

## Supporting Information for

### Increasing Materials' Response to Two-photon NIR Light via Self-immolative Dendritic Scaffold

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**Abbreviations.** DCM (dichloromethane), TFA (trifluoroacetic acid), EtOAc (ethyl acetate), DMF (dimethyl formamide), DMSO (dimethyl sulfoxide), DIEA (N, N-diisopropylethyl amine), RT (room temperature).

**General Methods and Instrumentation.** Flash chromatography was performed using a CombiFlash Companion system. <sup>1</sup>H NMR spectra were acquired using a Joel 500 MHz spectrometer or a Varian 400 MHz spectrometer. <sup>13</sup>C NMR spectra were acquired using a Varian spectrometer operated at 100 MHz. Purity of the new compounds and degradation of the dendrimers was monitored by an Agilent 1200 HPLC equipped with PDA and MSD detectors and a C18 column with 0.1 % formic acid/H<sub>2</sub>O and 0.1 % formic acid/acetonitrile as eluents at a flow rate of 0.4 mL/min. A Ti:Sapphire laser (Mai Tai HP, Spectra Physics) with ~100 fs pulse widths and 80 MHz repetition rate generated light for NIR irradiation. For dendrimers' degradation by NIR irradiation, 2.5 W (4 kW/cm<sup>2</sup>) of 740 nm light was focused into the solution using a EFL 33.0 mm lens. The 50  $\mu$ L quartz cells with 3 mm path length were used. The Amplex Red Glutamic Acid assay kit was purchased from Invitrogen and the analysis was performed according to the manufacturer's protocol. The fluorescence spectra were acquired on FluoroLog-3 spectrophotometer (JY Horiba, Inc.)

For all dendrimers, the irradiation experiments followed by injection into HPLC and Amplex Red analysis were performed in triplicate to obtain standard deviation values.

Compounds **1** and **3** were synthesized according to ref. 1.

**Compound 2.** Glutamate hydrochloride (0.26 g, 1.23 mmol) was dissolved in 10 mL dry DMF and 0.42 mL of DIEA (2.4 mmol) was added dropwise. Compound **1** was added and reaction was heated to 50°C and monitored by HPLC. After 2 hours the reaction mixture was cooled to RT and solvent was removed on rotary evaporator. The residue was dissolved in ethyl acetate (EtOAc) and the precipitate was filtered out. The solvent was removed on a rotary evaporator. The product was isolated by flash-chromatography (silica gel, Hexane/EtOAc gradient). Yield: 0.28 g (53%).

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.65 (s, 1H), 7.15 (s, 1H), 6.37 (s, 1H), 5.80 (d, J=10 Hz, 1H), 5.31 (s, 2H), 5.23 (s, 2H), 4.42 (m, 1H), 3.78 (s, 3H), 3.68 (s, 3H), 3.51 (s, 3H), 2.43 (m, 2H), 2.24 (m, 1H), 2.05 (m, 1H) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 173.12, 171.90, 160.13, 154.07, 148.57, 127.42, 112.38, 111.10, 108.48, 103.98, 95.08, 61.64, 56.66, 53.52, 52.73, 51.94, 29.88, 27.25 ppm.

HRMS: composition: C<sub>20</sub>H<sub>22</sub>BrNO<sub>10</sub>Na; mass measured: 538.0320; theor. mass: 538.0319; delta: 0.2 ppm.

Purity by HPLC: 96.77% at 254 nm; 98.77% at 320 nm; 98.86% at 280 nm.

**G0.** Compound **2** (0.1 g, 0.194 mmol) was dissolved in 2 mL of DCM/TFA (1/1) and stirred for 30 min. The solvents were removed on rotary evaporator. The residue was dissolved in 6 mL MeOH/H<sub>2</sub>O (5/1) and cooled to 0°C. 10 M NaOH solution was added dropwise until the reaction mixture turned yellow. The reaction was allowed to warm to RT and the progress was monitored by HPLC (C-18 Zorbax column, H<sub>2</sub>O/can gradient). After completion, the reaction mixture was neutralized with 10 M HCl to pH 5. MeOH was removed on rotary evaporator and aqueous fraction was extracted with EtOAc and dried over Na<sub>2</sub>SO<sub>4</sub>. After removal of EtOAc the product was purified by flash-chromatography (C-18, H<sub>2</sub>O/MeOH gradient). Yield: 0.038 g (40%). <sup>1</sup>H and <sup>13</sup>C NMR matched with the data reported in ref. 2.

HRMS: composition: C<sub>16</sub>H<sub>13</sub>NO<sub>9</sub>Br; mass measured: 441.9777; theor. mass: 441.9779; delta: 0.5 ppm.

**Compound 4.** Compound **3** (2 g, 2.8 mmol) was dissolved in a mixure of 10 mL dry DMF anf 0.93 mL (6.72 mmol) Et<sub>3</sub>N at RT. A solution of glutamate hydrochloride (1.42 g, 6.72 mmol) in 5 mL dry DMF was added dropwise and the reaction mixture was placed in oil bath at 50°C. The progress of reaction was followed by HPLC (C-18 Zorbax column, H<sub>2</sub>O/can gradient). After 3 hours the reaction mixture was cooled to RT and 20 mL EtOAc was added. The inorganic precipitate was filtered out. The solvents were removed on rotary evaporator. The product was isolated by flash-chromatography (silica gel, Hexane/EtOAc gradient). Yield: 1.8 g (85%).

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.20 (s, 2H), 5.61 (m, 2H), 5.02 (s, 2H), 4.36 (t, J=5 Hz, 2H), 3.73 (s, 6H), 3.65 (s, 6H), 3.6-3.45 (m, 4H), 3.14-2.90 (m, 6H), 2.50 (m, 4H), 2.40 (s, 3H), 2.23 (m, 2H), 2.01 (m, 2H), 1.64 (m, 9H) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 173.19, 172.37, 155.92, 135.87, 131.06, 130.72, 129.33, 62.43, 53.39, 52.64, 51.94, 47.33, 46.28, 35.53, 34.99, 30.04, 28.53, 27.67, 21.00 ppm.

HRMS: composition: C<sub>35</sub>H<sub>52</sub>N<sub>4</sub>O<sub>16</sub>Na; mass measured: 807.3269; theor. mass: 807.3271; delta: 0.2 ppm.

Purity by HPLC: 95.8% at 254nm; 95.5 at 280 nm.

**Compound 5.** Compound **4** (0.35 g, 0.446 mmol) was stirred in 2 mL of DCM and TFA (1/1) mixture for 30 min at RT. The solvents were removed on rotary evaporator, the residue was redissolved in 5 mL DMF and 2 mL of Et<sub>3</sub>N. Compound **1** (0.312 g, 0.670 mmol) was added and the reaction the mixture was stirred overnight at 50°C. After completion of reaction the mixture was cooled to RT, diluted with EtOAc

and inorganic precipitate was filtered out. The solution was dried over  $\text{Na}_2\text{SO}_4$ . The solvents were removed on rotary evaporator. The product was isolated by flash-chromatography on silica gel with Hexane/EtOAc gradient. Yield: 0.306 g (63%).

$^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 7.84 (s, 1H), 7.16 (m, 3H), 6.36 (s, 1H), 5.30 (m, 4H), 5.02 (s, 4H), 4.36 (t,  $J$ =5 Hz, 2H), 3.72 (s, 6H), 3.64 (s, 6H), 3.52 (s, 3H), 3.70-3.45 (m, 4H), 3.16-3.04 (m, 6H), 2.39 (m, 4H), 2.32 (s, 3H), 2.04 (m, 2H), 1.85 (m, 2H) ppm.

$^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 173.15, 172.39, 160.45, 154.18, 149.38, 145.33, 135.96, 135.84, 130.23, 129.29, 129.20, 128.00, 112.69, 110.67, 108.52, 104.01, 95.24, 62.59, 62.33, 56.79, 53.37, 51.91, 46.78, 35.06, 35.03, 34.41, 30.02, 27.62, 20.96 ppm.

HRMS: composition:  $\text{C}_{43}\text{H}_{53}\text{BrN}_4\text{O}_{20}\text{Na}$ ; mass measured: 1047.2331; theor. mass: 1047.2329; delta: 0.2 ppm.

Purity by HPLC: 96.52% at 254 nm; 97.68% at 320 nm; 96.71% at 280 nm.

**Compound 6.** Compound **4** (1.2 g, 1.5 mmol) was stirred in 10 mL DCM/TFA (1/1) for 30 min at RT. Solvents were removed on rotary evaporator, the residue was dissolved in 3 mL DMF and 5 mL  $\text{Et}_3\text{N}$  was added. Compound **3** (0.5 g, 0.7 mmol) in 5 mL DMF was added and the reaction was stirred at 50°C overnight. The solvents were removed on rotary evaporator and the product was isolated by flash-chromatography on silica gel with Hexane/EtOAc gradient. Yield: 1.2 g (95%).

$^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 7.19 (m, 6H), 5.01 (m, 12H), 4.35 (s, 4H), 3.72 (s, 12H), 3.64 (s, 12H), 3.70-3.25 (m, 12H), 3.14 – 2.78 (m, 18H), 2.37 (s, 9H), 2.39-2.25 (m, 8H), 2.20 (m, 4H), 1.97 (m, 4H), 1.45 (s, 9H) ppm.

$^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 173.05, 172.26, 162.68, 156.02, 153.99, 145.31, 135.72, 130.67, 130.19, 129.61, 129.18, 62.30, 53.30, 52.44, 51.76, 47.64-46.05, 36.50, 35.72-34.60, 31.42, 29.92, 28.38, 27.39, 20.84 ppm.

HRMS: composition:  $\text{C}_{81}\text{H}_{114}\text{N}_{10}\text{O}_{36}\text{Na}$ ; mass measured: 1825.7284; theor. mass: 1825.7289; delta: 0.3 ppm.

Purity by HPLC: 95.22% at 254 nm; 95.21% at 280 nm.

**Compound 7.** Compound **6** (0.5 g, 0.28 mmol) was dissolved in 4 mL DCM/TFA (1/1) and stirred at RT for 30 min. The solvents were removed on rotary evaporator and the residue was dissolved in 10 mL DMF/ $\text{Et}_3\text{N}$  (3/2). Compound **1** (0.163 g, 0.33 mmol) was added and the mixture was stirred at 50°C overnight. The solvents were removed on rotary evaporator and the product was isolated by flash-chromatography on silica gel with EtOAc/MeOH gradient. Yield: 0.53 g (93%).

$^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 7.77 (s, 1H), 7.16 (m, 7H), 6.33 (s, 1H), 5.31 (s, 4H), 5.00 (m, 12H), 4.34 (s, 4H), 3.71 (s, 12H), 3.63 (s, 12H), 3.51 (s, 3H), 3.70-3.19 (m, 12H), 3.14 – 2.75 (m, 18H), 2.31 (s, 9H), 2.39-2.25 (m, 8H), 2.17 (m, 4H), 1.96 (m, 4H) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 173.03, 172.27, 160.29, 156.27, 155.87, 154.10, 153.80, 135.80, 130.52, 130.04, 129.58, 129.20, 127.81, 112.57, 108.43, 103.89, 95.13, 62.22, 56.68, 53.31, 52.43, 51.75, 47.64-46.64, 35.69-34.47, 29.93, 27.35, 20.84 ppm.

HRMS: composition: C<sub>89</sub>H<sub>115</sub>BrN<sub>10</sub>O<sub>40</sub>Na<sub>2</sub>; mass measured: 1044.3126; theor. mass: 1044.3120; delta: 0.6 ppm.

Purity by HPLC: 95.29% at 254 nm; 97.09% at 320 nm; 965.19% at 280 nm.

**G1.** Compound **5** (0.29 g, 0.28 mmol) was reacted with TFA (1 mL) in DCM (1 mL) for 30 min. The solvents were removed on rotary evaporator. The residue was dissolved in 10 mL MeOH/H<sub>2</sub>O (10/1) and 10 M NaOH (0.4 mL) was added. The reaction mixture was stirred at 0°C for 10 hrs. After the reaction was completed, the solution was neutralized with 12 M HCl to pH 5. MeOH was removed on rotary evaporator and aqueous fraction was extracted with EtOAc. The product was purified by flash-chromatography using C-18 column and H<sub>2</sub>O/MeOH gradient. Yield: 0.168 g (65%).

<sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD):  $\delta$  = 7.87 (m, 1H), 7.22 (m, 2H), 6.84 (s, 1H), 6.25 (m, 1H), 5.35 (m, 2H), 4.98 (s, 4H), 4.19 (m, 2H), 3.64-3.57 (m, 4H), 3.31-2.99 (m, 6H), 2.39 (t, J=5 Hz, 4H), 2.32 (s, 3H), 2.14 (m, 2H), 1.87 (m, 2H).

HRMS: composition: C<sub>37</sub>H<sub>41</sub>BrN<sub>4</sub>O<sub>19</sub>Na; mass measured: 947.1439; theor. mass: 947.144; delta: 0.2 ppm.

Purity by HPLC: 97.21% at 254 nm; 97.36% at 320 nm; 97.71% at 280 nm.

**G2** was obtained from compound **7** (0.2 g, 0.098 mmol) according to the procedure described for **G1** above. Yield: 0.103 g (69%).

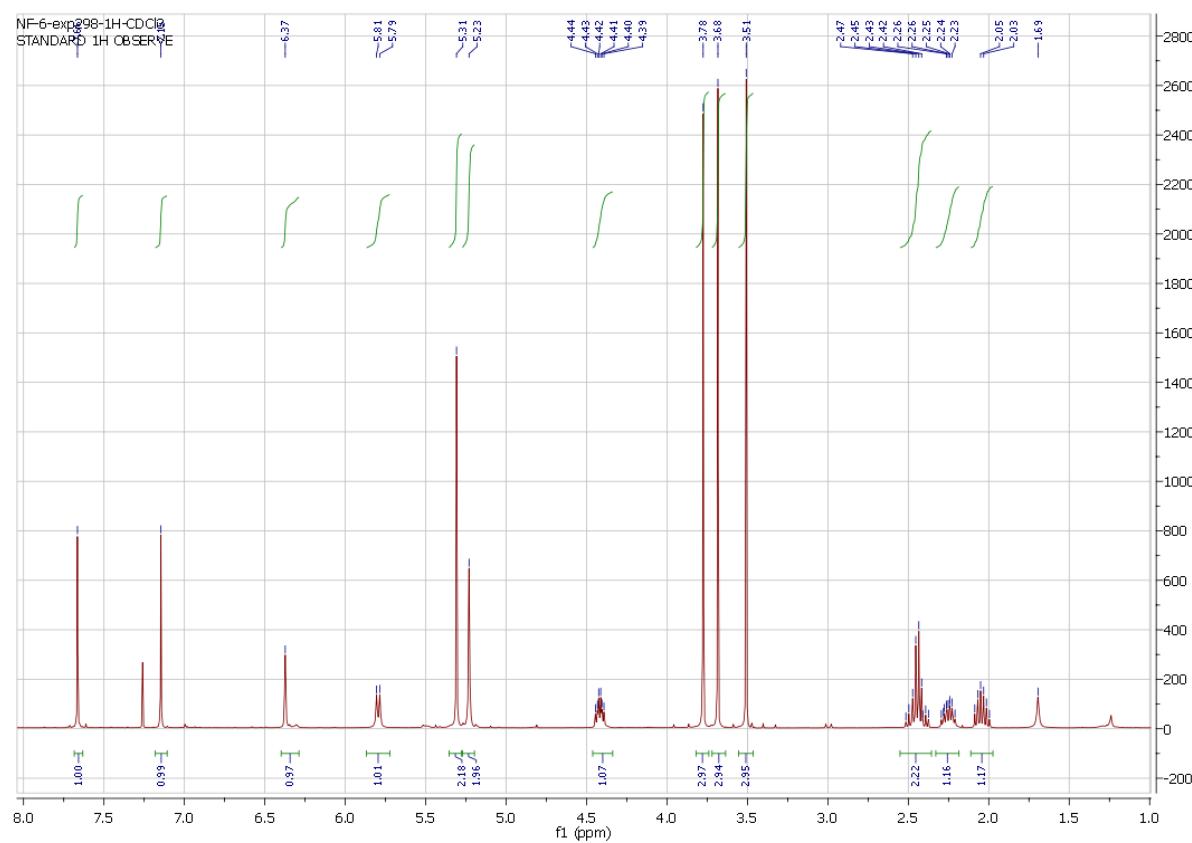
<sup>1</sup>H NMR (500 MHz, DMSO-<sub>d6</sub>):  $\delta$  = 7.86 (m, 1H), 7.19 (s, 7H), 6.91 (s, 1H), 6.14 (m, 1H), 5.28 (s, 2H), 4.92 (s, 12H), 3.98 (m, 4H), 3.05-2.81 (m, 18H), 2.29 (s, 9H), 2.36 – 2.17 (m, 8H), 1.94 (m, 4H), 1.76 (m, 4H) ppm.

HRMS: composition: C<sub>79</sub>H<sub>95</sub>BrN<sub>10</sub>O<sub>39</sub>Na<sub>2</sub>; mass measured: 966.2362; theor. mass: 966.2363; delta: 0.1 ppm.

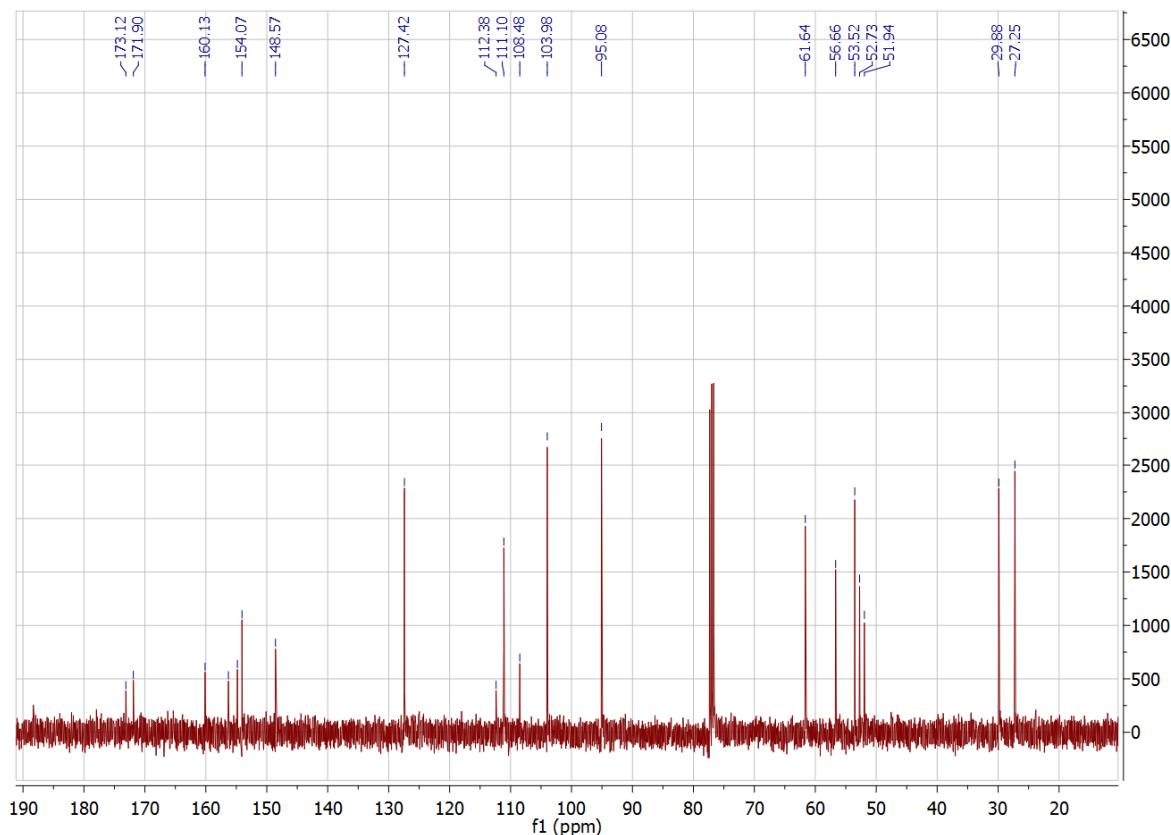
Purity by HPLC: 98.31% at 254 nm; 98.78% at 320 nm; 97.73% at 280 nm.

For **G1** and **G2**, the <sup>13</sup>C NMR spectra could not be acquired due to excessive peak broadening caused by hydrogen exchange.

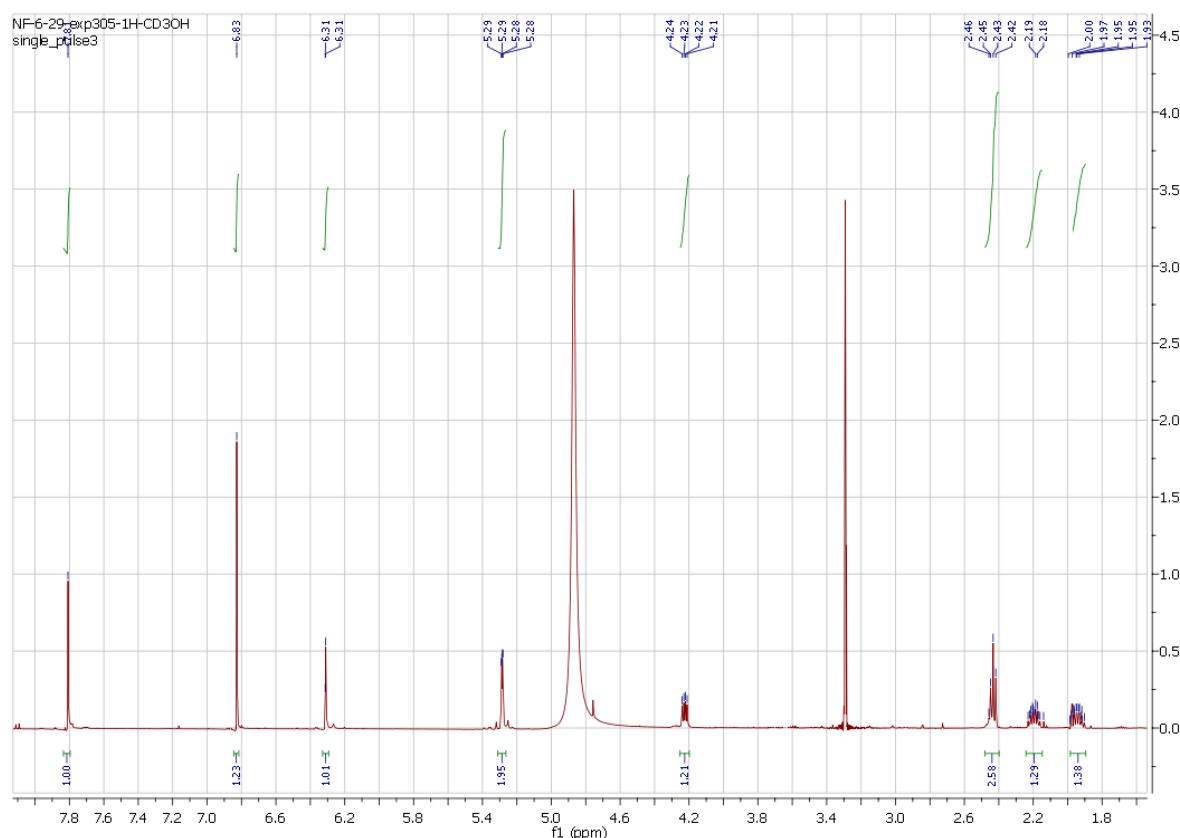
**Compound 2.**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz)



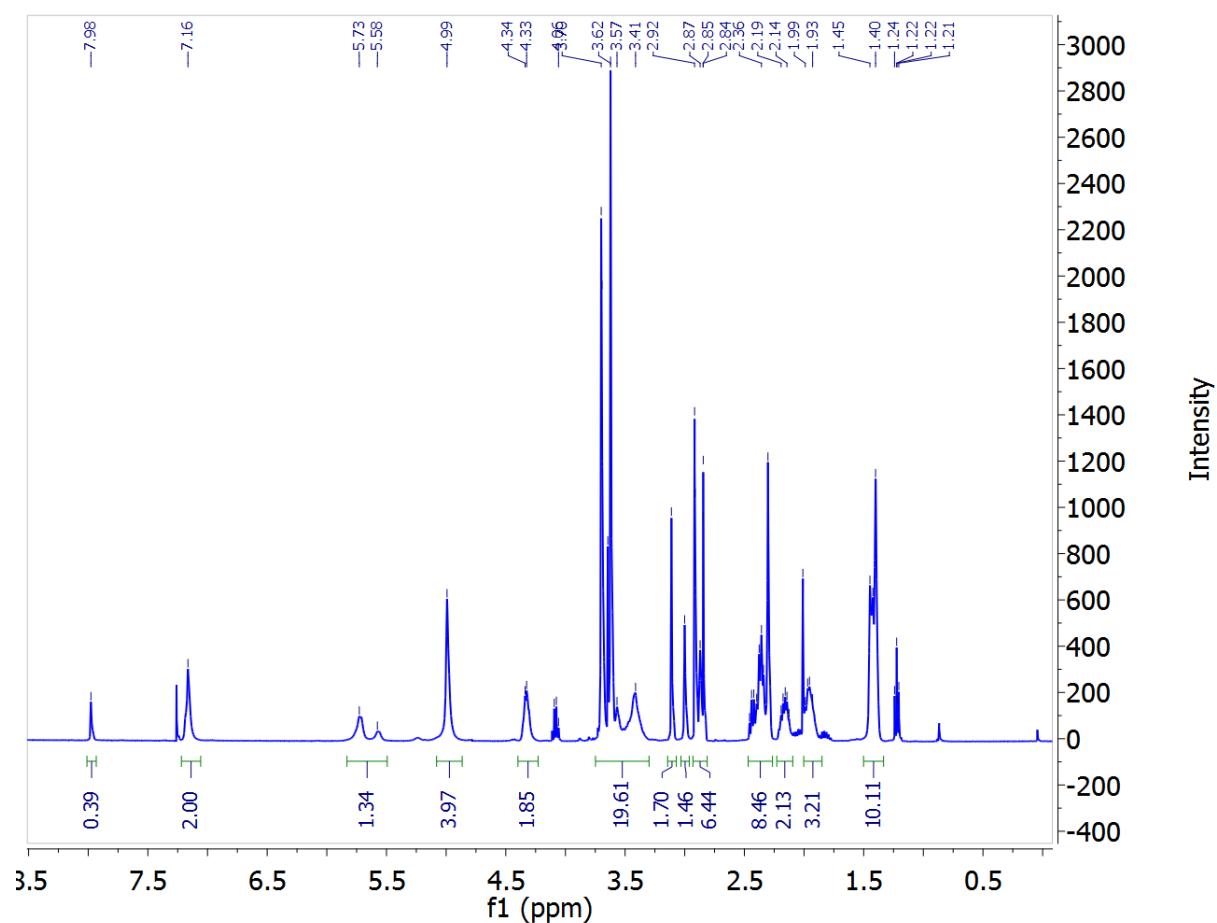
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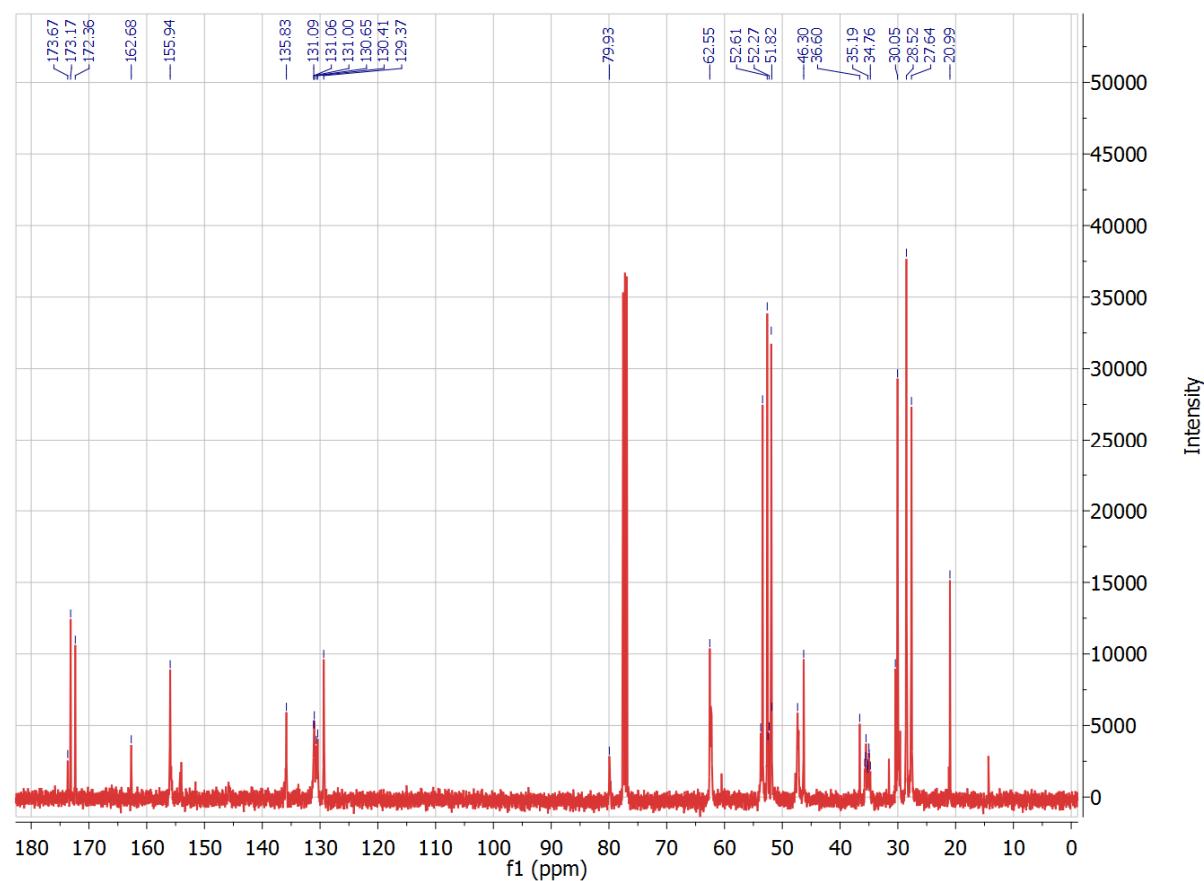
**G0.**  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ , 500 MHz)



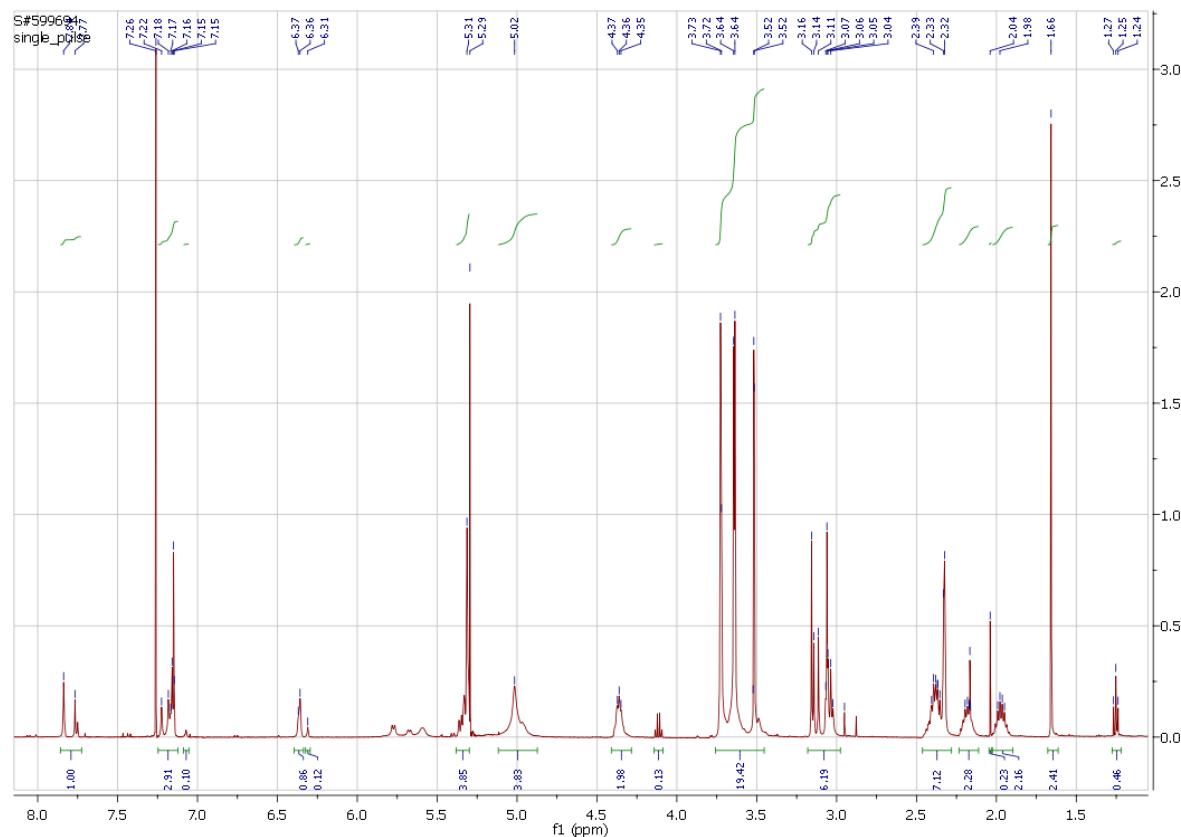
**Compound 4.**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )



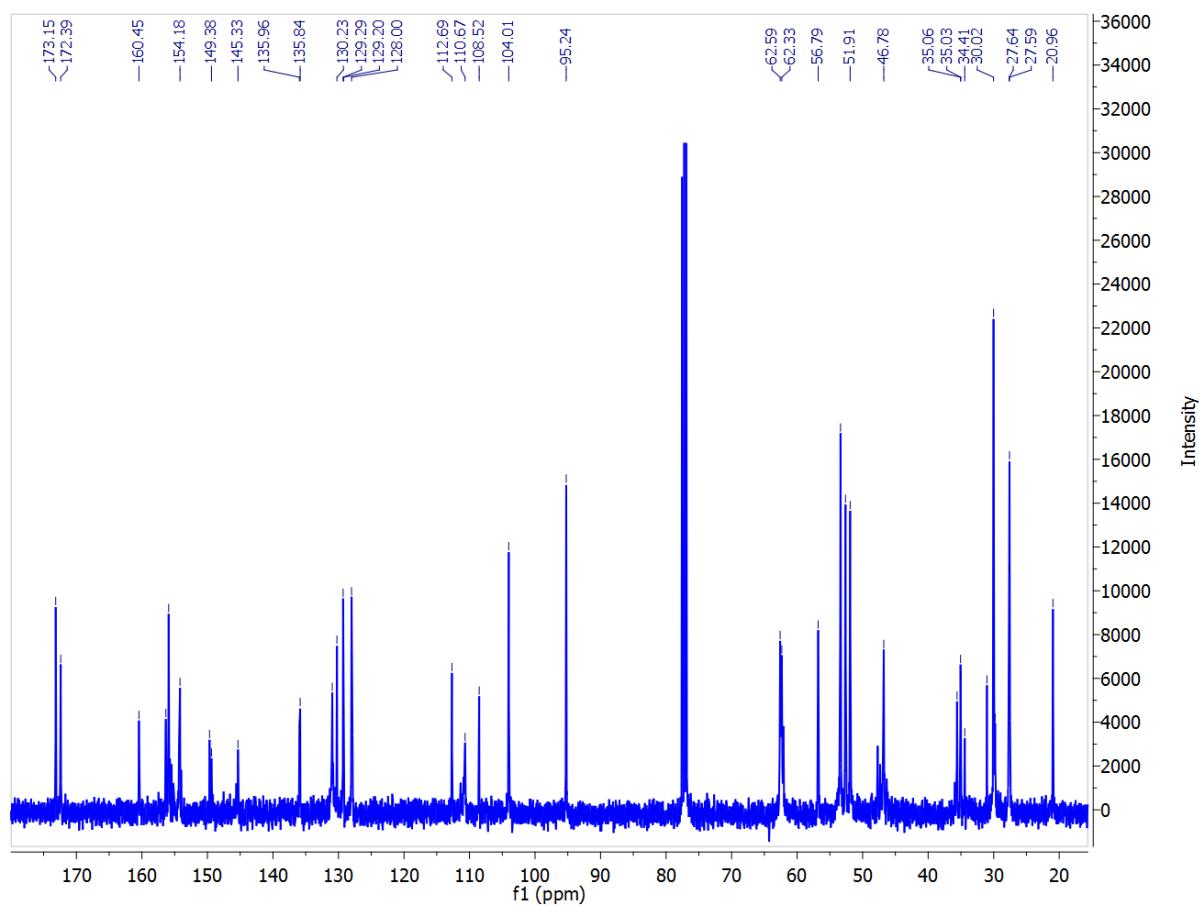
**Compound 4.**  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



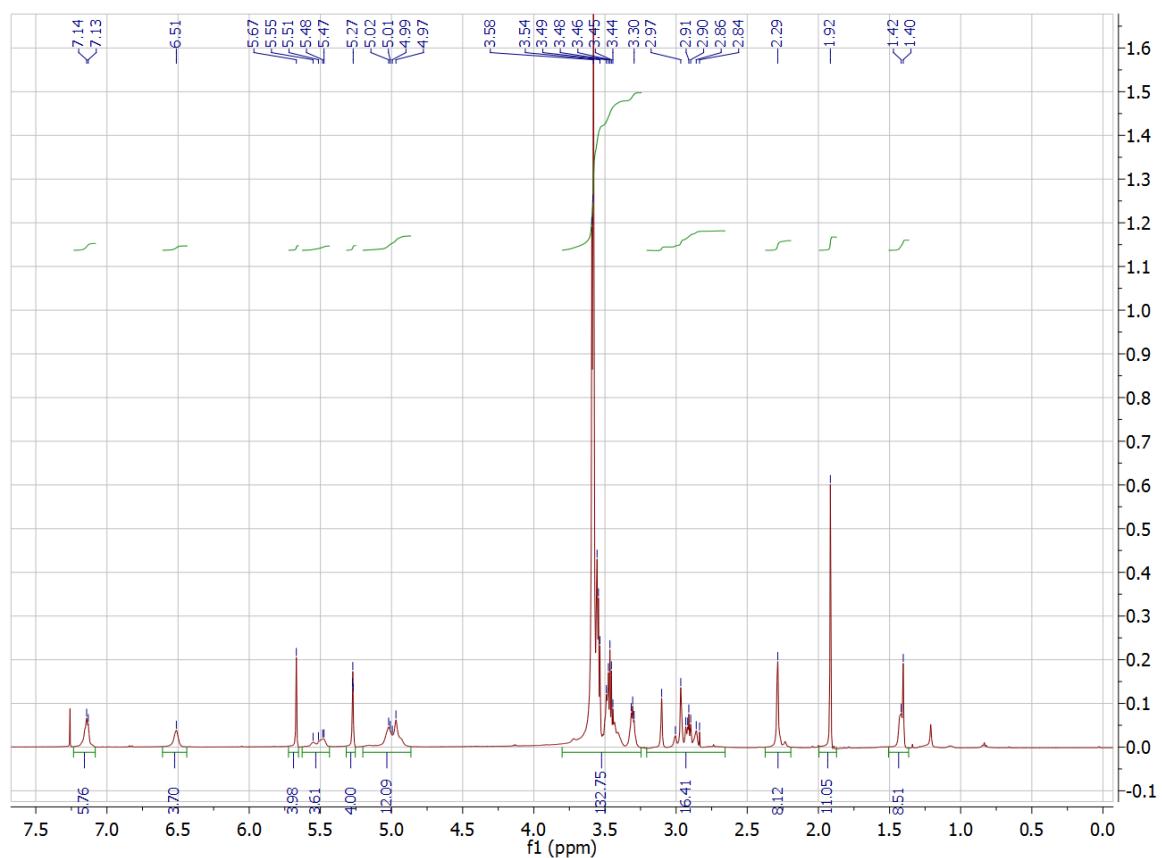
**Compound 5.**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )



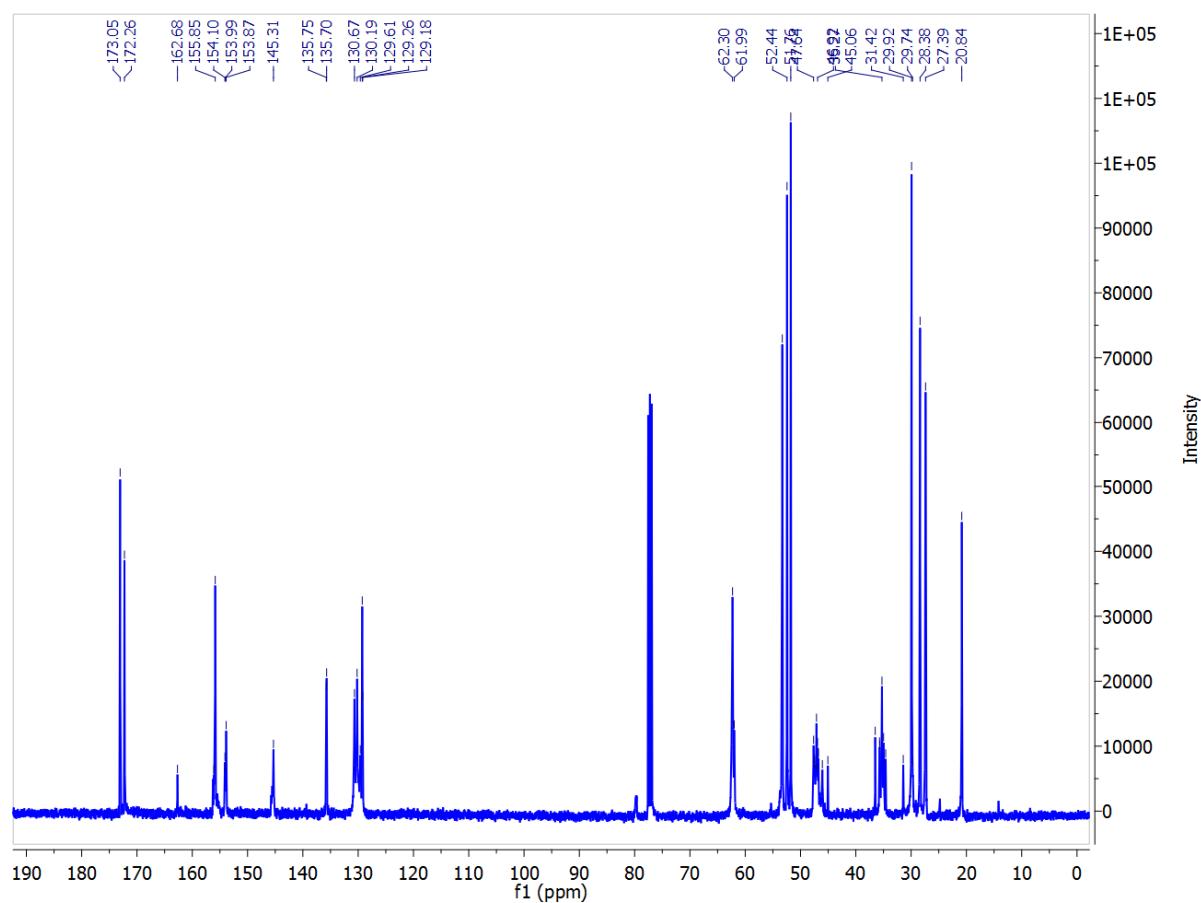
**Compound 5.**  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



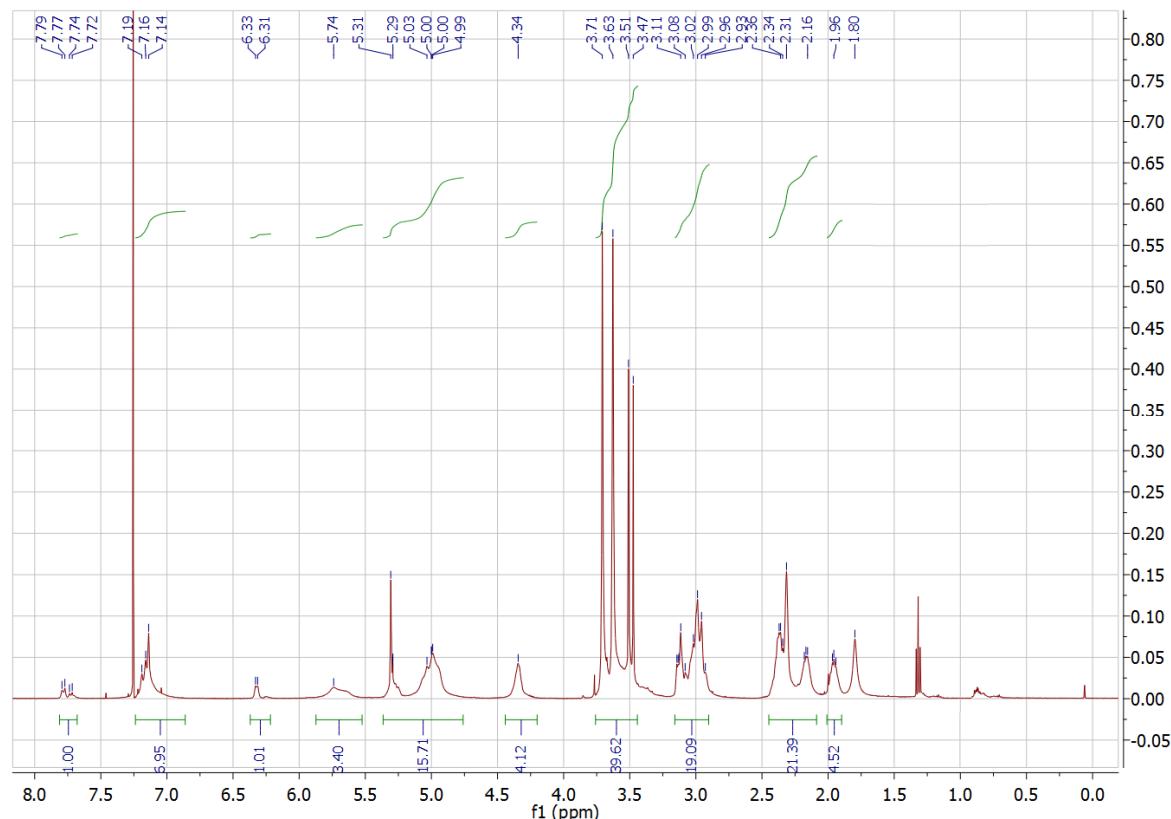
**Compound 6.**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )



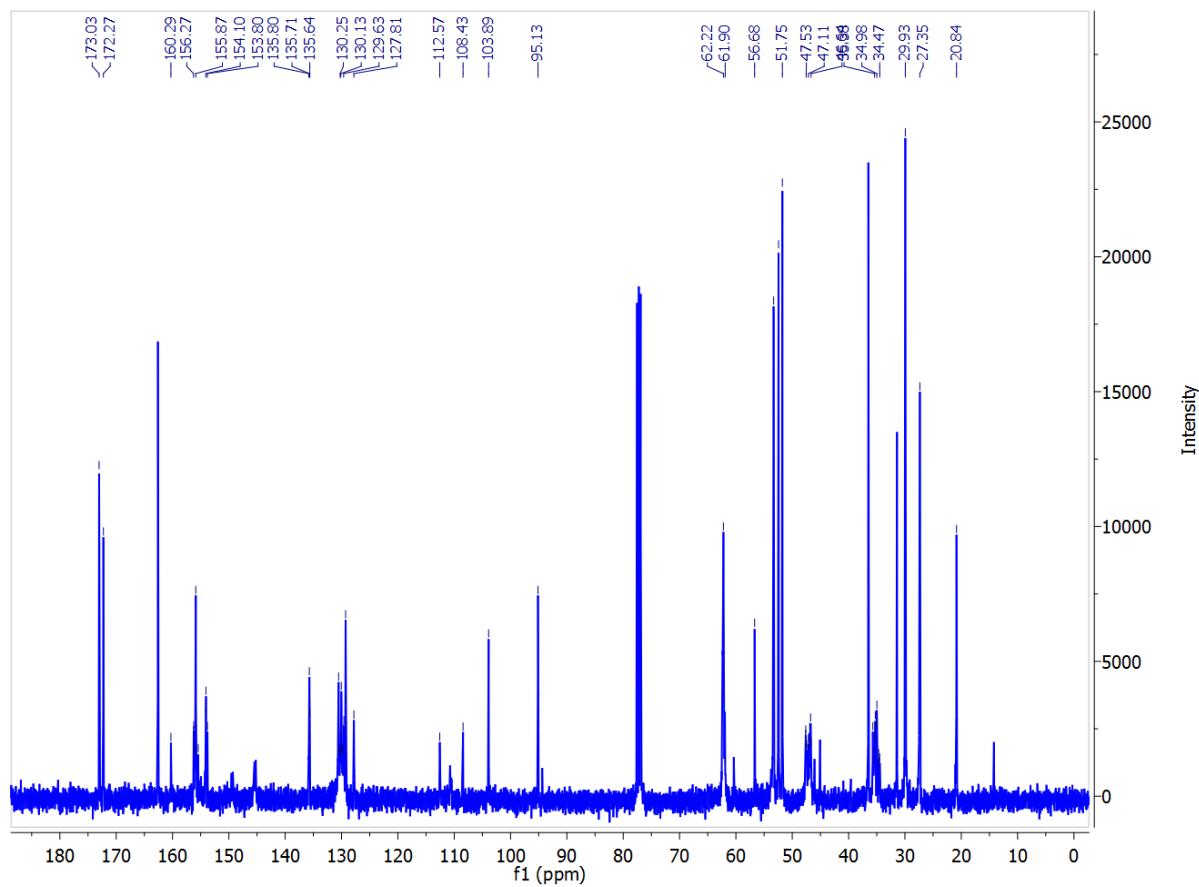
**Compound 6.**  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



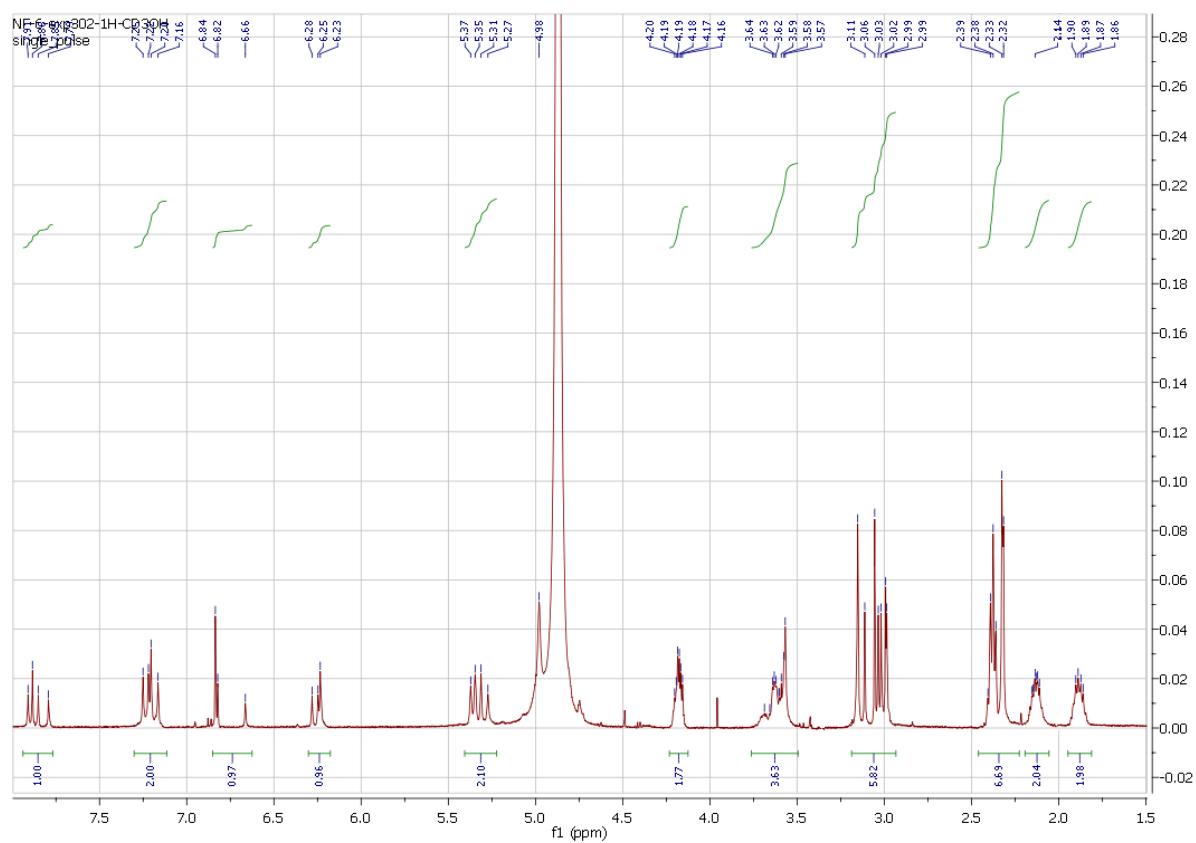
**Compound 7.**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )



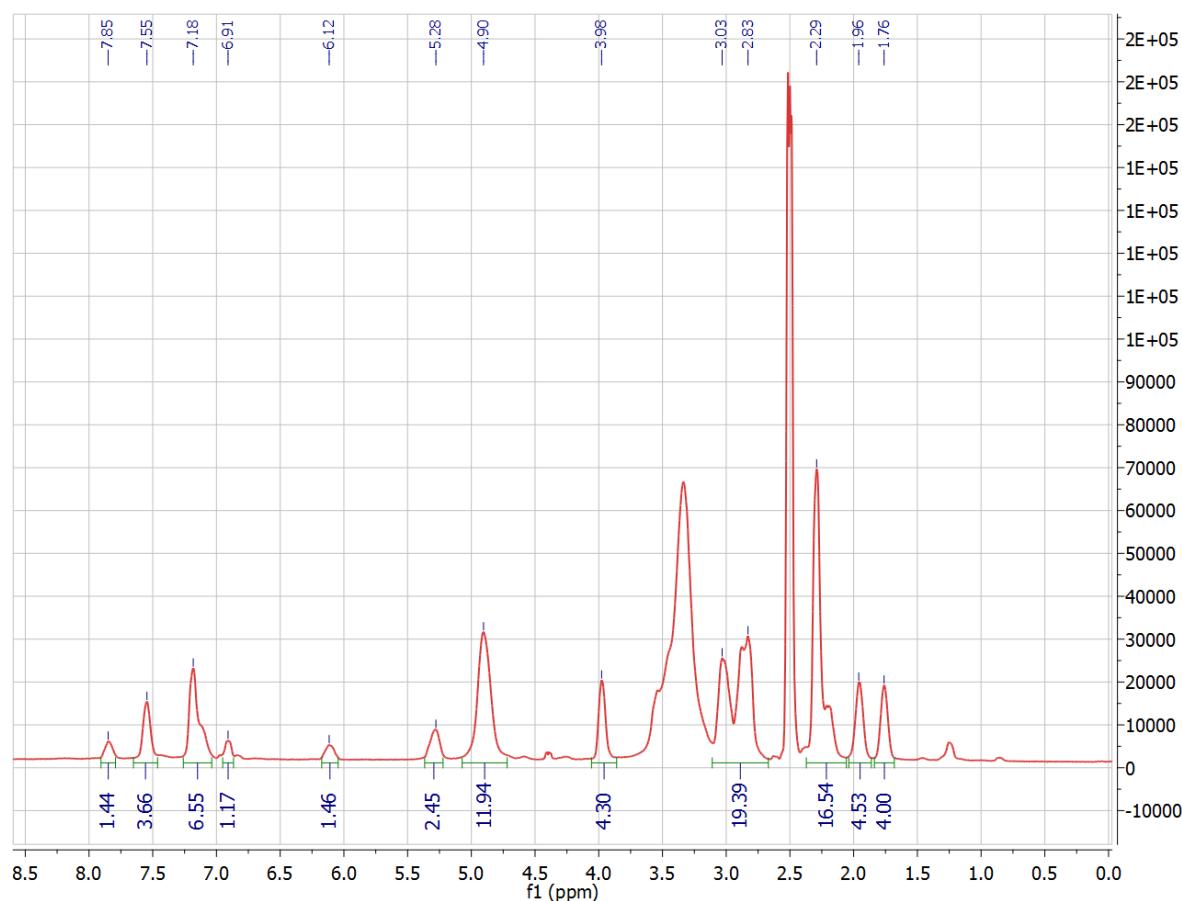
**Compound 7.**  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



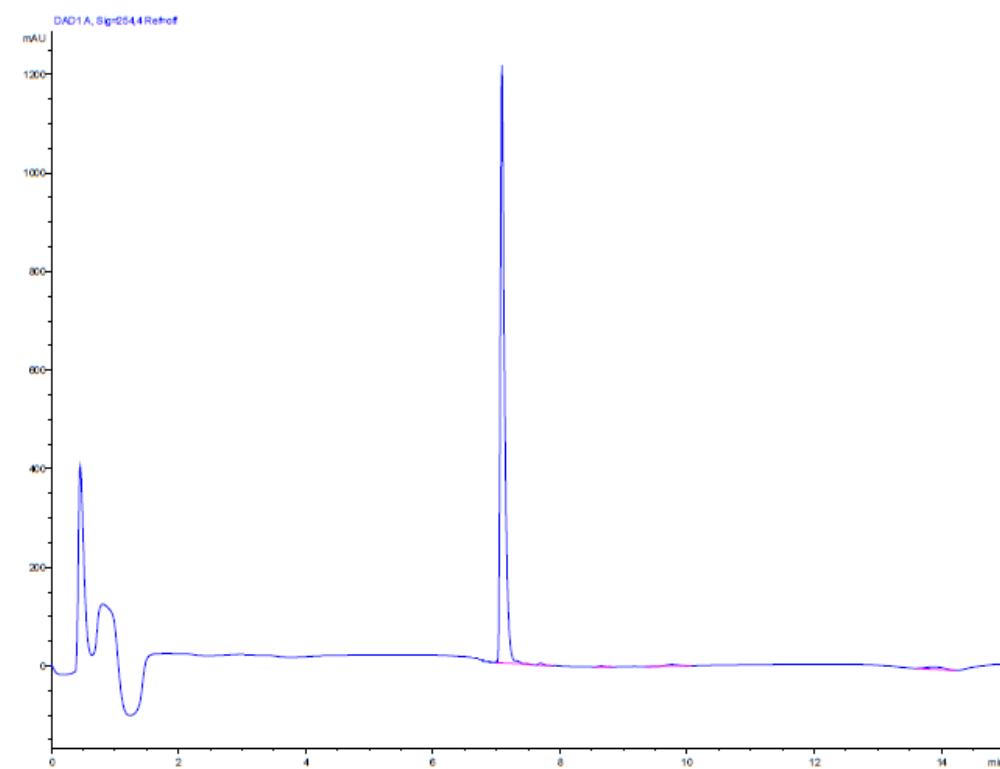
**G1.**  $^1\text{H}$  NMR (500 MHz,  $\text{CD}_3\text{OD}$ )



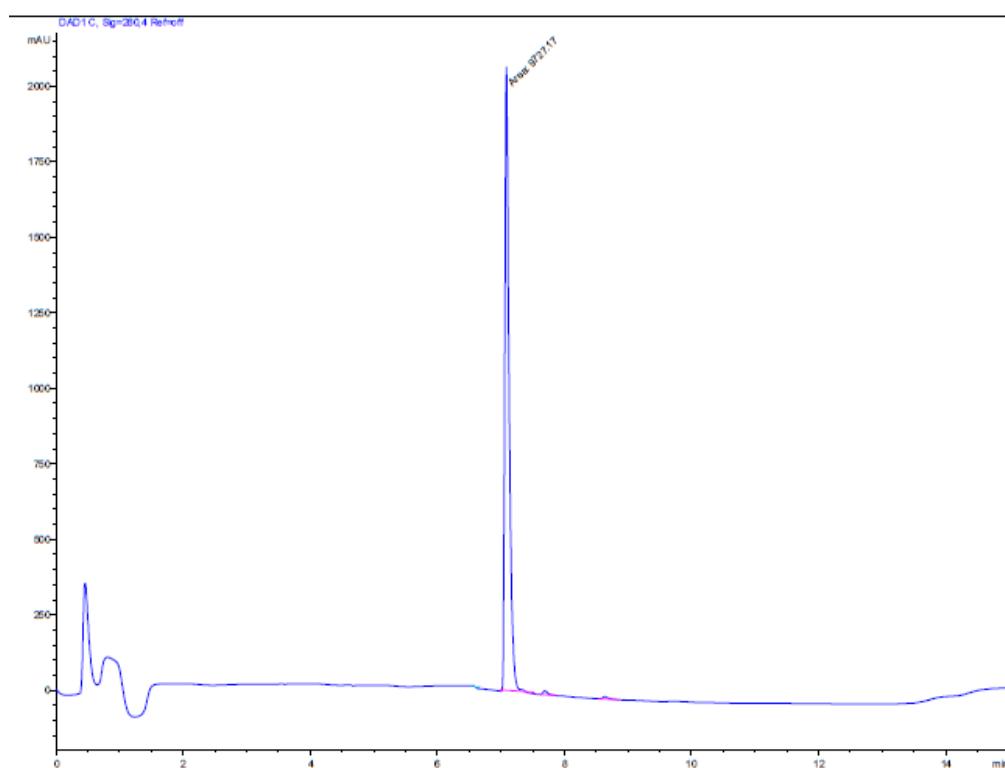
G2.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )



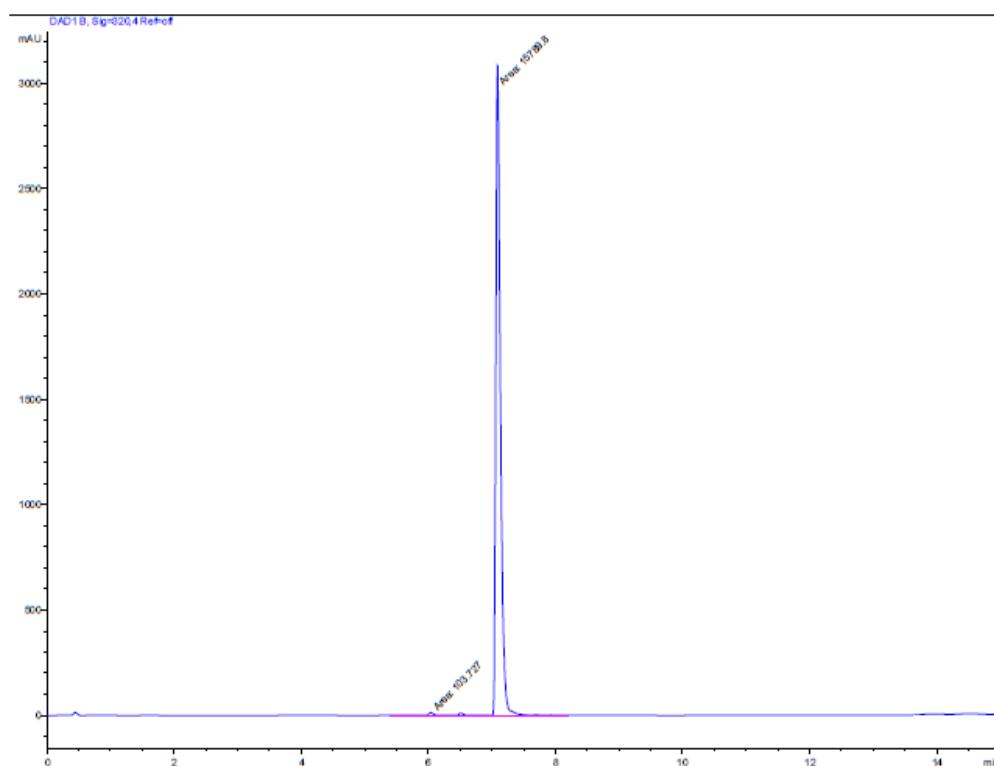
**Compound 2. HPLC trace (254 nm)**



**Compound 2. HPLC trace (280 nm)**



**Compound 2. HPLC trace (320 nm)**



## Compound 2. Integration of HPLC traces

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          Area Percent Report
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Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

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 1   7.087  BV R  0.0683  5638.49756 1212.57092  96.7723
 2   7.477  BB   0.0634   4.34185   1.06939   0.0745
 3   7.694  BB   0.0795   22.44258   4.26079   0.3852
 4   8.642  BB   0.1061   11.29974   1.64536   0.1939
 5   9.764  BV   0.2621   56.17157   3.34252   0.9641
 6  13.880  BB   0.3094  93.81101   4.94236   1.6101

Totals :           5826.56432 1227.83134

Signal 2: DAD1 B, Sig=320,4 Ref=off

Peak RetTime Type  Width      Area      Height     Area
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-----|-----|-----|-----|-----|-----|
 1   6.039  FM   0.1230  103.72677  14.05689   0.6488
 2   6.519  VV   0.1277   92.96394  11.32828   0.5815
 3   7.091  MF   0.0830  1.57898e4  3169.99609  98.7696

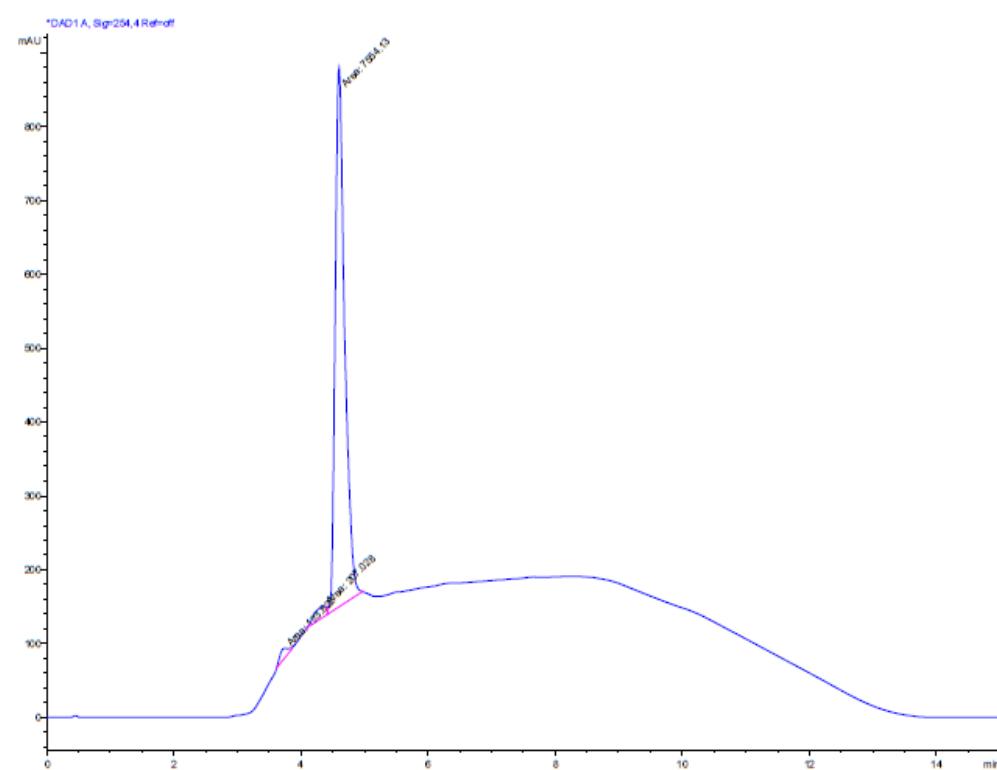
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Signal 3: DAD1 C, Sig=280,4 Ref=off

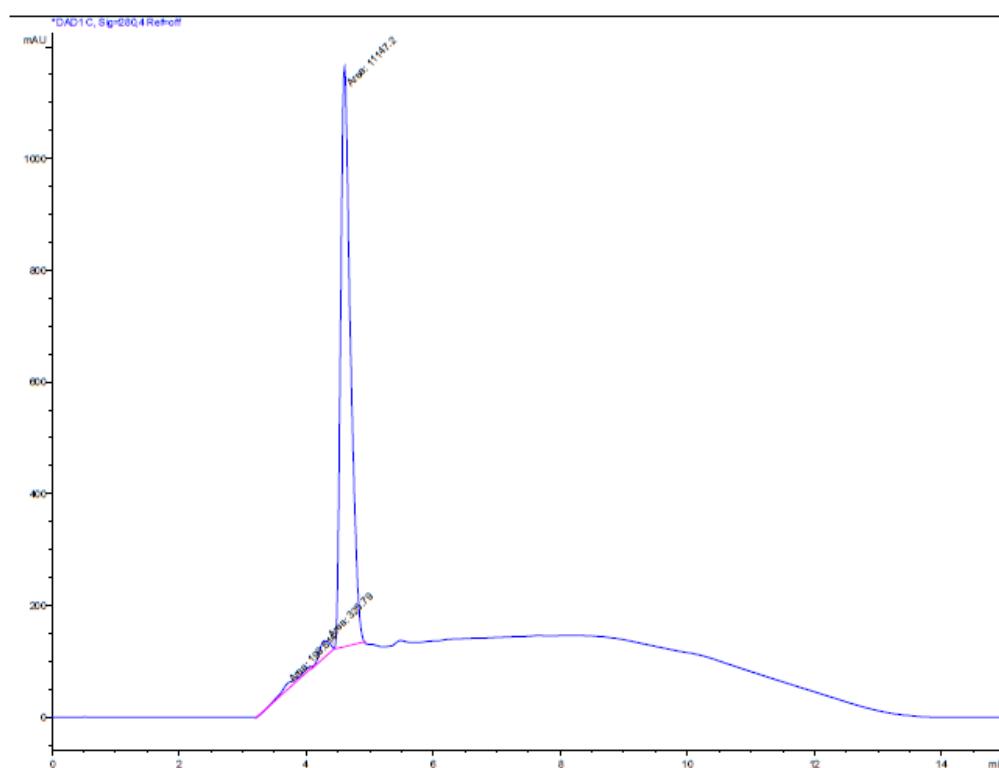
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 1   7.088  MM   0.0765  9727.17480 2119.04028  98.8601
 2   7.690  VV b  0.0947   66.47274   12.35918   0.6756
 3   8.642  BB   0.1045   45.68644   7.34858   0.4643

Totals :           9839.33399 2138.74805
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**Compound 4. HPLC trace (254 nm)**



**Compound 4. HPLC trace (280 nm)**



**Compound 4. Integration of HPLC traces.**

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          Area Percent Report
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Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
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 1  3.742 MM   0.1510  123.82155  13.66807  1.5703
 2  4.361 MF   0.2372  207.02815  14.54572  2.6256
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Totals :           7884.98349  758.57836

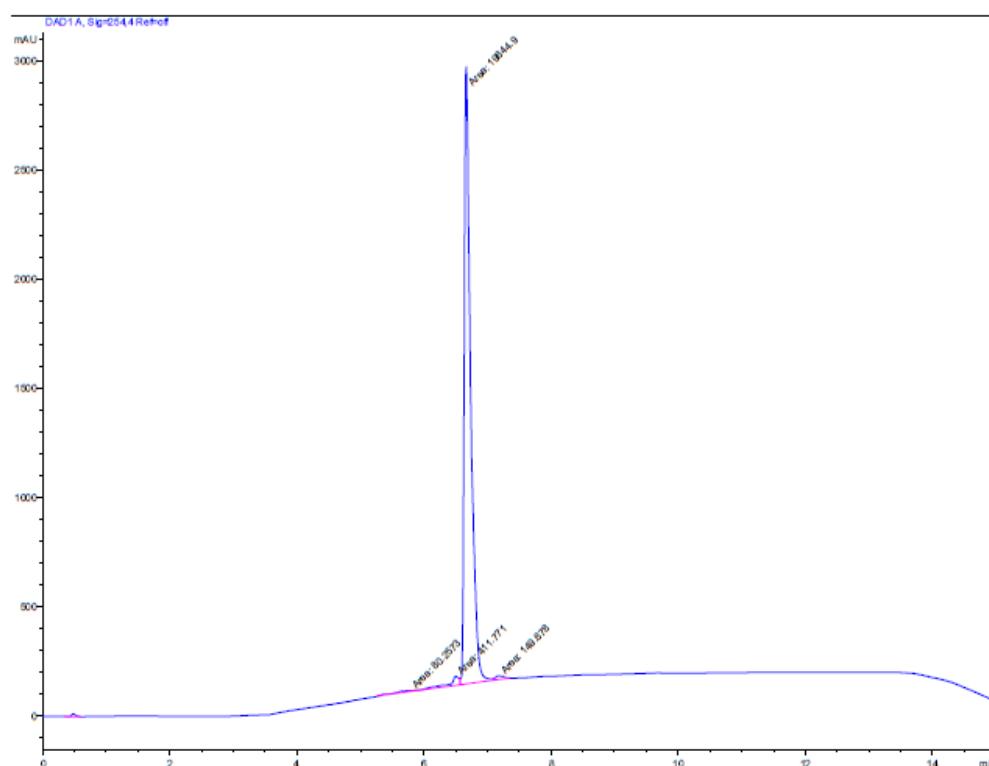
Signal 2: DAD1 C, Sig=280,4 Ref=off

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 2  4.304 MM   0.1850  325.79041  29.34939  2.7911
 3  4.606 MM   0.1783  1.11472e4  1041.94250  95.4988

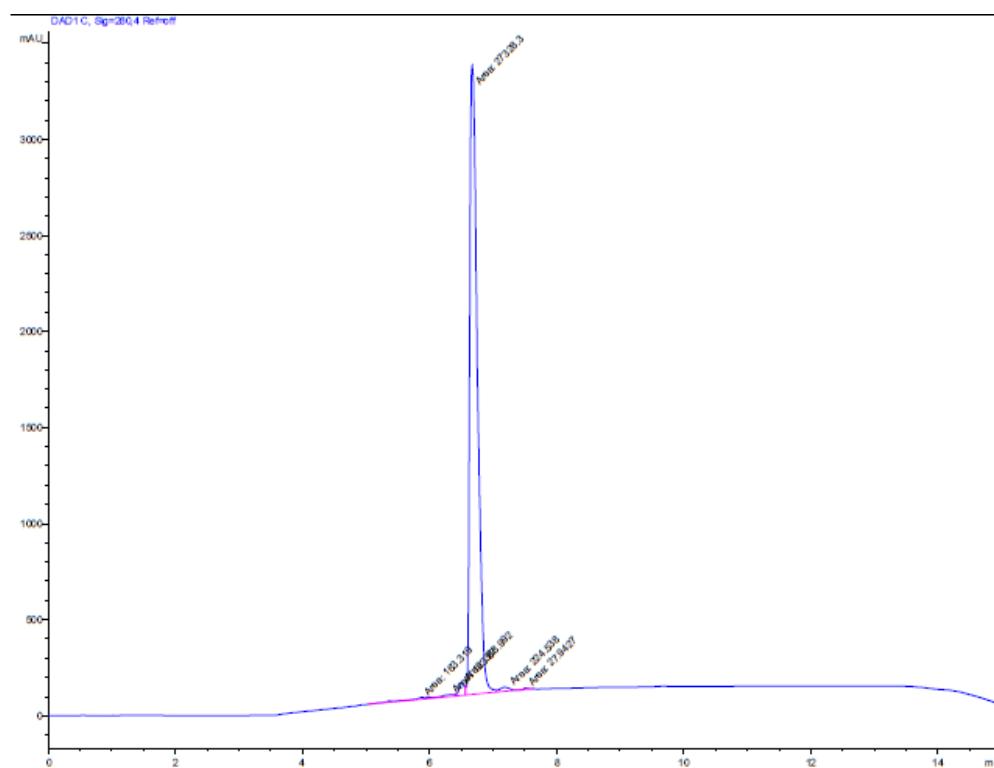
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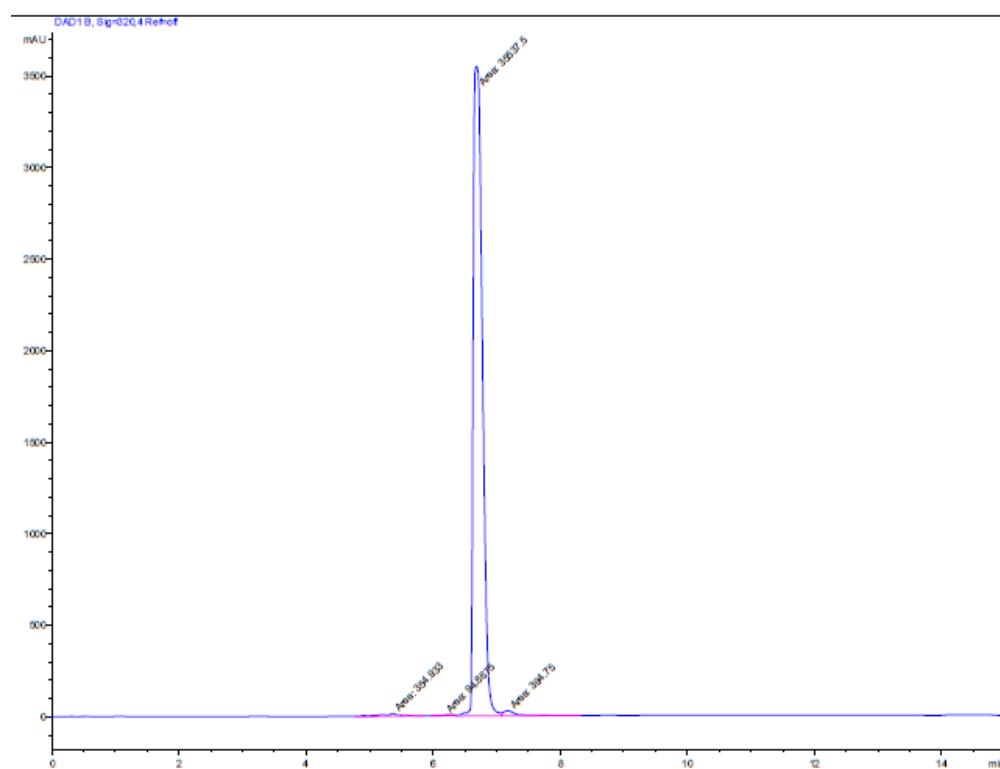
**Compound 5. HPLC trace (254 nm)**



**Compound 5. HPLC trace (280 nm)**



**Compound 5. HPLC trace (320 nm)**



### Compound 5. Integration of HPLC traces.

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          Area Percent Report
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Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width     Area      Height     Area
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 1  0.483  VB   0.0729  65.35059 12.94502  0.3211
 2  5.777  MF   0.4038  80.25726  3.31254  0.3943
 3  6.500  FM   0.1687  411.77136 40.68394  2.0232
 4  6.666  FM   0.1157  1.96449e4 2829.31128 96.5259
 5  7.176  FM   0.1469  149.67824 16.97940  0.7354

Totals :           2.03520e4  2903.23219

Signal 2: DAD1 B, Sig=320,4 Ref=off

Peak RetTime Type  Width     Area      Height     Area
# [min]       [min] [mAU*s] [mAU]     %
-----|-----|-----|-----|-----|-----|
 1  5.360  MM   0.3675  354.93338 16.09779  0.9756
 2  6.167  MF   0.2190  94.86752  7.22114  0.2608
 3  6.681  FM   0.1661  3.55375e4 3566.51978 97.6787
 4  7.173  FM   0.2498  394.75034 26.33272  1.0850

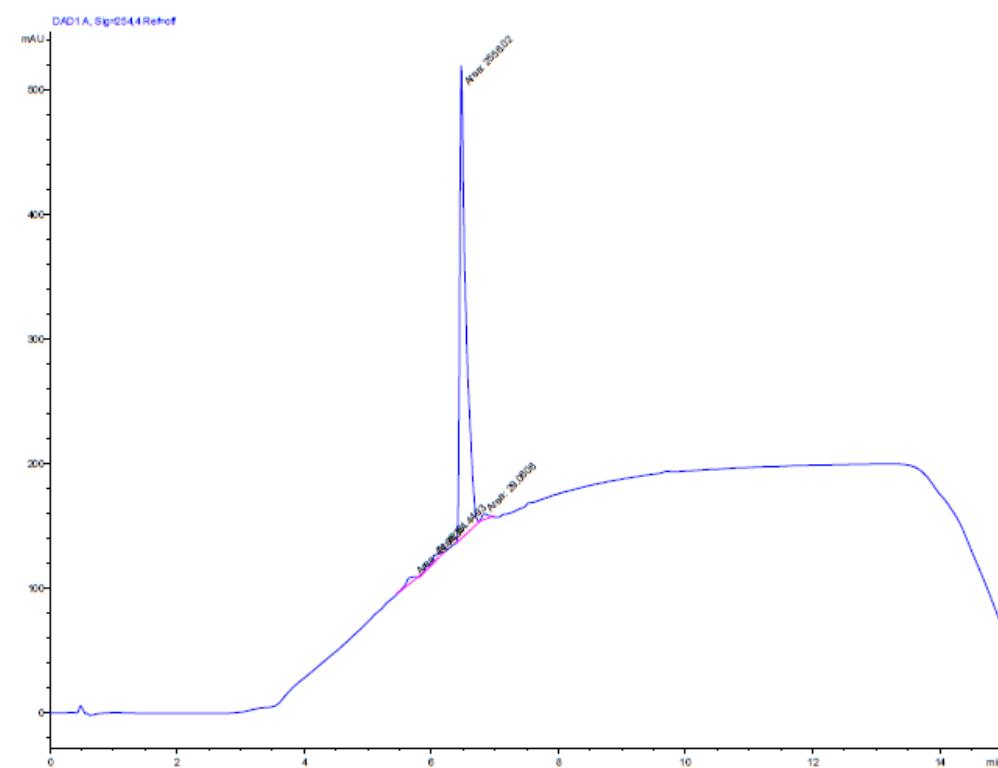
Totals :           3.63820e4  3616.17143

Signal 3: DAD1 C, Sig=280,4 Ref=off

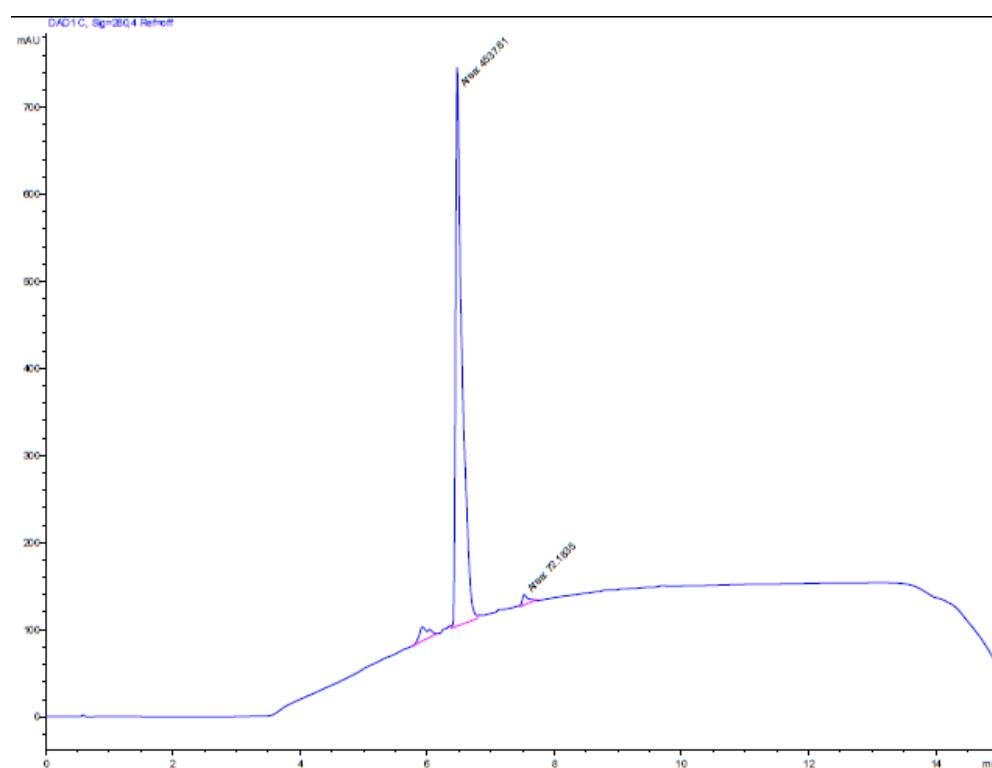
Peak RetTime Type  Width     Area      Height     Area
# [min]       [min] [mAU*s] [mAU]     %
-----|-----|-----|-----|-----|-----|
 1  5.877  MF   0.3279  163.31876 8.30139  0.5780
 2  6.325  FM   0.2284  123.86954 9.03941  0.4384
 3  6.503  FM   0.0979  388.99210 66.23907  1.3766
 4  6.677  FM   0.1349  2.73283e4 3375.37354 96.7135
 5  7.239  MF   0.1814  224.53767 20.63080  0.7946
 6  7.517  FM   0.0600  27.94269  7.76740  0.0989

Totals :           2.82570e4  3487.35160
```

**Compound 6. HPLC trace (254 nm)**



**Compound 6. HPLC trace (280 nm)**



## Compound 6. Integration of HPLC traces.

```
=====
          Area Percent Report
=====

Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|-----|
 1   5.727 MM    0.1874   44.76364   3.02000   1.6676
 2   6.047 MM    0.1731   54.44934   5.24206   2.0284
 3   6.475 MM    0.1118  2556.02197  381.11389  95.2213
 4   6.838 MM    0.1037   29.06061   4.66866   1.0826

Totals :           2684.29557  394.04461

Signal 2: DAD1 B, Sig=320,4 Ref=off

Signal 3: DAD1 C, Sig=280,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|-----|
 1   5.967 BB    0.2055  156.09244  12.87076   3.2752
 2   6.477 MM    0.1139  4537.61133  664.23846  95.2102
 3   7.530 MM    0.0960   72.18346  12.52650   1.5146

Totals :           4765.88722  689.63572

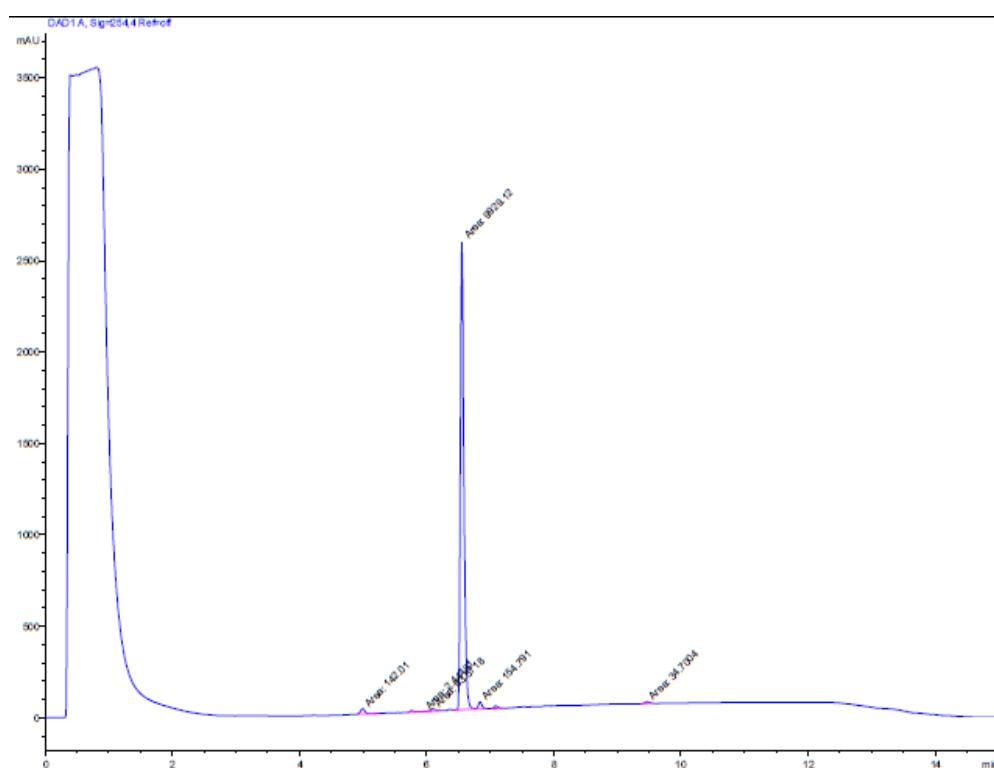
Signal 4: MSD1 TIC, MS File

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|-----|
 1   6.566 BB    0.2446  5.07602e6  2.73966e5  100.0000

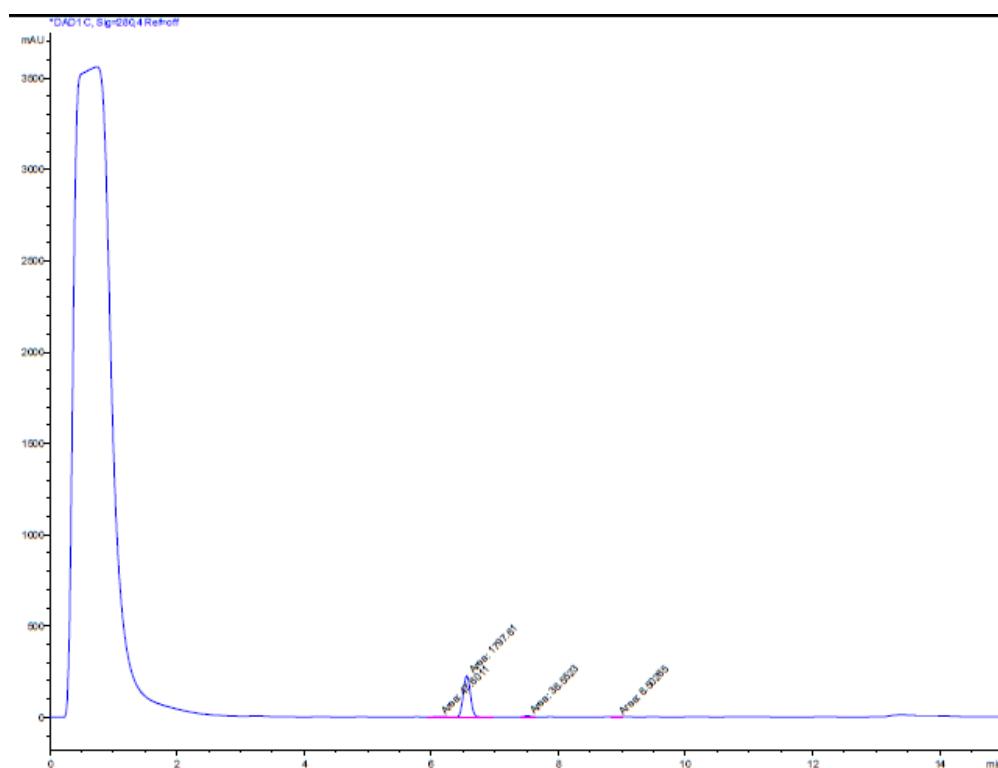
Totals :           5.07602e6  2.73966e5

=====
*** End of Report ***
```

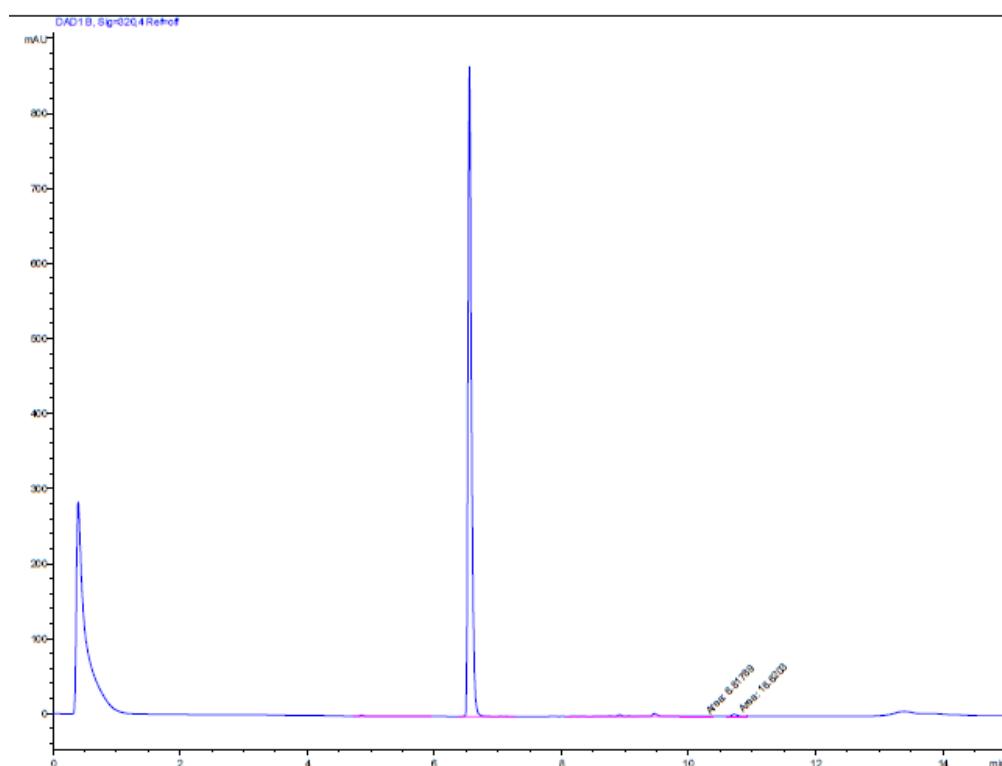
**Compound 7. HPLC trace (254 nm)**



**Compound 7. HPLC trace (280 nm)**



**Compound 7. HPLC trace (320 nm)**



### Compound 7. Integration of HPLC traces.

```
=====
          Area Percent Report
=====

Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width      Area      Height     Area
# [min]       [min] [mAU*s]   [mAU]     %
-----|-----|-----|-----|-----|-----|
 1  4.995 FM    0.0795 142.01025 29.76386  1.3629
 2  5.212 BV    0.0551  3.91492  1.11138  0.0376
 3  5.766 BB    0.0569 33.42433  8.67672  0.3208
 4  5.945 MF T  0.0773 2.44491 5.27159e-1 0.0235
 5  6.095 FM T  0.0789 63.57177 13.42993  0.6101
 6  6.554 MF    0.0645 9929.12207 2566.26416 95.2911
 7  6.840 FM    0.0647 154.79086 39.89325  1.4855
 8  7.086 BB    0.0572  55.79729 14.40436  0.5355
 9  9.465 MM    0.0630 34.70042  9.17674  0.3330

Totals :           1.04198e4 2683.24757

Signal 2: DAD1 B, Sig=320,4 Ref=off

Peak RetTime Type  Width      Area      Height     Area
# [min]       [min] [mAU*s]   [mAU]     %
-----|-----|-----|-----|-----|
 1  4.866 BV    0.2177 22.28537 1.40016  0.6595
 2  6.559 BB    0.0885 3280.91870 677.80762 97.0903
 3  8.921 VV    0.1580 25.80622  2.38902  0.7637
 4  9.474 VV    0.1189 26.79691  3.96449  0.7930
 5  10.230 MF   0.2962  6.81789 3.83645e-1 0.2018
 6  10.754 MF   0.0959 16.62031  2.88862  0.4918

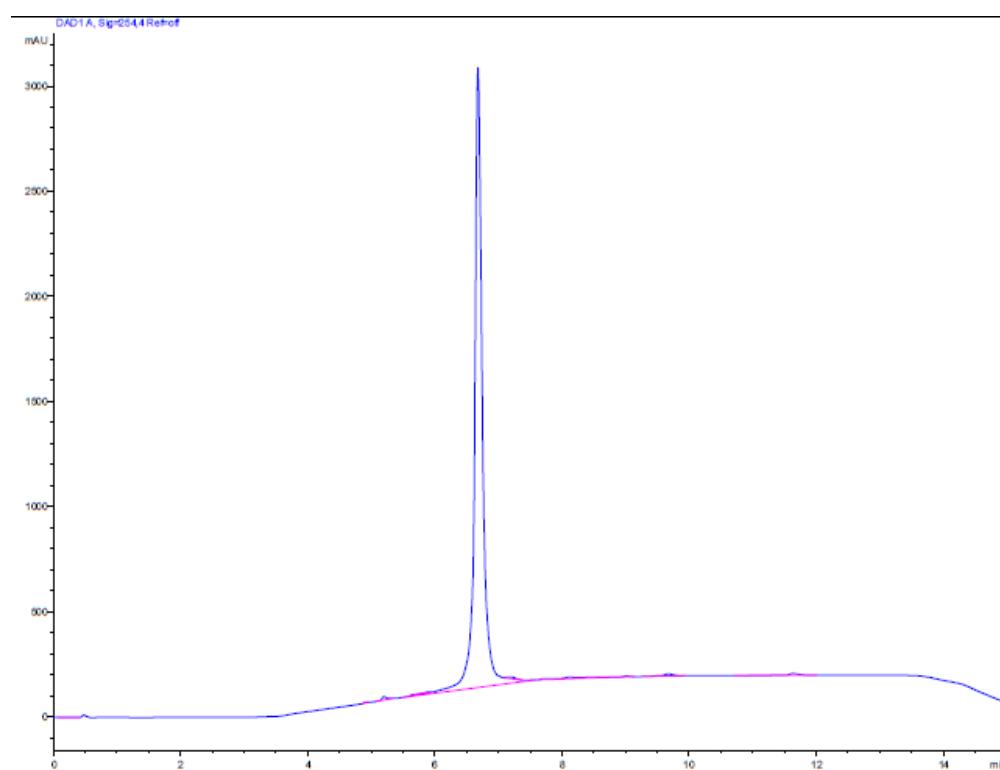
Totals :           3379.24540 688.83354

Signal 3: DAD1 C, Sig=280,4 Ref=off

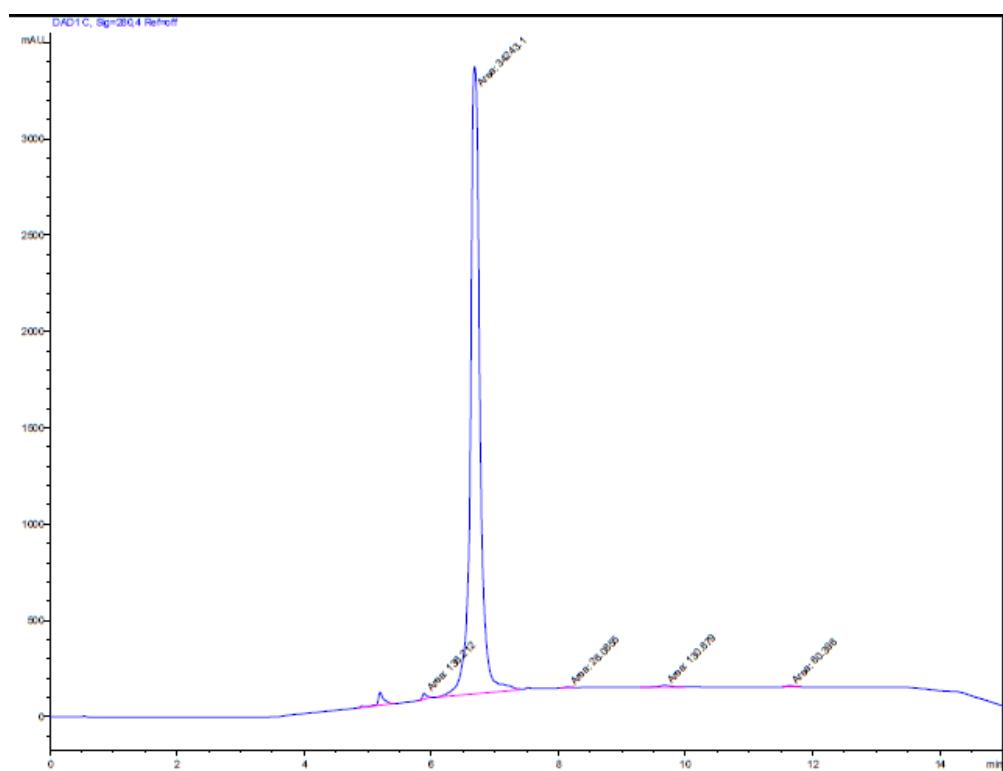
Peak RetTime Type  Width      Area      Height     Area
# [min]       [min] [mAU*s]   [mAU]     %
-----|-----|-----|-----|-----|
 1  6.113 MM    0.1756 47.60110 4.51731  2.5209
 2  6.560 MM    0.1314 1797.60681 228.07796 95.1990
 3  7.517 MM    0.1100 36.55231  5.53989  1.9358
 4  8.924 MM    0.0874  6.50265  1.24025  0.3444

Totals :           1888.26287 239.37542
```

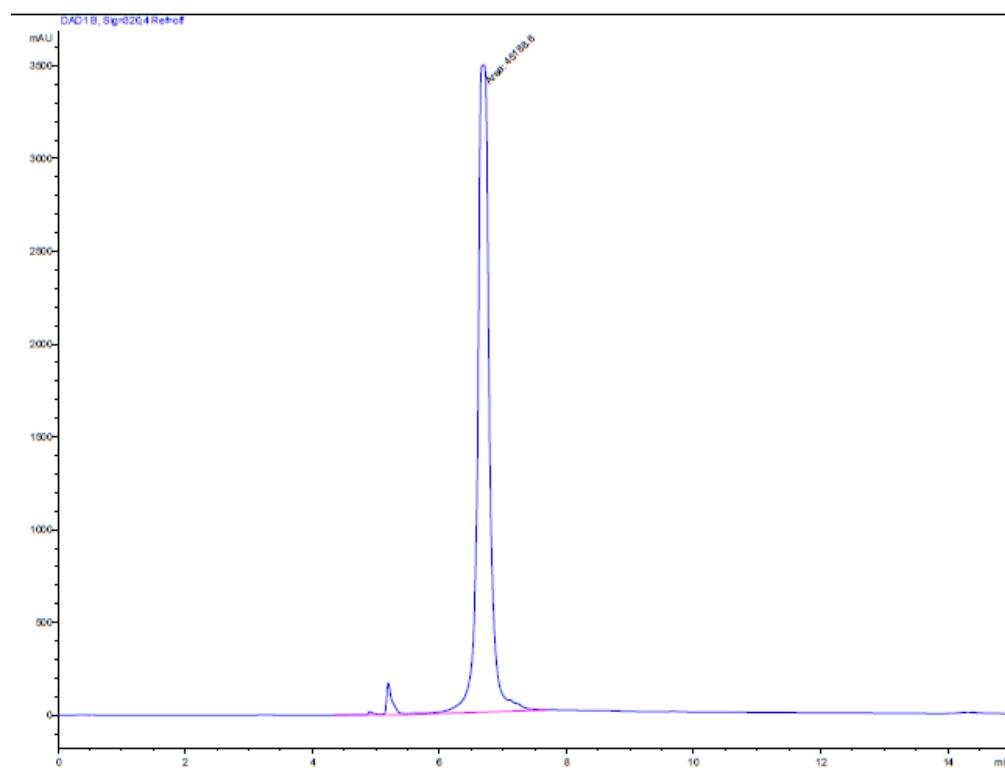
**G1. HPLC trace (254 nm)**



**G1. HPLC trace (280 nm)**



**G1. HPLC trace (320 nm)**



## G1. Integration of HPLC traces.

```
=====
          Area Percent Report
=====

Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width      Area      Height     Area
#   [min]        [min]    [mAU*s]    [mAU]      %
-----|-----|-----|-----|-----|-----|
 1   0.313  BV    0.1686   18.37831   1.36913   0.0703
 2   4.915  BB    0.0511   11.90346   3.73861   0.0455
 3   5.198  BB    0.0741   91.52818  17.75304   0.3502
 4   5.660  BV T  0.0535   14.56638   4.30759   0.0557
 5   5.891  VV T  0.0927   20.83250   3.08836   0.0797
 6   6.679  VV R  0.1265   2.54079e4  2949.79663  97.2127
 7   7.182  VV T  0.1476   82.89394   8.51596   0.3172
 8   7.725  BV    0.1011   29.98930   4.42265   0.1147
 9   7.890  VV    0.0906   23.02784   3.50618   0.0881
10   8.131  VV    0.2017   96.51692   6.71503   0.3693
11   8.548  VV    0.2108   54.76419   3.21435   0.2095
12   9.013  VB    0.2111   67.22124   4.14828   0.2572
13   9.673  BV    0.1715   131.66780  10.55944   0.5038
14  11.634  BB    0.1405   85.21042   8.68275   0.3260

Totals :           2.61364e4  3029.81800

Signal 2: DAD1 B, Sig=320,4 Ref=off

Peak RetTime Type  Width      Area      Height     Area
#   [min]        [min]    [mAU*s]    [mAU]      %
-----|-----|-----|-----|-----|
 1   4.436  BB    0.1192   9.24432   1.36189   0.0199
 2   4.926  BV    0.1049  110.15494  17.62743   0.2373
 3   5.215  VV    0.1234  1106.07068  154.75166   2.3830
 4   6.686  MM    0.2159  4.51886e4  3488.65259  97.3597

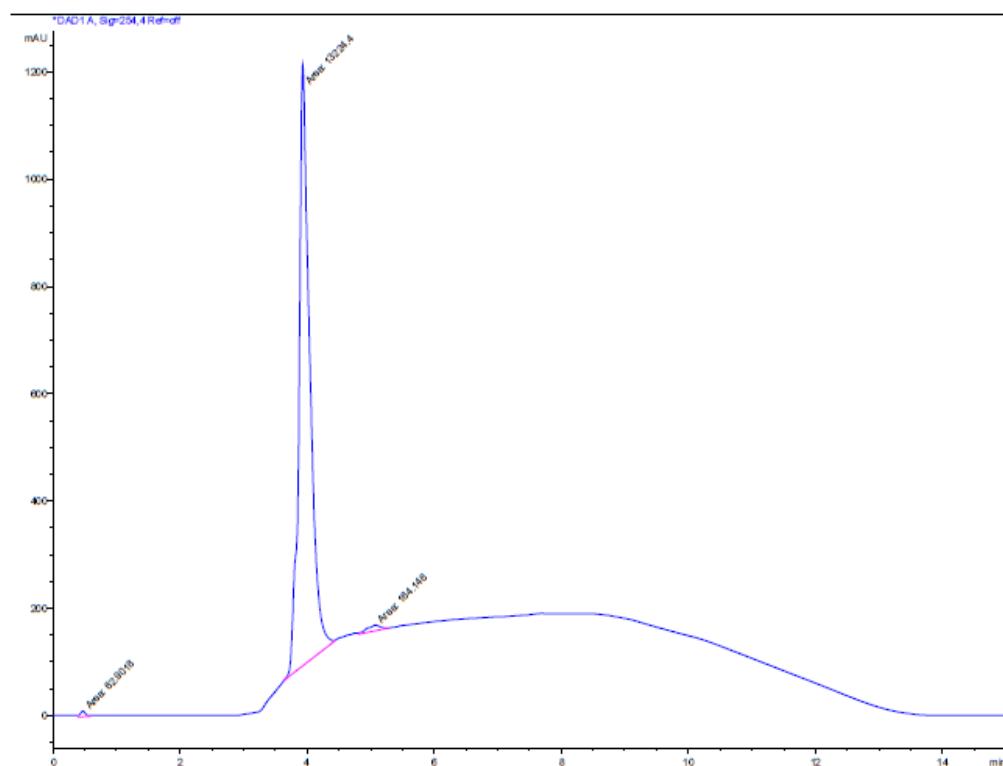
Totals :           4.64141e4  3662.39358

Signal 3: DAD1 C, Sig=280,4 Ref=off

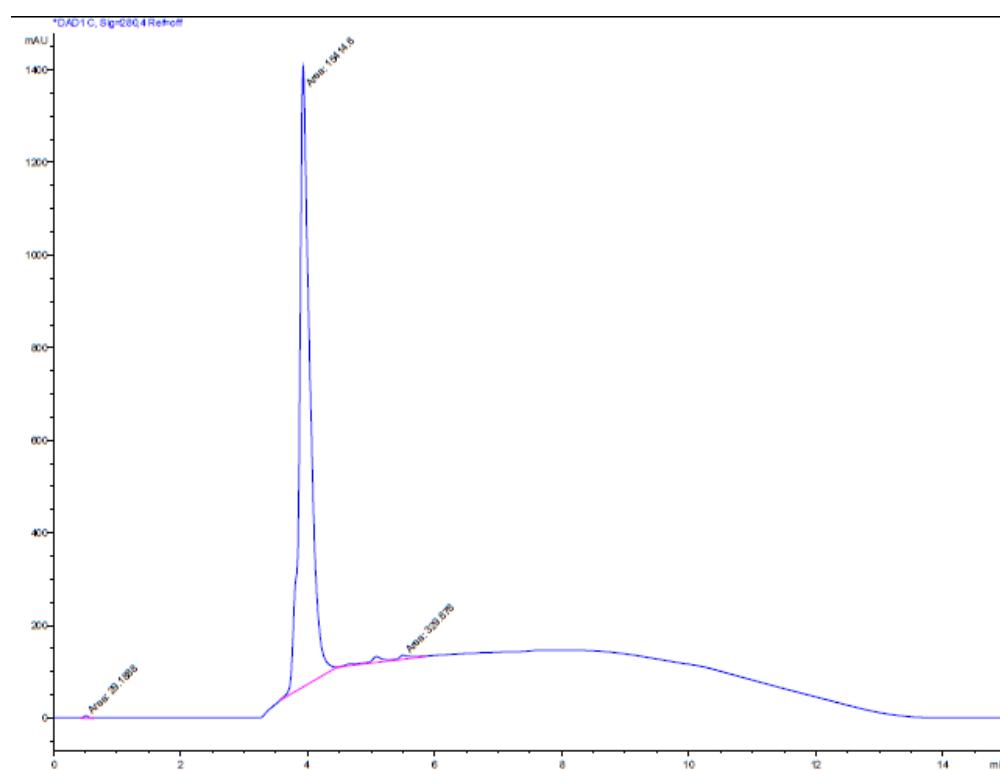
Peak RetTime Type  Width      Area      Height     Area
#   [min]        [min]    [mAU*s]    [mAU]      %
-----|-----|-----|-----|-----|
 1   5.217  BB    0.1277  449.18762  59.75112   1.2817
 2   5.891  MM    0.0695  136.21172  32.66082   0.3887
 3   6.685  MM    0.1719  3.42431e4  3320.40356  97.7100
 4   8.152  MM    0.0940  26.08551   4.62395   0.0744
 5   9.671  MM    0.2180  130.67891  9.99114   0.3729
 6  11.637  MM    0.1353  60.39601   7.44215   0.1723

Totals :           3.50457e4  3434.87274
```

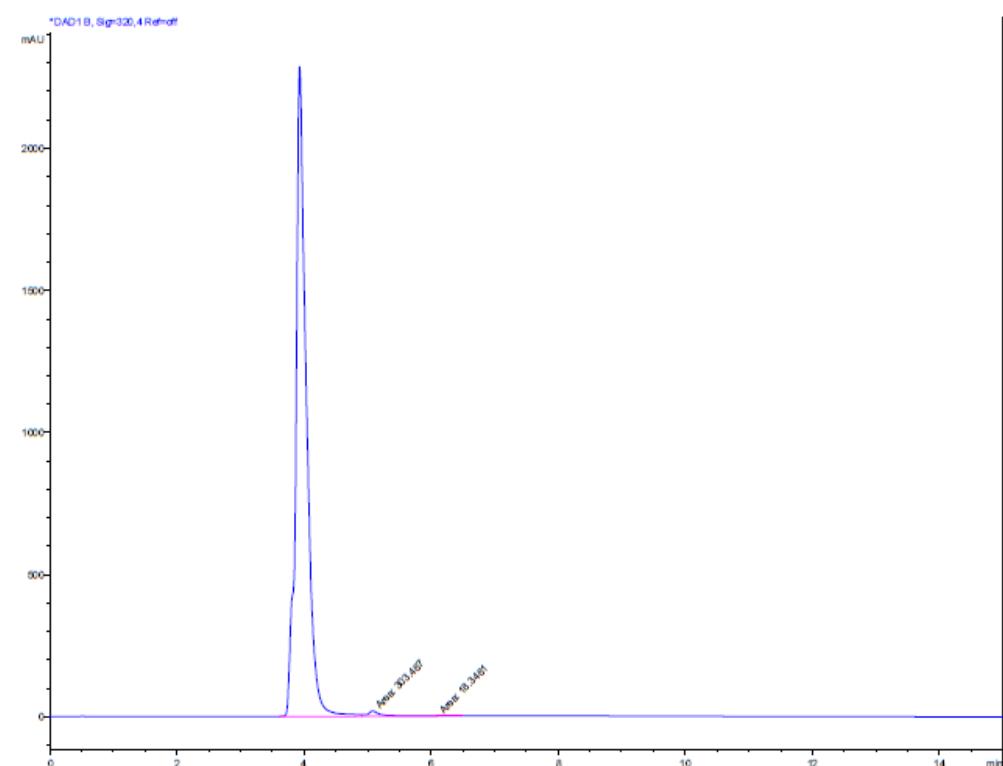
**G2. HPLC trace (254 nm)**



**G2. HPLC trace (280 nm)**



**G2. HPLC trace (320 nm)**



## G2. Integration of HPLC traces.

```
=====
          Area Percent Report
=====

Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:     :      1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|-----|
 1   0.472 MM    0.1004   62.90178  10.44576  0.4676
 2   3.932 MM    0.1966  1.32244e4  1121.18335 98.3121
 3   5.078 MM    0.2512   164.14577  10.89043  1.2203

Totals :           1.34514e4  1142.51954

Signal 2: DAD1 B, Sig=320,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|
 1   3.949 BV    0.1935  2.60469e4  2089.13403 98.7796
 2   5.081 MF    0.2641   303.46747  19.15088  1.1509
 3   6.088 FM    0.2384   18.34607   1.00418  0.0696

Totals :           2.63688e4  2109.28909

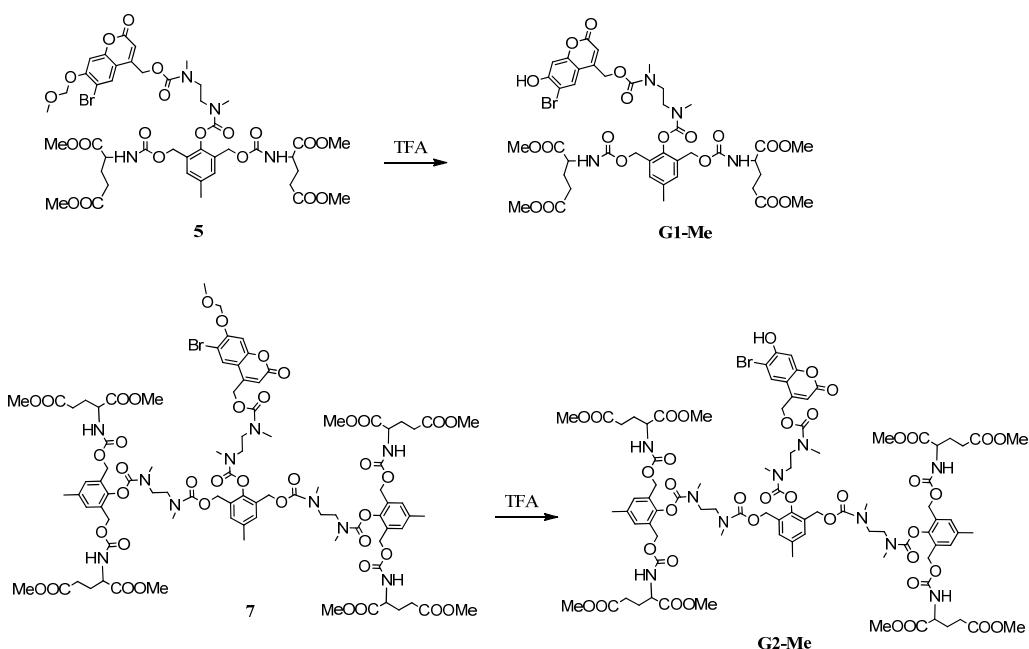
Signal 3: DAD1 C, Sig=280,4 Ref=off

Peak RetTime Type  Width      Area      Height      Area
#   [min]       [min]   [mAU*s]   [mAU]      %
-----|-----|-----|-----|-----|
 1   0.515 MM    0.0925   29.18877   5.25780  0.1850
 2   3.935 MM    0.1898  1.54146e4  1353.60974 97.7249
 3   5.507 MM    0.5934   329.67557   9.25875  2.0901

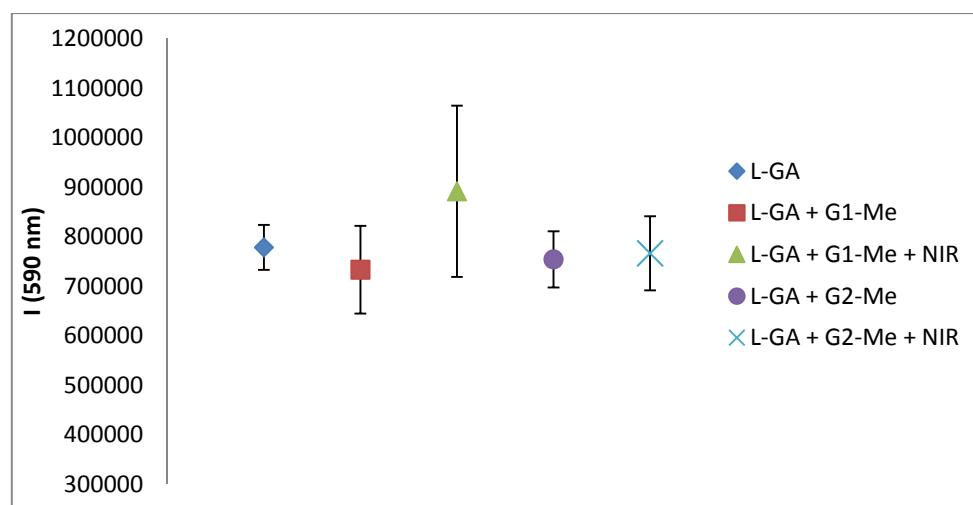
Totals :           1.57735e4  1368.12629
```

**Control experiment to evaluate the influence of the byproducts of dendrimers' degradation on the Amplex Red assay:**

Compounds **5** and **7** were treated with TFA in dry DCM for 30 min to remove the methoxymethyl protecting group (Scheme S1). The solvents were removed on rotary evaporator. The 1 mM solutions of **G1-Me** and **G2-Me** in acetonitrile/PBS pH 7.4 (1/2) were irradiated with NIR light for 30 min. After that, the irradiated and the non-irradiated control solutions were diluted to 3.5  $\mu$ M (for **G1-Me**) and 1.75  $\mu$ M (for **G2-Me**) with PBS pH 7.4 and incubated with 2 nM of *L*-glutamic acid at 37°C for 96 hours before analysis using the Amplex Red assay. The amount of *L*-glutamic acid detected in the presence of both irradiated and non-irradiated solutions of **G1-Me** and **G2-Me** was the same as in the pristine solution of *L*-glutamic acid (Figure S1). The experiment was performed in triplicates.



**Scheme S1.** Synthesis of **G1-Me** and **G2-Me**.



**Figure S1.** Detection of *L*-glutamic acid in the presence of **G1-Me** and **G2-Me**.

References:

1. N. Fomina, McFearin, C. L., Sermsakdi, M., Morachis, J. M., Almutairi, A., *Macromolecules*, 2011, **44**, 8590-8597.
2. T. Furuta, S. S. H. Wang, J. L. Dantzer, T. M. Dore, W. J. Bybee, E. M. Callaway, W. Denk and R. Y. Tsien, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 1193-1200.