

Supplementary Information

Photo-driven anti-Markovnikov alkyne hydration in self-assembled hollow complexes

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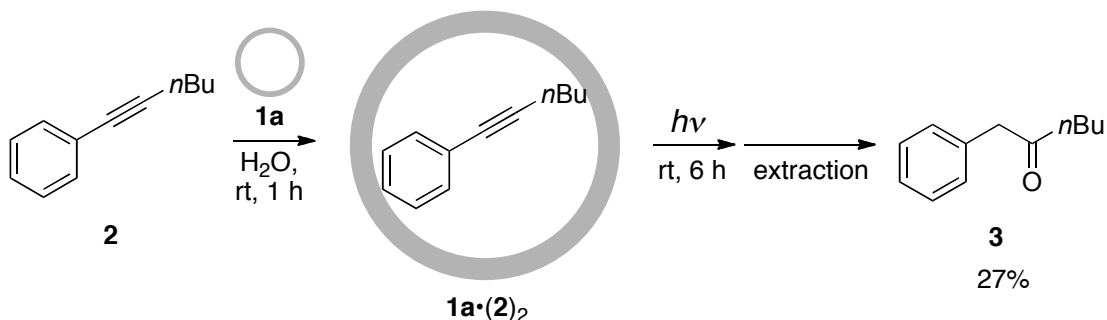
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 - physical data.

Materials and Instrumentations: ^1H NMR spectra were recorded on a Bruker DRX-500 (500 MHz) or a Bruker Avance 500 equipped with CP-TCI cryoprobe at 300 K. TMS or $\text{CHCl}_2\text{CHCl}_2$ (CDCl_3 solution) in a capillary served as an external standard ($\delta = 0$ ppm for TMS or $\delta = 5.90$ ppm for $\text{CHCl}_2\text{CHCl}_2$). GC-MS spectra were obtained on an Agilent 5973 inert Mass selective Detector equipped with a 6890N Network GC system and an EI source. Photoirradiation was carried out with a SEN LIGHTS CORP. HL400JH-3 400W high-pressure mercury lamp. Solvents and reagents were purchased from TCI Co., Ltd., WAKO Pure Chemical Industries Ltd., and Sigma-Aldrich Co. Deuterated H_2O was acquired from Cambridge Isotope Laboratories, Inc. and used as supplied for the complexation reactions and NMR measurements.

General procedure for photo-hydration of alkynes within cage 1: Arylalkynes (0.80 mmol) was suspended in a water solution (2.0 mL) of cage **1** (4.0×10^{-2} mmol; 20 mM) and the solution was stirred for 1 h at room temperature. After removing excess substrates by filtration, the formation of the host-guest complex was confirmed by ^1H NMR. The solution was degassed by freeze-pump-thaw method and filled with Ar, then irradiated with high-pressure mercury lamp (400 W) for 6 h at room temperature. After extraction with CDCl_3 , the yields were determined by ^1H NMR analysis of the crude mixture. *The product structures were identified by ^1H NMR, GC-MS and comparison with authentic samples.*

Photo-hydration of 1-phenyl-1-hexyne (**2**) within cage **1a**.



Physical data of 1a·(2)₂: ¹H NMR (500 MHz, D₂O, 300 K): $d = -0.98$ (t, $J = 7.5$ Hz, 3H, **2**), 0.22 (sextet, $J = 7.0$ Hz, 2H, CH₂, **2**), 0.37 (quintet, $J = 6.0$ Hz, 2H, CH₂, **2**), 1.21 (t, $J = 6.0$ Hz, 2H, CH₂, **2**), 2.84 (s, 72H, CH₃, **1a**), 3.22 (s, 24H, CH₂, **1a**), 5.37 (d, $J = 7.5$ Hz, 2H, CH, **2**), 6.03 (t, $J = 7.0$ Hz, 2H, CH, **2**), 6.95 (t, $J = 7.0$ Hz, 1H, CH, **2**), 8.81 (d, $J = 6.0$ Hz, 24H, PyH_a, **1a**), 9.41 (d, $J = 6.0$ Hz, 24H, PyH_b, **1a**).

Physical data of 3: ¹H NMR (500 MHz, CDCl₃, 300 K): $d = 0.86$ (t, $J = 7.5$ Hz, 3H, CH₃), 1.26 (sextet, $J = 7.5$ Hz, 2H, CH₂), 1.53 (quintet, $J = 8.0$ Hz, 2H, CH₂), 2.44 (t, $J = 7.5$ Hz, 2H, CH₂), 3.68 (s, 2H, CH₂), 7.20 (d, $J = 7.0$ Hz, 2H, CH), 7.26 (br, 1H, CH), 7.33 (t, $J = 7.5$ Hz, 2H, CH).
MS (EI) m/z : 176 [M]⁺, 91 [M-C₄H₉CO]⁺, 85 [M-C₆H₅CH₂]⁺.

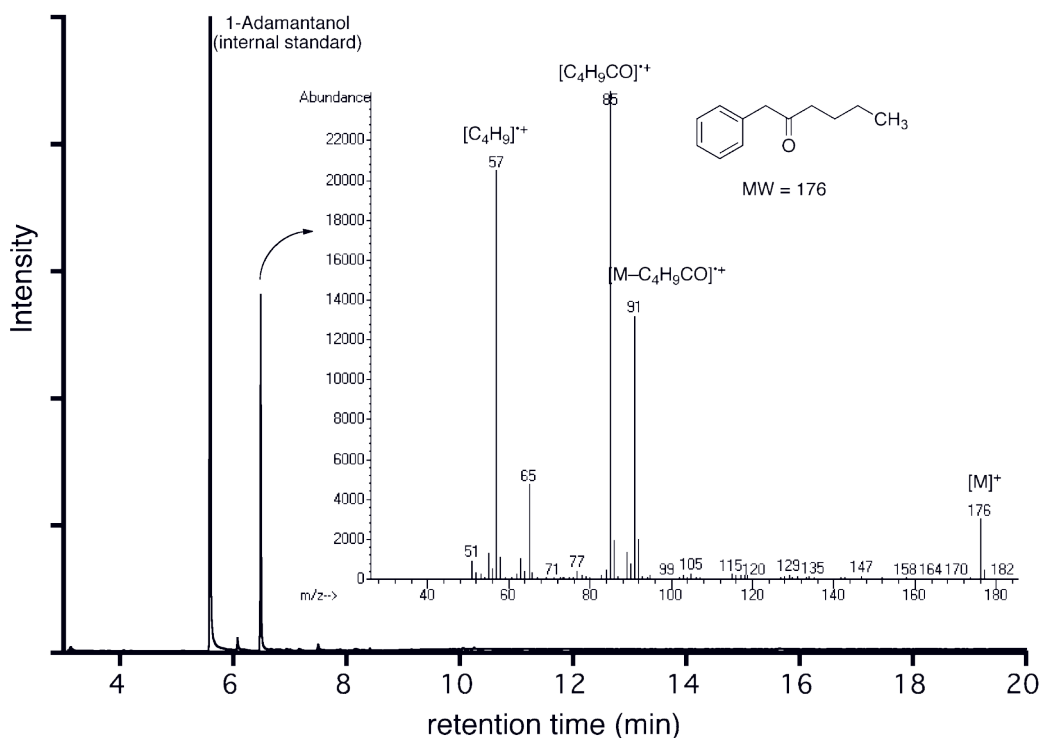


Fig. S1. GC and EI-MS spectra of **3** extracted from the reaction mixture (1-adamantanol: internal standard for GC-MS analysis).

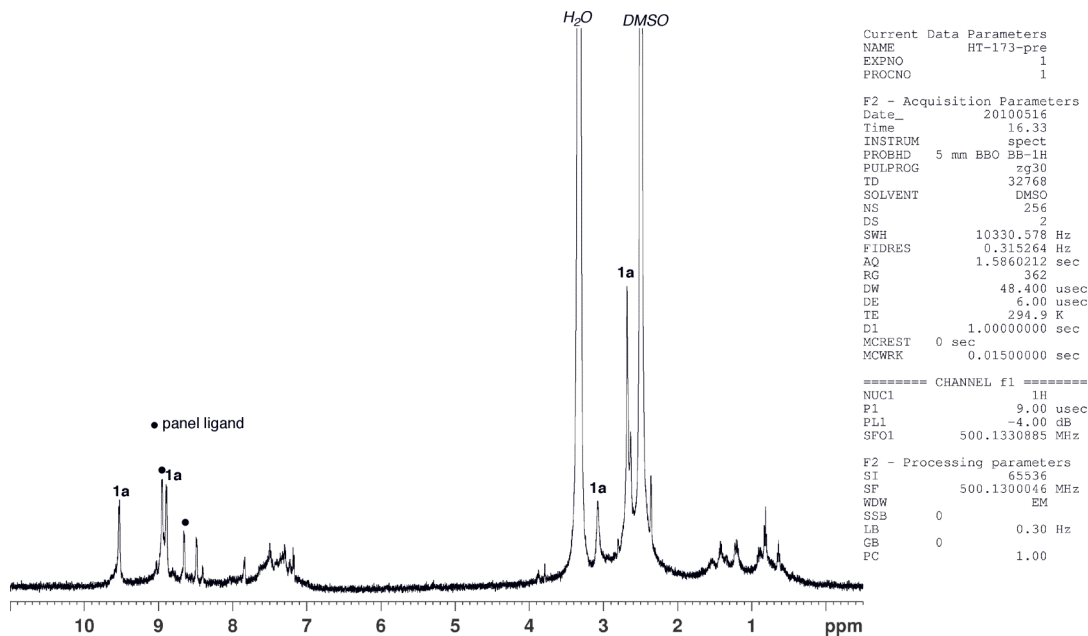
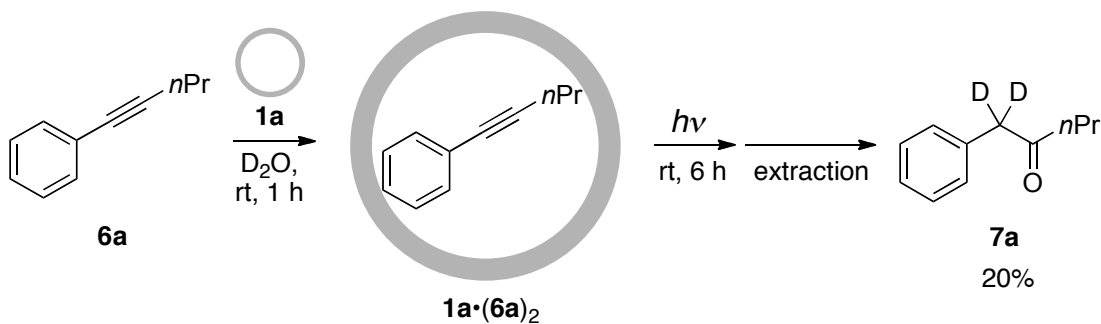


Fig. S2. ^1H NMR spectrum of the precipitated by-products after UV-light irradiation to inclusion complex $1\mathbf{a}\cdot(2)_2$. The precipitate was redissolved in $\text{DMSO-}d_6$. The broad signals could not be identified, but the Markovnikov adduct was not contained in the spectrum. Panel ligand of cage $1\mathbf{a}$: 2,4,6-tri(4-pyridyl)-1,3,5-triazine.

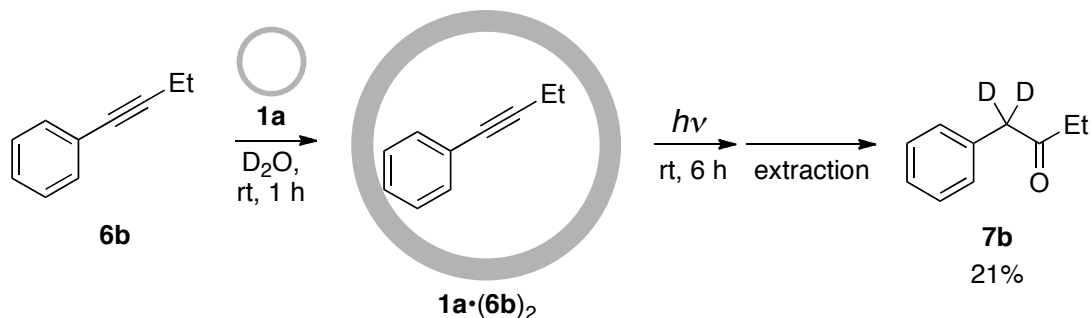
Photo-hydration of 1-phenyl-1-pentyne ($6\mathbf{a}$) within cage $1\mathbf{a}$.



Physical data of $1\mathbf{a}\cdot(6\mathbf{a})_2$: ^1H NMR (500 MHz, D_2O , 300 K): δ = -0.56 (t, J = 8.0 Hz, 3H, $6\mathbf{a}$), 0.44 (br, 2H, CH_2 , $6\mathbf{a}$), 1.22 (br, 2H, CH_2 , $6\mathbf{a}$), 2.81 (s, 72H, CH_3 , $1\mathbf{a}$), 3.19 (s, 24H, CH_2 , $1\mathbf{a}$), 5.12 (d, J = 7.0 Hz, 2H, CH , $6\mathbf{a}$), 5.84 (t, J = 7.0 Hz, 2H, CH , $6\mathbf{a}$), 6.66 (br, 1H, CH , $6\mathbf{a}$), 8.76 (d, J = 4.5 Hz, 24H, PyH_a , $1\mathbf{a}$), 9.36 (d, J = 4.0 Hz, 24H, PyH_b , $1\mathbf{a}$).

Physical data of $7\mathbf{a}$: ^1H NMR (500 MHz, CDCl_3 , 300 K): δ = 0.86 (t, J = 7.5 Hz, 3H, CH_3), 1.57 (sextet, J = 7.0 Hz, 2H, CH_2), 2.42 (t, J = 7.5 Hz, 2H, CH_2), 7.21 (d, J = 7.0 Hz, 2H, CH), 7.26 (br, 1H, CH), 7.33 (t, J = 7.5 Hz, 2H, CH). MS (EI) m/z : 164 $[\text{M}]^+$, 93 $[\text{M}-\text{C}_3\text{H}_7\text{CO}]^+$, 71 $[\text{M}-\text{C}_6\text{H}_5\text{CD}_2]^+$.

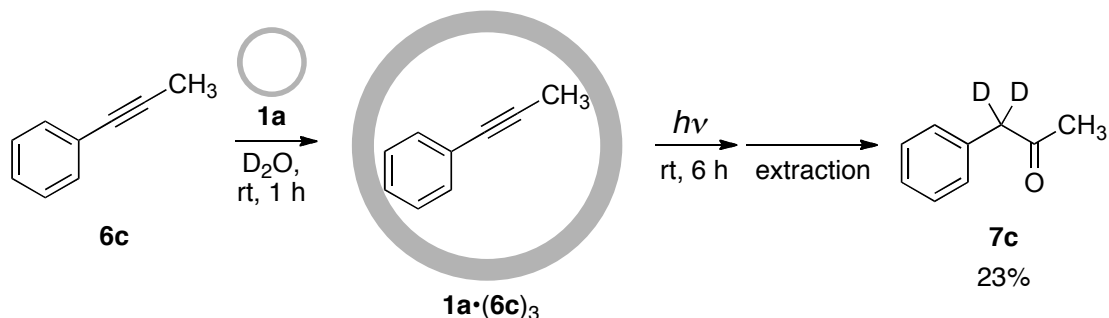
Photo-hydration of 1-phenyl-1-butyne (**6b**) within cage **1a**.



Physical data of 1a·(6b)₂: ¹H NMR (500 MHz, D₂O, 300 K): *d* = -0.22 (t, *J* = 7.5 Hz, 3H, **6b**), 1.09 (q, *J* = 7.5 Hz, 2H, CH₂, **6b**), 2.76 (s, 72H, CH₃, **1a**), 3.16 (s, 24H, CH₂, **1a**), 5.06 (d, *J* = 7.5 Hz, 2H, CH, **6b**), 5.84 (t, *J* = 7.5 Hz, 2H, CH, **6b**), 6.18 (t, *J* = 7.5 Hz, 1H, CH, **6b**), 8.73 (d, *J* = 5.5 Hz, 24H, PyH_a, **1a**), 9.35 (d, *J* = 5.0 Hz, 24H, PyH_b, **1a**).

Physical data of 7b: ¹H NMR (500 MHz, CDCl₃, 300 K): *d* = 1.02 (t, *J* = 7.5 Hz, 3H, CH₃), 2.47 (quartet, *J* = 7.5 Hz, 2H, CH₂), 2.42 (t, *J* = 7.5 Hz, 2H, CH₂), 7.21 (d, *J* = 7.5 Hz, 2H, CH), 7.26 (br, 1H, CH), 7.33 (t, *J* = 7.0 Hz, 2H, CH). MS (EI) *m/z*: 159 [M]⁺, 93 [M-CH₃CO]⁺, 57 [M-C₆H₅CD₂]⁺.

Photo-hydration of 1-phenyl-1-pentyne (**6c**) within cage **1a**.



Physical data of 1a·(6c)₃: ¹H NMR (500 MHz, D₂O, 300 K): *d* = 1.05 (s, 3H, CH₃, **6c**), 2.81 (s, 72H, CH₃, **1a**), 3.20 (s, 24H, CH₂, **1a**), 5.18 (d, *J* = 8.0 Hz, 2H, CH, **6c**), 5.67 (t, *J* = 7.5 Hz, 2H, CH, **6c**), 5.84 (t, *J* = 7.5 Hz, 1H, CH, **6c**), 8.75 (d, *J* = 4.0 Hz, 24H, PyH_a, **1a**), 9.41 (d, *J* = 4.0 Hz, 24H, PyH_b, **1a**).

Physical data of 7c: ¹H NMR (500 MHz, CDCl₃, 300 K): *d* = 2.15 (s, 3H, CH₃), 7.21 (d, *J* = 7.5 Hz, 2H, CH), 7.26 (br, 1H, CH), 7.34 (t, *J* = 7.0 Hz, 2H, CH). MS (EI) *m/z*: 136 [M]⁺, 93 [M-CH₃CO]⁺.

Photochemical reactions of benzyl ketone **3** (without cage **1a**).

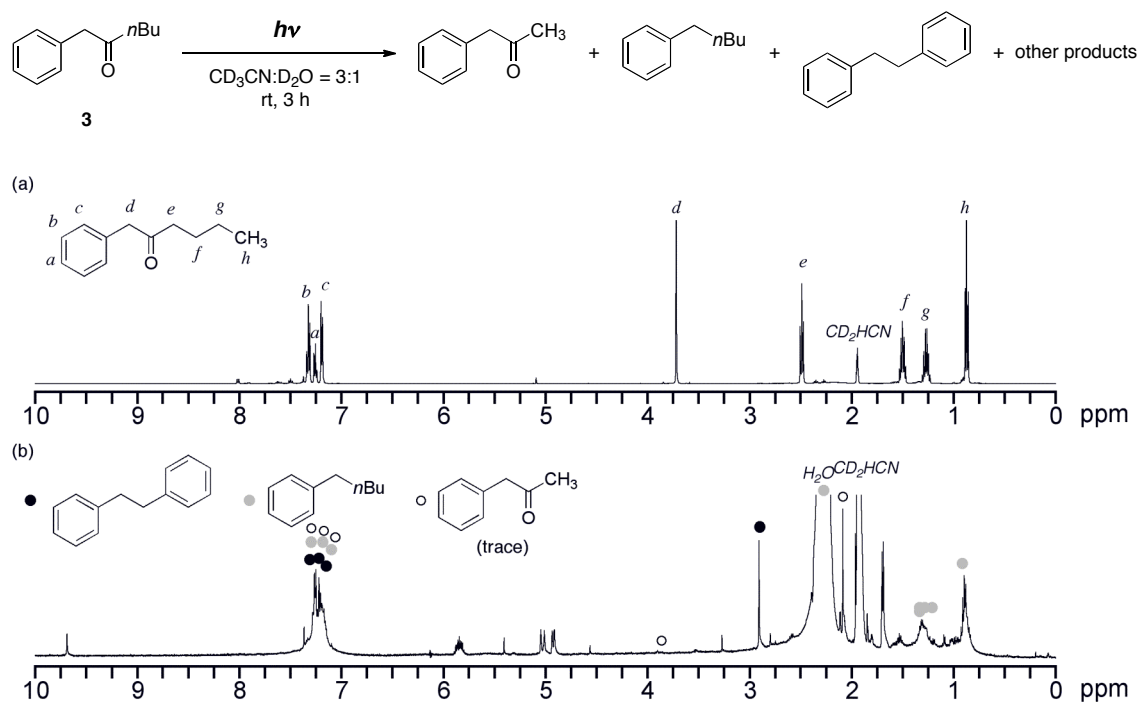
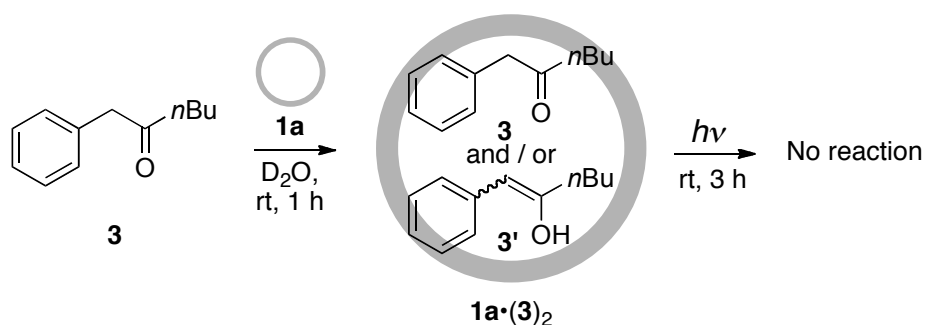


Fig. S3. ¹H NMR spectra (CD₃CN) of (a) benzyl ketone **3** and (b) the photoproducts. The photoproducts were identified by NMR and GC-MS.

Photochemical reactions of benzyl ketone **3** within cage **1a**.



Physical data of 1a•(3)₂: ¹H NMR (500 MHz, D₂O, 300 K): *d* = -1.13 (sextet, *J* = 6.5 Hz, 2H, CH₂, **3**), -0.91 (t, *J* = 6.5 Hz, 3H, CH₃, **3**), -0.73 (quintet, *J* = 6.5 Hz, 2H, CH₂, **3**), 0.38 (t, *J* = 7.5 Hz, 3H, CH₃, **3'**), 0.58 (t, *J* = 6.5 Hz, 2H, CH₂, **3**), 0.89 (sextet, *J* = 7.5 Hz, 2H, CH₂, **3'**), 1.15 (quintet, *J* = 7.5 Hz, 2H, CH₂, **3'**), 1.67 (s, 2H, CH₂, **3**), 2.00 (t, *J* = 7.5 Hz, 2H, CH₂, **3'**), 2.88 (s, 72H, CH₃, **1a**), 3.28 (s, 24H, CH₂, **1a**), 5.25 (d, *J* = 7.0 Hz, 2H, CH, **3**), 5.47 (t, *J* = 7.0 Hz, 2H, CH, **3**), 5.74 (t, *J* = 6.0 Hz, 1H, CH, **3**), 5.96 (t, *J* = 7.5 Hz, 2H, CH, **3'**), 6.18 (t, *J* = 7.0 Hz, 1H, CH, **3'**), 6.26 (d, *J* = 7.5 Hz, 2H, CH, **3'**), 8.87 (d, *J* = 5.0 Hz, 24H, PyH_a, **1a**), 9.50 (d, *J* = 5.0 Hz, 24H, PyH_b, **1a**).

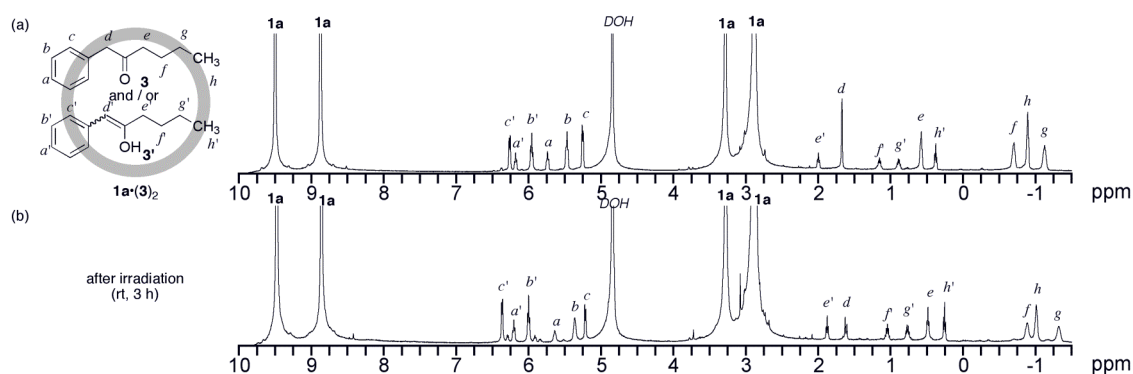
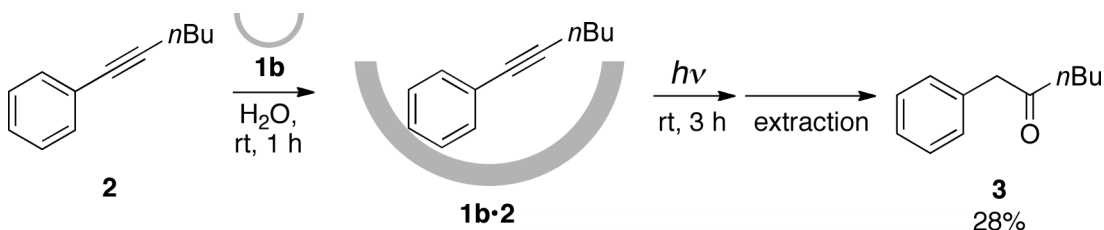


Fig. S4. ^1H NMR spectra (D_2O) of $1\mathbf{a}\cdot(\mathbf{3})_2$ (a) before and (b) after photoirradiation. Enol tautomer ($\mathbf{3}'$) of benzyl ketone $\mathbf{3}$ also existed in cage $1\mathbf{a}$. Although the ratio of $\mathbf{3}$ and $\mathbf{3}'$ changed and the benzylic protons (H_d) were deuterated during photoirradiation, the total amount of $\mathbf{3}$ and $\mathbf{3}'$ remained unchanged.

Photo-hydration of $\mathbf{2}$ within cage $1\mathbf{b}$.



Physical data of $1\mathbf{b}\cdot\mathbf{2}$: ^1H NMR (500 MHz, D_2O , 300 K): $d = -0.20$ (br, 5H, CH_2 and CH_3 , $\mathbf{2}$), -0.03 (br, 2H, CH_2 , $\mathbf{2}$), 0.46 (br, 2H, CH_2 , $\mathbf{2}$), $2.99\text{--}3.10$ (m, 24H, CH_2 , $\mathbf{1b}$), 5.15 (br, 2H, CH , $\mathbf{2}$), 5.42 (br, 2H, CH , $\mathbf{2}$), 5.69 (br, 1H, CH , $\mathbf{2}$), 7.93 (t, $J = 7.0$ Hz, 8H, PyH , $\mathbf{1b}$), 7.97 (t, $J = 7.0$ Hz, 4H, PyH , $\mathbf{1b}$), 9.05 (br, 8H, PyH , $\mathbf{1b}$), 9.16 (d, $J = 8.0$ Hz, 4H, PyH , $\mathbf{1b}$), 9.24 (d, 4H, $J = 5.5$ Hz PyH , $\mathbf{1b}$), 9.33 (d, $J = 5.5$ Hz, 8H, PyH , $\mathbf{1b}$), 9.71 (s, 4H, PyH , $\mathbf{1b}$), 10.56 (s, 8H, PyH , $\mathbf{1b}$).

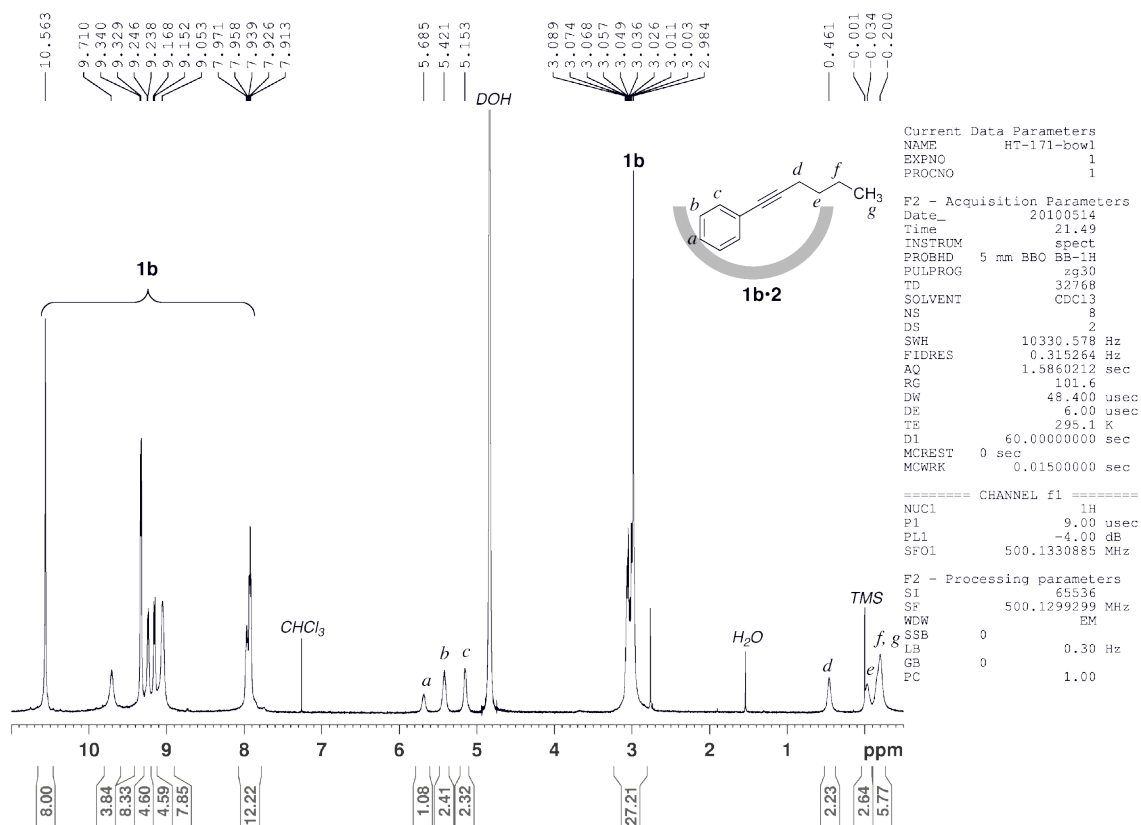


Fig. S5. ¹H NMR spectrum of **1b•2** (D₂O).

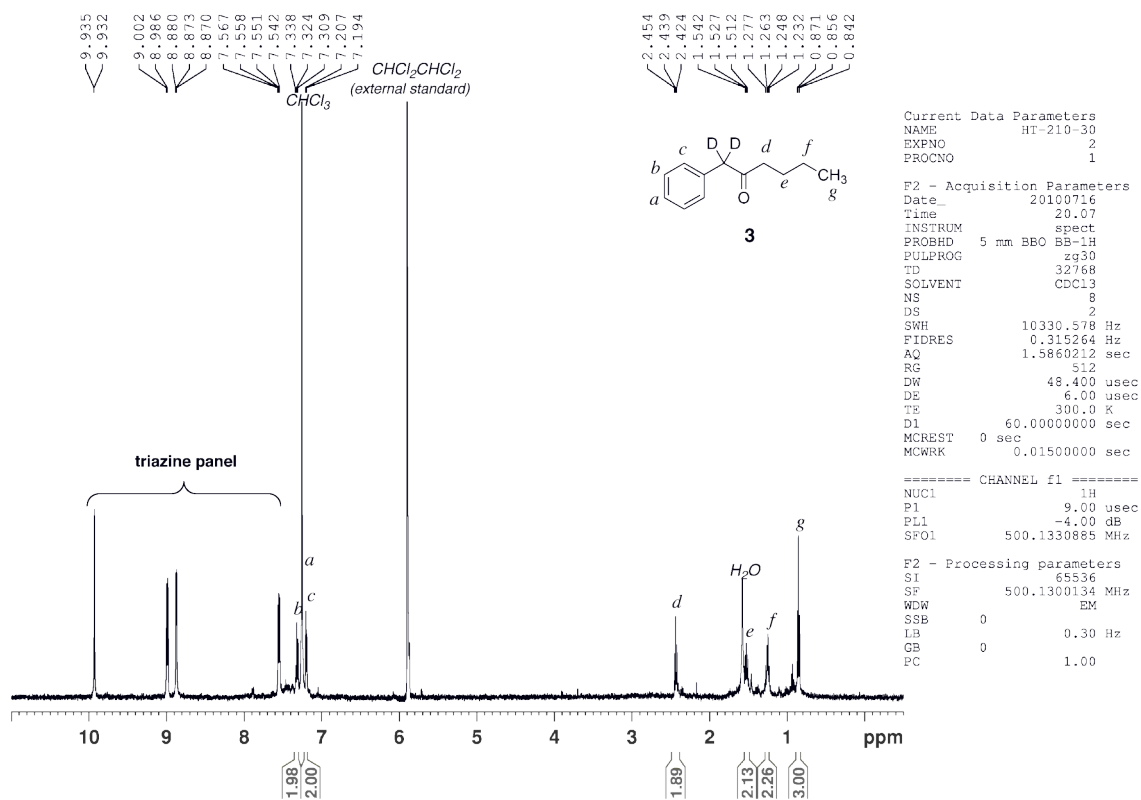
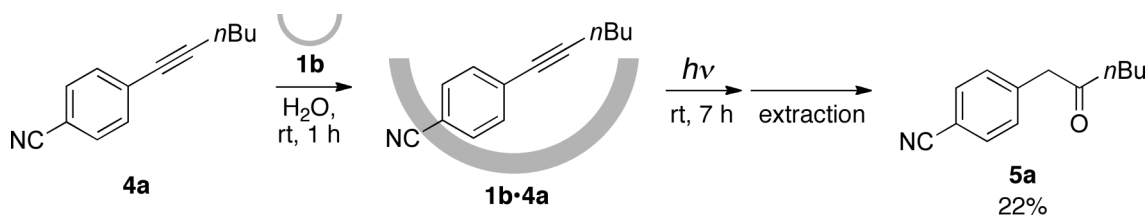


Fig. S6. ¹H NMR spectrum of **3** extracted from cage **1b** with CDCl₃ (reacted in D₂O solution).

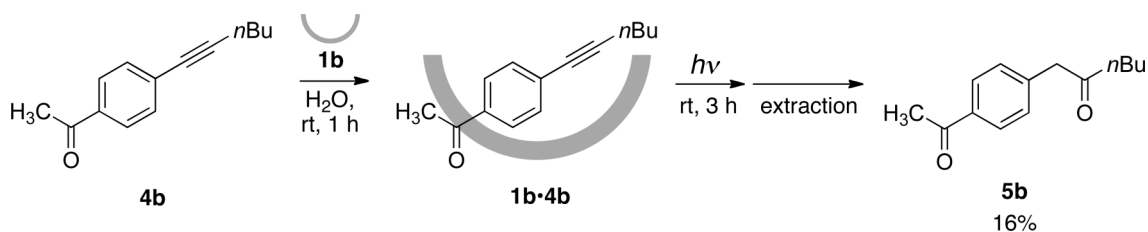
Photo-hydration of 4a within cage 1b.



Physical data of 1b·4a: ¹H NMR (500 MHz, D₂O, 300 K): *d* = 0.12 (br, 3H, CH₃, 4a), 0.31 (br, 2H, CH₂, 4a), 0.40 (br, 2H, CH₂, 4a), 1.06 (br, 2H, CH₂, 4a), 2.99–3.10 (m, 24H, CH₂, 1b), 5.40 (br, 2H, CH, 4a), 5.87 (br, 2H, CH, 4a), 7.99 (t, *J* = 7.0 Hz, 8H, PyH, 1b), 8.04 (t, *J* = 7.0 Hz, 4H, PyH, 1b), 9.11 (d, *J* = 6.0 Hz, 8H, PyH, 1b), 9.20 (d, *J* = 8.0 Hz, 4H, PyH, 1b), 9.24 (d, 4H, *J* = 5.5 Hz PyH, 1b), 9.39 (d, *J* = 5.5 Hz, 8H, PyH, 1b), 9.76 (s, 4H, PyH, 1b), 10.61 (s, 8H, PyH, 1b).

Physical data of 5a: ¹H NMR (500 MHz, CDCl₃, 300 K): *d* = 0.89 (t, *J* = 7.5 Hz, 3H, CH₃), 1.29 (sextet, *J* = 7.5 Hz, 2H, CH₂), 1.49 (quintet, *J* = 7.5 Hz, 2H, CH₂), 2.48 (t, *J* = 8.0 Hz, 2H, CH₂), 3.77 (s, 2H, CH₂), 7.31 (d, *J* = 9.0 Hz, 2H, CH), 7.62 (d, *J* = 9.0 Hz, 2H, CH). MS (EI) *m/z*: 201 [M]⁺, 116 [M–C₄H₉CO]⁺, 85 [C₄H₉CO]⁺, 57 [C₄H₉]⁺.

Photo-hydration of 4b within cage 1b.



Physical data of 1b·4b: ¹H NMR (500 MHz, D₂O, 300 K): *d* = 0.44 (br, 5H, CH₂ and CH₃, 4b), 0.69 (br, 2H, CH₂, 4b), 1.16 (br, 2H, CH₂, 4b), 1.53 (br, 2H, CH₃, 4b), 2.99–3.11 (m, 24H, CH₂, 1b), 5.30 (br, 2H, CH, 4b), 5.79 (br, 2H, CH, 4b), 8.02 (t, *J* = 7.0 Hz, 8H, PyH, 1b), 8.17 (t, *J* = 7.0 Hz, 4H, PyH, 1b), 8.98 (d, *J* = 6.0 Hz, 8H, PyH, 1b), 9.24 (d, *J* = 8.0 Hz, 4H, PyH, 1b), 9.31 (d, 4H, *J* = 5.5 Hz PyH, 1b), 9.44 (d, *J* = 5.0 Hz, 8H, PyH, 1b), 9.60 (s, 4H, PyH, 1b), 10.62 (s, 8H, PyH, 1b).

Physical data of 5b: ¹H NMR (500 MHz, CDCl₃, 300 K): *d* = 0.89 (t, *J* = 7.0 Hz, 3H, CH₃), 1.28 (sextet, *J* = 7.5 Hz, 2H, CH₂), 1.56 (quintet, *J* = 7.5 Hz, 2H, CH₂), 2.47 (t, *J* = 7.5 Hz, 2H, CH₂), 2.60 (s, 3H, CH₃), 3.75 (s, 2H, CH₂), 7.30 (d, *J* = 8.0 Hz, 2H, CH), 7.93 (d, *J* = 7.5 Hz, 2H, CH). MS (EI) *m/z*: 218 [M]⁺, 203 [M–CH₃]⁺, 57 [C₄H₉]⁺.