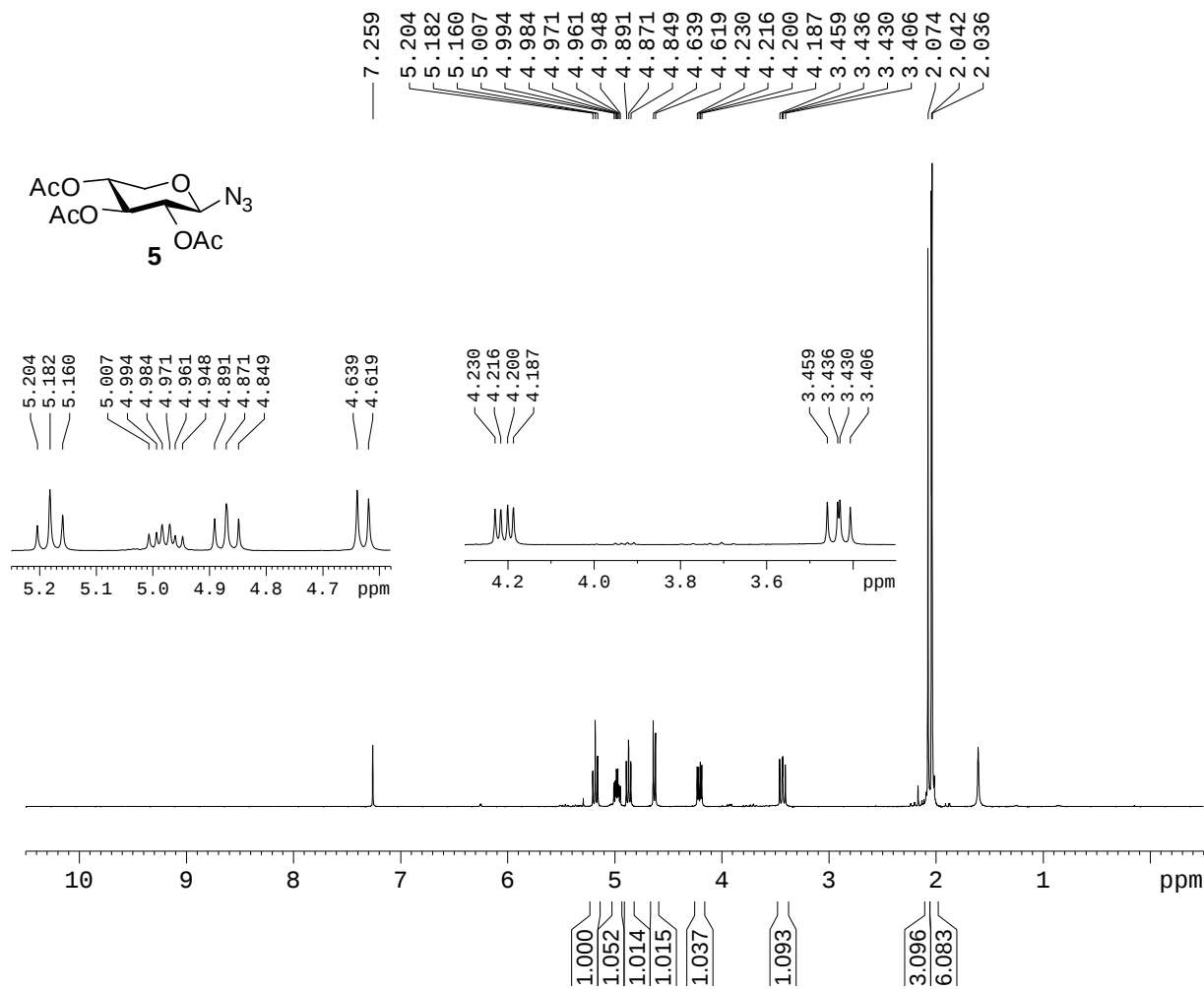
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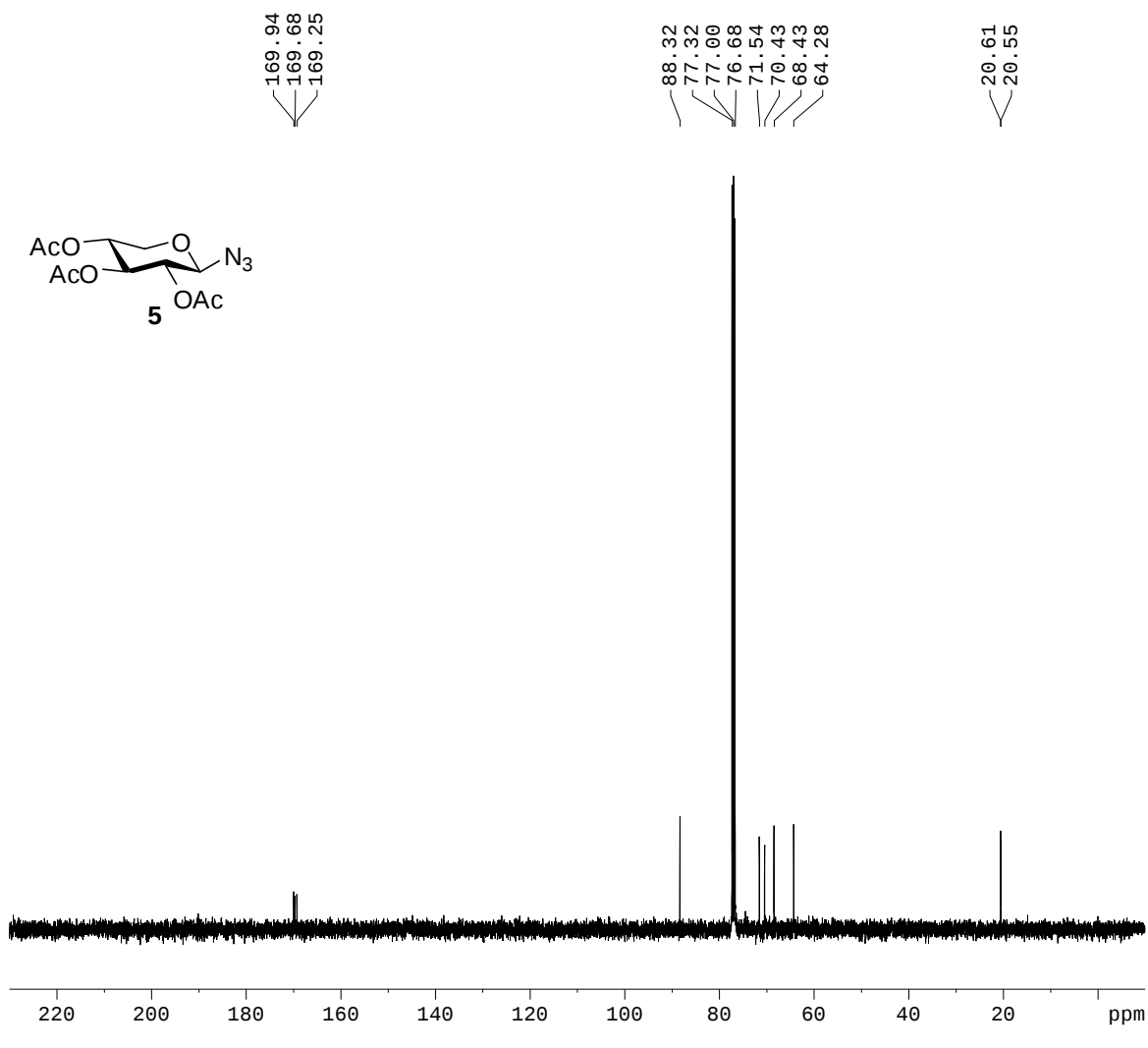
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CX 20.00 cm
CY 10.50 cm
F1P 5.373 ppm
F1 2150.09 Hz



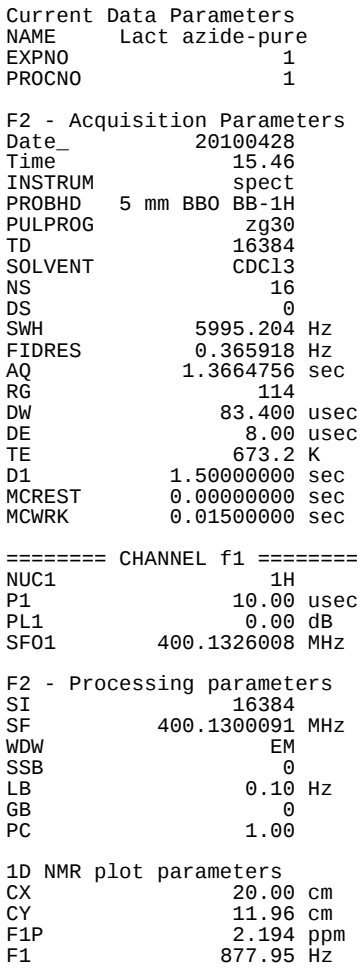
Current Data Parameters
NAME nsp-triazole-clc1
EXPNO 2
PROCNO 1

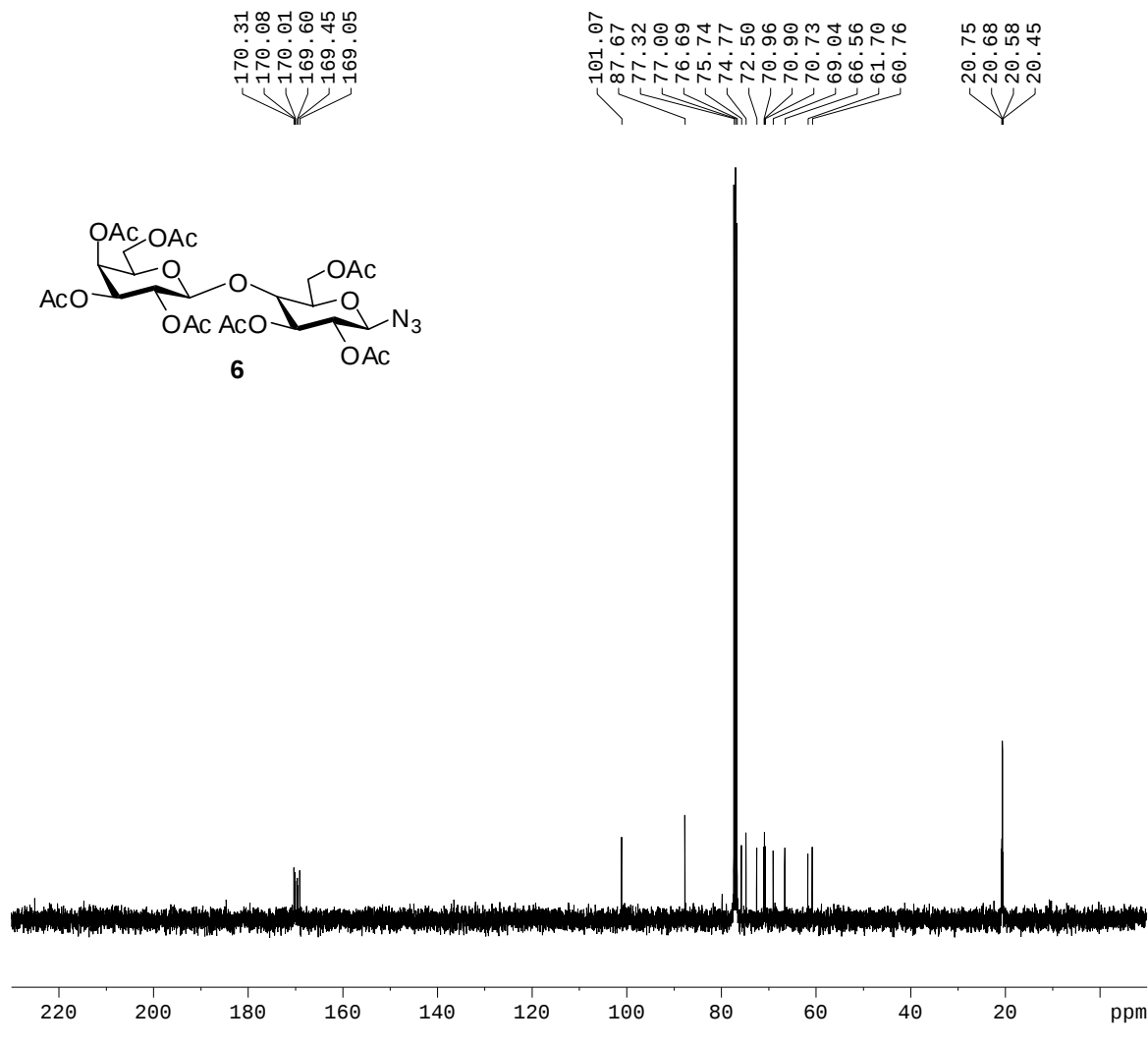
F2 - Acquisition Parameters
Date_ 20081227
Time 22.21
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 182
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 4096
DW 19.900 usec
DE 6.50 usec
TE 302.9 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127466 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





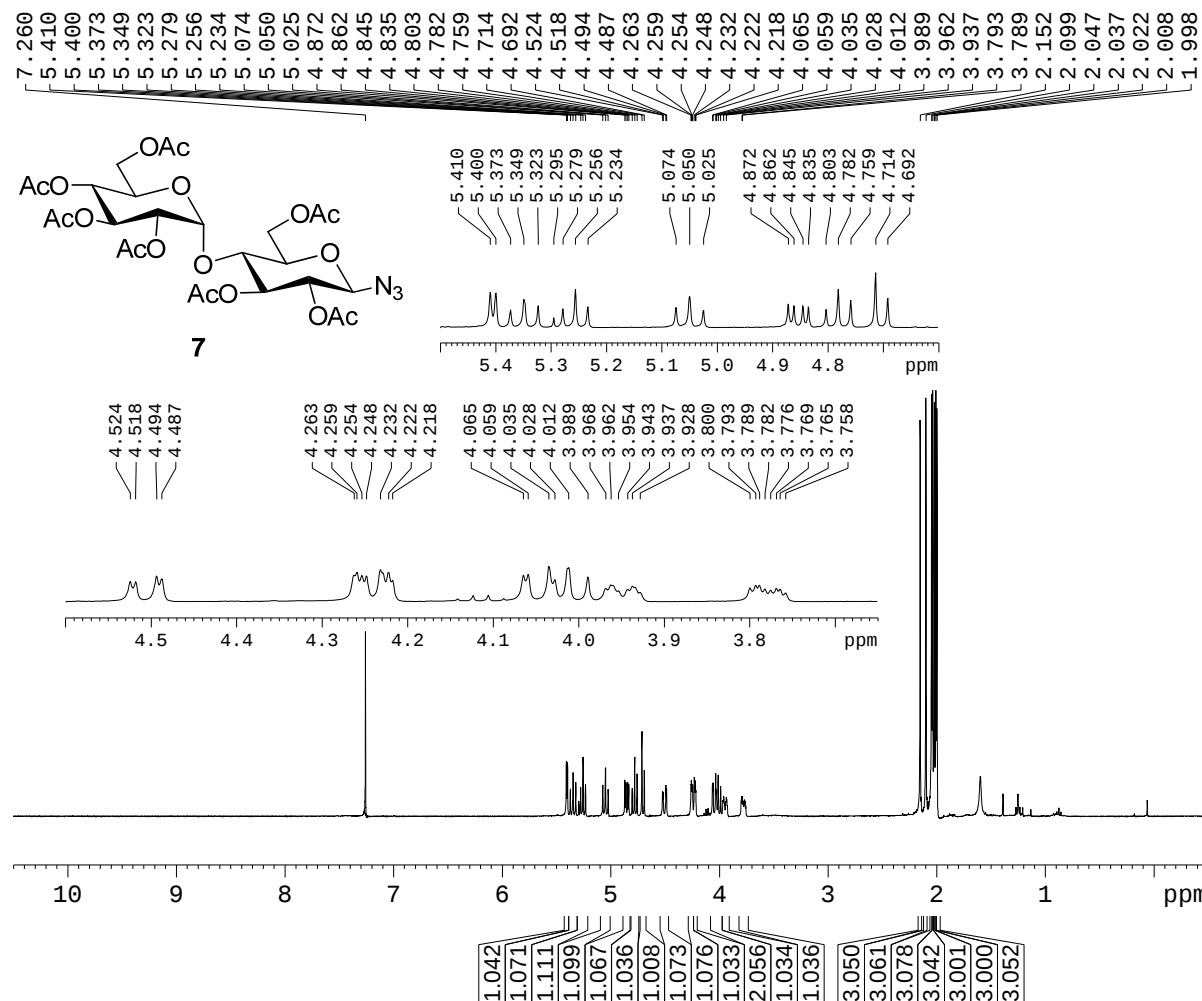
Current Data Parameters
NAME Lact azide-pure
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20100428
Time 15.47
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 97
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 8192
DW 19.900 usec
DE 8.00 usec
TE 673.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6243390 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



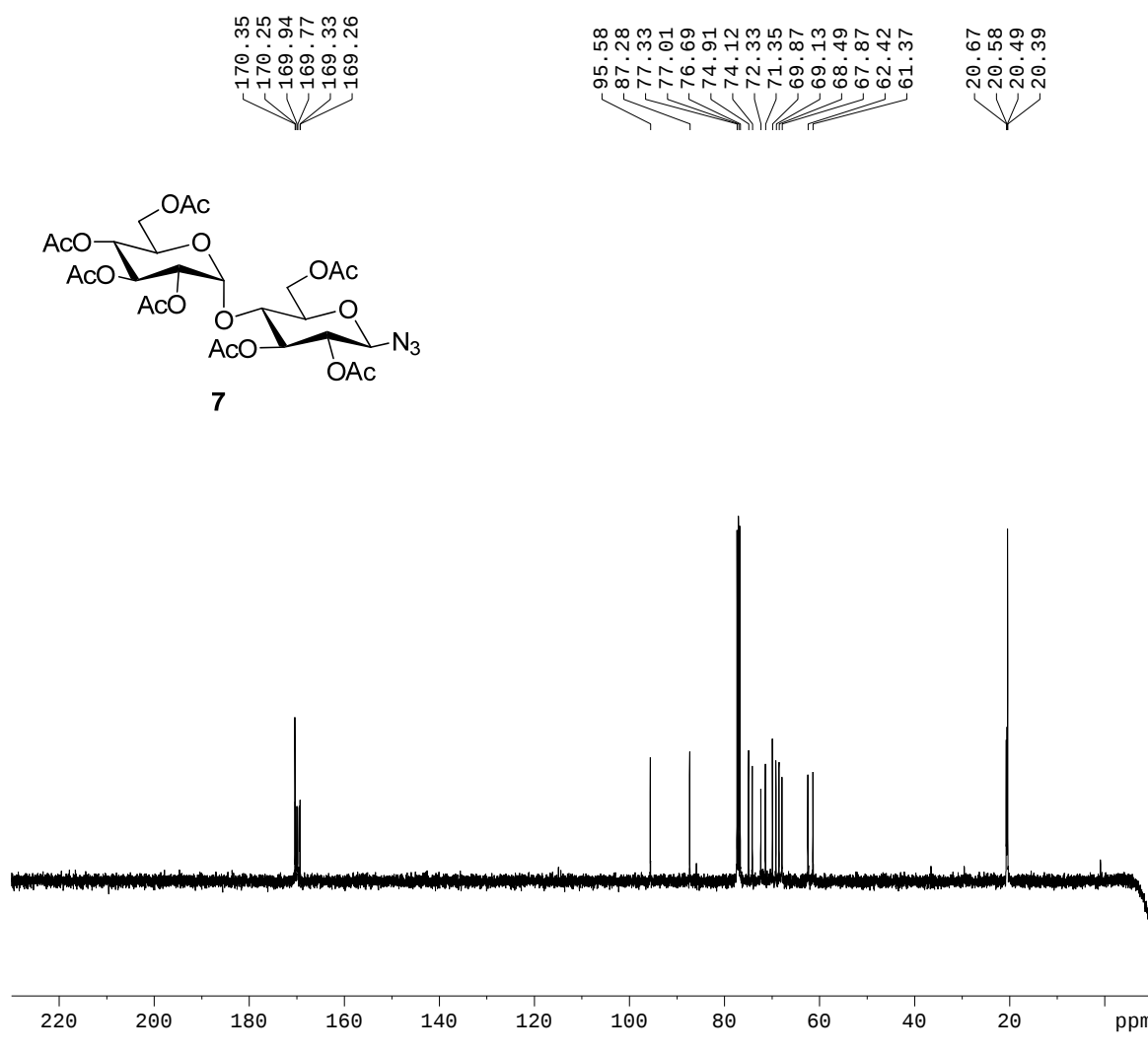
Current Data Parameters
NAME Maltose azide-pure
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20100331
Time 16.24
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 322.5
DW 83.400 usec
DE 6.50 usec
TE 297.8 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.70 usec
PL1 3.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.42 cm
F1P 5.482 ppm
F1 2193.37 Hz



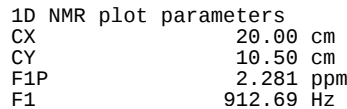
Current Data Parameters
NAME pure
EXPNO 2
PROCNO 1

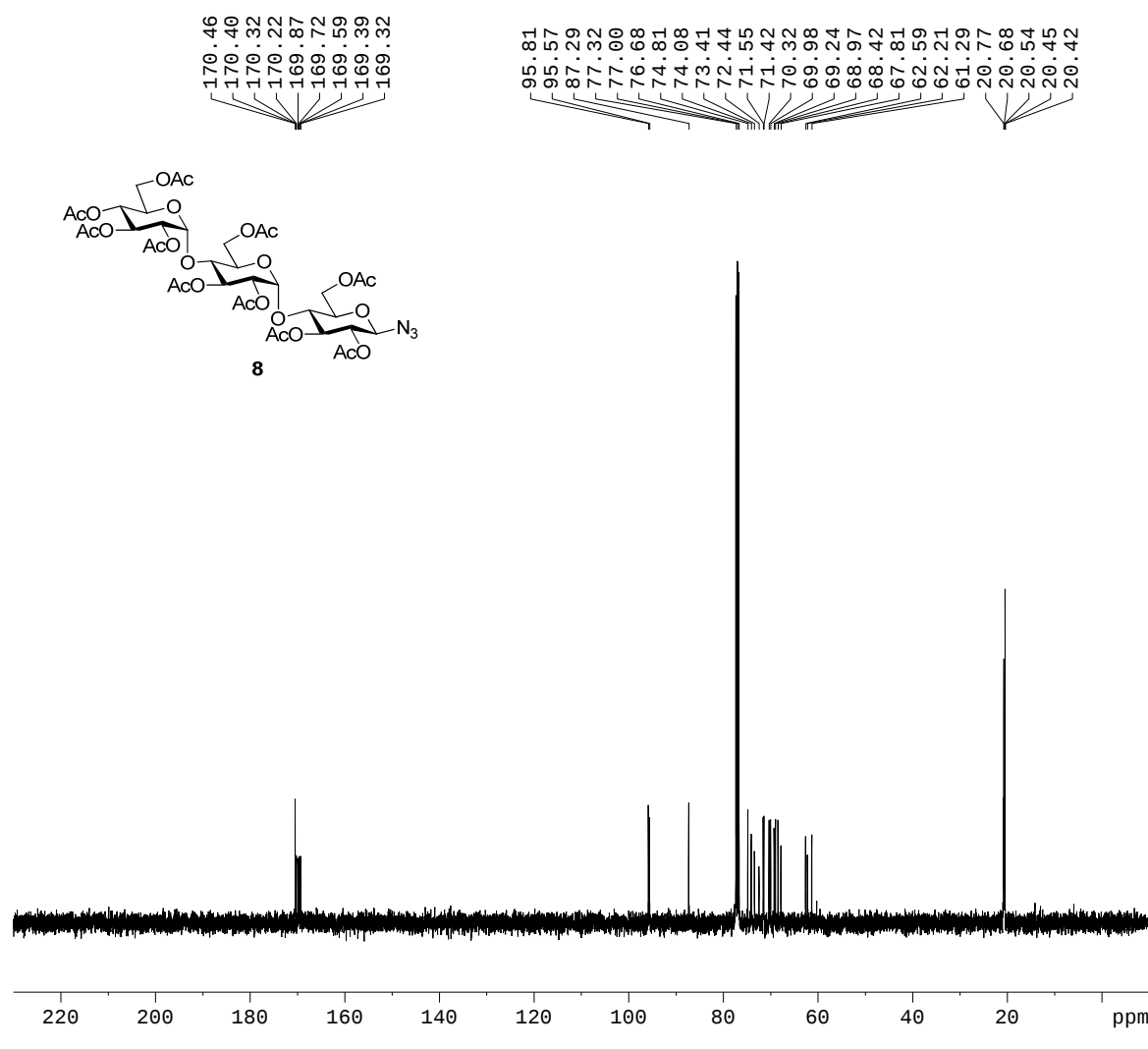
F2 - Acquisition Parameters
Date_ 20100410
Time 15.39
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 93
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 181
DW 19.900 usec
DE 8.00 usec
TE 673.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6243390 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127533 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





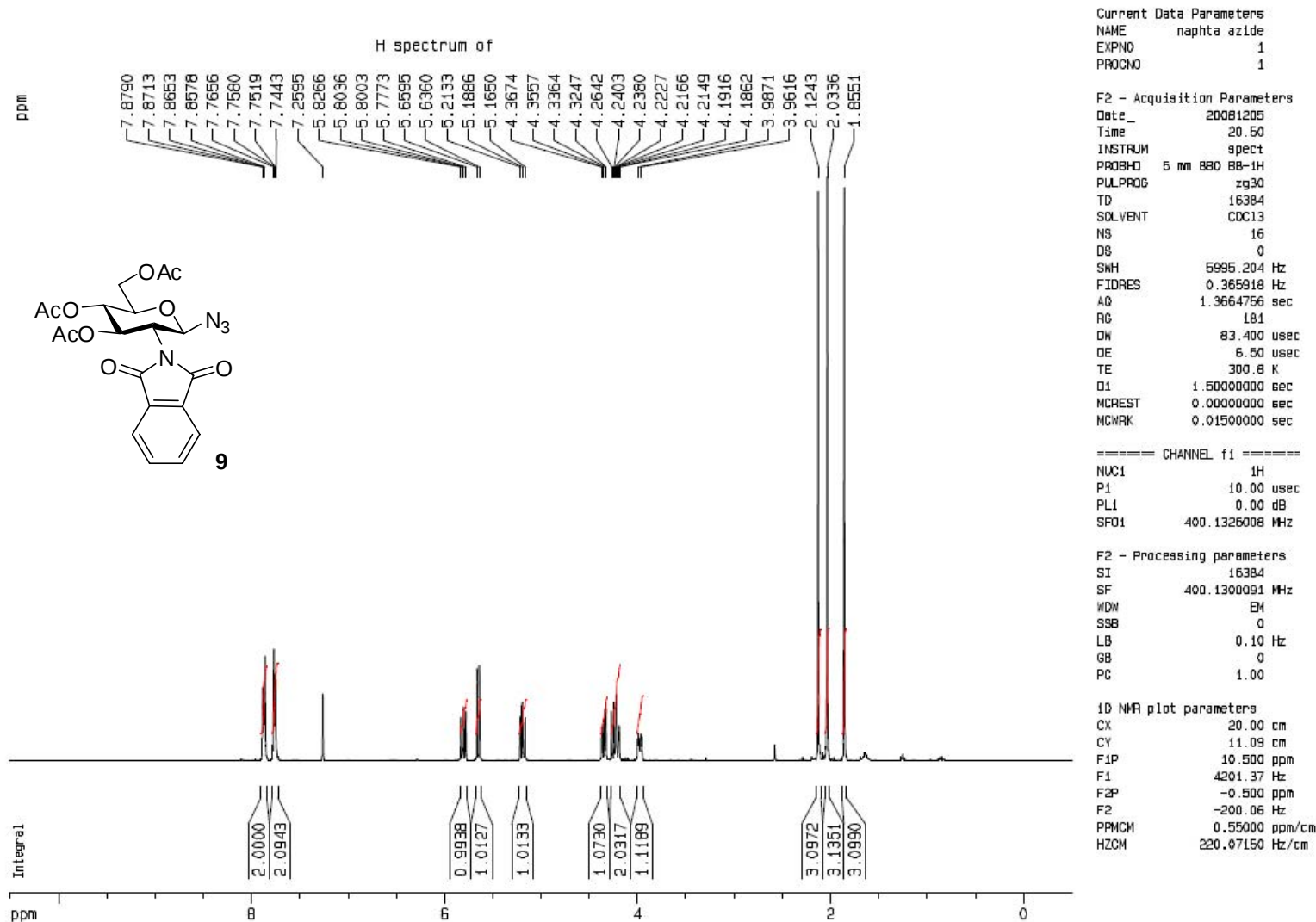
Current Data Parameters
NAME maltose azide
EXPNO 2
PROCNO 1

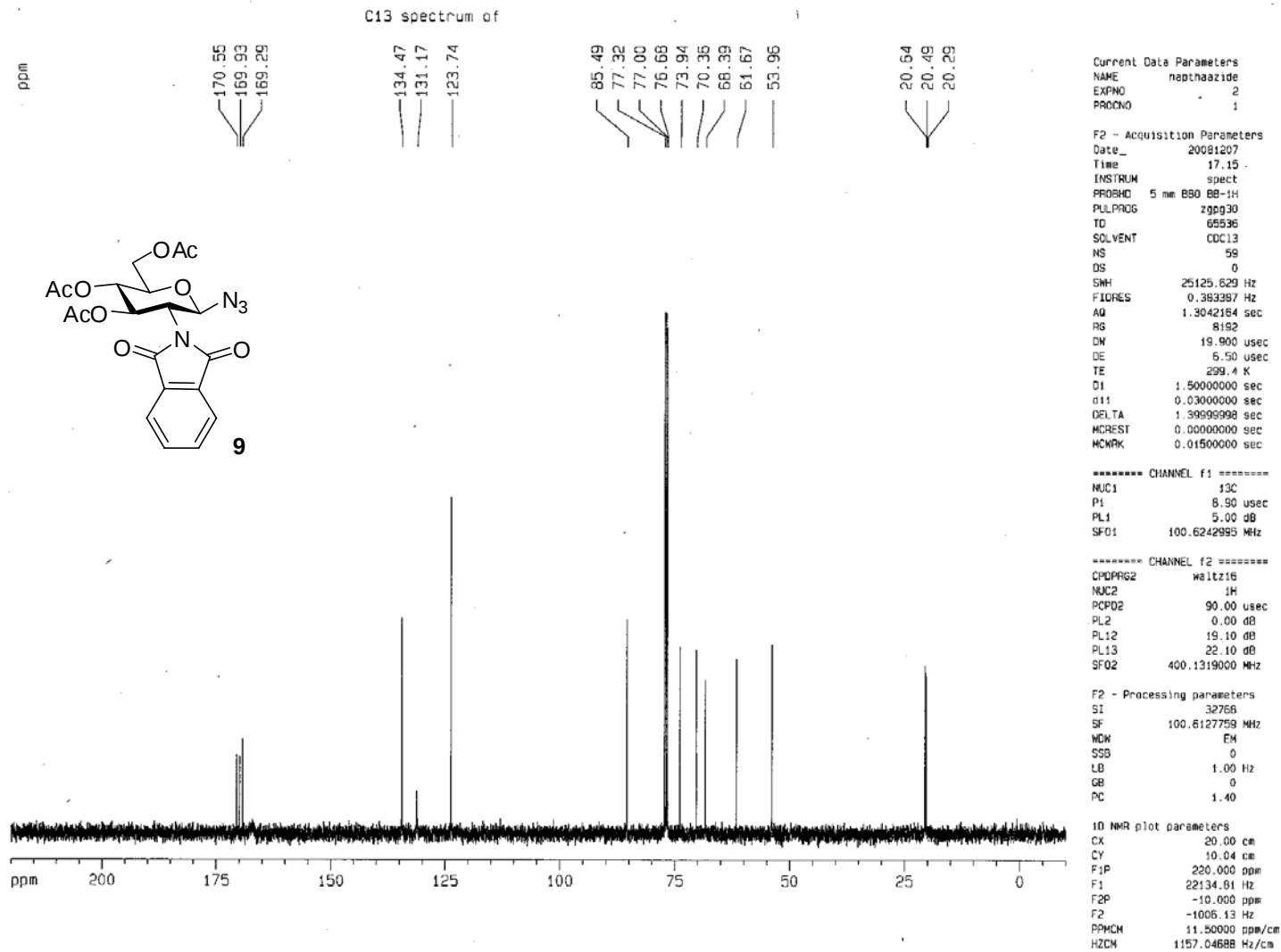
F2 - Acquisition Parameters
Date_ 20090323
Time 19.35
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 75
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 8192
DW 19.900 usec
DE 6.50 usec
TE 298.5 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

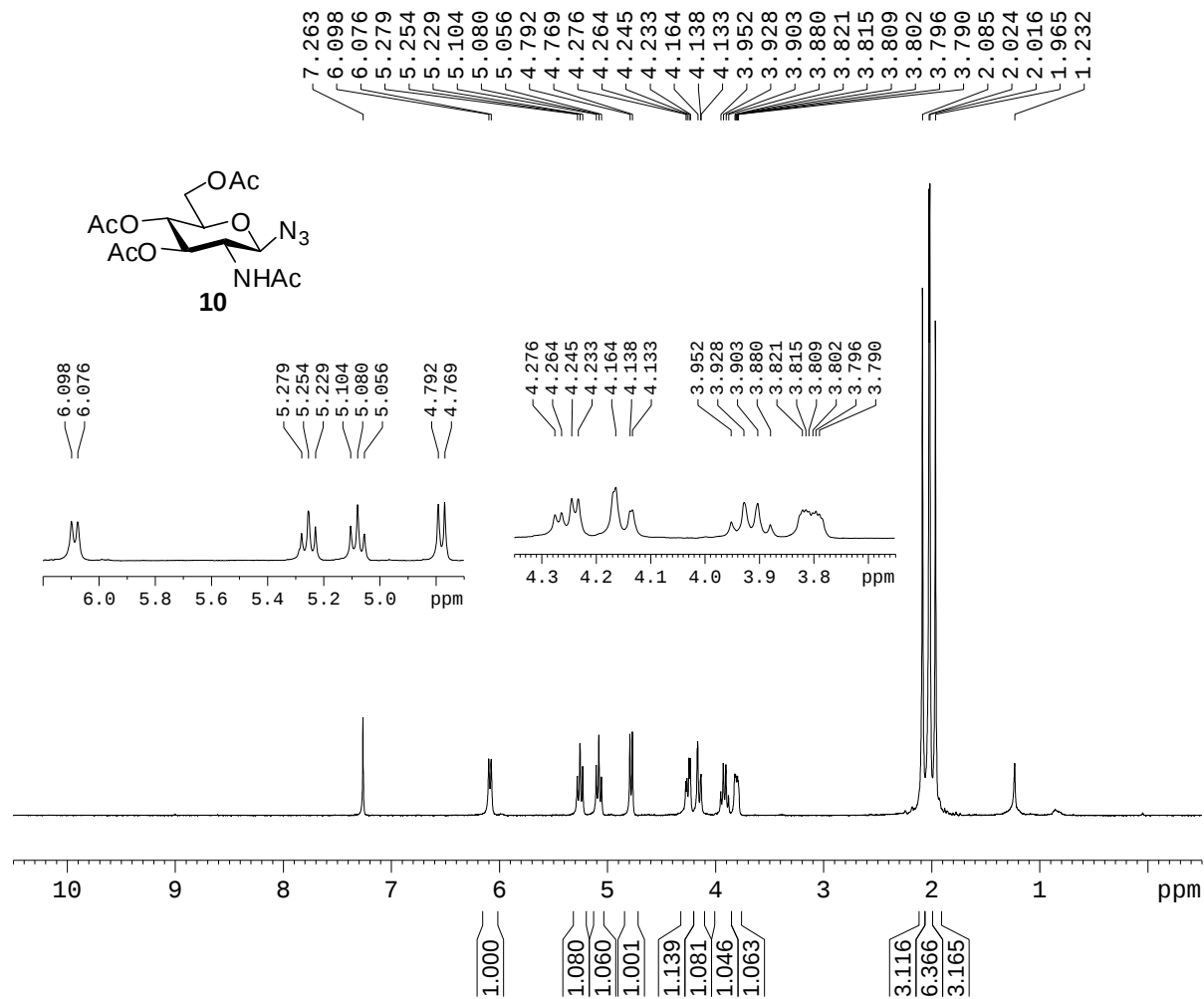
===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127528 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

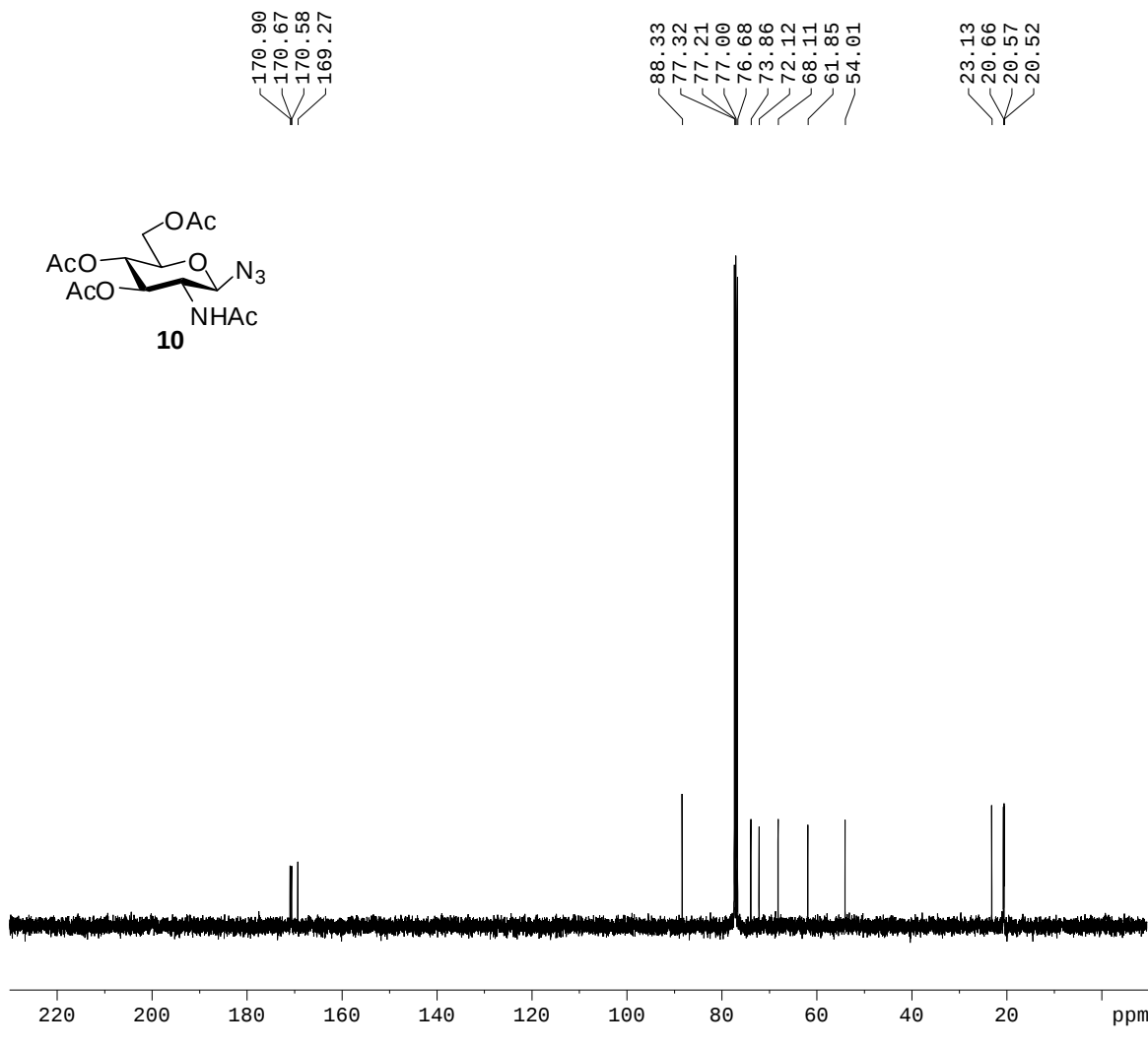






NAME NH AC azide C13
EXPNO 1
PROCNO 1
Date_ 20091218
Time 21.49
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT $CDCl_3$
NS 1
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 90.5
DW 83.400 usec
DE 6.50 usec
TE 297.2 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz
SI 16384
SF 400.1300074 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00



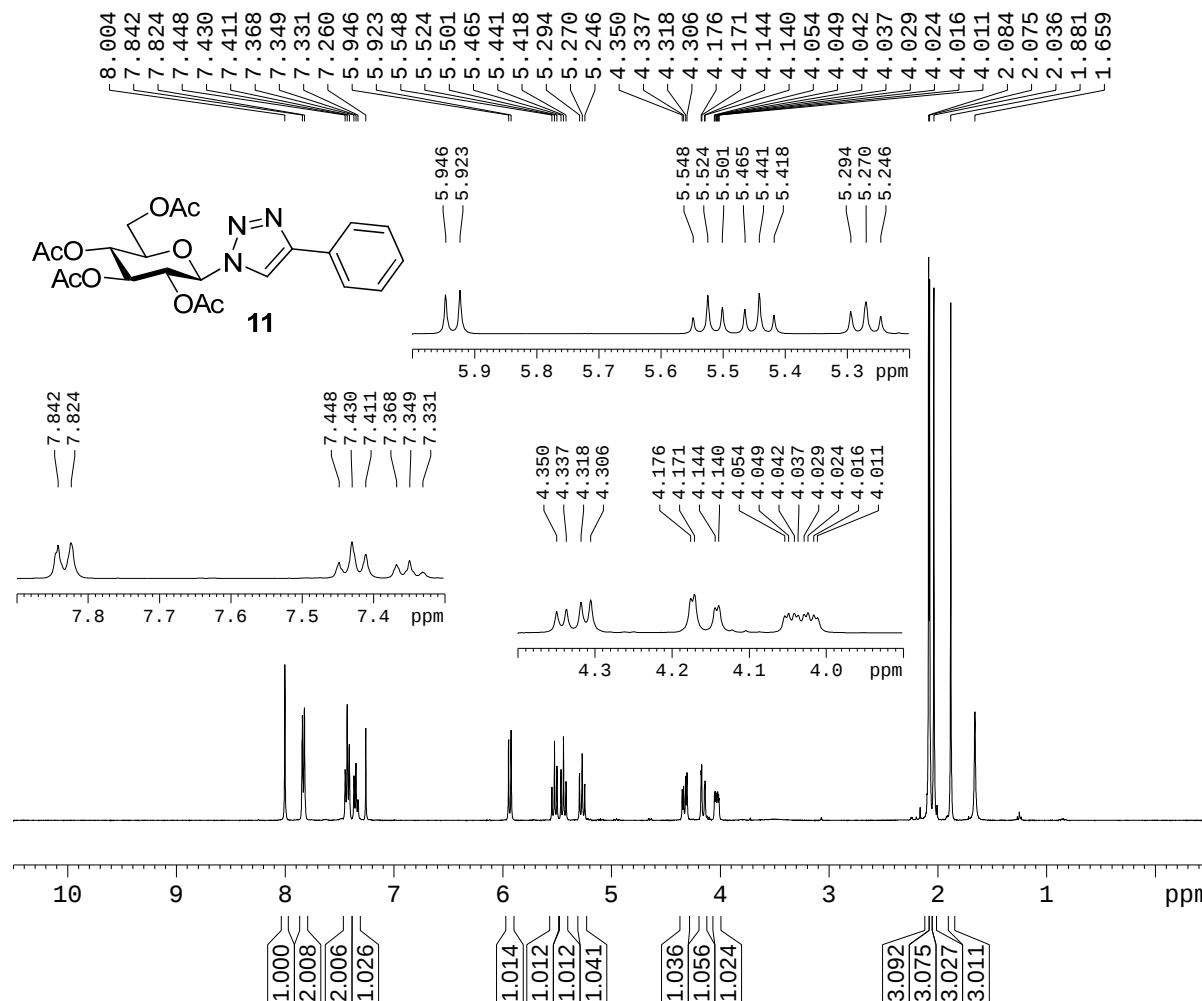
Current Data Parameters
NAME NH AC azide C13
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20091218
Time 21.50
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 98
DS 0
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 8192
DW 19.950 usec
DE 6.50 usec
TE 297.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6243390 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127513 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



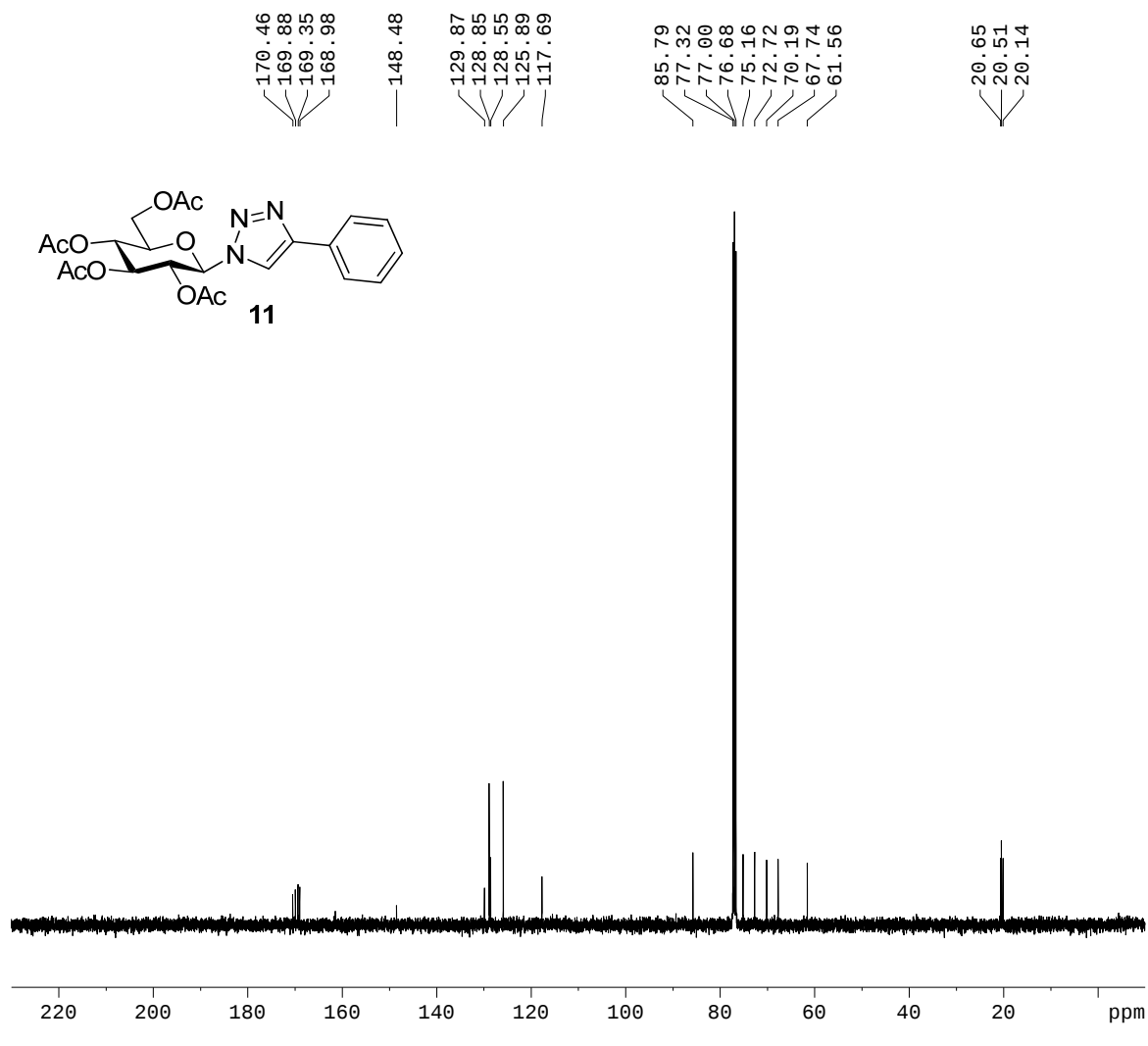
Current Data Parameters
NAME glucose triazole pui
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090411
Time 20.58
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 161.3
DW 83.400 usec
DE 6.50 usec
TE 297.6 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.50 cm
F1P 2.155 ppm
F1 862.43 Hz



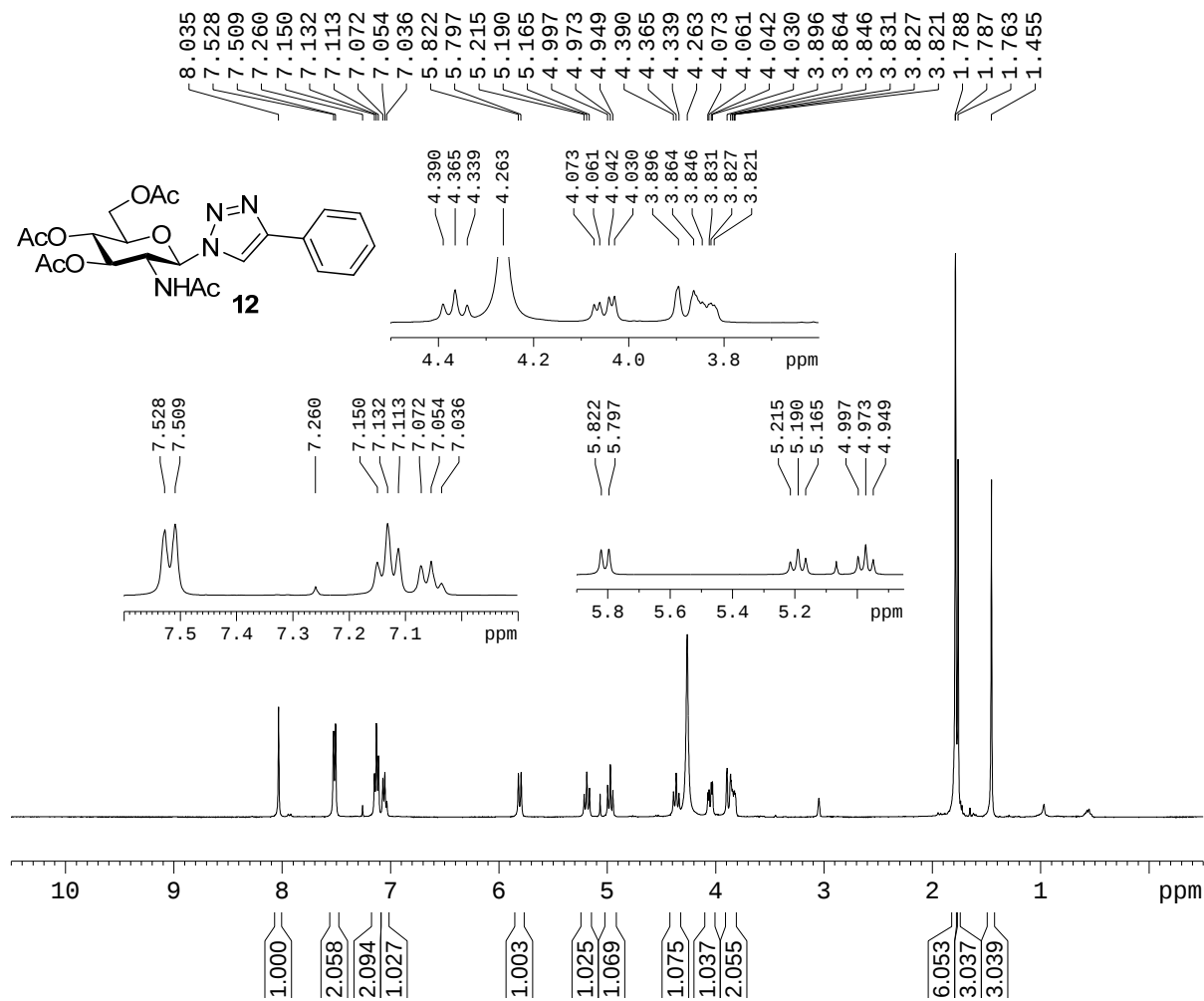
Current Data Parameters
NAME dr-12
EXPNO 9
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090307
Time 21.22
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 151
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 4096
DW 19.900 usec
DE 6.50 usec
TE 299.0 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127672 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



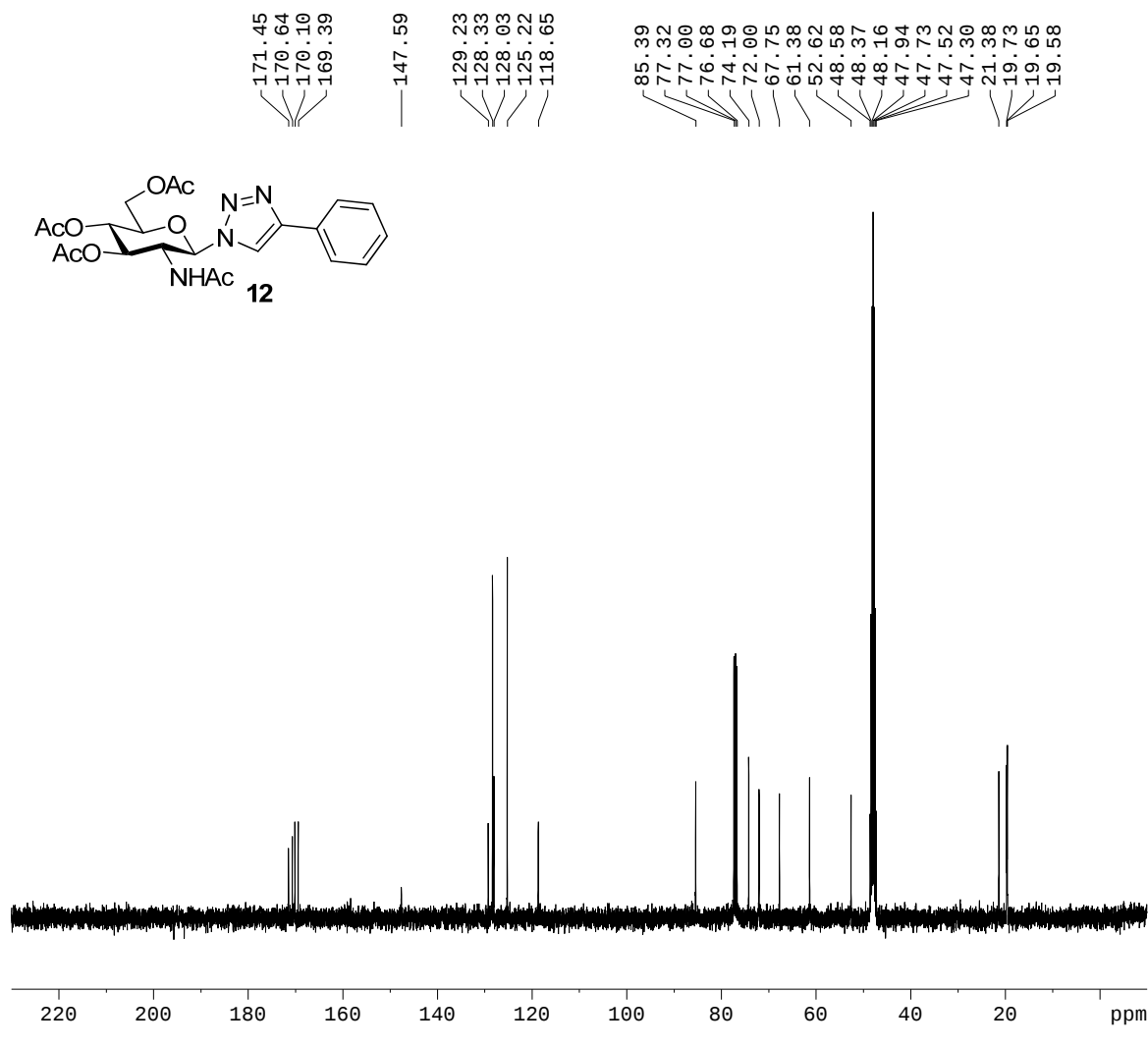
Current Data Parameters
NAME NHAc
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090405
Time_ 17.49
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT MeOD
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 57
DW 83.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1301123 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.50 cm
F1P 5.882 ppm
F1 2353.48 Hz



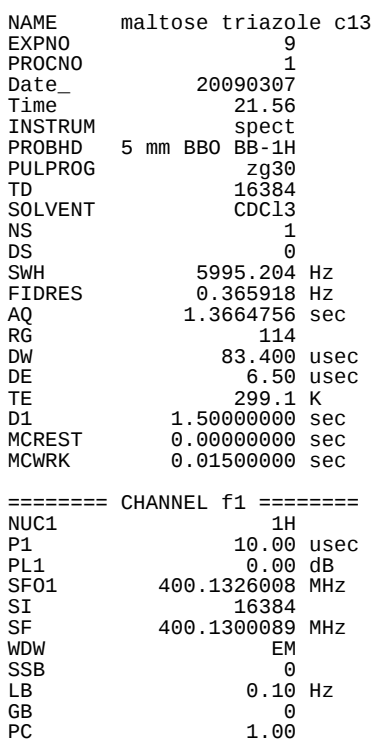
Current Data Parameters
NAME NHAc C13
EXPNO 4
PROCNO 1

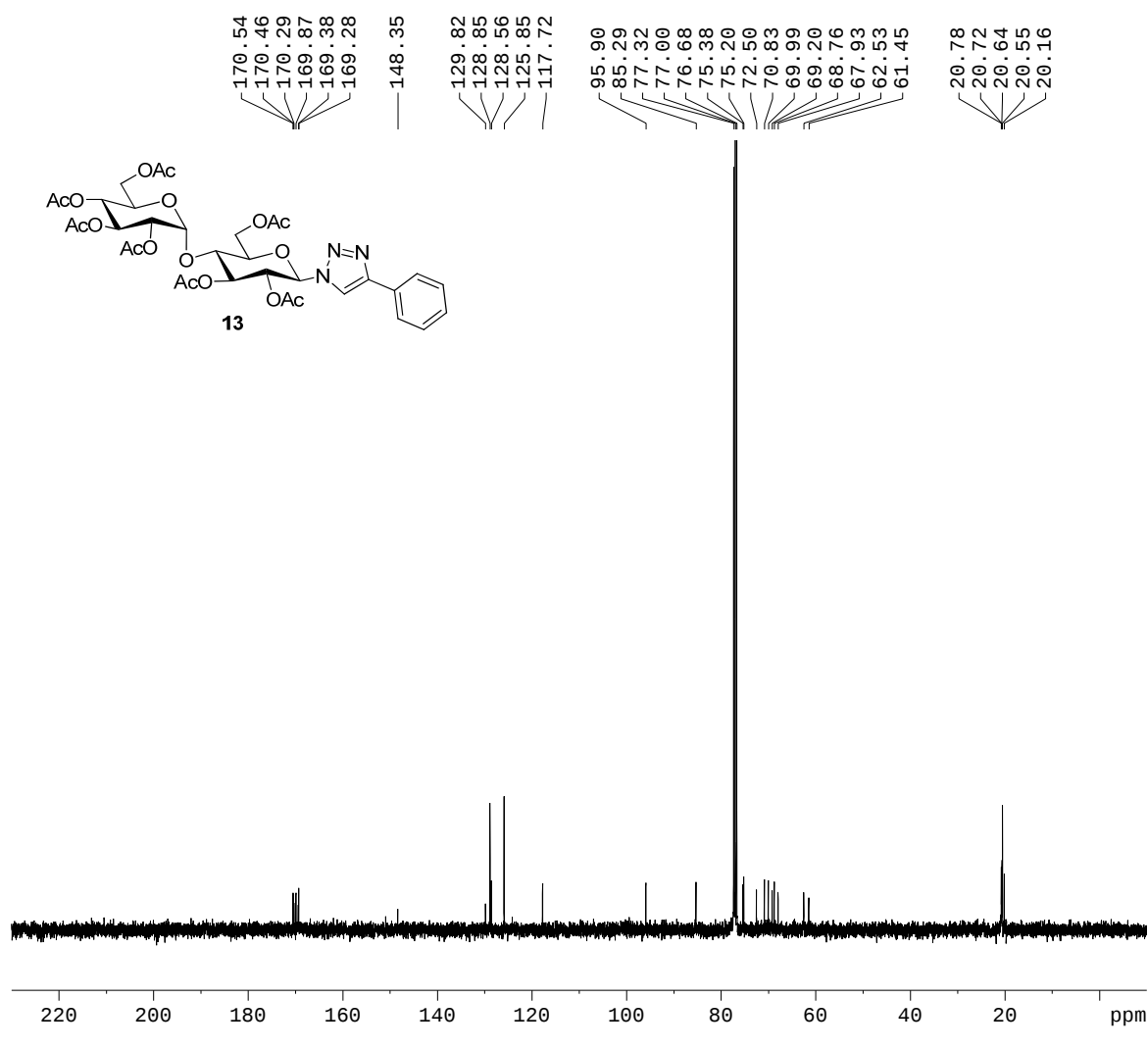
F2 - Acquisition Parameters
Date_ 20090405
Time 18.28
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 108
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 8192
DW 19.900 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127934 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





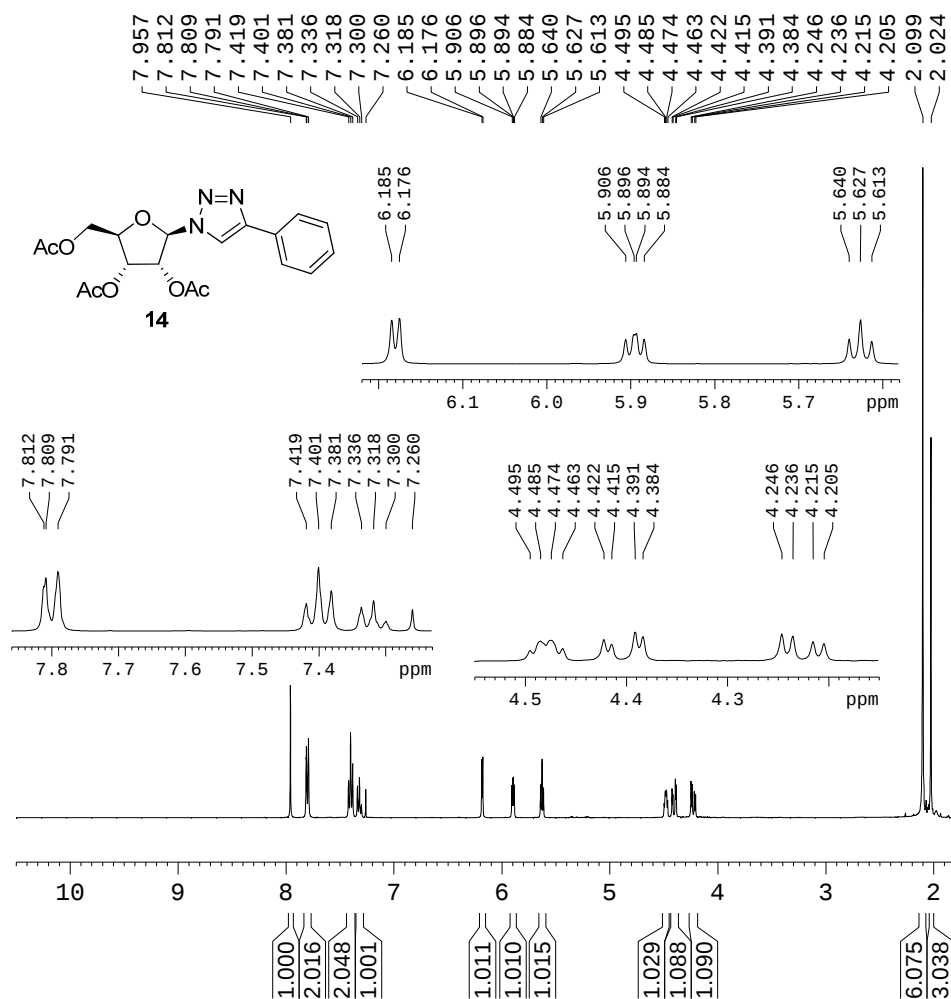
Current Data Parameters
NAME maltose triazole c13
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090307
Time 21.58
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 158
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 8192
DW 19.900 usec
DE 6.50 usec
TE 299.1 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127497 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



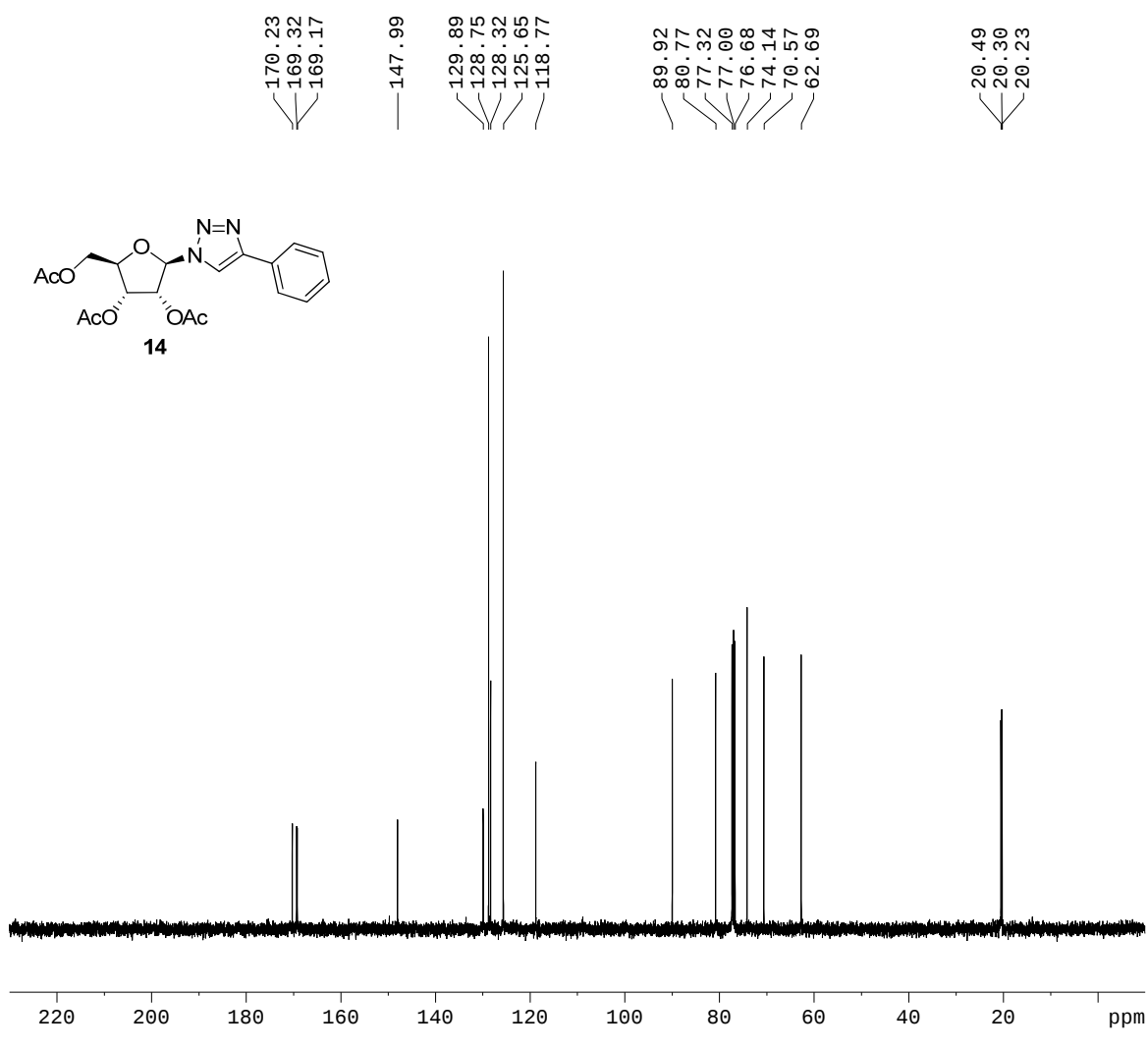
Current Data Parameters
NAME ribose triazole
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090314
Time 16.19
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 40.3
DW 83.400 usec
DE 6.50 usec
TE 299.9 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1300088 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.50 cm
F1P 8.128 ppm
F1 3252.12 Hz



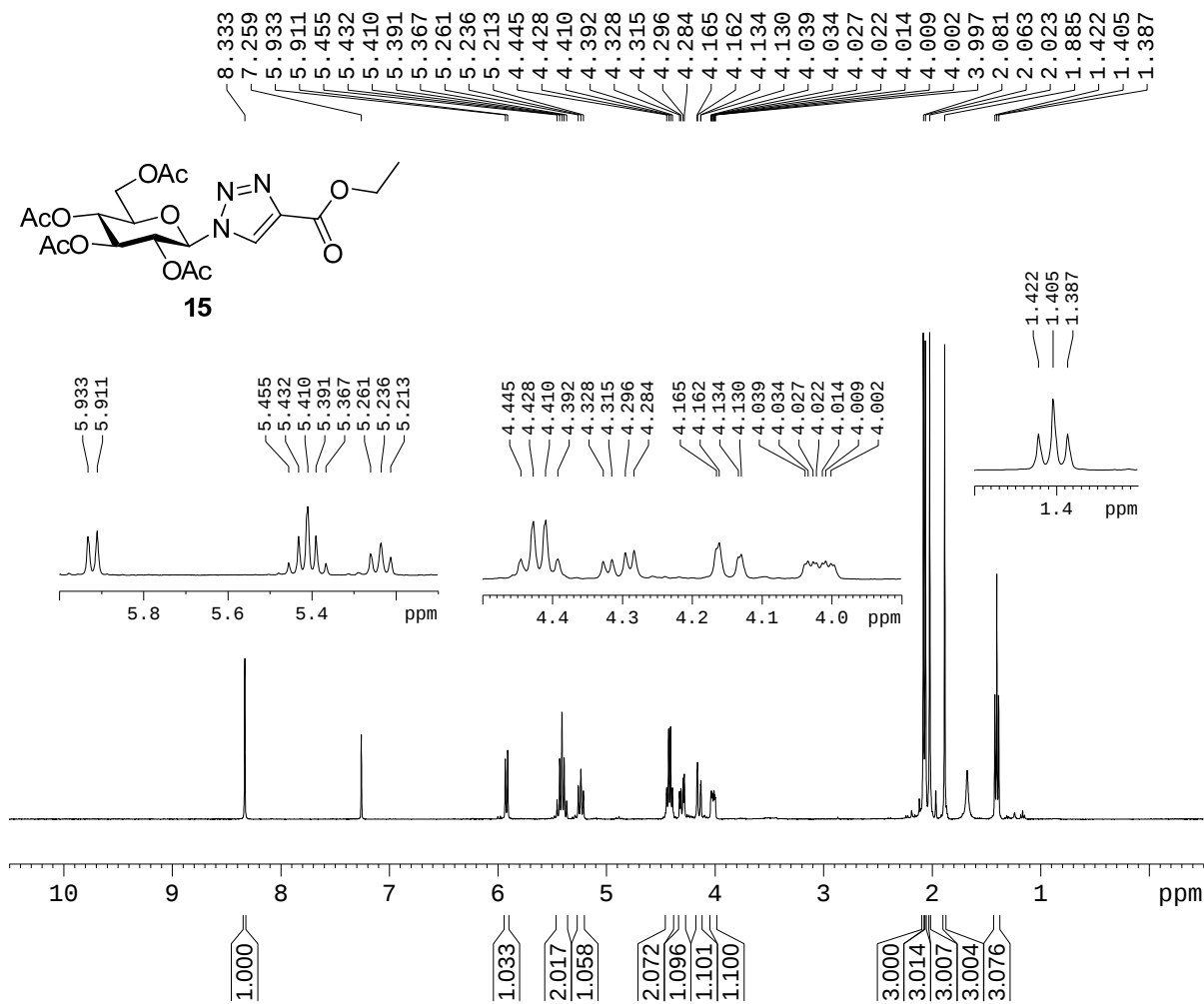
Current Data Parameters
NAME PTHALIC-12PH-INDOL
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090321
Time 20.57
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 68
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 4096
DW 19.900 usec
DE 6.50 usec
TE 300.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127604 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



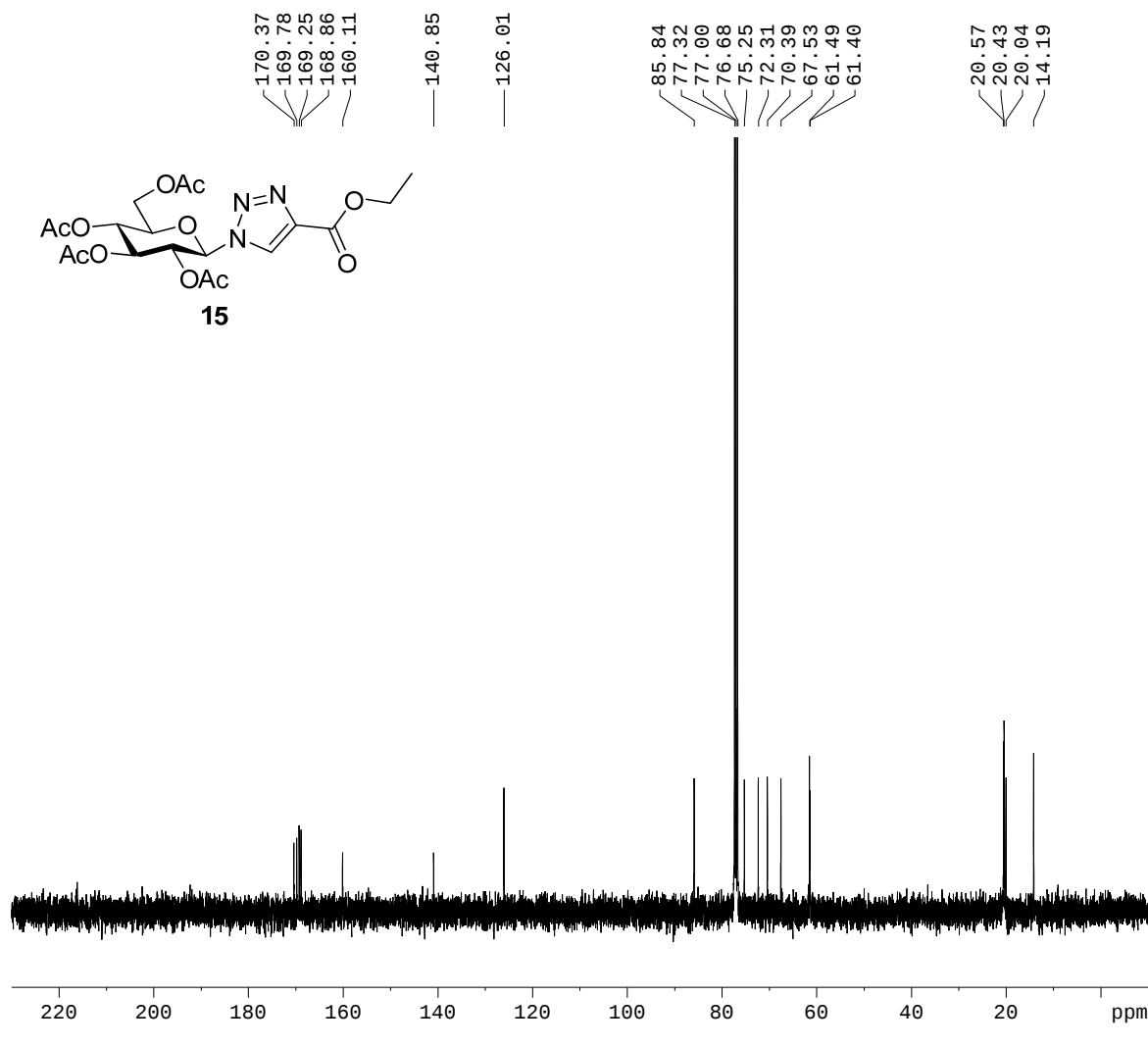
Current Data Parameters
NAME NHAc
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090405
Time 18.17
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 161.3
DW 83.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.50 cm
F1P 2.215 ppm
F1 886.46 Hz



Current Data Parameters
NAME propargy ester triazol
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters

Date_ 20090404
Time 20.39
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 83
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 8192
DW 19.900 usec
DE 6.50 usec
TE 297.8 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====

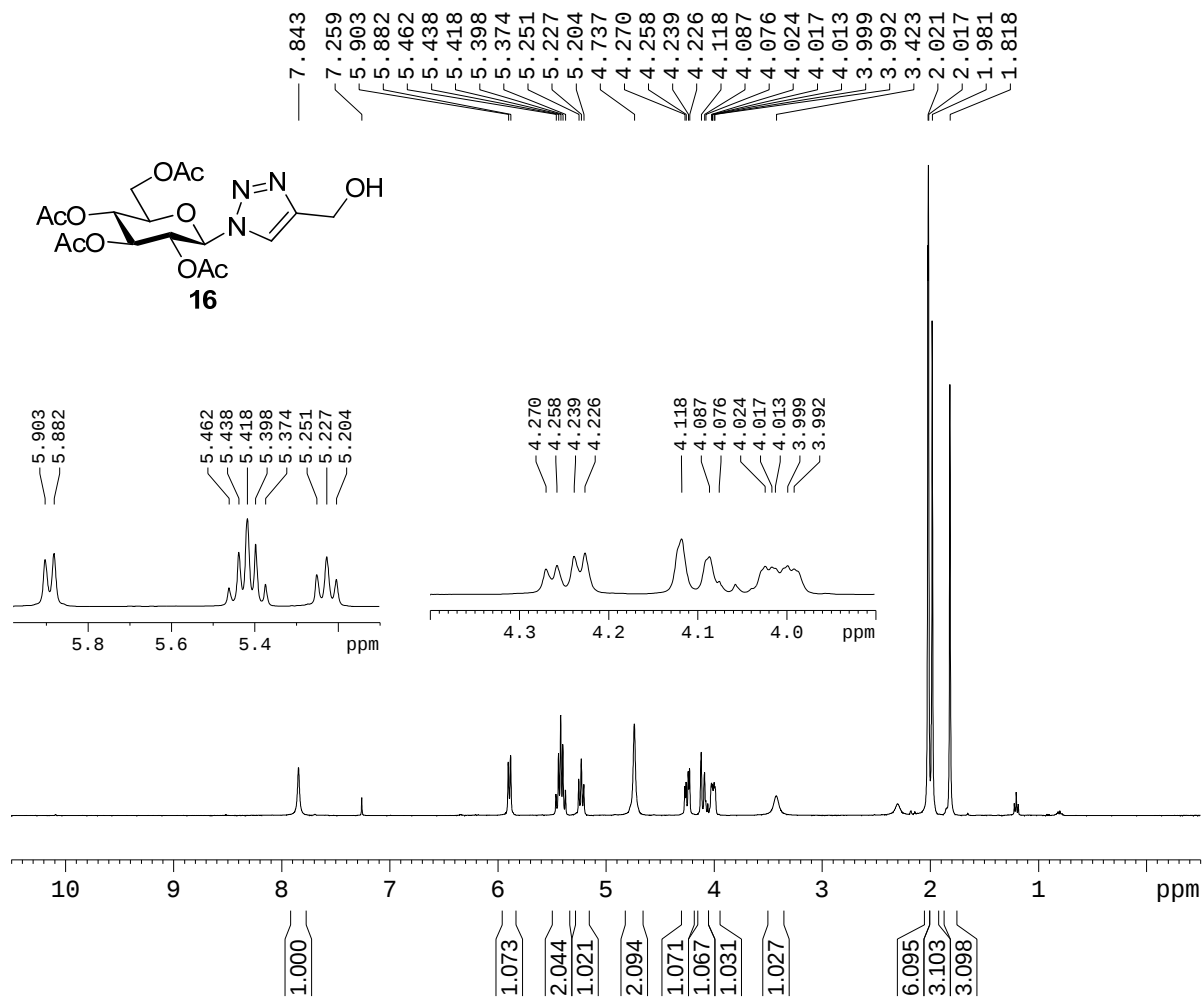
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6242995 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters

SI 32768
SF 100.6127520 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



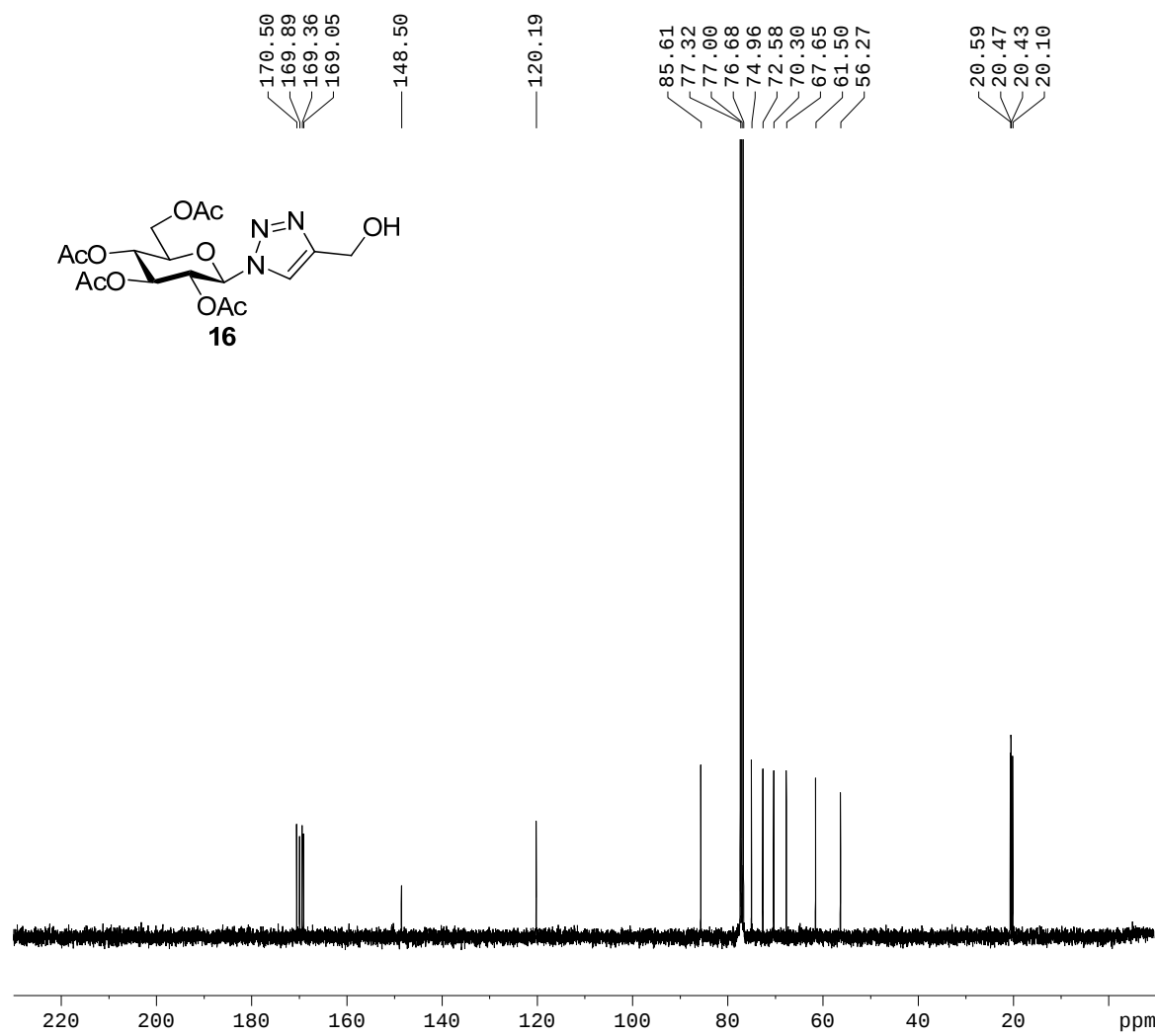
Current Data Parameters
NAME -propargy triazole
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090326
Time 20.50
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 45.3
DW 83.400 usec
DE 6.50 usec
TE 298.7 K
D1 1.50000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 0.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.50 cm
F1P 6.065 ppm
F1 2426.88 Hz



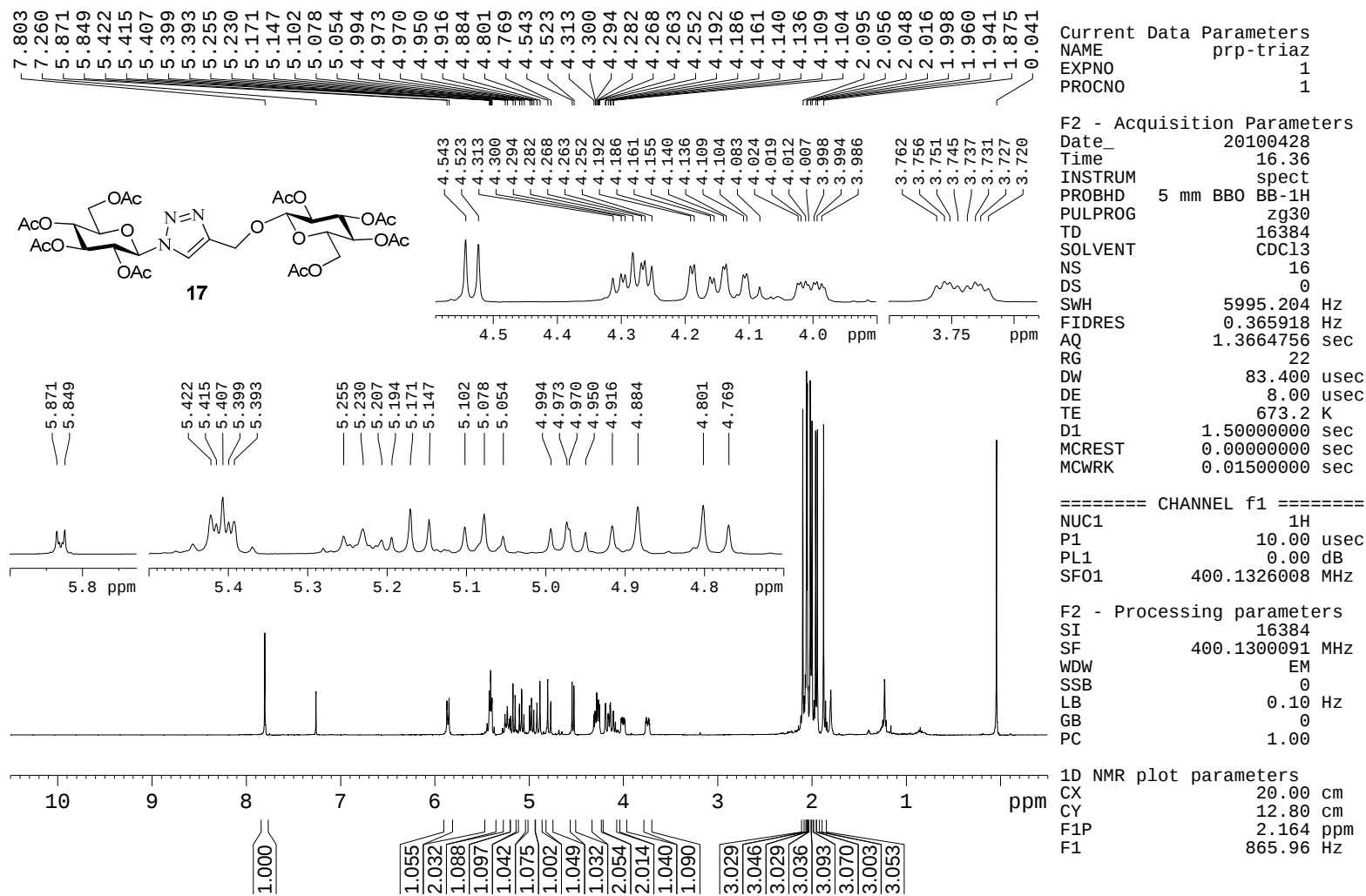
Current Data Parameters
NAME prp-alcohol-sug
EXPNO 1
PROCNO 1

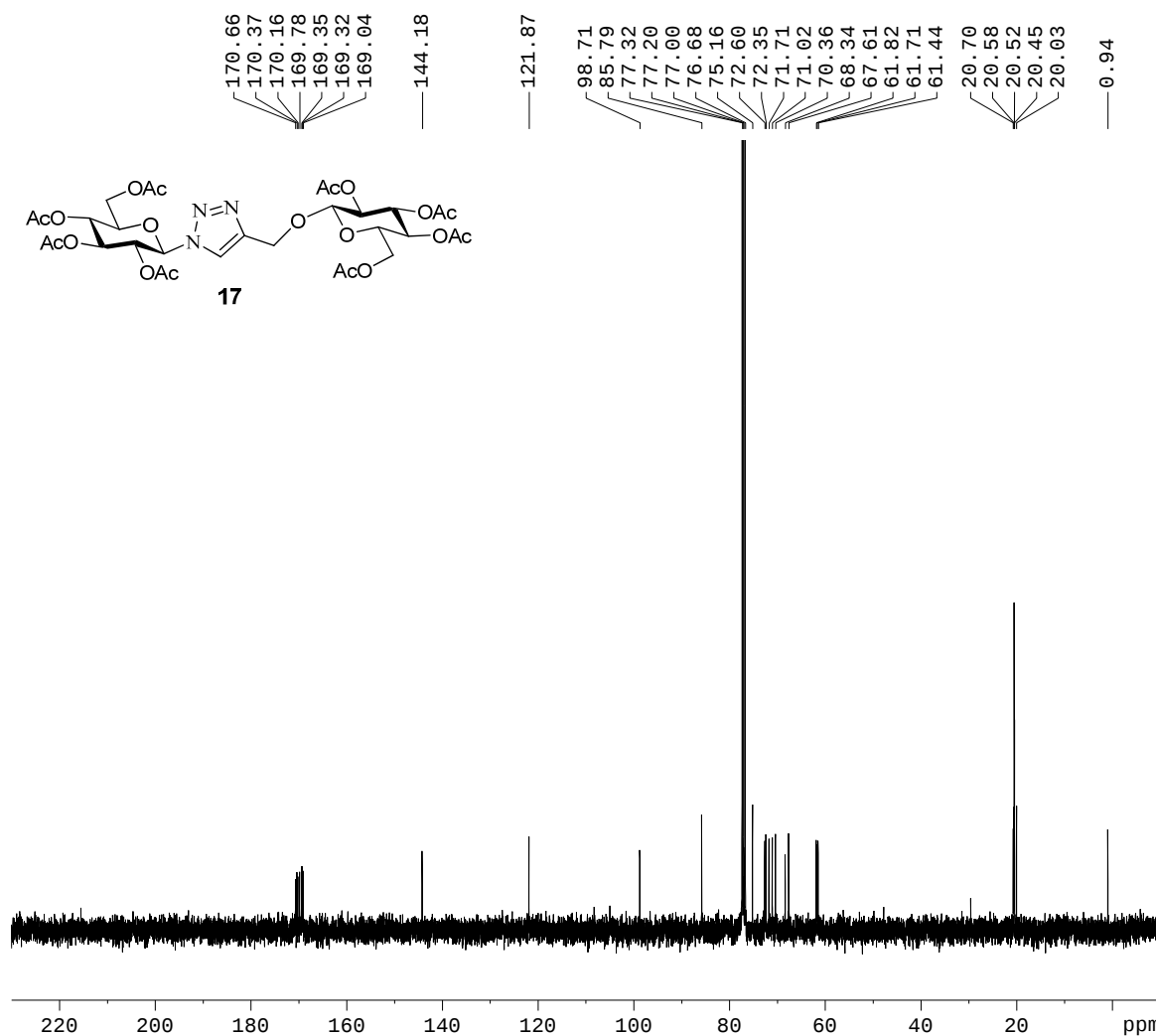
F2 - Acquisition Parameters
Date_ 20100107
Time 21.48
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 207
DS 0
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 4096
DW 19.950 usec
DE 6.50 usec
TE 297.1 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6243390 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127528 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





Current Data Parameters
NAME prp-triaz
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20100428
Time 16.39
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 84
DS 0
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 4096
DW 19.900 usec
DE 8.00 usec
TE 673.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.90 usec
PL1 5.00 dB
SF01 100.6243390 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 0.00 dB
PL12 19.10 dB
PL13 22.10 dB
SF02 400.1319000 MHz

F2 - Processing parameters
SI 32768
SF 100.6127509 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40