

Chemical Communications

## Dynamic Clicked Surfaces Based on Functionalised Pillar[5]arene

Huacheng Zhang,<sup>a</sup> Nathan L. Strutt,<sup>a,b</sup> Ragnar S. Stoll,<sup>a</sup> Hao Li,<sup>a</sup> Zhixue Zhu,<sup>a</sup> and J. Fraser Stoddart\*<sup>a,b</sup>

<sup>a</sup>Department of Chemistry, Northwestern University, 2145 Sheridan Road, Evanston, IL, 60208-3113, USA.

<sup>b</sup>Graduate School of EEWS (WCU), Advanced Institute of Science and Technology (KAIST),

373-1, Guseong Dong, Yuseong Gu, Daejeon 305-701, Republic of Korea.

# **Electronic Supplementary Information**

General Methods	<b>S2</b>
Compounds Synthesised	<b>S3</b>
Synthesis of 1	<b>S3</b>
Figure S1—S9	S4—S8
Table S1	<b>S8</b>
References	<b>S9</b>

* To Whom Correspondence should be Addressed		
Professor J Fraser Stoddart		
Department of Chemistry		
Northwestern University		
2145 Sheridan Road		
Evanston, IL 60208-3113, USA		
Tel: (+1)-847-491-3793		
Fax: (+1)-847-491-1009		
E-mail: stoddart@northwestern.edu		

#### **General Methods**

Reagents were purchased from Aldrich and used without further purification. 4-hydroxy-4'-ethynyl azobenzene<sup>S1</sup> (2) and azido-pillar[5]arene<sup>S2</sup> (3) were synthesised according to the literature procedures. The THF was dried using a commercial solvent purification system (SG Water, Inc.). The H<sub>2</sub>O used was triply distilled. The UV lamp is a Blak-Ray series, model B100A/R and 365 nm longwave UV, purchased from UVP, LLC. UV spectra were recorded with TU-1800pc UV-Vis Spectrophotometer at 298 K. The stock solutions ( $10^{-2} \text{ mol} \cdot \text{L}^{-1}$ ) of 1 were prepared with triply distilled H<sub>2</sub>O. All sample solutions for the investigation of the CAC were freshly prepared by diluting the stock solutions according to literature procedures.<sup>S3</sup>

1D and 2D nuclear magnetic resonance (NMR) spectra were recorded at 298 K on Bruker Avance III 500 spectrometer with working frequencies of 500 MHz for <sup>1</sup>H and 125 MHz for <sup>13</sup>C nuclei. Chemical shifts are reported in ppm relative to the signals corresponding to the residual non-deuterated solvent (CDCl<sub>3</sub>:  $\delta$  7.26 ppm; Tetrahydrofuran-d<sub>8</sub>:  $\delta$  1.73 and 3.58 ppm; D<sub>2</sub>O:  $\delta$  4.79 ppm), and coupling constants were recorded in Hertz (Hz). All <sup>13</sup>C NMR spectra were recorded with the simultaneous decoupling of <sup>1</sup>H nuclei. The following abbreviations were used to explain the multiplicities: s, singlet; d, doublet; t, triplet; b, broad peaks; m, multiplet or overlapping peaks. Electrospray Ionization (ESI) mass spectra were obtained on an Agilent LC-TOF high-resolution mass spectrometer.

Negative-stained TEM and Cryo-TEM were measured on a Hitachi H-8100 electron microscope (100—200 kV) equipped with slow scan CCD and using cold cathode field emission as the gun. The samples for negative-stained TEM were prepared by dropping a droplet of the orange yellow sample solution onto a TEM grid (copper grid, 300 meshes, coated with carbon film), immediately staining with 2% uranyl acetate in H<sub>2</sub>O (about 2  $\mu$ L) and allowing to air-dry. SEM images were obtained on a FEI Quanta 600 SFEG scanning electron microscope (0.2—30 kV) equipped with both solid-state BSED and low vacuum LFSED as detectors. Atomic force microscopy was performed by using a multimode Nanoscope IIIA system operated in tapping mode using silicon cantilevers. DLS measurements were carried out with a Zetasizer Nano ZS instrument purchased from Malvern Instruments Ltd. at 298 K using a 633 nm 'red' laser. The mean hydrodynamic radius was calculated with Zetasizer software.

#### **Compounds Synthesised**



#### Synthesis of 1

1: 4-hydroxy-4'-ethynyl azobenzene (2, 25 mg, 0.11 mmol) and 3 (90 mg, 0.11 mmol), tetrakis(acetonitrile) copper(I) hexafluorophosphate (4.1 mg, 0.011 mmol) and tris[(1-benzyl-1H-1,2,3-triazol-4-yl)methyl] amine (TBTA) were dissolved in Me<sub>2</sub>CO (20 mL). The mixture was then stirred under reflux for 2 days. The solution was then poured into H<sub>2</sub>O (200 mL). The aqueous phase was extracted (3 × 200 mL) with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried (Mg<sub>2</sub>SO<sub>4</sub>) and the solvent was removed in vacuum. The mixture was subjected to column chromatography (SiO<sub>2</sub>, gradient elution from 2% up to 20% MeOH in CH<sub>2</sub>Cl<sub>2</sub>) to afford 1 (108.3 mg) in 94% yield. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.16 (s, 1H), 7.97 (t, *J* = 10 Hz, 4H), 7.91 (d, *J* = 10 Hz, 2H), 7.01 (d, *J* = 5 Hz, 2H), 6.86-6.74 (m, 8H), 6.44 (t, *J* = 5 Hz, 2H), 4.35 (d, *J* = 5 Hz, 2H), 3.83-3.78 (m, 12H), 3.72-3.59 (m, 26H), 3.43 (s, 3H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  = 159.2, 152.3, 151.6, 151.2, 151.0, 150.7, 132.1, 128.9, 128.6, 128.4, 128.1, 127.8, 127.6, 126.2, 125.1, 123.2, 121.6, 115.9, 115.2, 114.6, 114.4, 114.2, 114.1, 113.8, 67.2, 55.9, 50.1, 29.9. HR-MS (ESI): C<sub>60</sub>H<sub>61</sub>N<sub>5</sub>O<sub>11</sub> calcd for *m/z* = 1028.4440, found *m/z* = 1028.4443 [*M* + H]<sup>+</sup>.

Figure S1—S9



Fig. S1 The critical assembly concentration (CAC) of 1 in the 1/2 (v/v) mixture of H<sub>2</sub>O and THF was determined in three separate trials, giving an average value of  $0.62 \pm 0.03$  mM by detection of the maximum absorption band in the UV-vis spectra.



**Fig. S2** Size distributions of **1** cast on a TEM grid from a  $1:2 \text{ H}_2\text{O}$  / THF mixture (2 mM) in visible light atmosphere (left), irradiated by UV (365 nm) for 1 h (middle) and then exposed in visible light for another 1 h (right).



Fig. S3 The orange-yellow sample solution was dropped onto a silicon plate and allowed to air-dry. Air tapping mode AFM images of assemblies of 1 (upper left, planar image with scale bar =  $15.3 \mu m$ ; upper right, steric image; bottom, a line profile extracted along the marked white trace in the upper left image).



**Fig. S4** DLS size distributions of **1** (molar ratio = 1:1) in the 1:2  $H_2O$  / THF mixture (2 mM) before irradiating by UV (solid line), after irradiating by UV and exposing in visible light (dash line).



Fig. S5 2D NMR NOESY spectrum of 1 at ambient temperature. Inset: proposed structure of lamellar assemblies formed by 1.



**Fig. S6** <sup>1</sup>H-NMR spectra (500 MHz, r. t.) of 1:1 **2** and DMpillar[5]arene mixture (2 mM) in 1:2 (v/v)  $D_2O$  / THF-d<sub>8</sub> mixed solvent in comparison with the individual **2** and the individual DMpillar[5]arene.



**Fig. S7** The sample solution, **1** in the 1/2 (v/v) mixture of water and THF (2 mM), was irradiated by the UV lamp for 1 hour at rt. One droplet of the solution was cast on a silicon plate and carefully air-dried in the absence of light. Immediately, AFM detection with air tapping mode was performed. Air tapping mode AFM images of assemblies of **1** (upper left, planar image with scale bar = 7.6 mm; upper right, steric image; bottom, a line profile extracted along the marked white trace in the upper left image).



**Fig. S8** Micro-morphology of **1** in the 1:2  $H_2O$  / THF mixture (2 mM) after exposing in visible light once again for 1 h: negative stained TEM image (a) and gold sputtering SEM images (b, scale bar = 5  $\mu$ m).



Fig. S9 Air tapping mode AFM images of assemblies of 1 which were exposed to visible light (upper left, planar image with scale bar =  $15.3 \mu m$ ; upper right, steric image; bottom, a line profile extracted along the marked white trace in the upper left image).

### Table S1

Table S1. Morphologies and sizes of aggregates obtained by TEM, SEM, ESEM and AFM.

Conditions	Morphology	Size (D / nm)
Visible light	Hollow spheres (TEM)	150~600 (TEM)
	Hollow spheres (SEM)	180~700 (SEM)
	Hollow spheres (ESEM)	300~1000 (ESEM)
	Hollow spheres (AFM)	300~1000 (AFM)
UV (365 nm)	Solid spheres (TEM)	300~650 (TEM)
	Solid spheres (SEM)	200~600 (SEM)
	Solid spheres (AFM)	300~700 (AFM)
Visible light	Hollow spheres (TEM)	250~1700 (TEM)
	Hollow spheres (SEM)	300~1500 (SEM)
	Hollow spheres (AFM)	300~1800 (AFM)

#### References

<sup>S1</sup> A. Loudet, J. Han, R. Barhoumi, J. P. Pellois, R. C. Burghardt, K. Burgess, *Org. Biomol. Chem.*, 2008, 6, 4516.

<sup>52</sup> N. L. Strutt, R. S. Forgan, J. S. Spruell, Y. Y. Botros, J. F. Stoddart, J. Am. Chem. Soc., 2011, 133, 5668.

<sup>53</sup> (a) P. Mukerjee, M. J. Gumkowski, C. C. Chan, R. Sharma, J. Phys. Chem., 1990, **94**, 8832; (b) B. W.

Liu, Y. Chen, B. E. Song, Y. Liu, Chem. Commun., 2011, 47, 4418.