

Supporting Information for

Rational design of a nanometer-sized covalent octahedron

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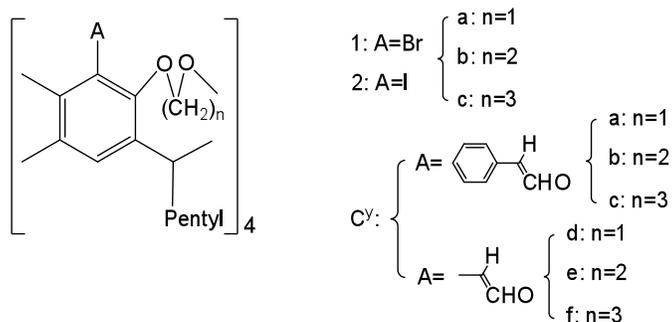
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Table of Content

- | | |
|--|--------------|
| 1. Experimental procedures for the synthesis of cavitands and nanocapsules | Page S2-S11 |
| 2. ^1H NMR, ^{13}C NMR, MALDI TOF MS and GPC traces | Page S12-S25 |

1. Experimental Procedures

1.1 General: Reagents and chromatography solvents were purchased from Aldrich and used without further purification except that chloroform was passed through K_2CO_3 prior to use. THF was dried over Na/benzophenone and distilled under argon. 1H NMR spectra recorded in $CDCl_3$ were referenced to residual $CHCl_3$ at $\delta_H = 7.26$. ^{13}C NMR spectra recorded in $CDCl_3$ were referenced to $^{13}CDCl_3$ at $\delta_C = 77.5$ ppm. Mass spectra were recorded on an Applied Biosystems Voyager DE-Pro mass spectrometer (MALDI-TOF). External standards were used for calibration and 2,4,6-trihydroxyacetophenone (THAP) as matrix. Gel permeation chromatography (GPC) was performed on a Thermo SpectraSYSTEM HPLC system equipped with dual wavelength UV/Vis detector (280 nm), Eppendorf CH-30 column heater and two Jordi GPC columns (cross linked DVB; 103 Å pore size; MW cutoff ~ 25,000; 7.8mm × 30cm) with $CH_2Cl_2/1\% NEt_3$ as mobile phase at a flow of 1 mL/min. Approximate molecular weights of analytes were determined from a semilogarithmic calibration plot.

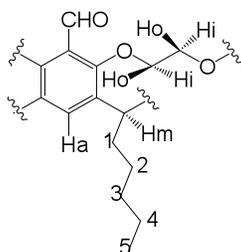


| Components | C^a | C^b | C^c [1] |
|--|-----------------|-----------------|-----------------|
| $H_2N\text{-} \text{---} \text{NH}_2$ L^a | Complex Mixture | $C^b_2L^a_4$ | Complex Mixture |
| $H_2N\text{-} \text{---} \text{NH}_2$ L^b | $C^a_4L^b_8$ | $C^b_6L^b_{12}$ | Complex Mixture |
| $H_2N\text{-} \text{---} \text{---} \text{---} \text{NH}_2$ L^c | $C^a_4L^c_8$ | $C^b_6L^c_{12}$ | Complex Mixture |
| $H_2N\text{-} \text{---} \text{---} \text{---} \text{NH}_2$ L^d | $C^a_4L^d_8$ | $C^b_2L^c_4$ | Complex Mixture |
| T | $C^a_6T_{12}$ | $C^b_6T_{12}$ | NA |

Table 1: TFA-catalyzed condensation products

1.2 Cavitand C^e

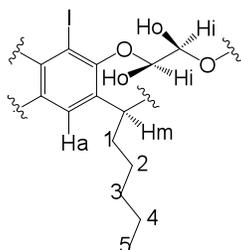
Tetrabromocavitand **1b**^[2] (520 mg, 0.53 mmol) was dried under vacuum overnight at 100 °C and then dissolved in 20 mL THF under argon. THF (10 mL) was added to a second dried flask, which was charged with PrMgBr/THF solution (2.6 mL, 2 mol/L) and anhydrous LiCl (216 mg, 5.10 mmol). The suspension was stirred under argon until all LiCl was dissolved. Then, the resulting solution was cannulated to the solution of **1c** at -78 °C. After the Br/MgBr exchange, which was monitored by ¹H NMR spectroscopy, was complete, anhydrous DMF (0.43 mL, 6.33 mmol) was injected to the solution. After 1 hr, the reaction was quenched with aqueous HCl solution (10 mL, 1M). The product was extracted with ethyl acetate (3×30 mL). The combined organic solution was washed with sat. NaCl solution, dried over anhydrous Na₂SO₄ and concentrated under vacuum. The product was further purified by silica gel column chromatography with THF/CH₂Cl₂=2/98 as the mobile phase. The white solid was obtained in 25% yield.



¹H NMR (400 MHz, CDCl₃, 25 °C): δ_H 10.35 (s, 4H, -CHO), 7.56 (s, 4H, H_a), 5.35 (t, *J*=8.1 Hz, 4H, H_m), 4.51 (m, 8H, H_o), 3.67 (m, 8H, H_i), 2.10 (m, 8H, H₁), 1.42-1.18 (m, 24H, H₂, H₃ and H₄), 0.88 (t, *J*=7.1Hz, 12H, H₅). ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ_C 190.24, 155.85, 137.37, 130.17, 125.13, 74.42, 34.50, 33.32, 32.28, 27.87, 23.07, 14.53. Ms (Maldi-tof) *m/z*: 1007.4911 (M+Na⁺, 100%); Calcd for C₆₀H₇₂O₁₂+Na⁺: 1007.4916.

1.3 Cavitand 2b

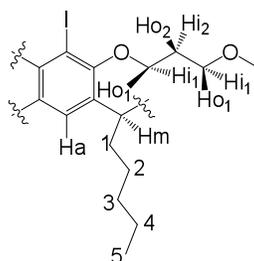
Tetrabromocavitand **1b** (530 mg, 0.54 mmol) was dried overnight under vacuum at 100 °C and then dissolved in 30 mL THF under argon. At -78 °C, BuLi/Hexane (1.6 mL, 2.5M) was injected into the above solution. After 3 hours, I₂ (1.3 g, 5.12 mmol) was added to the solution. The reaction was quenched with water. The product was extracted with ethyl acetate (3×30 mL) and then the combined organic solution was washed with Na₂S₂O₃ solution (20 mL, 10%), water and sat. NaCl solution, dried over anhydrous Na₂SO₄ and concentrated in vacuum. The product was further purified by silica gel column chromatography with CH₂Cl₂/Hexane=1/1 as mobile phase. The slightly yellow solid was obtained in 81% yield.



^1H NMR (500 MHz, CDCl_3 , 25 °C): δ_{H} 7.29 (s, 4H, H_a), 5.27 (t, $J=8.2$ Hz, 4H, H_m), 4.46 (m, 8H, H_o), 3.76 (m, 8H, H_i), 2.04 (m, 8H, H_1), 1.38-1.15 (m, 24H, H_2 , H_3 and H_4), 0.86 (t, $J=7.2$ Hz, 12H, H_5). ^{13}C NMR (125 MHz, CDCl_3 , 25 °C): δ_{C} 154.88, 136.98, 125.25, 90.36, 70.89, 36.05, 35.24, 32.31, 27.87, 23.05, 14.54. Ms (Maldi-tof) m/z : 1399.1543 ($\text{M}+\text{Na}^+$, 100%); Calcd for $\text{C}_{56}\text{H}_{68}\text{O}_{8}\text{I}_4+\text{Na}^+$: 1399.0984.

1.4 Cavitand 2c

Cavitand **1c**^[2] (1.03 g, 0.99 mmol) was dried overnight under vacuum at 100 °C and then dissolved in 40 mL THF under argon. THF (10 mL) was added to a second dried flask, which was charged with PrMgBr/THF solution (5 mL, 2 mol/L) and anhydrous LiCl (420 mg, 9.91 mmol). The suspension was stirring under argon until all LiCl was dissolved. Then, the resulting solution was cannulated to the solution of **1c** at -78 °C. After the Br/MgBr exchange, which was monitored by ^1H NMR spectroscopy, was complete, I_2 (2.52 g, 9.92 mmol) was added at the same temperature. The reaction was quenched with water. The product was extracted with ethyl acetate (3×50 mL). The combined organic solution was washed with $\text{Na}_2\text{S}_2\text{O}_3$ solution (50 mL, 10%), water and sat. NaCl solution, dried over anhydrous Na_2SO_4 and concentrated in vacuum. The product was further purified by silica column with Hexane/ $\text{EtOAc}=1/4$ as the mobile phase. The slightly yellow product was obtained in 70%.

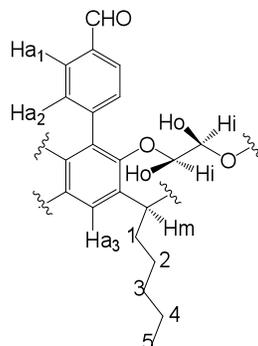


^1H NMR (400 MHz, CDCl_3 , 25 °C): δ_{H} 7.04 (s, 4H, H_a), 4.91 (t, $J=7.5$ Hz, 4H, H_m), 4.67 (m, 8H, H_o_2), 3.94 (m, 8H, H_i_2), 2.32 (m, 4H, H_o_1), 2.08 (m, 4H, H_i_1), 1.93 (m, 8H, H_1), 1.36-1.14 (m, 24H, H_2 , H_3 and H_4), 0.86 (t, $J=6.9$ Hz, 12H, H_5). ^{13}C NMR (100 MHz, CDCl_3 , 25 °C): δ_{C} 157.18, 134.89, 125.78, 90.23, 70.29, 36.46, 36.22, 34.45, 29.85, 27.74, 23.03, 14.59. Ms (Maldi-tof) m/z : 1455.1230 ($\text{M}+\text{Na}^+$, 100%); Calcd for $\text{C}_{60}\text{H}_{76}\text{O}_8\text{I}_4 + \text{Na}^+$: 1455.1611.

1.5 Cavitand C^b

A Schlenk flask was charged with cavitand **2b** (176.5 mg, 177.4 μmol), potassium (4-formylphenyl)trifluoroborate (225.6 mg, 1.06 mmol), $\text{Pd}(\text{OAc})_2$ (17.3 mg, 71 μmol), PPh_3 (37.2 mg, 142 μmol), and K_2CO_3 (294 mg, 2.13 mmol) and was evacuated and refilled with argon 3 times. After adding THF (10 mL) and deionized water (1 mL), the flask was sealed and kept stirring at 100 °C overnight. Then, the solution was acidified with aqueous HCl (3 mL, 1 M) and extracted with EtOAc (3×10 mL). The combined organic layers were washed with sat. NaCl solution, dried over anhydrous Na_2SO_4 and concentrated in vacuum. The pure product was obtained as a white solid in 65.4% yield

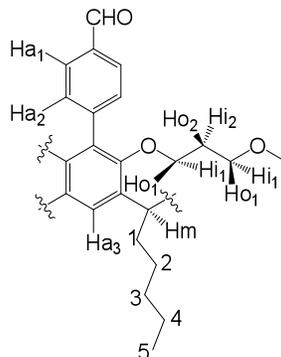
by using silica gel column with EtOAc/CH₂Cl₂=5/95 as the mobile phase.



¹H NMR (300 MHz, CDCl₃, 25 °C): δ_H 10.03 (s, 4H, -CHO), 7.89 (d, *J*=8.0, 8H, Ha₁), 7.63 (s, 4H, Ha₃), 7.34 (d, *J*=8.0 Hz, 8H, Ha₂), 5.26 (t, *J*=8.4, 4H, Hm), 3.84 (m, 8H, Ho), 3.41 (m, 8H, Hi), 2.23 (m, 8H, H1), 1.47-1.18 (m, 24H, H2, H3 and H4), 0.91 (t, *J*=6.8 Hz, 12H, H5). ¹³C NMR (125 MHz, CDCl₃, 25 °C): δ_C 192.18, 151.88, 142.38, 136.83, 136.02, 131.04, 130.58, 129.76, 125.16, 72.76, 35.22, 34.41, 32.37, 28.09, 23.16, 14.60. Ms (Maldi-tof) *m/z*: 1311.6413 (M+Na⁺, 100%); Calcd for C₈₄H₈₈O₁₂+Na⁺: 1311.6167.

1.6 Cavitand C^c

A Schlenk flask was charged with cavitand **2c** (103.15 mg, 72 μmol), potassium (4-formylphenyl)trifluoroborate (92 mg, 432.2 μmol), Pd(OAc)₂ (3.5 mg, 14.4 μmol), PPh₃ (7.5 mg, 28.8 μmol), and K₂CO₃ (119.3 mg, 864.4 μmol) and was evacuated and refilled with argon 3 times. After adding THF (10 mL) and deionized water (1mL), the flask was sealed and kept stirring at 100 °C overnight. Then, the solution was acidified with aqueous HCl (3 mL, 1M) and extracted with EtOAc (3×10 mL). The combined organic layers were washed with sat. NaCl solution, dried over anhydrous Na₂SO₄ and concentrated in vacuum. The pure product was then obtained as a white solid in 71% yield by using silica column with EtOAc/CH₂Cl₂=1/35 as the mobile phase.



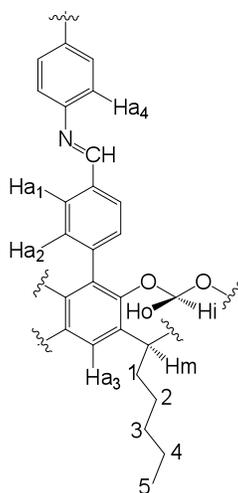
¹H NMR (300 MHz, CDCl₃, 25 °C): δ_H 10.04 (s, 4H, -CHO), 7.88 (d, *J*=8.0, 8H, Ha₁), 7.54 (d, *J*=8.0 Hz, 8H, Ha₂), 7.46 (s, 4H, Ha₃), 5.01 (t, *J*=8.0, 4H, Hm), 3.79 (m, 8H, Ho₂), 3.52 (m, 8H, Hi₂), 2.12 (m, 8H, H1), 1.87 (m, 4H, Ho₁), 1.54 (m, 8H, Hi₁), 1.44-1.20 (m, 24H, H2, H3 and H4), 0.90 (t, *J*=6.8 Hz, 12H, H5). ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ_C 192.54, 154.37, 143.57, 135.54, 134.38, 132.15, 129.50, 128.39, 125.72, 71.79, 36.53, 34.68, 32.49, 30.32, 27.99, 23.18, 14.62. Ms (Maldi-tof) *m/z*: 1367.6341 (M+Na⁺, 100%); Calcd for C₈₈H₉₆O₁₂+Na⁺: 1367.6799.

1.7 General procedure for synthesis of nanocapsules (Procedure A)

Stock solutions of the tetraformylcavitand (10 mg/mL), of the di- or triamine linker (10 mg/mL) and of TFA (1% v/v) in CDCl_3 were prepared. Then, the solutions of cavitand and linker were mixed in stoichiometric ratio followed by the addition of catalytic amount TFA solution. Several molecular sieves were added to above solution. After 1-2 hours, the solution was diluted to 0.5 ml with CDCl_3 , if necessary, and transferred into a NMR tube. The reaction was monitored by ^1H NMR spectroscopy. After completion, the solvent was removed.

1.7.1 Tetracavitand capsule $\text{C}^{\text{a}}_4\text{L}^{\text{b}}_8$

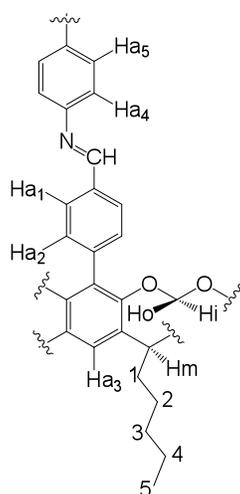
From $\text{C}^{\text{a}[1]}$ (3.58 mg, 2.91 μmol), 1,4-phenylenediamine (0.63 mg, 5.81 μmol) and TFA (4 μL , 1%v/v in CDCl_3) according to **procedure A**. Based on GPC, the purity is 90% and the rest are hexacavitand-aggregates.



^1H NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{b}}_8$: δ_{H} 8.53 (s, 8H, -CHN-), 8.51 (s, 8H, -CHN-), 7.94 (d, $J=7.5$ Hz, 16H, Ha₁), 7.83 (d, $J=7.5$ Hz, 16H, Ha₁), 7.38 (s, 8H, Ha₃), 7.35 (s, 8H, Ha₃), 7.29 (s, 16H, Ha₄), 7.27 (d, $J=8.6$ Hz, 16H, Ha₂), 7.24 (s, 16H, Ha₄), 7.08 (d, $J=8.6$ Hz, 16H, Ha₂), 5.48-5.30 (m, 12H, Ho), 4.97-4.78 (m, 16H, Hm), 4.64 (m, 4H, Hi), 4.51-4.30 (m, 12H, Hi), 3.56 (m, 4H, Ho), 2.47-2.2 (m, 32H, H1), 1.57-1.34 (m, 96H, H2, H3 and H4), 1.04-0.86 (m, 48H, H5). ^{13}C NMR (125 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{b}}_8$: δ_{C} 159.92, 159.08, 153.33, 153.08, 152.86, 152.77, 150.68, 149.78, 139.06, 138.99, 138.92, 138.77, 137.74, 137.69, 135.78, 135.70, 131.03, 130.76, 129.64, 129.17, 128.81, 122.57, 122.33, 122.26, 120.81, 120.67, 101.57, 101.41, 99.43, 37.70, 37.61, 32.59, 32.58, 32.55, 31.09, 30.89, 30.43, 28.23, 28.20, 28.16, 23.24, 23.21, 23.18, 14.64, 14.63. Dosy NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{b}}_8$: $D=2.50 \times 10^{-10}$ m^2/s . Ms (Maldi-tof) of $\text{C}^{\text{a}}_4\text{L}^{\text{b}}_8$ m/z : 5511.6259 ($\text{M}+\text{H}^+$, 100%); Calcd for $\text{C}_{368}\text{H}_{352}\text{O}_{32}\text{N}_{16} + \text{H}^+$: 5511.6503.

1.7.2 Tetra-cavitand capsule $\text{C}^{\text{a}}_4\text{L}^{\text{c}}_8$

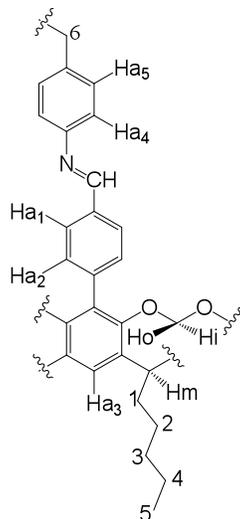
From $\text{C}^{\text{a}[1]}$ (2.33 mg, 1.88 μmol), benzidine (0.69 mg, 3.77 μmol) and TFA (2 μL , 1%v/v in CDCl_3) according to **procedure A**. Based on GPC, the purity is 80% and the rest are hexacavitand-aggregates.



^1H NMR (300 MHz, CDCl_3 , 25 °C) of $\text{C}^{\text{a}}_4\text{L}^{\text{c}}_8$: δ_{H} 8.48 (s, 8H, -CHN-), 8.45 (s, 8H, -CHN-), 7.88 (d, $J=8.6$ Hz, 16H, Ha₁), 7.79 (d, $J=8.6$ Hz, 16H, Ha₁), 7.60 (d, $J=8.6$ Hz, 16H, Ha₅), 7.54 (d, $J=8.6$ Hz, 16H, Ha₅), 7.32 (s, 8H, Ha₃), 7.30 (s, 8H, Ha₃), 7.25 (d, $J=8.4$ Hz, 16H, Ha₄), 7.20 (d, $J=8.4$ Hz, 16H, Ha₄), 7.14 (d, $J=7.8$ Hz, 16H, Ha₂), 7.04 (d, $J=7.8$ Hz, 16H, Ha₂), 5.35-5.28 (m, 12H, Ho), 4.85-4.79 (m, 16H, Hm), 4.70 (m, 4H, Hi), 4.37-4.29 (m, 12H, Hi), 3.65 (m, 4H, Ho), 2.37-2.24 (m, 32H, H1), 1.49-1.32 (m, 96H, H2, H3 and H4), 0.96-0.86 (m, 48H, H5). ^{13}C NMR (125 MHz, CDCl_3 , 25 °C) of $\text{C}^{\text{a}}_4\text{L}^{\text{c}}_8$: δ_{C} 160.39, 160.27, 159.56, 153.30, 153.07, 152.95, 152.82, 151.82, 151.70, 151.01, 139.02, 138.95, 138.80, 137.86, 137.79, 137.63, 135.75, 135.70, 135.62, 131.00, 130.77, 129.65, 129.23, 129.14, 128.87, 128.16, 128.09, 127.77, 122.04, 121.92, 121.89, 120.82, 120.71, 115.91, 101.44, 101.16, 99.77, 37.69, 37.64, 32.59, 31.06, 30.98, 30.53, 28.19, 23.22, 14.65. Dosy NMR (500 MHz, CDCl_3 , 25 °C) of $\text{C}^{\text{a}}_4\text{L}^{\text{c}}_8$: $D=2.14 \times 10^{-10}$ m²/s. Ms (Maldi-tof) of $\text{C}^{\text{a}}_4\text{L}^{\text{c}}_8$ m/z: 6119.9712 (M+H⁺, 100%); Calcd for $\text{C}_{416}\text{H}_{384}\text{O}_{32}\text{N}_{16} + \text{H}^+$: 6119.9100.

1.7.3 Tetra-cavitand capsule $\text{C}^{\text{a}}_4\text{L}^{\text{d}}_8$

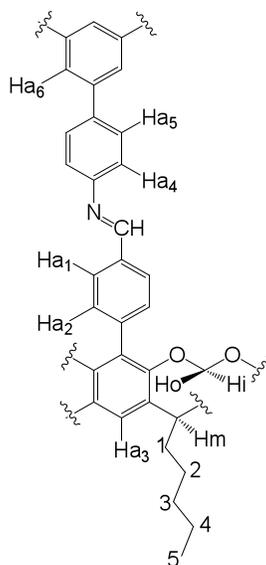
From $\text{C}^{\text{a}[1]}$ (2.35 mg, 1.90 μmol), 1,4-ethylenedianiline (0.81 mg, 3.81 μmol) and TFA (2 μL , 1%v/v in CDCl_3) according to **procedure A**. Based on GPC, the purity is 80% and the rest are di- and hexacavitand-aggregates.



^1H NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{d}}_8$: δ_{H} 8.50 (s, 8H, -CHN-), 8.47 (s, 8H, -CHN-), 7.92 (d, $J=7.7$ Hz, 16H, Ha_1), 7.85 (d, $J=7.7$ Hz, 16H, Ha_1), 7.37 (s, 8H, Ha_3), 7.36 (s, 8H, Ha_3), 7.34 (d, $J=8.2$ Hz, 16H, Ha_5), 7.29 (d, $J=8.2$ Hz, 16H, Ha_5), 7.24-7.16 (m, 48H, Ha_4 and Ha_2), 7.12 (d, $J=7.6$ Hz, 16H, Ha_2), 5.41-5.27 (m, 12H, Ho), 4.95-4.86 (m, 16H, Hm and Hi), 4.42-4.29 (m, 12H, Hi), 3.88 (m, 4H, Ho), 2.94 (s, 16H, H_6), 2.91 (s, 16, H_6), 2.46-2.27 (m, 32H, H_1), 1.61-1.36 (m, 96H, H_2 , H_3 and H_4), 1.03-0.92 (m, 48H, H_5). ^{13}C NMR (125 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{d}}_8$: δ_{C} 159.88, 159.52, 153.22, 153.06, 152.98, 152.87, 150.57, 150.24, 140.70, 140.52, 139.04, 139.00, 138.97, 138.81, 137.74, 137.67, 135.76, 135.73, 130.91, 130.71, 129.98, 129.72, 129.64, 129.45, 129.40, 129.30, 128.77, 121.59, 121.52, 120.77, 120.66, 115.70, 101.37, 100.16, 38.46, 38.43, 37.67, 37.62, 32.59, 31.01, 30.60, 28.19, 23.24, 23.21, 14.64. Dosy NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$) of $\text{C}^{\text{a}}_4\text{L}^{\text{d}}_8$: $D=2.06 \times 10^{-10}$ m^2/s . Ms (Maldi-tof) of $\text{C}^{\text{a}}_4\text{L}^{\text{d}}_8$ m/z : 6345.1584 ($\text{M}+\text{H}^+$, 100%); Calcd for $\text{C}_{432}\text{H}_{416}\text{O}_{32}\text{N}_{16}+\text{H}^+$: 6345.1495.

1.7.4 Hexa-cavitand capsule $\text{C}^{\text{a}}_6\text{T}_{12}$

From $\text{C}^{\text{a}[1]}$ (2.41 mg, 1.90 μmol), 1,3,5-tris(4-aminophenyl)benzene (0.91 mg, 2.61 μmol) and TFA (2 μL , 1%v/v in CDCl_3) according to **procedure A**. Based on GPC, the purity is >95%.

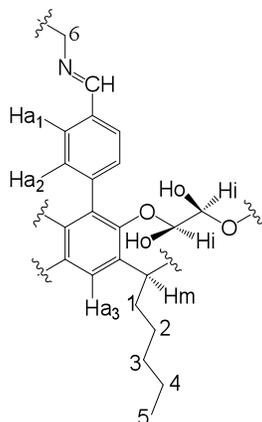


^1H NMR (300 MHz, CDCl_3 , 25 $^\circ\text{C}$): δ_{H} 8.53 (s, 24H, -CHN-), 7.92 (d, $J=8.3$ Hz, 48H, Ha_1), 7.82 (s, 24H, Ha_3), 7.76 (d, $J=8.3$ Hz, 48H, Ha_5), 7.39 (s, 24H, Ha_6), 7.32 (d, $J=8.3$ Hz, 48H, Ha_4), 7.19 (d, $J=8.3$ Hz, 48H, Ha_2), 5.24 (m, 24H, Ho), 4.89 (t, $J=7.8$ Hz, 24H, Hm), 4.25 (m, 24H, Hi), 2.39 (m, 48H, H_1), 1.58-1.35 (m, 144H, H_2 , H_3 and H_4), 0.98 (t, $J=6.7$ Hz, 72H, H_5). ^{13}C NMR (125 MHz, CDCl_3 , 25 $^\circ\text{C}$): δ_{C} 160.48, 153.03, 152.10, 142.30, 139.21, 138.94, 138.14, 135.58, 130.68, 129.53, 128.90, 124.54, 125.02, 121.85, 120.72, 101.05, 37.59, 32.59, 30.84, 28.20, 23.21. Dosy NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$): $D=1.85 \times 10^{-10}$ m^2/s . Ms (Maldi-tof) m/z : 9779.8018 ($\text{M}+\text{H}^+$, 100%); Calcd for $\text{C}_{672}\text{H}_{600}\text{O}_{48}\text{N}_{24}+\text{H}^+$: 9780.5500

1.7.5 Hemicarcerand $\text{C}^{\text{b}}_2\text{L}^{\text{a}}_4$

From C^{b} (2.08 mg, 1.61 μmol), 1,2-ethylenediamine (0.19 mg, 3.23 μmol) and TFA (2 μL ,

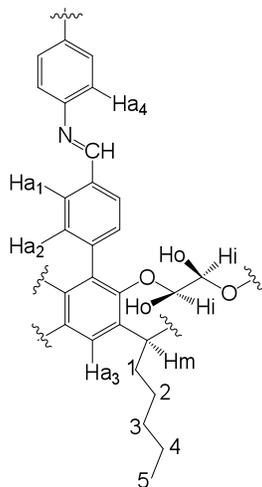
1%v/v in CDCl₃) according to **procedure A**. NMR shows that the purity is >95%.



¹H NMR (500 MHz, CDCl₃, 25 °C): δ_H 8.46 (s, 8H, -CHN-), 7.83 (d, *J*=8.6 Hz, 16H, Ha₁), 7.65 (s, 8H, Ha₃), 7.23 (d, *J*=8.6 Hz, 48H, Ha₁), 5.28 (t, *J*=7.8 Hz, 8H, Hm), 3.97-3.72 (m, 32H, Ho and H6), 3.39 (m, 16H, Hi), 2.24 (m, 16H, H1), 1.50-1.18 (m, 48H, H2, H3 and H4), 0.90 (t, *J*=7.1 Hz, 24H, H5). ¹³C NMR (125 MHz, CDCl₃, 25 °C): δ_C 161.86, 151.89, 137.93, 136.31, 135.75, 130.63, 130.08, 128.23, 124.37, 72.53, 62.95, 34.95, 34.14, 32.19, 27.94, 22.92. Ms (Maldi-tof) *m/z*: 2676.5396 (M+H⁺, 100%); Calcd for C₁₇₆H₁₉₂O₁₆N₈ +H⁺: 2676.4652

1.7.6 Hexa-cavitand capsule C^b₆L^b₁₂

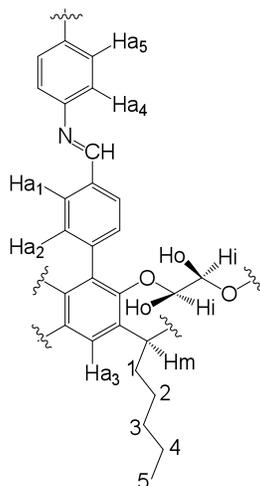
From C^b (2.32 mg, 1.80 μmol), 1,4-phenylenediamine (0.40 mg, 3.6 μmol) and TFA (2.5 μL, 1%v/v in CDCl₃) according to **procedure A**. NMR shows that the purity is >95%.



¹H NMR (300 MHz, CDCl₃, 25 °C): δ_H 8.53 (s, 24H, -CHN-), 7.93 (d, *J*=8.3 Hz, 48H, Ha₁), 7.67 (s, 24H, Ha₃), 7.31 (d, *J*=8.3 Hz, 48H, Ha₁), 7.26 (s, 48H, Ha₄), 5.30 (t, *J*=7.2 Hz, 24H, Hm), 3.89 (m, 48H, Ho), 3.46 (m, 48H, Hi), 2.26 (m, 48H, H1), 1.52-1.22 (m, 144H, H2, H3 and H4), 0.92 (t, *J*=6.4 Hz, 72H, H5). ¹³C NMR (125 MHz, CDCl₃, 25 °C): δ_C 159.81, 152.10, 150.45, 139.22, 136.68, 135.88, 130.82, 130.74, 128.88, 124.81, 122.32, 72.78, 35.20, 34.50, 32.47, 28.22, 23.19, 14.63. Dosy NMR (500 MHz, CDCl₃, 25 °C): D=2.11*10⁻¹⁰ m²/s. Ms (Maldi-tof) *m/z*: 8603.1035 (M+H⁺, 100%); Calcd for C₅₇₆H₅₇₆O₄₈N₂₄ +H⁺: 8603.3647

1.7.7 Hexa-cavitand capsule C^bL_{12}

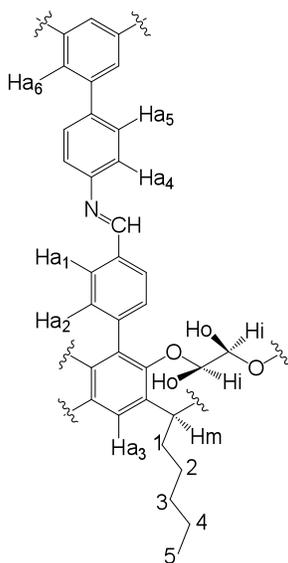
From C^b (2.07 mg, 1.61 μmol), benzidine (0.59 mg, 3.22 μmol) and TFA (2 μL , 1%v/v in CDCl_3) according to **procedure A**. NMR shows that the purity is >95%.



^1H NMR (300 MHz, CDCl_3 , 25 $^\circ\text{C}$): δ_{H} 8.55 (s, 24H, -CHN-), 7.94 (d, $J=7.8$ Hz, 48H, Ha₁), 7.75-7.60 (m, 72H, Ha₃ and Ha₂), 7.41-7.25 (m, 96H, Ha₄ and Ha₅), 5.31 (t, $J=8.1$ Hz, 24H, Hm), 3.91 (m, 48H, Ho), 3.48 (m, 48H, Hi), 2.26 (m, 48H, H1), 1.49-1.28 (m, 144H, H2, H3 and H4), 0.92 (t, $J=6.8$ Hz, 72H, H5). ^{13}C NMR (125 MHz, CDCl_3 , 25 $^\circ\text{C}$): δ_{C} 160.09, 152.10, 151.51, 139.22, 138.86, 136.69, 135.87, 130.82, 128.88, 128.11, 124.82, 121.89, 72.82, 35.24, 34.45, 32.48, 28.22, 23.20, 14.64. Dosy NMR (500 MHz, CDCl_3 , 25 $^\circ\text{C}$): $D=1.73 \times 10^{-10}$ m^2/s . Ms (Maldi-tof) m/z : 9516.8574 ($\text{M}+\text{H}^+$, 100%); Calcd for $\text{C}_{648}\text{H}_{624}\text{O}_{48}\text{N}_{24}+\text{H}^+$: 9516.7676

1.7.8 Hexa-cavitand capsule C^bT_{12}

From C^b (1.88 mg, 1.46 μmol), 1,3,5-tris(4-aminophenyl)benzene (0.85 mg, 2.43 μmol) and TFA (2 μL , 1%v/v in CDCl_3) according to **procedure A**. NMR shows that the purity is >95%.

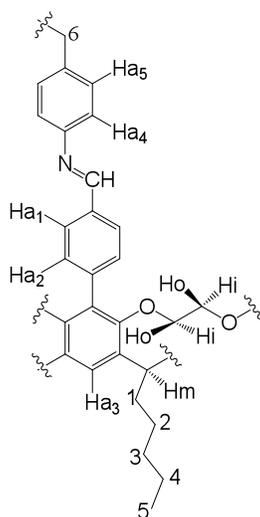


^1H NMR (300 MHz, CDCl_3 , 25 $^\circ\text{C}$): δ_{H} 8.52 (s, 24H, -CHN-), 7.95 (d, $J=8.2$ Hz, 48H, Ha₁),

7.79 (s, 24H, Ha₃), 7.74 (d, J=8.2 Hz, Ha₂), 7.67 (s, 24H, Ha₆), 7.38-7.27 (m, 96H, Ha₄ and Ha₅), 5.32 (t, J=7.5 Hz, 24H, Hm), 3.87 (m, 48H, Ho), 3.68 (m, 48H, Hi), 2.28 (m, 48H, H1), 1.54-1.31 (m, 144H, H2, H3 and H4), 0.94 (t, J=6.8 Hz, 72H, H5). ¹³C NMR (125 MHz, CDCl₃, 25 °C): δ_C 160.37, 152.11, 151.98, 142.50, 139.45, 139.29, 136.72, 135.82, 131.03, 130.85, 128.90, 128.65, 125.23, 124.76, 121.87, 72.65, 35.12, 34.56, 32.51, 28.26, 23.20, 14.84. Dosy NMR (500 MHz, CDCl₃, 25 °C): D=1.80*10⁻¹⁰ m²/s. Ms (Maldi-tof) m/z: 10117.4707 (M+H⁺, 100%); Calcd for C₆₉₆H₆₄₈O₄₈N₂₄+H⁺: 10117.4104

1.7.9 Hemicarcerand capsule C^b₂L^d₄

From C^b (2.12 mg, 1.65 μmol), 1,2-ethylenedianiline (0.70 mg, 3.30 μmol) and TFA (2 μL, 1%v/v in CDCl₃) according to **procedure A**. NMR shows that the purity is 95% and the rest are larger aggregates.



¹H NMR (500 MHz, CDCl₃, 25 °C) of C^b₂L^d₄: δ_H 8.44 (s, 8H, -CHN-), 7.89 (d, J=7.7 Hz, 16H, Ha₂), 7.69 (s, 8H, Ha₃), 7.29 (d, J=7.7 Hz, 48H, Ha₂), 7.04 (d, J=8.3 Hz, 16H, Ha₄), 6.96 (d, J=8.3 Hz, 16H, Ha₅), 5.31 (t, J=8.3 Hz, 8H, Hm), 3.90 (m, 16H, Ho), 3.45 (m, 16H, Hi), 2.98 (s, 16H, H6), 2.25 (m, 16H, H1), 1.46-0.90 (t, J=7.1 Hz, 24H, H5). ¹³C NMR (125 MHz, CDCl₃, 25 °C) of C^b₂L^d₄: δ_C 161.86, 151.89, 137.93, 136.31, 135.75, 130.63, 130.08, 128.23, 124.37, 72.53, 62.95, 34.95, 34.14, 32.19, 27.94, 22.92. Ms (Maldi-tof) of C^b₂L^d₄ m/z: 3284.6484 (M+H⁺, 100%); Calcd for C₂₂₄H₂₂₄O₁₆N₈+H⁺: 3284.7094

Reference:

- [1] C. B. Aakeroy, P. D. Chopade, *Org. Lett.* **2010**, *13*, 1-3.
- [2] R. C. Helgeson, K. Paek, C. B. Knobler, E. F. Maverick, D. J. Cram, *J. Am. Chem. Soc.* **1996**, *118*, 5590-5604.

2. NMR Spectra, Maldi-TOF Mass Spectra and GPC Chromatograms

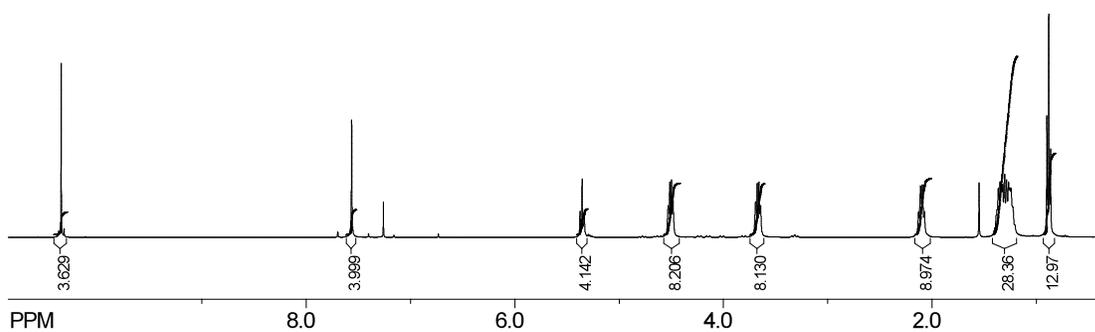


Figure S-1: ¹H NMR spectrum (400 MHz, CDCl₃, 25 °C) of C^e

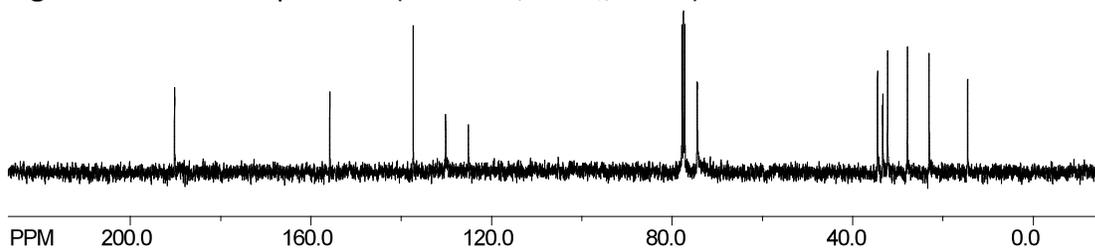


Figure S-2: ¹³C NMR spectrum (100 MHz, CDCl₃, 25 °C) of C^e

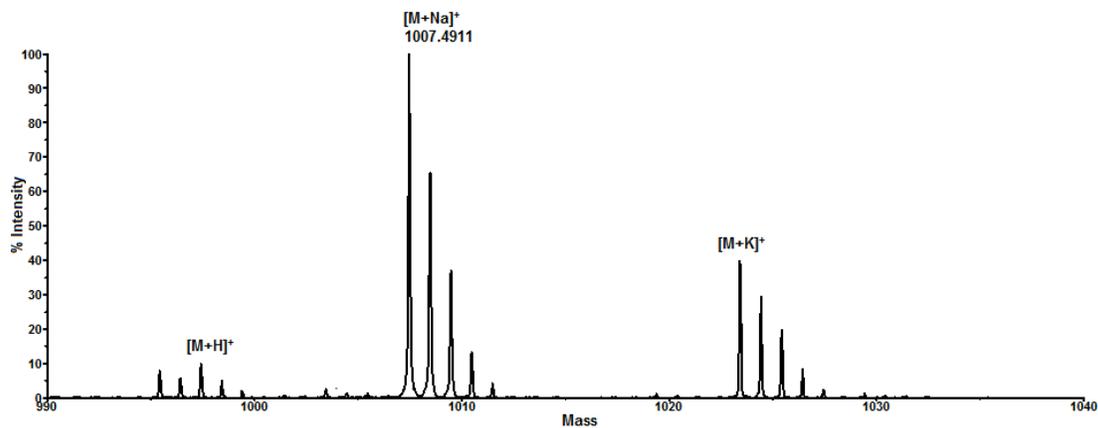


Figure s-3: Maldi-tof Mass Spectrum of C^e.

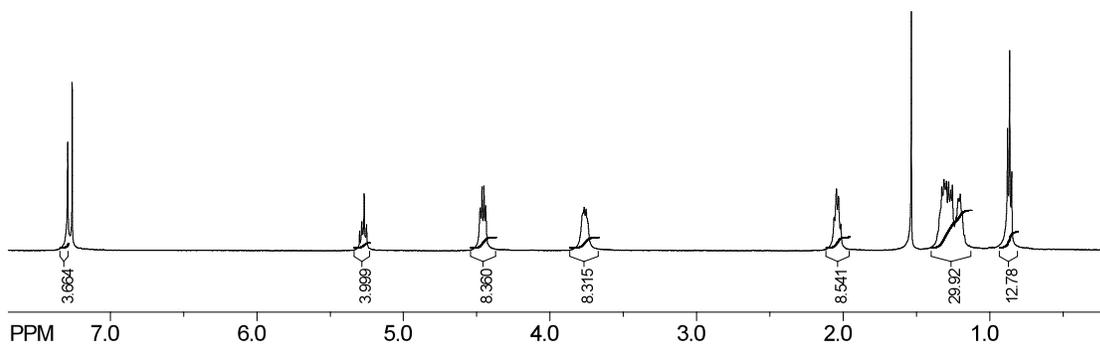


Figure S-4: ^1H NMR spectrum (500 MHz, CDCl_3 , 25 $^\circ\text{C}$) of **2b**.

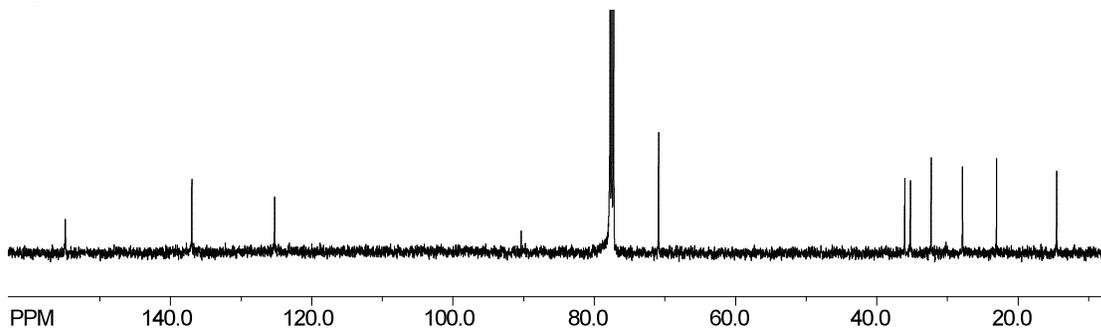


Figure S-5: ^{13}C NMR spectrum (125 MHz, CDCl_3 , 25 $^\circ\text{C}$) of **2b**

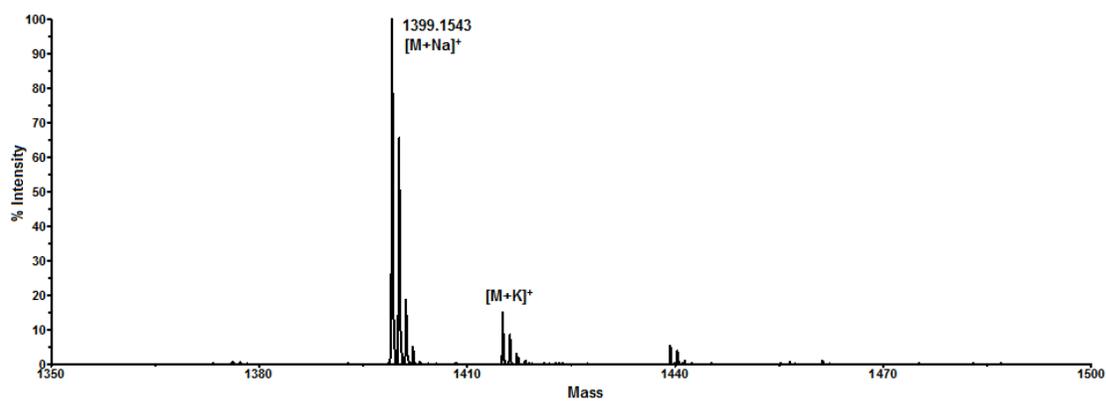


Figure S-6: Maldi-tof Mass Spectrum of **2b**.

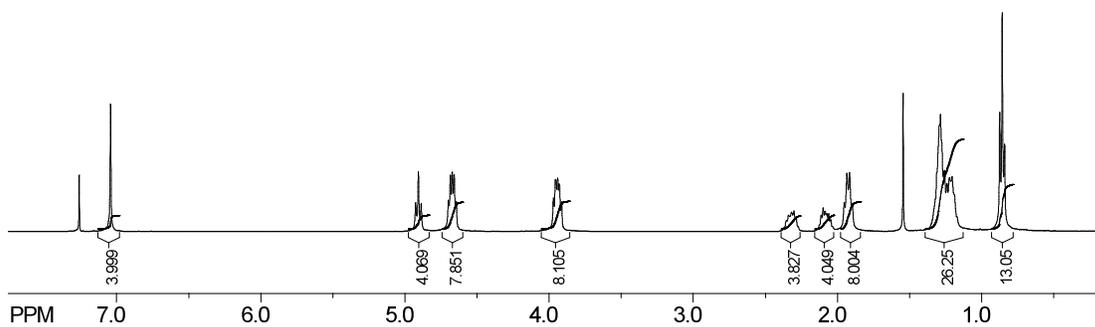


Figure S-7: ^1H NMR spectrum (400 MHz, CDCl_3 , 25 $^\circ\text{C}$) of **2c**.

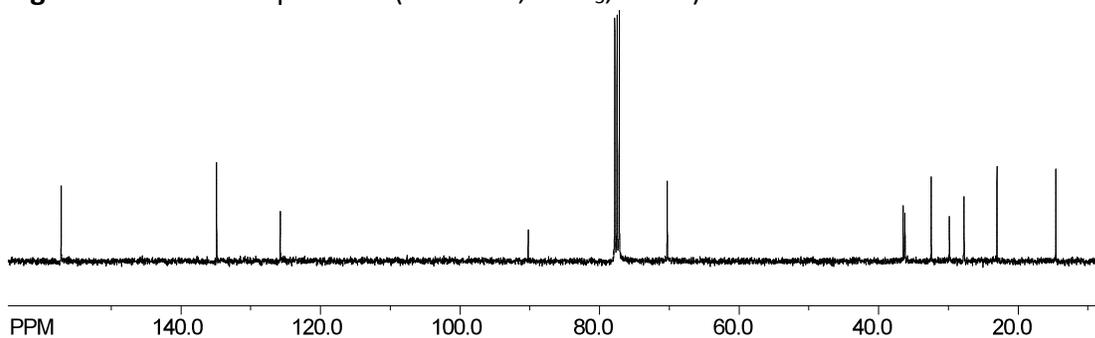


Figure S-8: ^{13}C NMR spectrum (100 MHz, CDCl_3 , 25 $^\circ\text{C}$) of **2c**.

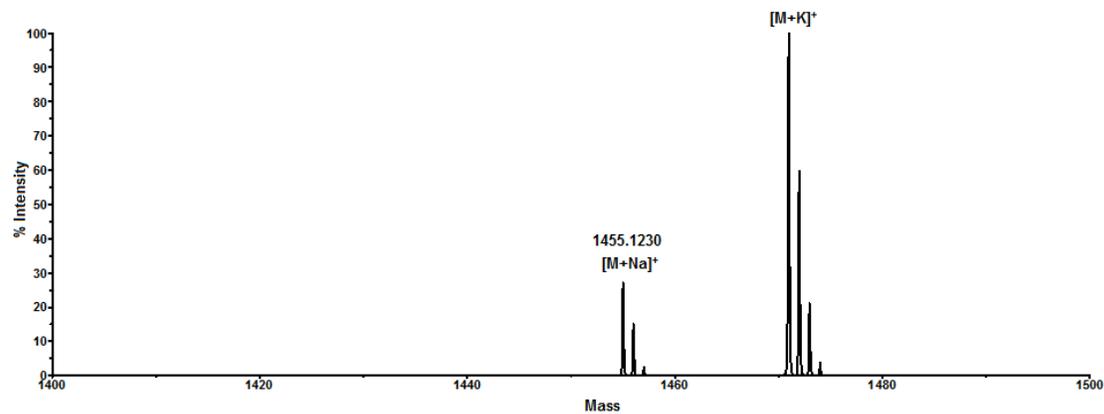


Figure S-9: Maldi-tof Mass Spectrum of **2c**.

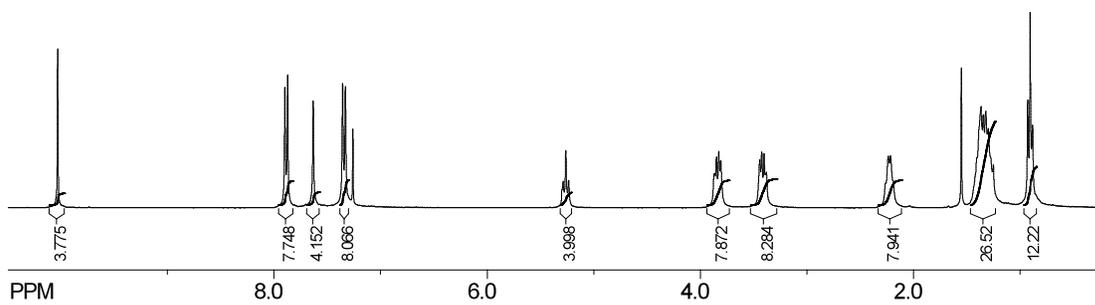


Figure S-10: ¹H NMR spectrum (300 MHz, CDCl₃, 25 °C) of **C^b**.

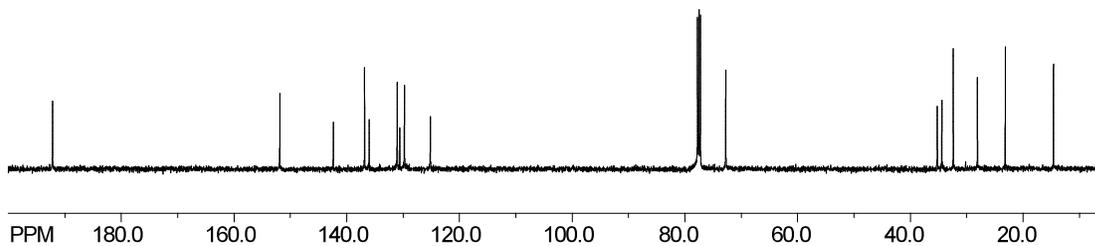


Figure S-11: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of **C^b**.

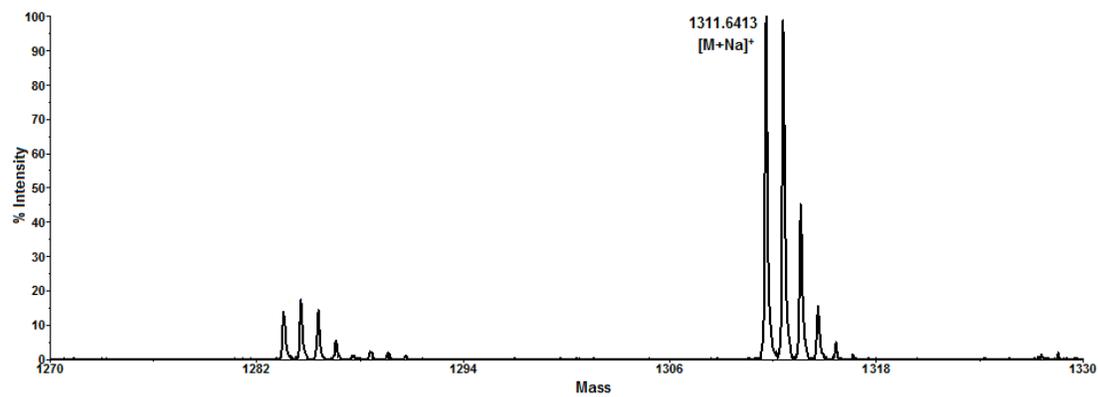


Figure S-12: Maldi-tof Mass Spectrum of **C^b**.

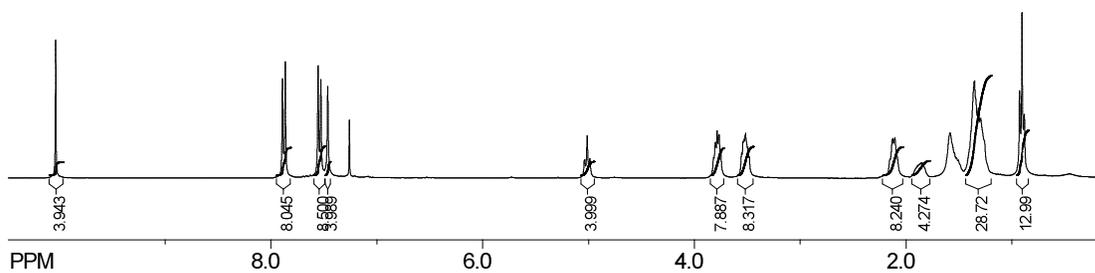


Figure S-13: ^1H NMR spectrum (300 MHz, CDCl_3 , 25 $^\circ\text{C}$) of C^c .

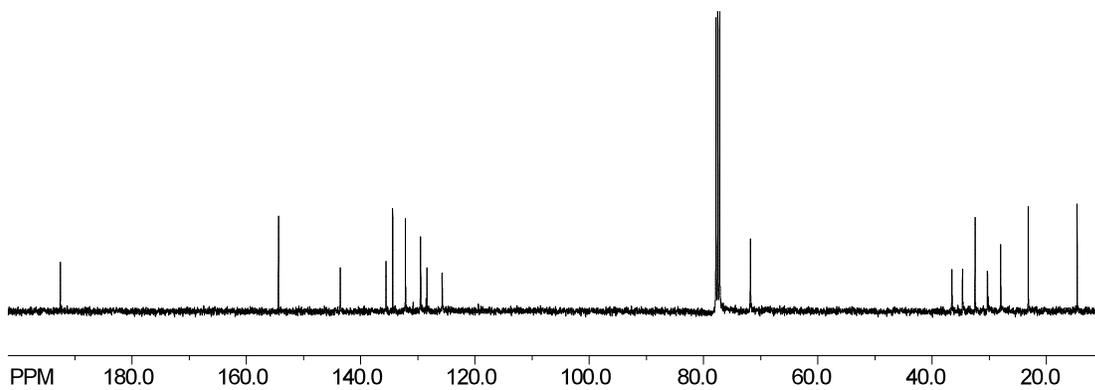


Figure S-14: ^{13}C NMR spectrum (100 MHz, CDCl_3 , 25 $^\circ\text{C}$) of C^c .

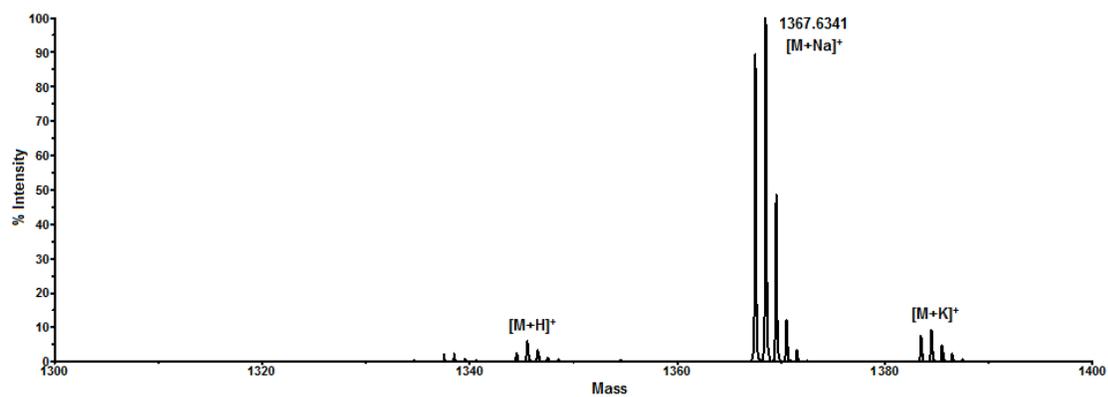


Figure S-15: Maldi-tof Mass Spectrum of C^c .

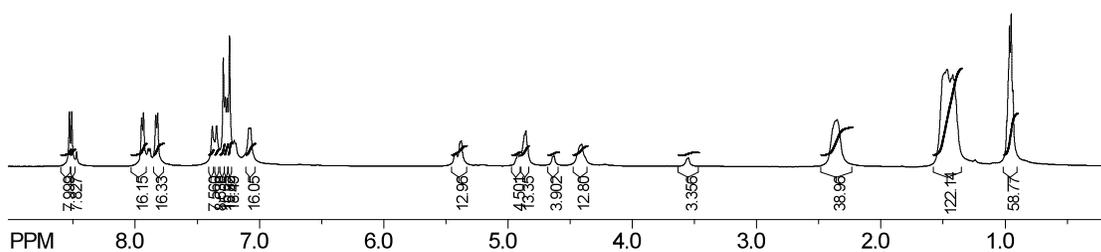


Figure S-16: ¹H NMR spectrum (500 MHz, CDCl₃, 25 °C) of C^a₄L^b₈

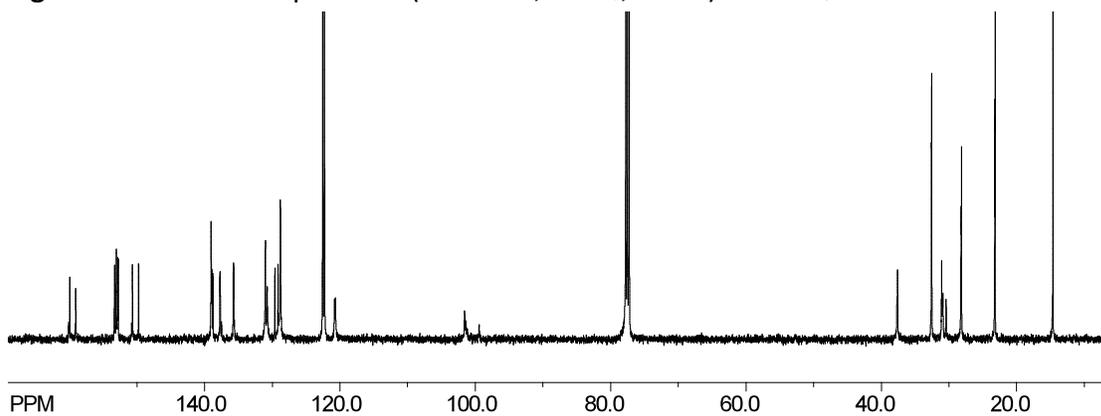


Figure S-17: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of C^a₄L^b₈

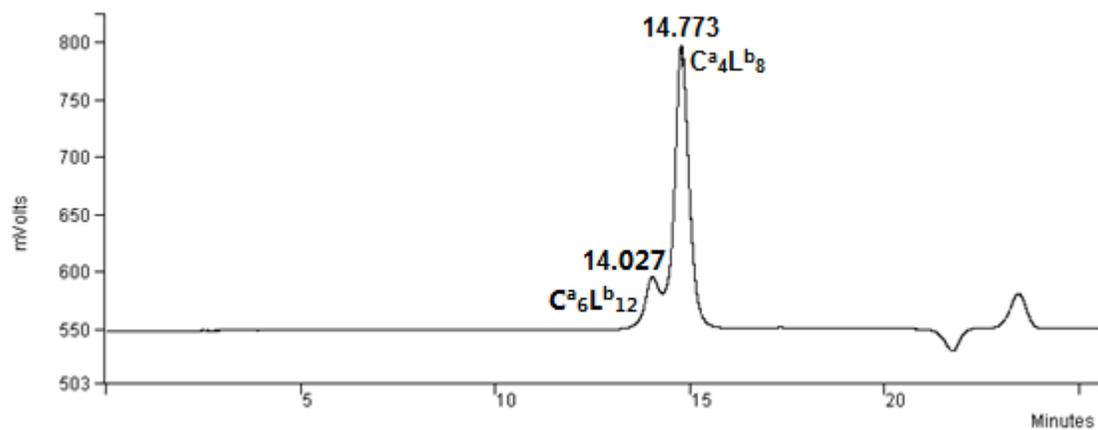


Figure S-18: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of C^a₄L^b₈

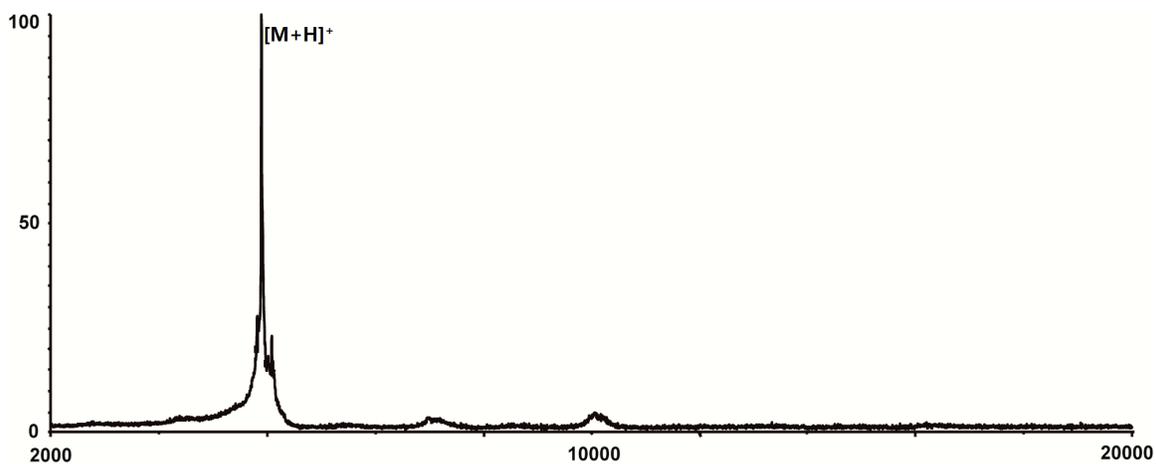


Figure S-19: Maldi-tof Mass Spectrum of C^a₄L^b₈

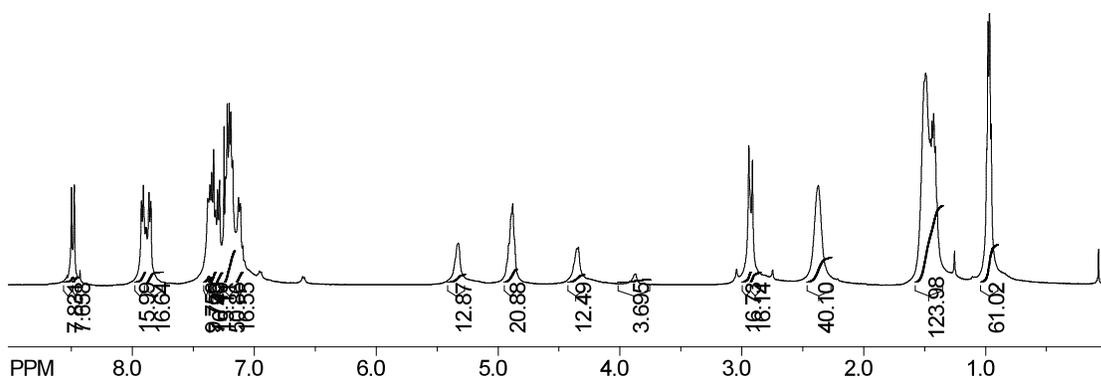


Figure S-24: ¹H NMR spectrum (500 MHz, CDCl₃, 25 °C) of C₄L^d₈

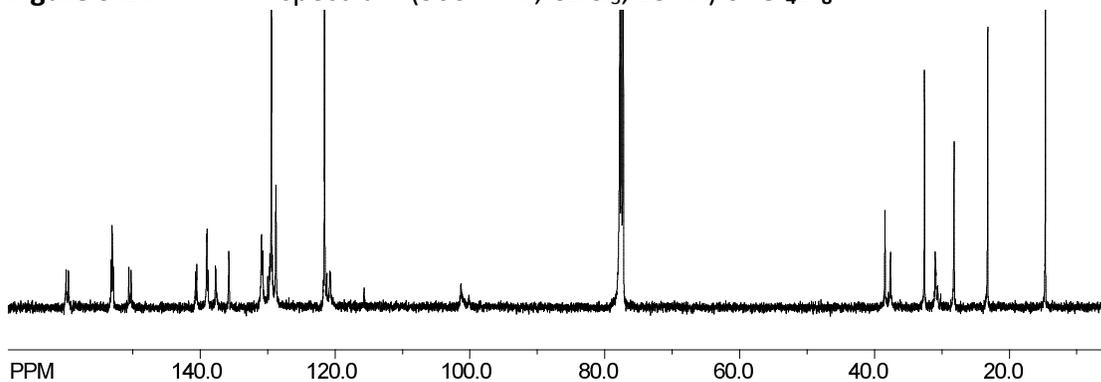


Figure S-25: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of C^a₄L^d₈

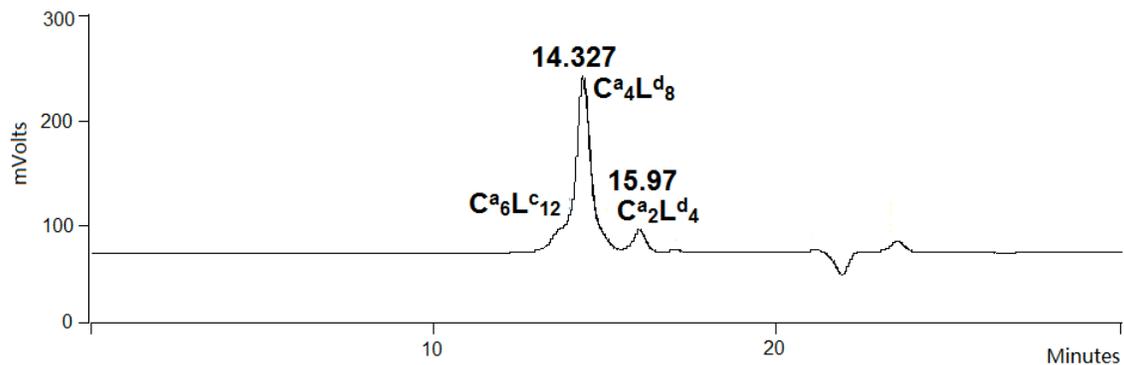


Figure S-26: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of C^a₄L^d₈

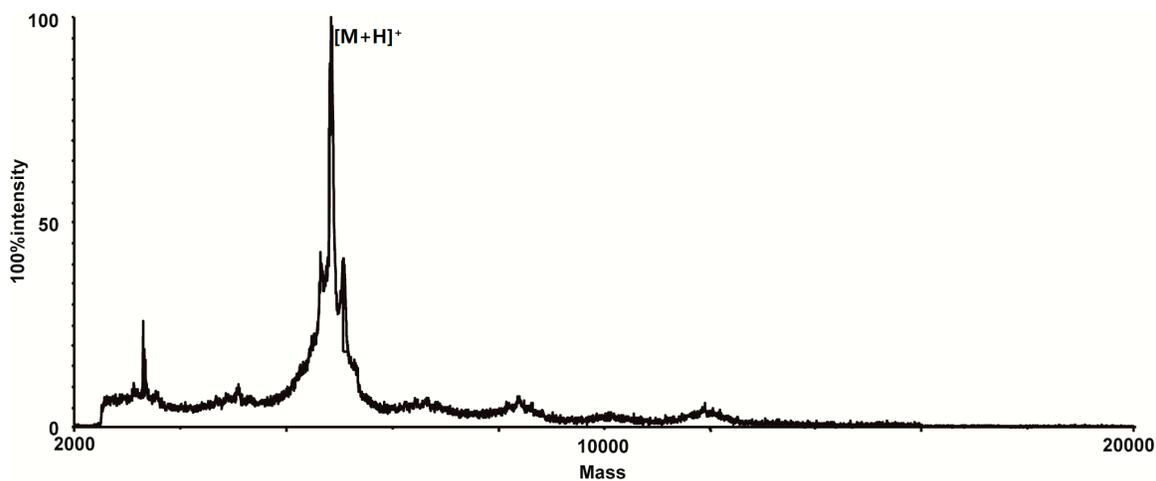


Figure S-27: Maldi-tof Mass spectrum of C^a₄L^d₈

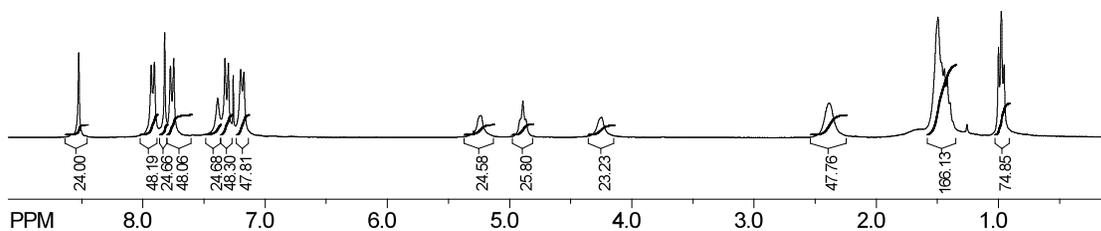


Figure S-28: ¹H NMR spectrum (300 MHz, CDCl₃, 25 °C) of C^a₆T₈

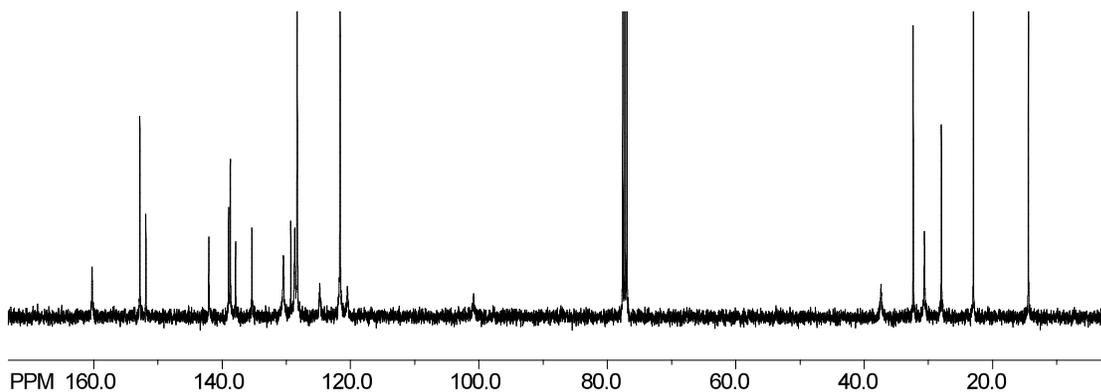


Figure S-29: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of C^a₆T₈

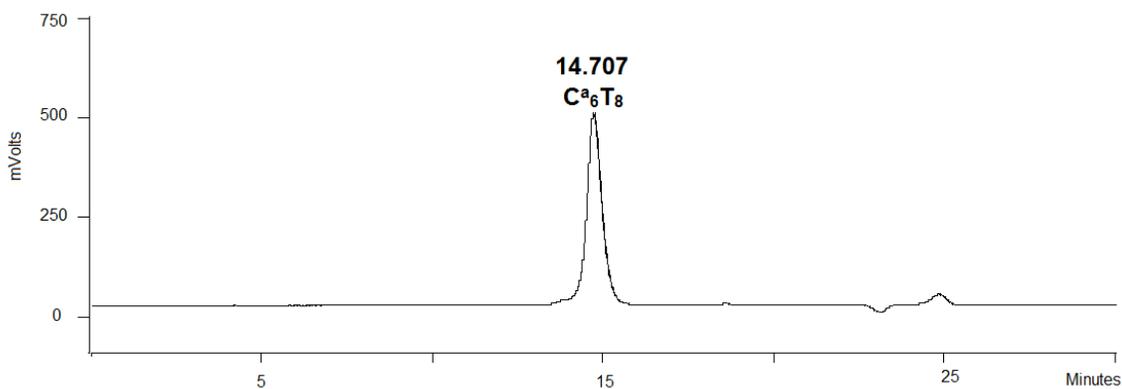


Figure S-30: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 0.9 ml/min) of C^a₆T₈

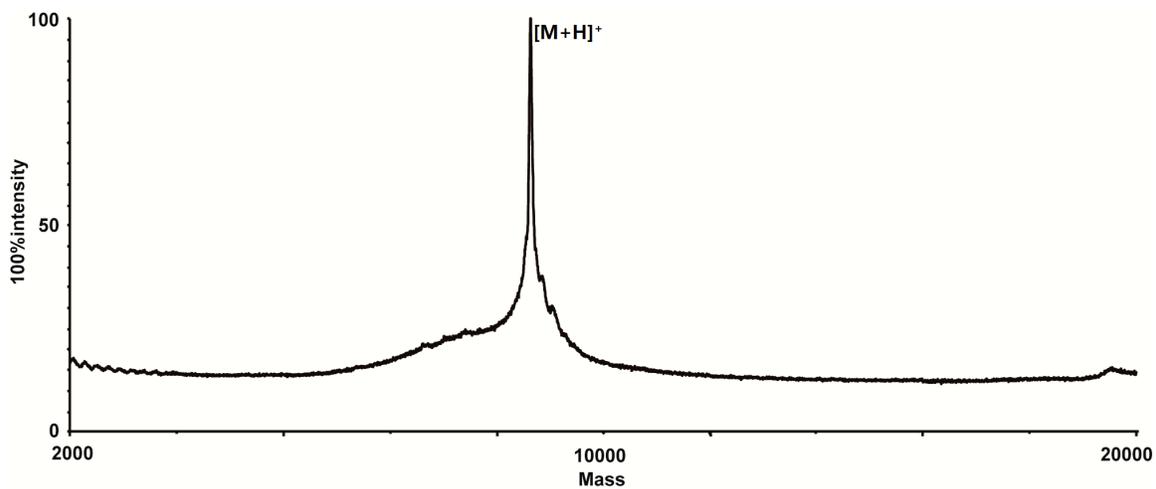


Figure S-31: Maldi-tof Mass spectrum of C^a₆T₈

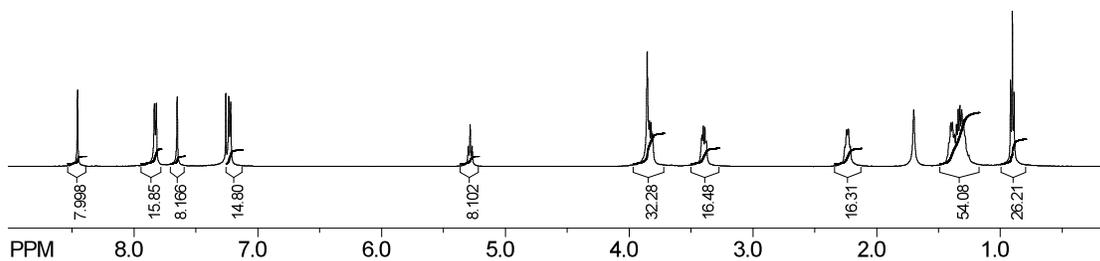


Figure S-32: ¹H NMR spectrum (300 MHz, CDCl₃, 25 °C) of **C^b₂L^a₄**

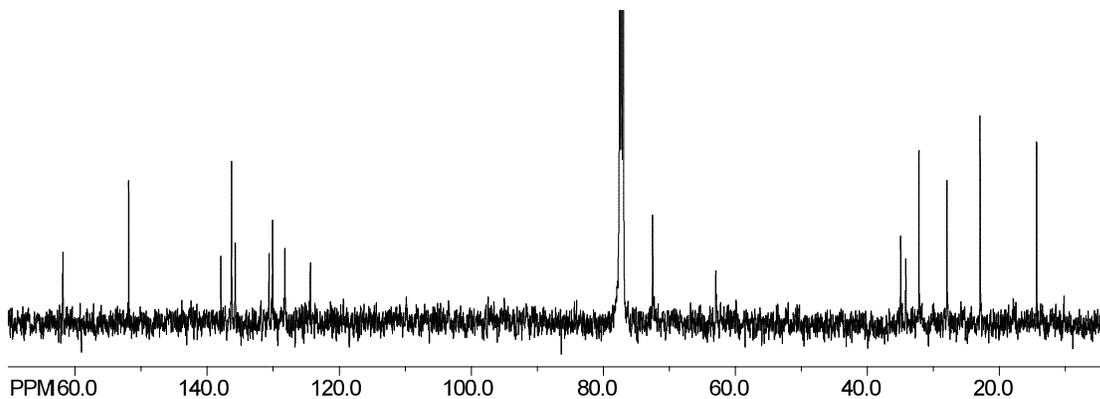


Figure S-33: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of **C^b₂L^a₄**

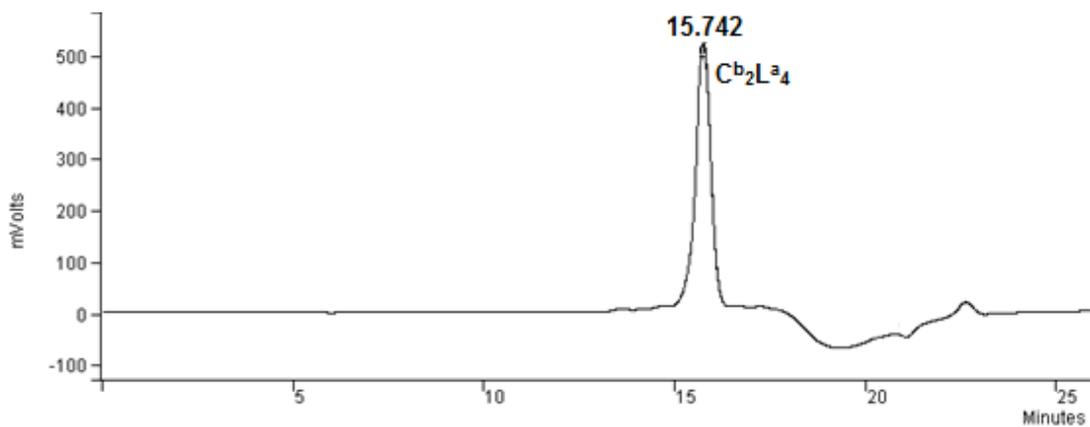


Figure S-34: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of **C^b₂L^a₄**

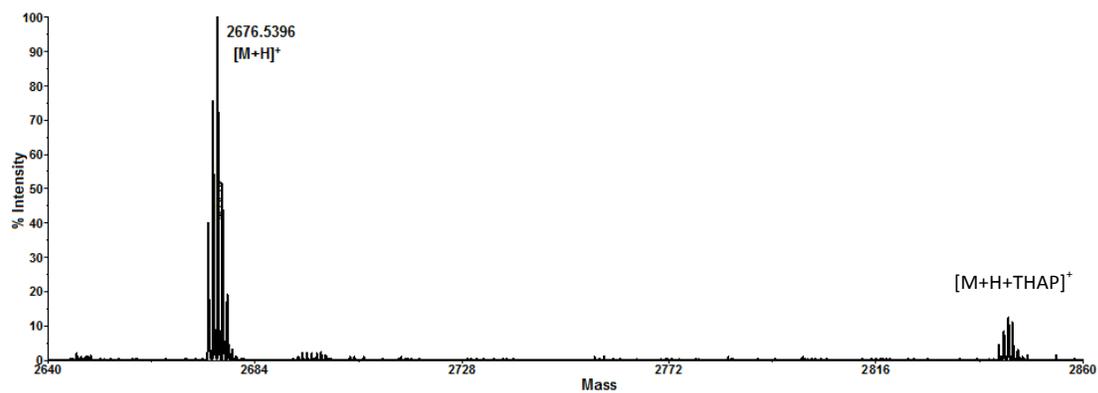


Figure S-35: Maldi-tof Mass spectrum of **C^b₂L^a₄**

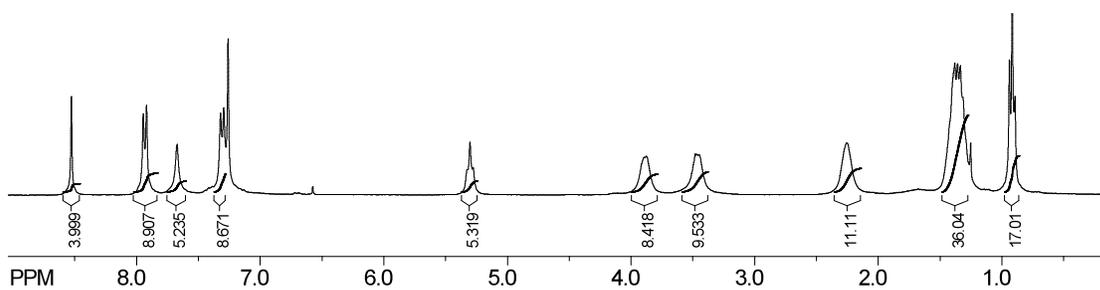


Figure S-36: ^1H NMR spectrum (300 MHz, CDCl_3 , 25 °C) of C_6L_{12}

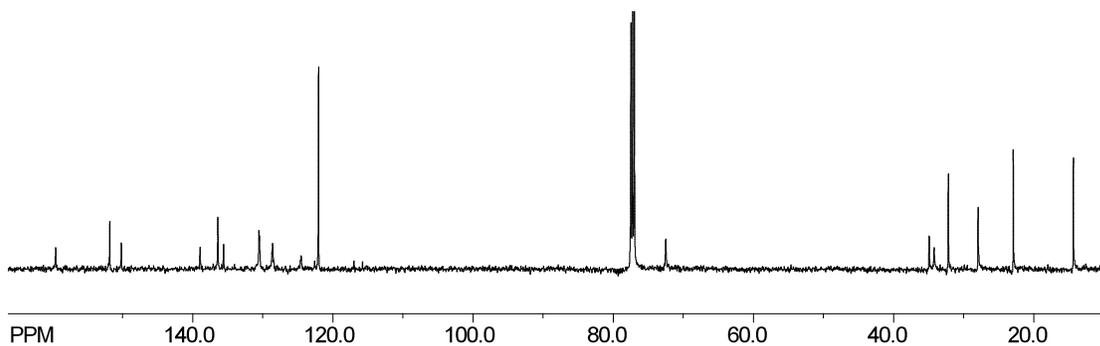


Figure S-37: ^{13}C NMR spectrum (125 MHz, CDCl_3 , 25 °C) of C_6L_{12}

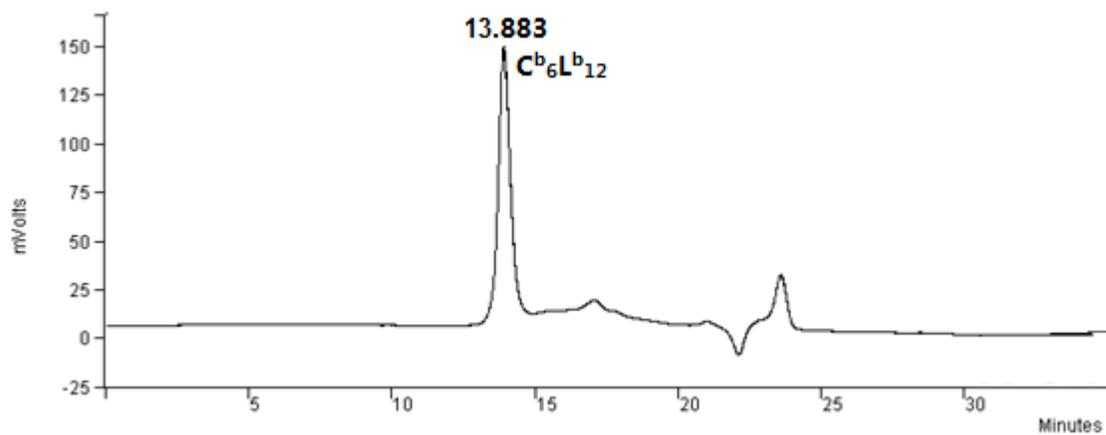


Figure S-38: GPC chromatogram (CH_2Cl_2 +1% TEA, 60 °C, 280 nm, 1 ml/min) of C_6L_{12}

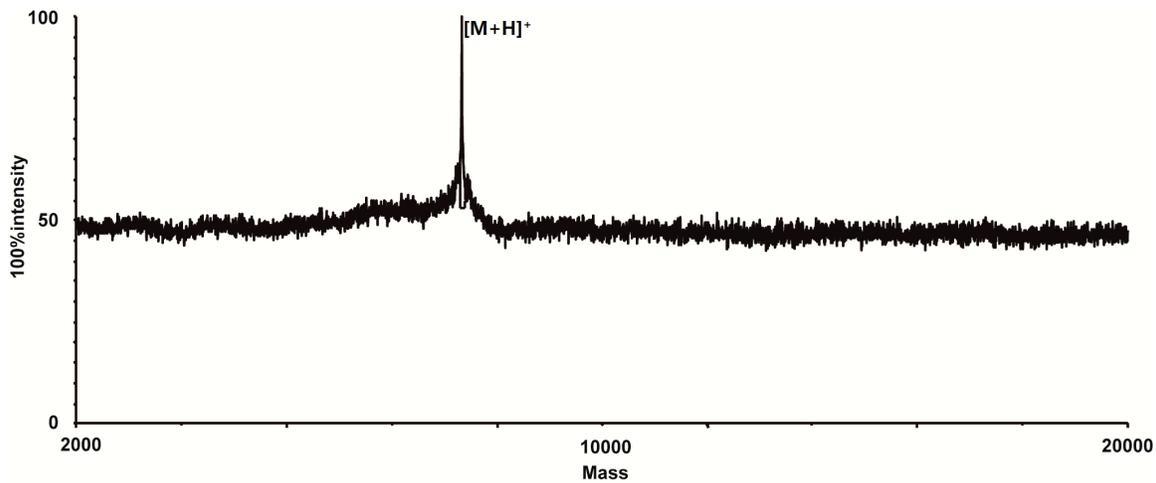


Figure S-39: Maldi-tof Mass spectrum of C_6L_{12}

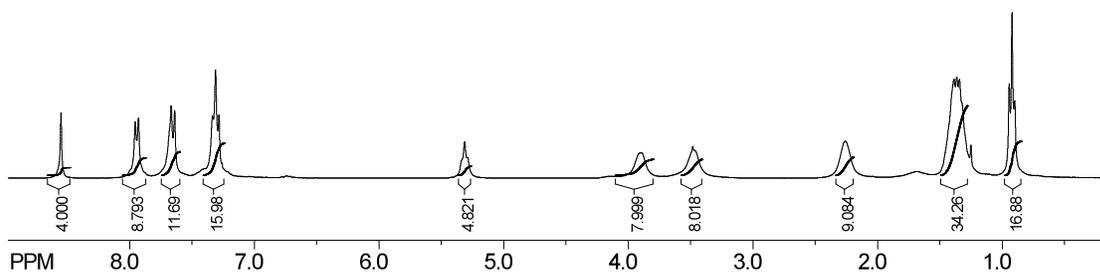


Figure S-40: ¹H NMR spectrum (300 MHz, CDCl₃, 25 °C) of C^b₆L^c₁₂

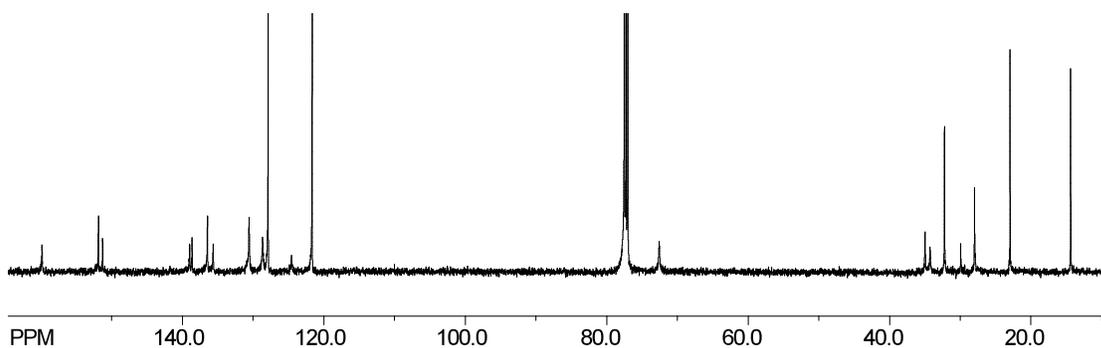


Figure S-41: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of C^b₆L^c₁₂

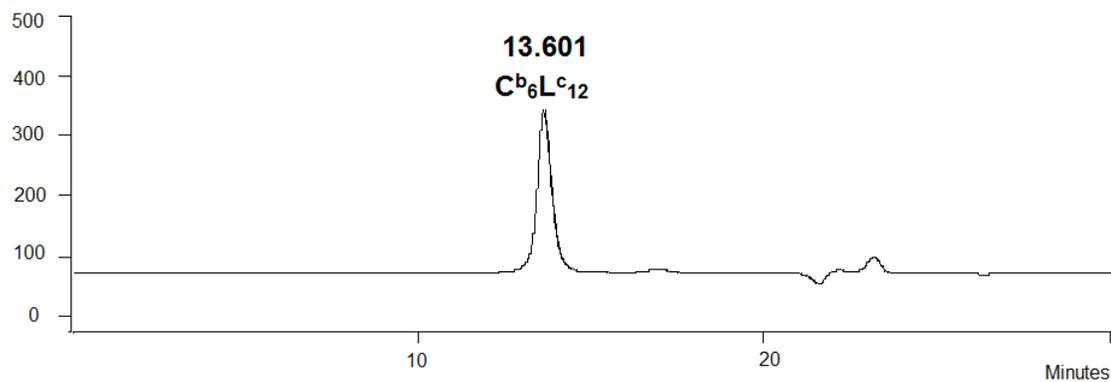


Figure S-42: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of C^b₆L^c₁₂

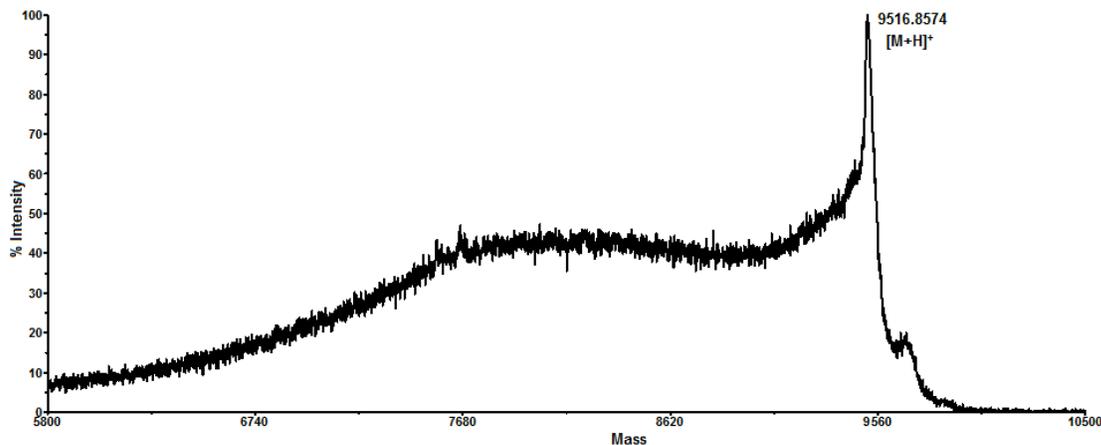


Figure S-43: Maldi-tof Mass spectrum of C^b₆L^c₁₂

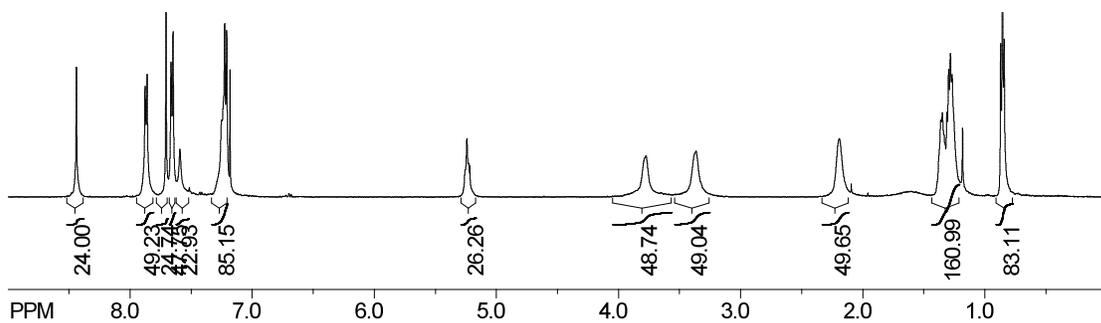


Figure S-44: ¹H NMR spectrum (300 MHz, CDCl₃, 25 °C) of C^b₆T₁₂

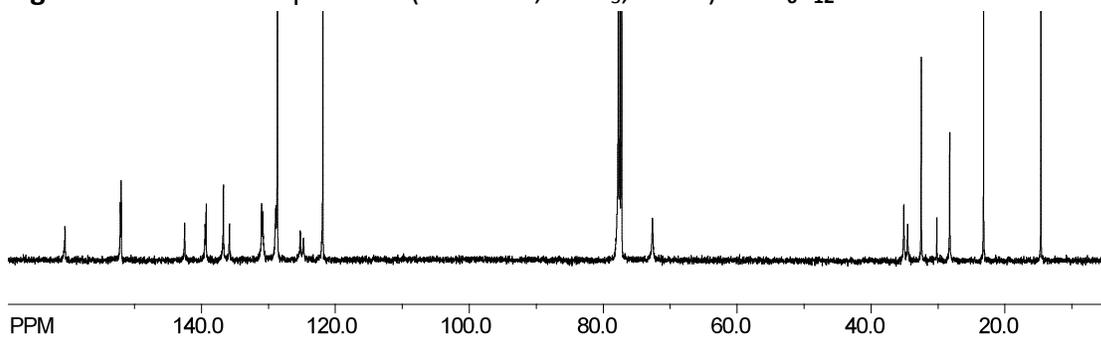


Figure S-45: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of C^b₆T₁₂

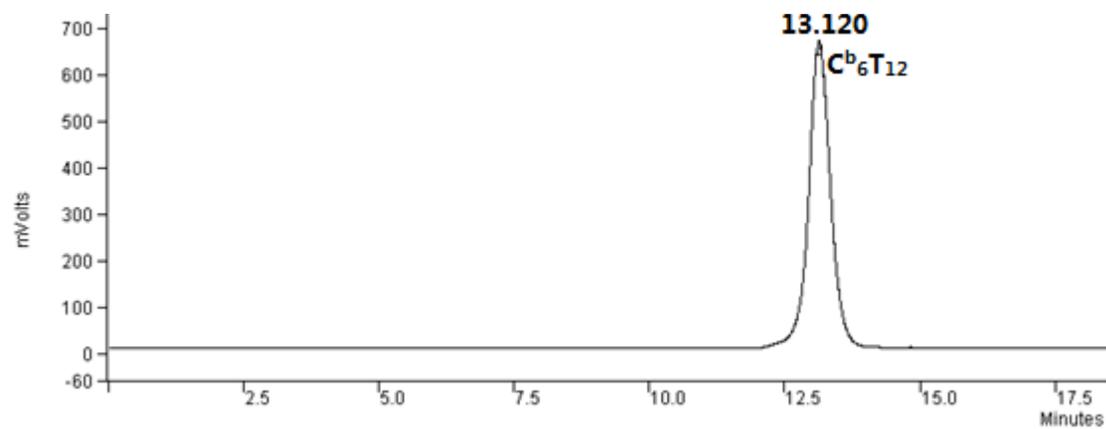


Figure S-46: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of C^b₆T₁₂

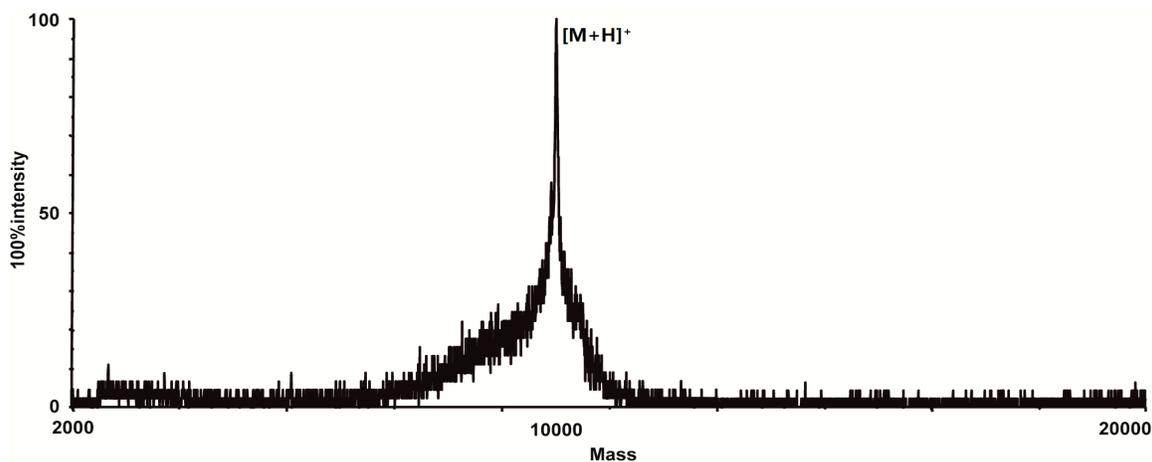


Figure S-47: Maldi-tof Mass spectrum of C^b₆T₁₂

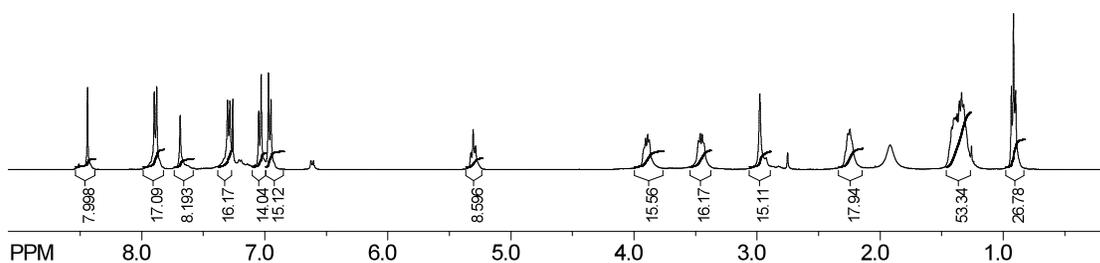


Figure S-48: ¹H NMR spectrum (500 MHz, CDCl₃, 25 °C) of $C^b_2L^d_4$

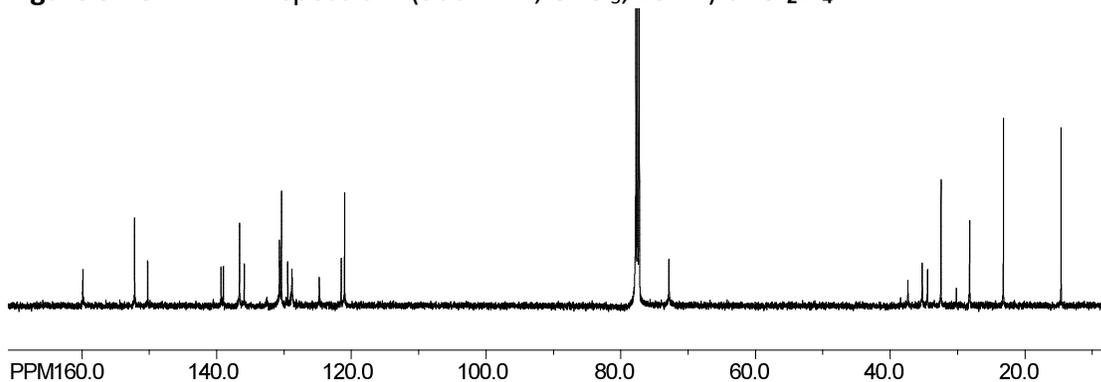


Figure S-49: ¹³C NMR spectrum (125 MHz, CDCl₃, 25 °C) of $C^b_2L^d_4$

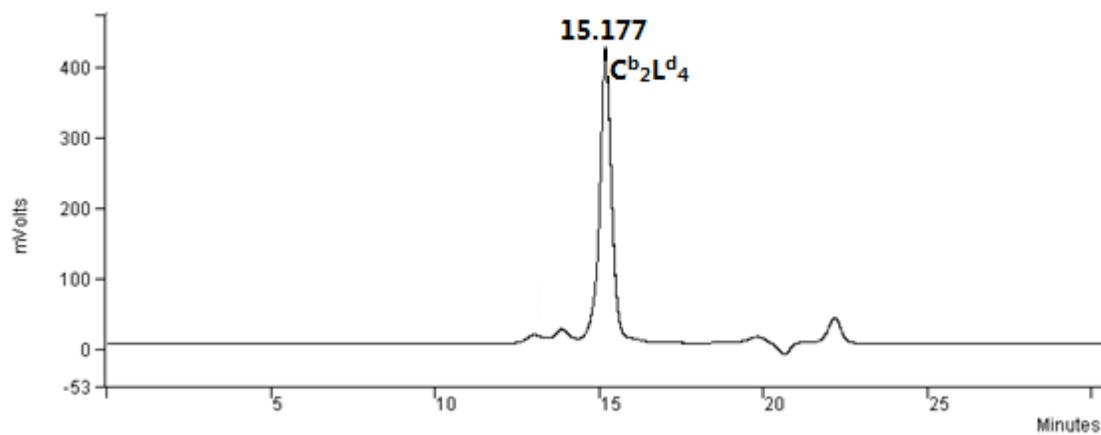


Figure S-50: GPC chromatogram (CH₂Cl₂+1% TEA, 60 °C, 280 nm, 1 ml/min) of $C^b_2L^d_4$

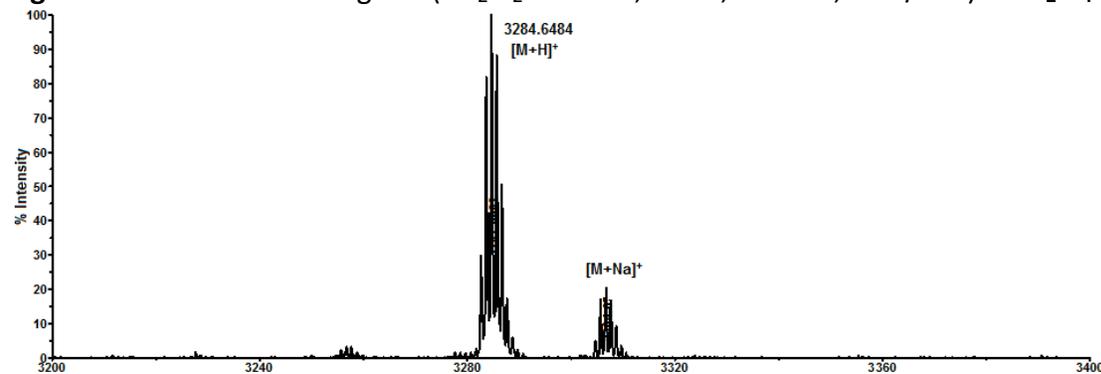


Figure S-51: Maldi-tof Mass spectrum of $C^b_2L^d_4$