Supplementary Information

Highly Enantioselective Recognition of a Wide Range of Carboxylic Acids Based on Enantioselectively Aggregation-Induced Emission

Dong-Mi Li and Yan-Song Zheng Department of Chemistry, Huazhong University of Science and Technology, Wuhan 430074, P. R. China. zyansong@hotmail.com

Materials and Methods

Materials. All reagents and solvents were chemical pure (CP) grade or analytical reagent (AR) grade and were used as received unless otherwise indicated. $CDCl_3$ for NMR solvent was stirred for 20 min over anhydrous K_2CO_3 before used.

Measurements. ¹H NMR and ¹³C NMR spectra were measured on a Bruker AV 400 spectrometer at 298 K in CDCl₃. 2D NOESY spectra were measured on a Varian NMR System 600 at 600 MHz at 298 K in CDCl₃. Fluorescent emission spectra were collected on Shimadzu RF-5301 fluorophotometer at 298 K. Field emission scanning electron microscopy (FE-SEM) images were taken on a FEI Sirion200 electron microscope operating at 5 kV or 10 kV. Mass spectrum was measured on Bruker BioApex FTMS instrument.

¹H NMR titration was carried out by addition of concentrated solution of chiral acid **4** into the solution of receptor **3** in CDCl₃. To keep constant concentration of sensor **3** and account for dilution effects during titration, the solution of **4** was prepared with the solution of **3** at its initial concentration as a solvent. The association constants were calculated by nonlinearly curve fitting in Origin 6.1 using following equation (*Chem Eur J.* 1998, 4, 845):

$$\delta_{obs} = \delta_{\mathbf{H}} + \frac{([\mathbf{H}] + [\mathbf{G}] + 1/K_a) - \sqrt{([\mathbf{H}] + [\mathbf{G}] + 1/K_a)^2 - 4[\mathbf{H}][\mathbf{G}]}}{2[\mathbf{H}]} \cdot (\delta_{com} - \delta_{\mathbf{H}})$$

 δ_{obs} : chemical shift of the proton of methine group connected with the amino group of (*R*,*R*)-**3** after enantiomer of **4** was gradually added;

 $\delta_{\rm H}$: chemical shift of the proton of methine group connected with the amino group of (*R*,*R*)-**3** without **4**;

 δ_{com} : chemical shift of the proton of methine group connected with the amino group of (*R*,*R*)-3 in complex of 3-4;

- [H]: molar concentration of (R,R)-3;
- [G]: molar concentration of enantiomer of 4 added during titration.

Syntheses of (R,R)-3 and (S,S)-3

(*R*,*R*)-**3**: synthesis of (*R*,*R*)-N-Boc-3. То the flask were added (*1R*, *2R*)-N-Boc-N'-chloroacetyl-1,2-cyclohexanediame (*1R*,*2R*)-**1** (1 3.5 mmol). g, (E)- α -(p-nitrophenyl)- β -(p-hydroxyphenyl) acrylonitrile 2 (0.77 g, 2.9 mmol), anhydrous K₂CO₃ (0.24 g, 1.75 mmol), KI (0.096 g, 0.58 mmol) and acetonitrile (60 mL) in order. The mixture was refluxed for about 3 h until one of reactants disappeared (monitored by TLC, dichloromethane : methanol V / V 15 : 1). The reaction mixture was cooled to room temperature and the resultant yellow precipitates were collected by filtering. The solid was dissolved into 1,2-dichloroethane and washed with water until the organic phase had pH 6. Upon dried over anhydrous Na_2SO_4 and filtered, the organic phase was evaporated to give a vellow solid. Remove of the Boc group of N-Boc-3. The obtained yellow solid was dissolved in dichloromethane (10 mL) and the solution was cooled to 0 °C. Trifluoroacetic acid (8.56 mL, 115.2 mmol) was dropped into the solution over 0.5 h. The solution was continued to stir for about 20 minutes at 0 °C (monitored by TLC, dichloromethane : methanol V / V 15 : 1). After washed two times with water, the solution was adjusted to pH7-8 by 20 % (weight) of Na₂CO₃ aqueous solution and extracted with chloroform. The combined organic phase was dried over anhydrous Na₂SO₄, filtered and evaporated to afford a yellow solid (1.02 g, 85 %). Mp 125 – 127 °C; $[\alpha]^{20}$ – 32.5 (c, 1.0, THF); IR (KBr) 3364, 3269, 3096, 2928, 2856, 2213, 1660, 1588, 1564, 1514, 1443, 1430, 1378, 1341; ¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, J = 8.8 Hz, 2H), 7.97 (d, J = 8.8 Hz, 2H), 7.82 (d, J = 8.8 Hz, 2H), 7.62 (s, 1H), 7.06 (d, J = 8.8 Hz, 2H); 6.44 (d, J = 8.8 Hz, 1H), 4.63, 4.58 (2d, J = 14.8 Hz, 2H), 3.69 – 3.56 (m, 1H), 2.54 -2.43 (m, 1H), 2.05 - 1.98 (m, 2H), 1.75 (d, J = 10.4 Hz, 2H), 1.60 - 1.15 (m, 6H); ^{13}C NMR (100MHz, CDCl₃) & 167.4, 159.6, 147.7, 144.4, 140.7, 132.0, 127.1, 126.5, 124.4, 117.4, 115.3, 107.5, 67.4, 56.0, 55.3, 35.6, 32.4, 25.0; ESI⁺ MS *m*/*z* calcd for C₂₃H₂₄N₄O₄ 420 [M], Found 421.1 [(M+1)]⁺.

(*S*,*S*)-**3**: synthesized according to the same procedure for (*S*,*S*)-**3**. Mp 124 – 126 °C; $[\alpha]^{20}_{D}$ –33.6 (*c*, 1.0, THF); IR (KBr) 3366, 3270, 3099, 2933, 2859, 2215, 1661, 1588, 1565, 1515, 1446, 1430, 1342 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, *J* = 8.8 Hz, 2H), 7.97 (d, *J* = 8.8 Hz, 2H), 7.82 (d, *J* = 8.8 Hz, 2H), 7.62 (s, 1H), 7.06 (d, *J* = 8.8 Hz, 2H); 6.43 (d, *J* = 8.8 Hz, 1H), 4.63, 4.58 (2d, *J* = 14.8 Hz, 2H), 3.68 – 3.57 (m, 1H), 2.52 – 2.42 (m, 1H), 2.05 – 1.95 (m, 2H), 1.75 (d, *J* = 11.2 Hz, 2H), 1.45 – 1.15 (m, 6H); ¹³C NMR (100MHz, CDCl₃) δ 167.4, 159.6, 147.7, 144.4, 140.7, 132.0, 127.1, 126.5, 124.4, 117.5, 115.3, 107.5, 67.4, 56.0, 55.3, 35.6, 32.4, 25.0; ESI⁺ MS *m*/*z* calcd for C₂₃H₂₄N₄O₄ 420 [M], Found 421.1 [(M+1)]⁺.

Supporting Figures



Figure S1. ¹H NMR spectrum of (R,R)-3 in CDCl₃.



Figure S2. ¹³C NMR spectrum of (R,R)-3 in CDCl₃.



Figure S3. IR spectrum of (R,R)-3.

Electronic Supplementary Material (ESI) for Chemical Communications This journal is The Royal Society of Chemistry 2011



Figure S4. MS spectrum of (*R*,*R*)**-3**.



Figure S5. ¹H NMR spectrum of (*S*,*S*)-**3** in CDCl₃.



Figure S6. ¹³C NMR spectrum of (*S*,*S*)-3 in CDCl₃.



Figure S7. IR spectrum of (*S*,*S*)**-3**.

Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2011



Figure S8. MS spectrum of (*S*,*S*)**-3**.



Figure S9. Changes of the fluorescence spectra of (R,R)-**3** (5.0×10^{-4} M) with *n*-hexane added in 1,2-dichloroethane. $\lambda_{ex} = 320$ nm, ex/em slits = 5/3 nm. Inset: curve of the fluorescence intensity vs *n*-hexane percentage measured at 550 nm.



Figure S10. Photos of a mixture of (R,R-3) and enantiomers of acids in solvent under daylight.





Figure S11. The fluorescence spectra of the mixture of acid enantiomer and (*R*,*R*)-**3** in solvent(s). C) 5×10-3 M in ethanol. D) 5×10-3 M in 1,2-dichloroethane and n-hexane 1.2:0.8. E) 2.0×10-3 M in 1,2-dichloroethane and n-hexane 2.0:2.5. F) 2×10-3 M in 1,2-dichloroethane. G) 5×10^{-3} M in 1,2-dichloroethane. H, I) 2.5×10⁻³ M in 1,2-dichloroethane. J) [(*R*,*R*)-**3**] = 2 [**13**] = 5×10^{-3} M in ethanol and water 1:1. K) 2.2×10⁻³ M in ethanol and water 1:3.5. L) 2.5×10⁻³ M in ethanol and water 1:3.5. M) 3.3×10^{-3} M in ethanol and water 1:1. N) 2.8×10^{-3} M in ethanol and water 1:2.5. O) 5×10^{-3} M in 1,2-dichloroethane and *n*-hexane 1.7 : 1.1. P) 2.2×10^{-3} M in 1,2-dichloroethane and ethanol 4:0.5. Q) 5×10^{-3} M in 1,2-dichloroethane and *n*-hexane 1.2 : 0.8. R) 5.3×10^{-3} M in 1,2-dichloroethane and *n*-hexane 1.0 : 0.9. [(*R*,*R*)-**3**] = [acid] unless indicated.



(c)

Figure S12. (a) and (b) FE-SEM images of precipitates from the mixture of (R,R)-3 and (S)-4 in 1,2-dichloroethane. (c) FE-SEM images of the solid obtained by evaporation of solution of (R,R)-3 and (R)-4 in 1,2-dichloroethane.



Figure S13. Job plot for ¹H NMR titration of (R,R)-3 ([(R,R)-3] = 0.005 M) with (R)-4 in CDCl₃.



Figure S14. Chemical shift of (R,R)-3 ([(R,R)-3] = 0.005 M) with concentration of (R)-4 added. The red curve is the result by fitting.



Figure S15. Job plot for ¹H NMR titration of (R,R)-3 ([(R,R)-3] = 0.005 M) with (S)-4 in CDCl₃.



Figure S16. Chemical shift of (R,R)-3 ([(R,R)-3] = 0.005 M) with concentration of (S)-4 added. The red curve is the result by fitting.



Figure S17. ¹H NMR spectrum of (*S*)-4 (a); of (*R*,*R*)-3 (b); (c) of a 1:1 mixture of (*R*)-4 and (*R*,*R*)-3 (c); of a 1:1 mixture of (*S*)-4 and (*R*,*R*)-3 (d). [(R,R)-3] = [(R)-4] = [(S)-4] = 5 mM in CDCl₃. And the difference of chemical shift of the mixture of (*R*,*R*)-3 and enantiomers of 4 vs that of free ones ($\Delta\delta$ ppm) that was put in the table.



Figure S18. 2D NOESY spectrum of a 1:1 mixture of (*S*)-4 and (*R*,*R*)-3 in CDCl₃. It was measured in 5 h at 25 °C. [(R,R)-3] = [(S)-4] = 10 mM.



Figure S19. 2D NOESY spectrum of a 1:1 mixture of (*R*)-4 and (*R*,*R*)-3 in CDCl₃. It was measured in 5 h at 25 °C. [(R,R)-3] = [(R)-4] = 10 mM.



Figure S20. (a) Portion of the 2D NOESY spectrum of a 1:1 mixture of (R,R)-3 and (S)-4, and (b) that of (R)-4.