Supporting Information

Graphene Oxide-Protected DNA Probes for Multiplex MicroRNA Analysis in Complex Biological Samples Based on Cyclic Enzymatic Amplification Method [‡]

Liang Cui, Xiaoyan Lin, Ninghang Lin, Yanling Song, Zhi Zhu, Xi Chen, Chaoyong James Yang*

The Key Laboratory of Analytical Science, the Key Laboratory for Chemical Biology of Fujian Province, State Key Laboratory of Physical Chemistry of Solid Surfaces, and Department of Chemical Biology, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen 361005 (China) Fax: +86 592 2189959; E-mail: cyyang@xmu.edu.cn

Experimental Section

Materials

DNase I, HPLC-purified RNA, RNase inhibitor, and DEPC-treated water were purchased from Takara Biotechnology Co. Ltd. (Dalian, China). The DNA probes were synthesized on a PolyGen Column 12 DNA synthesizer and the reagents were purchased from Glen Research (Sterling, VA, USA). All DNA/RNA sequences are listed in Tables S1. Graphene oxide (GO) was synthesized from natural graphite powder by a modified Hummers method.¹ Cell lysate from lung carcinoma cell line A549 and MCF-10A was obtained according to the manufacturer's protocol.

Fluorescence measurements

Fluorescence measurements were carried out on a RF-5301-PC Fluorescence Spectrophotometer (Shimadzu, Japan). For P7a, excitation and emission wavelengths were set at 560 and 582 nm, respectively, with a 5 nm bandwidth. The emission spectra were obtained by exciting the samples at 560 nm and scanning the emission from 570 to 650 nm in steps of 1 nm. For P7e and P7i, excitation and emission wavelengths were set at 490 and 520 nm, 643 and 670nm, respectively, with same a bandwidth of 5 nm. All experiments were conducted in 20mM Tris-HCl (pH 8.0) buffer containing 5 mM MgCl₂ and 50 mM NaCl. The amplified detection of target were performed in 200 μ L solution consisting of 50 nM probe, 20 units of DNase I and varying concentrations of target miRNA at RT for 20 min.

Gel electrophoresis

A 20% navitve PAGE analysis of the products from the cyclic enzymatic amplification reaction was carried out in 1×TBE (89mM Tris base, 89mM Boric acid, 2 mM EDTA, pH 8.3) for about 2 hours. After Stains-All staining, gels were scanned.

Name	Sequence
P7a	5' -AAC TAT ACA ACC TAC TAC CTC A -TMR -3'
P7e	5'- ACT ATA CAA CCT CCT ACC TCA - FAM -3'
P7i	5'- AAC AGC ACA AAC TAC TAC CTC A - Cy5 -3'
P-mir21	5'-T CAA CAT CAG TCT GAT AAG CTA -TMR -3'
Let-7a	5' - UGA GGU AGU AGG UUG UAU AGU U -3'
Let-7e	5'- UGA GGU AG <u>G</u> AGG UUG UAU AGU -3'
Let- 7i	5'- UGA GGU AGU AG <u>U</u> UUG U <u>GC U</u> GU U -3'

Table S1. Sequences of oligonucleotides used in this study



Figure S1. Change of fluorescence intensity as a function of target RNA concentration



Figure S2. Detection of different concentrations of microRNA target in the absence of DNase I. Experiment was performed with 1×10^{-7} M probe and different concentrations of target. The curves from top to down contain target microRNA at the concentration of 2.5×10^{-7} , 5×10^{-8} , 1×10^{-8} , 5×10^{-9} and 0 M, respectively.



Figure S3. (A) Response of CEAM to different concentrations of target in cell media. (B) The expression profiles of mir-21 from A549 and MCF-10A.

References:

1. W. S. Hummers and R. E. Offeman, J. Am. Chem. Soc., 1958, 80, 1339.