# **Electronic Supplementary Information**

# Design and synthesis of catenated rings based on toroidal DNA structures

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## **Materials and Methods:**

**Reagents:** *Taq* DNA Polymerase, T4 DNA ligase, Nuclease BAL-31, Sac I and Cre recombinase were purchased from New England Biolabs (Ipswich, MA). Human Topoisomerase I was obtained from TopoGEN (Columbus, OH). pGH plasmid that contains our newly designed DNA sequence was provided by Generay Biotech (Shanghai, China) and single strand oligonucleotides were produced by Sigma-Aldrich (Singapore).

**PCR amplification of Linear DNA 1:** A reported standard PCR amplification process<sup>1-2</sup> was followed in our studies to obtain Linear DNA 1 using oligonucleotides 5'GTGGATCCTCGTCGCAAAAC3' and 5'CCGGATCCATGGTTAACCC3' as forward and reverse primers and pGH plasmid as template.

**Preparations of Linear DNA 2:** A mixture containing 2 pmol Linear DNA 1, 5 U of Sac I, 10 mM Bis-Tris-Propane-HCl, 10 mM MgCl<sub>2</sub>, 1 mM Dithiothreitol was incubated 37 °C for 2 hours. The resultant cohesive end-containing Linear DNA 2 was purified using the QIAquick Purification Kits (Qiagen).

**Preparations of Circular DNA 1:** A mixture containing 2 pmol Linear DNA 2, 60 mM Tris–HCl (pH 7.6), 25 mM NaCl, 13 mM MgCl<sub>2</sub>, 10 mM DTT, 1mM ATP, 25 mg/ml BSA and 10 U of T4 DNA ligase was incubated at 16  $^{\circ}$ C for 16 hours. The resultant mixture was further incubated with BAL-31 at 30  $^{\circ}$ C for 2 hours, which were then analyzed using 2.5% agarose gel electrophoresis. The DNA product that is corresponding to Band 1 in Lane 5 of Figure 2B was purified next using Mini Prep Cell (Bio-rad) before AFM examination.

**Reaction of relaxed form of Circular DNA 1 with Human Topoisomerase I:** A mixture containing 0.2 pmol circular DNA 1, 1 U of Human Topoisomerase I, 10 mM Tris–HCl (pH 7.9), 150 mM NaCl, 0.1% bovine serum albumin (BSA), 0.1 mM spermidine and 5% glycerol was incubated at 37 °C for 1 hour followed by analysis using 2% agarose gel electrophoresis.

Formation of catenated DNA: A mixture containing 0.2 pmol Circular DNA 1, 50 mM Tris-HCl, 33 mM NaCl, 10 mM MgCl<sub>2</sub> and 1 U of Cre recombinase was incubated at 37  $^{\circ}$ C for 12 hours. The resultant reaction mixture was further analyzed using 2.5% agarose gel electrophoresis.

### AFM studies of our synthesized catenated DNA

(*a)* Immobilization of DNA samples on micas<sup>3-5</sup>: A buffer (PH 7.0) containing 40 mM HEPES and 10 mM MgCl<sub>2</sub> was placed on freshly cleaved mica, which was further kept at room temperature for 5 min. The pre-prepared mica was rinsed with dd water thoroughly and dried under the argon flow. A 20  $\mu$ L solution that contains DNA sample was deposited on the mica, which was subsequently incubated for 5 min, washed with H<sub>2</sub>O, dried under the argon flow and then stored in vacuum

cabinet.

(*b*) *AFM examination*: AFM images of DNA were obtained in Tapping Mode<sup>TM</sup> on a Multimode<sup>TM</sup> AFM (Veeco, Santa Barbara, CA) in connection with a Nanoscope  $V^{TM}$  controller. Antimony (n) doped Si cantilevers with nominal spring constants between 20 and 80 N/m were selected. Scan frequency was 1.9 Hz per line and the modulation amplitude was in a nanometer range <sup>6-7</sup>. All DNA sample determinations were carried out in air at room temperature.



#### pGH plasimd (4563 bp in length)

#### Nucleotide sequence of Linear DNA 1 (PCR product from pGH plasimd)

5 ' GTGGATCCTCGCCGAAAACGAGCTCAGTTGGGTAATTTTTAGGGTTTTCCCAGTTTTGACGTTGTTTTTCGACGGAATTCCCTTTTTACGACTCACTTTTTGCCTTGACTAGA 3 ' CACCTAGGAGCAGCGTTTTGCTCGAGTCAACCCATTAAAAATCCCCAAAAGGGTCAAAAACTGCAACAAAAAGCTGCCTTAAGGGAAAAATGCTGAGTGAAAAACGGAACTGATCT GGGTTTTTACCCCATGGTCTATTTTGGTCTTTTGCCATAACTTTTTATAGCATACATTTTACGAGTTTTATAAGCTGTTTTTGCCATAACTTTTTGCCATAACTTTTTATAGCAT CCCAAAAATGGGGTACCAGATAAAAACCAGAAAACGGTATTGAAAAATATCGTATGTAAAATGCTCAAAATATTCGACAAAAACGGTATTGAAAAACGGTATTGAAAAATATCGTA ACATTTTACGAGTTTTATAAGCTGTTTTTTGCCATAACTTTTTATAGCATACATTTTACGAGTTTTATAAGCTGTTTTTCATGAGGCTTCTTTTATAGGTTTTTGTCATGATTTTA TGTAAAATGCTCAAAATATTCGACAAAAACGGTATTGAAAAATATCGTAAGAATGCTCAAAATATTCGACAAAAAGTACTCCGAAGAAAAATATCCAAAAACAGTACTAAAAA TACCATAGAAAAAACGGTATTGAAAAATATCGTATGTAAAATGCTCCAAAATATTCGACAAAAAGTACTCCGAAGAAAATATCCAAAAACAGTACTAAAATTACCATAGAAAAAGCA CGGTGGCATTTTTCGGGGTTTTGCGCGGATGCCTTTTTGTTTATGGGCCTTTTTACATCAGGTTTTTTCCGCCCAGCAATGATTTTTGCCCCTTTTAGATTTTTCAATGATATTTTT GCCACCGTAAAAAGCCCCCAAAACGCGCCTACGGAAAAACAAATACCCGGAAAAATGTAGTCCAAAAAAGGCGAGTCGTTACTAAAAACGGGGAAAATCTAAAAAAGTTACTATAAAAA AGGCGTTTTTGACGTTTTCAGTTTTTCCGTGTCGCCCTTTTTTCCCTTTTTTGCGCATTTTTTCGGCACTTTTTTGCATATATTTTTTGGAGTTGTTTTGATCCGTTTTGATTTTCA TCCGCAAAAACTGCAAAAAGTCAAAAAGGCACAGCGGGAAAAAAGGGGAAAAAACGCGTAAAAAAGCCGTGAAAAAACGTATATAAAAAAACCTCAACAAAACTAGGCAAAAACTAGGCAAAAAAGT Lox P site 1 GTGCGTTTTTGGCCATTTTGCCTAGTTTTTTTTTTTTGTATAACTTCGTATAGCATACATTATACGAAGTTAT<sup>T</sup>TTGCTATTTTTGTTAATTTTTGCCAATTTTCGTATTTTCGCTATTTTTG CACGCAAAAACCGGTAAAACGGATCAAAAAATATTGAAGCATATCGTATGTAATATGCTTCAATAAAACGATAAAAAACAATTAAAAACGGTTAAAAACCGATAAAAAGCGATAAAAAAC GAGACGATAAAAAAAAACAATTAAAAAACGGTTAAAAAGCTCCATAAAAAGCGATAAAAAAACCGTAAAAAAACGACGTAAAAAAACCGGTAAAAAAACGGACAAAAAACGAGTGGGTA AAGGAGAGAAGATTTTGGGTCTCAGTTTTGATACCCGACGATTTTGACACCACGATTTTTGCAGGCGTTTTGGGCTCAGTTTTGATACCCGACGATTTTGACACCACGATTTTGCAGGCGCTCTAAAACCGTCCCGAGATCTTGGGCCGCTAAAACCGTGGGTGCTAAAAACGTCCGCGAGACCACGAGTCAAAAACTGTGGGCTGCTAAAAACG Lox P site 2 aggcgtttttgagccatttttgcgtatagcattttggatacgatgttttccatgátaacttcgtatagcatacattatacgaagttatttttgggctcagttttgatacccggatc TCCGCAAAAAACTCCGGTAAAAACCCATATCGTAAAAACCTATGCTACAAAAGGTACTATTGAAGCATATCGTATGTAATATGCTTCAATAAAAACCCCGAGTCAAAAACTATGGGCCTAG TTTTGACACCACGATTTTTGCAGGCGTTTTTGAGCCATTTTGCGTATAGCATTTTGGATACGATGTTTTCCATGTTTTAGTGGTTTTTGACGTAGCTTTTCGCCATGGTAGTCAT AAAACTGTGGTGCTAAAAACGTCCGCAAAAACCTCGGTAAAAACGCATATCGTAAAAACCTATGCTACAAAAGGTACAAAAATCACCGAAAAACTGCAGCAGTAACAACGG

TTTTATAGCTGTTTTTGTGTGAGATTTTTATCCGCTCACTTTTCGAATTCCTTTTTACGAGCCGGATTTTTGCGGTGTGGGCTTTTTGTCCGTTTTCCTGTTTGAGCTCGGGGTTAA AAAATATCGACAAAAACACCCTTAAAAATAGGCGAGTGAAAAGCTTAAGGAAAAATGCCCGGGCCTAAAAACGCCCACCGAAAAACAGGCAAAAGGACAAACTCGAGCCCCAATT

CCATGGATCCGG 3' GGTACCTAGGCC 5'

Figure S1

### Nucleotide sequence of Linear DNA 2



Figure S2



Figure S3. AFM images of Non-Catenated Ring 1 and Non-Catenated Ring 2. A: AFM image of Non-Catenated Ring 1 (603 bp in length) that are corresponding to Band 4 in Lane 3 in Figure 3A (1  $\mu$ m x 1  $\mu$ m scans). B: AFM image of Non-Catenated Ring 2 (985 bp in length) that are corresponding to Band 3 in Lane 3 in Figure 3A (1  $\mu$ m x 1  $\mu$ m scans).



Figure S4. Schematic illustration of strand recombination of toroidal DNA in which two lox p sites are aligned in an anti-parallel fashion. This Cre recombinase-catalyzed reaction does not lead to any

catenated DNA structure. (Tindicates the lox p site)



Figure S5. Electrophoretic analysis of reactions between Circular DNA 2 and Cre recombinase. Lnae 1: molecular weight markers; Lane 2: Circular DNA 2 alone; Lane 3: reaction mixture of Circular DNA 2 and Cre recombinase.



Figure S6. Optimization of reaction yield for formation of catenated DNA structure. Land 1: molecular weight markers; Lane 2: Circular DNA 1 alone; Lane 3: reaction mixture of Circular DNA 1 and Cre recombinase. The reaction mixture loaded in Lane 3 was prepared as follows : 4 U (1 U each time) of Cre recombinase was added to a solution containing 0.2 pmol Circular DNA 1, 50 mM Tris-HCl, 33 mM NaCl and 10 mM MgCl<sub>2</sub> for four times within 20 minute intervals. After each addition of Cre recombinase, the resultant solution was further incubated at 37 °C. The reaction yield is  $\sim 16\%$ .

### **References for Supplementary Information:**

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