Supporting Information

for

Carbohydrate Functionalized Iron(III) complexes as Biomimetic Siderophores

Rohan Yadav^a and Raghavendra Kikkeri^{a,*}

^a Department of Chemistry, Indian Institute of Science Education and Research, Pune-411021

*Correspondence should be address to R.K. (<u>rkikkeri@iiserpune.ac.in</u>).

Table of Contents:

- 1. General Information
- 2. General Procedure
- 3. Synthesis of ligand 10 and 13
- 4. Synthesis of ligand 19 and 20.
- 5. Synthesis of dendrons 25 and 26.
- 6. Ligand titration with iron(III) ion.
- 7. Tubidimetric analysis
- 8. Imaging experiment
- 9. Bacterial growth and 96-well growth promotion assay
- 10. LB plate assay
- **11. Additional Reference**

12. ¹H, ¹³C-NMR, Mass Spectrometry Data of all the Compounds

1. General Information

All chemicals used were reagent grade and used as supplied except where noted. Analytical thin layer chromatography (TLC) was performed on Merck silica gel 60 F254 plates (0.25 mm). Compounds were visualized by UV irradiation or dipping the plate in CAN solution followed by

heating. Column chromatography was carried out using force flow of the indicated solvent on Fluka Kieselgel 60 (230–400 mesh). ¹H and ¹³C NMR spectra were recorded on Bruker 500 MHz (or 125 MHz for 13C) and Jeol 400 MHz (or 100 MHz for 13C) using residual solvents signals as an internal reference (CDCl₃ δ H, 7.26 ppm, δ c 77.3 ppm and CD₃OH δ H 3.31 ppm, δ c 49.0 ppm). The chemical shifts (δ) are reported in *ppm* and coupling constants (*J*) in Hz. IR spectra were recorded on a Perkin Elmer 1600 FTIR spectrometer. Optical rotation measurements were conducted using a Perkin- Elmer 241 polarimeter. 2,3-dihydroxy benzoic acid and 2,3,4,5,6-pentafluorophenol were purchased from Sigma-aldrich. ConcavalinA was purchased from Sigma-aldrich. Professor Orndorff (North Carolina State University) for providing ORN178 and 208 *E. coli* stains.

2. General Procedures:

General Procedure A: Synthesis of catechol substituted sugar-monomers or tripods or nanopodes.

The Boc-protected amino-sugar or tripods or nonapodes (1.0 equiv) was dissolved in dichloromethane/trifluoroacetic acid (3:1, 10 mL) and stirred at room temperature for 1 h. The solvent was evaporated *in vacuo* and the residue redissolved in anhydrous dichloromethane (20 mL). 2,3-*O*-benzyl-benzoic acid (1.5 equiv), *N*,*N*'-Diisopropylcarbodiimide (1.5 eq) and HOBT (0.1 eq) were added, the pH was adjusted to 8 with triethylamine and stirred at room temperature for 12 h. The solvent was evaporated *in vacuo* and purified by flash silica column chromatography.

General Procedure B: Deacetylation and hydrogenolysis of sugar derivatives.

To a solution of sugar-substituted tripod (1.0 equiv) in methanol (10 mL) was added sodium methoxide (6 to 20 eq) and stirred for 2 h at rt. The mixture was neutralized with amberlite– acidic resin, filtered and then concentrated *in vacuo*. The residue was dissolved once more in

methanol (10 mL), 10 % Pd/C (Pd/C, 10 mol % added) and hydrogen gas was bubbled through the solution for 12 h. The solution was filtered over celite and the filtrate concentrated *in vacuo* to afford the final compound.

General Procedure C: Synthesis of iron(III) complexes.

To a solution of dendron (10-11, 19-20, 25-26) (3.3 eq) in MeOH (10 mL) were added the appropriate metal salt (1 eq) and refluxed for 12 h. The solvent was evaporated to obtain black color oily residue, which was used as such for next step.¹



Figure 1. Peracetylated mannose and galactose tripode amine were synthesized using published procedures.¹

3. Synthesis of ligand 10 and 13



Scheme 1. Reagents and Conditions: (a) TFA/DCM; 2,3-bis(benzyolxy)-benzoic acid, DIC/HOBT/DCM; (b) NaOMe/MeOH; H₂/Pd-C/MeOH. (c) FeCl₃/MeOH.

2,3-bis(benzyolxy)-*N***-2,3,4,6-tetra***-O***-acetyl***-α***-D-mannopyranoside-ethylbenzamide 9** . General procedure A using 2-(*tert*-butoxycarbonylamino)ethoxy-2,3,4,6-tetra-*O*-acetyl-*α*-D-mannopyranoside **8** (0.2 g, 0.4 mmol), 2,3-bis(benzyolxy)-benzoic acid (0.2 g, 0.5 mmol), DIC (94 µL, 0.5 mmol) and HOBT (10 mg, 0.06 mmol). Purification by flash chromatography petroleum ether : ethyl acetate (60:40) yielded 2,3-bis(benzyolxy)-*N*-2,3,4,6-tetra-*O*-acetyl-*α*-D-mannopyranoside-ethylbenzamide (0.158 g, 56%). R_f = 0.5 (PE : EtOAc, 60:40); $[\alpha]_D^{r.t}$ = +33.1 (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.11 (t, 1H, *J* = 4.0 Hz), 7.72-7.62 (m, 2H), 7.50-7.30 (m, 9H), 7.18-7.10 (m, 2H), 5.32- 5.06 (m, 8H), 4.72 (s, 1H), 4.22 (dd, 1H, *J* = 5.2,

7.2 Hz); 4.00 (d, 1H, J = 10.5 Hz), 3.94-3.90 (m, 1H); 3.71-3.62 (m, 1H), 3.48-3.38 (m, 2H), 2.13 (s, 3H), 2.04 (s, 3H), 1.96 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 169.9, 169.6, 165.4, 151.6, 146.7, 136.4, 132.1, 128.9, 128.3, 127.7 127.2, 124.4, 123.3, 117.3, 97.5, 71.8, 69.4, 69.1, 68.4, 66.8, 66.2, 62.4, 39.1, 20.7, 20.6 HRMS (MALDI-ToF) (*m/z*); calcd. for C₃₇H₄₁NO₁₃Na 730.2476; found:730.2476 [M+Na]⁺.

2,3-bis(benzyolxy)-*N*-2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside-ethylbenzamide. 12.

General procedure A using 2-(*tert*-butoxycarbonylamino)ethoxy-2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranoside **11** (0.2 g, 0.4 mmol), 2,3-bis(benzyolxy)-benzoic acid (0.2 g, 0.5 mmol) DIC (95 µL, 0.5 mmol) and HOBT (10 mg, 0.06 mmol). Purification by flash chromatography petroleum ether : ethylacetate (60:40) yielded 2,3-bis(benzyolxy)-*N*-2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranoside-ethylbenzamide (0.136 g, 48%). R_f = 0.5 (PE : EtOAc, 60:40); $[\alpha]_D^{r.t}$ = +14.1 (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): 8.14 (t, 1H, *J* = 4.0 Hz), 7.72-7.62 (m, 2H), 7.50-7.30 (m, 9H), 7.18-7.10 (m, 2H), 5.3 (d, 1H, J = 2.8 Hz), 5.31-5.06 (m, 4H), 4.99-4.93 (m, 2H), 4.35 (d, 1H, J = 6 Hz), 4.20-4.06 (m, 3H), 3.89-3.83 (m, 1H), 3.67- 3.59 (m, 1H), 3.57-3.47 (m, 2H), 2.18 (s, 3H), 2.00 (s, 3H), 1.97 (s, 3H), 1.95 (s, 3H).¹³C NMR (100 MHz, CDCl₃): δ 170.2, 169.7, 169.6, 165.4, 151.8, 146.5, 136.4, 132.1, 129.2, 128.5, 127.9 127.4, 124.5, 122.9, 116.9, 101.2, 76.4, 71.2, 70.4, 68.7, 68.5, 67.1, 61.2, 39.4, 20.7, 20.6; HRMS (MALDI-ToF) (*m/z*); calcd. for C₃₇H₄₁NO₁₃Na 730.2476; found:730.2476 [M+Na]⁺.

2,3-bis(hydroxy)-*N*-*a*-**D**-mannopyranoside-ethylbenzamide 10. General procedure B with 2,3-*bis*(benzyolxy)-*N*- 2,3,4,6-tetra-*O*-acetyl-*a*-D-mannopyranoside-ethylbenzamide **9** (0.1 g, 0.14 mmol), sodium methoxide (45 mg, 0.8 mmol) and 10 % Pd/C (10 mg) yielded 2,3-bis(hydroxy)-*N*- *a*-D-mannopyranoside-ethylbenzamide **10** (32 mg, 64%). $[\alpha]_D^{r.t} = +39.8$ (c = 1.0, H₂O); ¹H NMR (400 MHz, MeOD): δ 7.26 (d, 1H, *J* = 9.2 Hz), 6.92 (d, 1H, *J* = 8.0 Hz), 6.73 (t, 1H, *J* = 8.0 Hz), 4.82 (s, 1H), 3.94-3.75 (m, 3H), 3.75-3.51 (m, 7H). ¹³C NMR (100 MHz, MeOD): δ 169.9, 148.3, 145.5, 118.1, 117.7, 115.6, 73.5, 71.2, 70.7, 67.1, 65.8, 61.4, 39.1. HRMS (MALDI-ToF) (*m/z*); calcd. for C₁₅H₂₁NO₉Na 382.1114; found:382.0784 [M+Na]⁺

2,3-bis(hydroxy)-*N*- β-D-galactopyranoside-ethylbenzamide 13. General procedure B with 2,3-bis(benzyolxy)-*N*- 2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside-ethylbenzamide 12 (90 mg,

0.12 mmol), sodium methoxide (40 mg, 0.75 mmol) and 10 % Pd/C (10 mg) yielded 2,3bis(hydroxy)-*N*- α -D-mannopyranoside-ethylbenzamide **13** (35 mg, 77%). [α]_D^{r.t} = +21.8 (c = 1.0, H₂O); ¹H NMR (400 MHz, MeOD): δ 7.26 (d, 1H, *J* = 9.2 Hz), 6.93 (d, 1H, *J* = 8.0 Hz), 6.73 (t, 1H, *J* = 8.0 Hz), 4.31 (d, 1H, *J* = 8.0 Hz), 4.05-3.95 (m, 2H), 3.84-3.47 (m, 8H). ¹³C NMR (100 MHz, MeOD): δ 170.1, 145.9, 144.5, 118.3,117.5, 103.8, 75.4, 73.5, 71.2, 68.9, 68.1, 61.1, 39.4. HRMS (MALDI-ToF) (*m*/*z*); calcd. for C₁₅H₂₁NO₉Na 382.1114; found:382.0784 [M+Na]⁺



6

Scheme 2. Reagents and Conditions: (a) TFA/DCM; 2,3-bis(benzyolxy)-benzoic acid, DIC/HOBT/DCM; (b) 1N NaOH/MeOH; pentafluorophenol (PFP)/DIC/DCM; (c) Comp 8 or 11/DCM/TEA; (d) NaOMe/MeOH; H₂/Pd-C/MeOH. (e) FeCl₃/MeOH.

2,3-bis(benzyolxy)-3-{*N*-{tris[3-[ethylcarboxyl-ethoxy)methyl]}methylamide}-3-ethyl

benzamide *tert*-butoxycarbonyl-3-{*N*-{tris[3-[ethylcarboxyl-ethoxy) (15). mmol) methyl]}methylamide}-3- β -alanine 14 (3 5.06 dissolved g, was in dichloromethane/trifluoroacetic acid (3:1, 10 mL) and stirred at room temperature for 1 h. The solvent was evaporated in vacuo, the residue was re-dissolved in anhydrous dichloromethane (25 mL) and the pH adjusted to 8 using triethylamine. 2,3-bis(benzyolxy)-benzoic acid (2.5 g, 7.6 mmol) and diisopropyl carbadiazine (1.2 ml, 7.6 mmol) were added to the above mixture and stirred for 12 h at rt. Finally, the mixture was concentrated in vacuo and purified by flash chromatography (DCM:MeOH; 95:5) to afford 2,3-bis(benzyolxy)-3-{N-{tris[3-[ethylcarboxy]ethoxy)methyl]} methylamide}-3-ethyl benzamide 15. (2.4 g, 60%). $R_f = 0.5$ (CH₂Cl₂/MeOH, 98:5); ¹H NMR (400 MHz, CDCl₃): δ 8.2 (t, 1H, J = 8.0 Hz), 7.62 (t, 2H, J = 4.0 Hz), 7.45-7.27 (m, 9H), 7.12 (t, 3H, J = 8.0 Hz), 5.12 (s, 2H), 5.08 (s, 2H), 4.12 (q, 6H, J = 8.0 Hz), 3.64 (s, 6H), 3.61 (t, 6H, J=8.0 Hz), 3.54 (q, 2H, J=8.0 Hz), 2.46 (t, 6H, J=8.0 Hz), 2.34 (t, 2H, J=8.0 Hz). 1.23 (t. 9H, J = 8.0 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 170.7, 169.9, 169.8, 165.4, 151.7, 149.9, 136.4, 132.2, 128.9, 128.8, 127.7, 127.1, 124.4, 123.2, 117.3, 70.8, 68.9, 65.9, 60.3, 60.1, 53.4, 41.8, 38.1, 35.4, 24.9. HRMS (MALDI-ToF) (*m/z*); calcd. for C₄₃H₅₆N₂O₁₃Na 831.3680; found:831.3682 [M+Na]⁺.

2,3-bis(benzyolxy)-2-carbonyl-3-{N-{tris[3-[pentafluoro phenyl carboxyl-ethoxy) methyl]}methylamide}-3-ethyl benzamide 16. To a solution of 2,3-bis(benzyolxy)-3-{N-{tris[3-[ethylcarboxyl-ethoxy)methyl]} methylamide}-3-ethyl benzamide 15. (1.5 g, 1.86 mmol) in methanol (10 mL) was added NaOH (aqueous, 1 N, 3 mL) The mixture was stirred at rt for 2 h, concentrated *in vacuo*, adjusted to pH 5 with HCl (aqueous, 1 N) and extracted with EtOAc. The organic layer was dried with Na₂SO₄, filtered and concentrated *in vacuo*. The residue was dissolved in DCM and 2,3,4,5,6-pentafluorophenol (1.69 g, 9.3 mmol) was added. After cooling to 0 $^{\circ}$ C, diisopropyl carbadiazine (1.6 mL, 10.2 mmol) was added and the reaction mixture was stirred for 12 h at room temperature, concentrated *in vacuo* and purified by silica column flash

chromatography (DCM:EtOAc; 92:8) to afford 8-*O*-benzyl-quinoline-2-carbonyl-3-{*N*-{tris[3-[pentafluoro phenyl carboxyl-ethoxy)methyl]} methylamide}-3- β -alanine (1.3 g, 59%). R_f = 0.5 (CH₂Cl₂/EtOAc, 90:10); ¹H NMR (400 MHz, CDCl₃): δ 8.27 (t, 1H, *J* = 8.0 Hz), 7.63 (t, 2H, *J* = 4.0 Hz) 7.45-7.27 (m, 9H), 7.14 (t, 2H, *J* = 8.0 Hz), 5.13 (s, 2H), 5.09 (s, 2H), 4.45 (br,1H), 3.73 (m, 12H), 3.54 (q, 2H, *J* = 8.0 Hz), 2.81 (t, 6H, *J* = 8.0 Hz), 2.32 (t, 2H, *J* = 8.0 Hz). ¹³C NMR (100MHz, CDCl₃): δ 170.4, 169.9, 169.8, 165.2, 152.4, 149.2, 142.3, 142.3, 140.1, 139.3, 136.7, 128.9, 127.2, 122.1, 118.7, 75.9, 75.6, 70.8, 65.1, 60.2, 54.2, 38.8, 35.2. HRMS (MALDI-ToF) (*m/z*); calcd. for C₅₅H₄₁N₂O₁₃F₁₅ 1222.2369; found: 1222.2365 [M]^{+.}

2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-O-acetyl-a-D-

mannopyranoside-ethoxy]methyl]methylamide}-3- ethyl benzamide (14). General procedure A using 2-(*tert*-butoxycarbonylamino)ethoxy-2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranoside 21 (0.2 g, 0.41 mmol) and 8-*O*-benzyl-quinoline-2-carbonyl-3-{*N*-{tris[3-[pentafluoro phenyl carboxyl-ethoxy) methyl]}methylamide}-3- β -alanine 11 (0.1 g, 0.082 mmol). Purification by flash chromatography (DCM:MeOH; 94:6) yielded 8-*O*-benzyl-quinoline-2-carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranoside-ethoxy]methyl]methylamide}-3- β -

alanine (0.087 g, 58%). $R_f = 0.5$ (CH₂Cl₂/MeOH, 92:8); $[\alpha]_D^{r.t} = +21.1$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, MeOD): δ 8.20 (t, 1H, J = 4.0 Hz), 7.72-7.62 (m, 2H), 7.50-7.30 (m, 9H), 7.18-7.10 (t, 2H), 5.26-5.21 (m, 9H), 5.13 (s, 2H), 5.09 (s, 2H), 4.23 (dd, 3H, J = 4.8, 8.0 Hz), 4.09 (dd, 3H, J = 2.4, 9.6 Hz), 4.02-4.01 (m, 3H), 3.73-3.71 (m, 6H), 3.65 (br.s, 6H), 3.61 (t, 6H, J = 6.0 Hz), 3.53-3.50 (m, 9H), 3.38 (t, 6H, J = 4.5 Hz), 2.62 (t, 2H, J = 6.9 Hz), 2.35 (t, 6H, J = 6.0 Hz), 2.10 (s, 9H), 2.03 (s, 9H), 2.01 (s, 9H), 1.91 (s, 9H); ¹³C NMR (100 MHz, MeOD): δ 170.2, 169.7, 169.6, 165.4, 151.8, 146.5, 136.4, 132.1, 129.2, 128.5, 127.9 127.4, 124.5, 122.9, 116.9, 101.4, 71.8, 70.4, 70.2, 69.7, 69.3, 69.0, 63.1, 40.1, 38.3, 31.0, 20.4; (MALDI-ToF) (*m/z*): [M+Na]⁺ calcd. for C₈₅H₁₁₃N₅O₄₀Na 1866.6860; found:1866.6934.

(vii) 2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranoside-ethoxy]methyl]methylamide}-3- β -alanine 18. General procedure A using 2-(*tert*-butoxycarbonylamino)ethoxy-2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranoside 22 (0.2 g, 0.41 mmol) and 8-*O*-benzyl-quinoline-2-carbonyl-3-{*N*-{tris[3-[pentafluoro phenyl carboxyl-ethoxy) methyl]}methylamide}-3- β -alanine 11 (100 mg, 0.08 mmol). Purification column chromatography yielded 2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranoside-ethoxy]methyl] methylamide}-3- β -alanine (0.064 g, 42%). R_f =

0.5 (CH₂Cl₂/MeOH, 92:8); $[\alpha]_D^{rt} = +18.7$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, MeOD): δ 8.20 (t, 1H, *J* = 4.0 Hz), 7.72-7.62 (m, 2H), 7.50-7.30 (m, 9H), 7.18-7.10 (t, 2H), 5.37 (d, 3H, *J* = 4.0 Hz), 5.27 (s, 2H), 5.14 (t, 5H, *J* = 8.0 Hz), 4.99 (dd, 3H, *J* = 4.0 Hz), 4.49 (d, 3H, *J* = 8.0 Hz), 4.12-4.10 (m, 6H), 3.92 (t, 3H, *J* = 6.0 Hz), 3.84 (d, 3H, *J* = 4.0 Hz), 3.65 (br.s, 18H), 3.42 (m, 9H), 3.34-3.31 (m, 2H), 2.39 (t, 6H, *J* = 6.0 Hz), 2.14 (s, 9H), 2.04 (s, 9H), 2.02 (s, 9H), 1.96 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 171.1, 170.0, 169.8, 169.7, 155.3, 149.4, 139.7, 138.6, 131.6, 129.3, 128.7, 128.1, 119.9, 112.2, 101.1, 70.6, 69.1, 68.7, 67.2, 66.9, 61.2, 59.7, 39.1, 37.0, 30.4, 20.6; (MALDI-ToF) (*m*/*z*): [M+Na]⁺ calcd. for C₈₅H₁₁₃N₅O₄₀Na 1866.6860; found:1866.6934.

2,3-bis(hydroxy)-2-carbonyl-3-{tris[3-[2-ethoxy-a-D-mannopyranoside-

ethoxy]methyl]methylamide}-3-*β*-alanine 19. General procedure B with 2,3-bis(benzyolxy)-2carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-*O*-acetyl-*α*-D-mannopyranoside-ethoxy]methyl] methylamide}-3-*β*-alanine 17 (70 mg, 0.038 mmol), sodium methoxide (12 mg, 0.27 mmol) and 10 % Pd/C (10 mg) yielded 2,3-bis(hydroxy)-2-carbonyl-3-{tris[3-[2-ethoxy-*α*-Dmannopyranoside-ethoxy]methyl]methylamide}-3-*β*-alanine 19 (19 mg, 43%). [*α*]_D^{r.t} = + 5.4(c = 1.0, H₂O); ¹H NMR (400 MHz, MeOD): δ 7.26 (d, 1H, *J* = 8.0 Hz), 6.92 (d, 1H, *J* = 8.0 Hz), 6.74 (d, 1H, *J* = 8.0 Hz),4.75 (s, 3H), 3.81 (d, 6H, *J* = 7.8 Hz), 3.72-3.67 (m, 12H), 3.64-3.54 (m, 18H), 3.53-3.49 (m, 8H), 2.62 (t, 2H, *J* = 6.9 Hz), 2.33 (t, 6H, *J* = 6.0 Hz); ¹³C NMR (100 MHz, MeOD): δ 170.1, 145.9, 144.5, 118.3,117.5, 101.2, 74.3, 71.6, 69.6, 68.2, 66.8, 62.3, 61.1, 39.9, 37.0; (MALDI-ToF) (*m*/*z*); [M+Na]⁺ calcd. for C₄₇H₇₇N₅O₂₈Na 1182.4653; found:1182.6038.

2,3-bis(hydroxy)-2-carbonyl-3-{tris[3-[2-ethoxy-β-D-galactopyranoside-

ethoxy]methyl]methylamide}-3-β-alanine 20 : General procedure B with 2,3-bis(benzyolxy)-2carbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside-ethoxy]methyl] methylamide}-3-β-alanine 18 (40 mg, 0.022 mmol), sodium methoxide (7 mg, 0.132 mmol) and 10% Pd/C (10 mg) yielded 2,3-bis(hydroxy)-2-carbonyl-3-{tris[3-[2-ethoxy-β-Dgalactopyranoside-ethoxy]methyl]methylamide}-3-β-alanine (13 mg, 54%). [α]_D^{r.t} = +13.9 (c = 1.0, H₂O); ¹H NMR (400 MHz, MeOD): δ 7.26 (d, 1H, *J* = 8.0 Hz), 6.92 (d, 1H, *J* = 8.0 Hz), 6.74 (d, 1H, *J* = 8.0 Hz), 4.29 (d, 3H, *J* = 7.5 Hz), 3.82-3.55 (m, 44H), 2.61 (t, 2H, *J* = 6.9 Hz), 2.33 (t, 6H, *J* = 6.0 Hz); ¹³C NMR (100 MHz, MeOD): δ 170.1, 145.9, 144.5, 118.3,117.5,103.5, 75.0, 72.6, 70.6, 68.5, 63.2, 48.9, 39.3, 35.8; (MALDI-ToF) (m/z); $[M+Na]^+$ calcd. for C₄₇H₇₇N₅O₂₈Na 1182.4653; found:1182.6038.



Scheme 3. Reagents and Conditions: (a) Comp 22 or 23/DCM/TEA; (d) NaOMe/MeOH; H₂/Pd-C/MeOH. (e) FeCl₃/MeOH.

5. Synthesis of ligand 25 and 26.

2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-carboxyl ethoxy]methyl] 3'-{tris[2'-ethoxy-2,3,4,6-tetra-O-acetyl- α -D-mannopyranoside-ethoxy]methyl]methylamide}-3- β -alanine 22. General procedure A using *tert*-butoxycarbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-O-acetyl- α -D-

mannopyranoside-ethoxy]methyl]methylamide}-3- β -alanine 23 (66 mg, 0.041 mmol), 2,3bis(benzyolxy)-2-carbonyl-3-{N-{tris[3-[pentafluoro phenyl carboxyl-ethoxy) methyl]}methylamide}-3- β -alanine **16** (10 mg, 8.1 μ mol) and flash silica column chromatography to afford 2,3-(benzyolxy)-2-carbonyl-3-{tris[3-carboxylethoxy]methyl]3' $tris[2'-ethoxy-2,3,4,6-tetra-O-acety]-\alpha$ -D-mannopyranoside-ethoxy]methyl]methylamide}-3- β alanine (15 mg, 65%). $R_f = 0.5$ (CH₂Cl₂/MeOH, 90:10); $[\alpha]_D^{r,t} = -9.1$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, MeOD): δ 7.59 (t, 1H, J = 8.4 Hz), 7.39 (m, 1H), 7.45-7.28 (m, 10H), 7.10 (t, 1H, J = 8.0 Hz), 5.45 (s, 2H), 5.21-5.26 (m, 27H), 4.23 (dd, 9H, J = 4.8, 7.2 Hz), 4.09 (dd, 9H, J = 2.4, 9.6 Hz), 4.03-4.01 (m, 14H), 3.70-3.62 (br.s, 54H), 3.55 (s, 28H), 3.45 (br.s, 32H), 2.45 (t, 28H, J = 6.0 Hz), 2.13 (s, 27H), 2.05 (s, 27H), 2.03 (s, 27H), 1.95 (s, 27H); ¹³C NMR (100 MHz, MeOD): δ 171.8, 171.6, 170.5, 167.0, 169.6, 166.2, 155.3, 149.7, 141.6, 140.1, 139.6, 129.9, 129.3, 128.6, 120.9, 120.1, 112.9, 102.2, 74.5, 72.2, 69.0, 68.9, 68.5, 67.2, 64.9, 62.3, 55.8, 39.8, 36.3, 20.8. MALDI-TOF (m/z): calcd. for C₂₂₅H₃₂₄N₁₈O₁₂₀Na 5220.9696; found: 5220.956 $[M+Na]^+$.

2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-carboxyl ethoxy]methyl] 3'-{tris[2'-ethoxy-2,3,4,6tetra-O-acetyl-\beta-D-galactopyranoside-ethoxy]methyl]methylamide}-3-\beta-alanine 24. General *tert*-butoxycarbonyl-3-{tris[3-[2-ethoxy-2,3,4,6-tetra-O-acetyl-β-Dprocedure А using galactopyranoside-ethoxy]methyl]methylamide}-3- β -alanine 23 (0.60 g, 0.039 mmol), 2,3bis(benzyolxy)-2-carbonyl-3-{*N*-{tris[3-[pentafluoro phenyl carboxyl-ethoxy) methyl]} methylamide}-3- β -alanine 16 (10 mg, 8.1 μ mol) and purification by flash silica column chromatography afforded 2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-carboxyl ethoxy]methyl] 3' $tris[2'-ethoxy-2,3,4,6-tetra-O-acety]-\beta-D-galactopyranoside-ethoxy]methyl]methylamide}-3-\beta$ alanine (15 g, 66%). $R_f = 0.5$ (CH₂Cl₂/MeOH, 90:10); $[\alpha]_D^{r.t} = -15.1$ (c =1.0, CHCl₃); ¹H NMR (400 MHz, MeOD): δ 7.61 (t, 1H, J = 8.4 Hz), 7.39 (m, 1H), 7.45-7.28 (m, 10H), 7.10 (t, 1H, J = 8.1 Hz), 5.38 (d, 9H, J = 4.0 Hz), 5.10 (dd, 10H, J = 4.0, 8.0 Hz), 4.48 (d, 9H, J = 4.0 Hz), 4.13 (d, 32H, J = 6.9 Hz), 3.87 (br.s, 9H), 3.66 (br.s, 78H), 3.39-3.36 (m, 24H), 2.43 (br, 32H), 2.13(s, 27H), 2.05 (s, 27H), 2.01 (s, 27H), 1.94 (s, 27H); ¹³C NMR (100 MHz, MeOD): δ 168.8, 167.0, 169.6, 166.2, 150.3, 149.4, 139.7, 133.6, 132.8, 131.6, 129.3, 128.6, 126.9, 97.5, 70.7, 70.3, 69.5, 69.2, 69.0, 68.9, 68.5, 63.7, 62.3, 53.3, 39.8, 35.3, 20.8; MALDI-TOF (*m/z*): calcd. for C₂₂₅H₃₂₄N₁₈O₁₂₀Na 5220.9696; found: 5220.956 [M+Na]⁺.

2,3-bis(hydroxyl)-2-carbonyl-3-{tris[3-carboxyl ethoxy]methyl] 3'-{tris[2'-ethoxy-\alpha-D-mannopyranoside-ethoxy]methyl]methylamide}-3-\beta-alanine 5. General procedure B using 2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-carboxylethoxy]methyl]3'-{tris[2'-ethoxy-2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranoside-ethoxy]methyl]methylamide}-3- β -alanine (15 mg, 2.8 µmol), sodium methoxide (10 mg, 2 µmol) and Pd/C (10 mg) yielded 2,3-bis(hydroxyl)-2-carbonyl-3-{tris[3-[2-ethoxy- α -D-mannopyranoside-ethoxy]methyl]methylamide}-3- β -alanine (10 mg, 69%). [α]_D^{r.t} = +51.2 (c = 1.0, H₂O); ¹H NMR (400 MHz, D₂O): δ 7.26 (d, 1H, *J* = 9.2 Hz), 6.92 (d, 1H, *J* = 8.0 Hz), 6.73 (t, 1H, *J* = 8.0 Hz), 4.85 (s, 9H), 3.81-3.52 (m, 112H), 3.46-3.31 (m, 37H), 2.45 (t, 18H, *J* = 8.0 Hz), 2.39-2.33 (m, 12H); ¹³C NMR (100 MHz, D₂O): δ 176.7, 173.8, 148.3, 145.5, 128.6, 119.6, 118.6, 115.5, 101.2, 74.3, 71.6, 69.6, 68.2, 66.8, 62.3, 61.1, 39.9, 37.0; (MALDI-ToF) (*m*/*z*); C₁₄₃H₂₄₅N₁₇O₈₅Na 3583.5269; found : 3583.5234 [M+Na]⁺.

2,3-bis(hydroxyl)-2-carbonyl-3-{*tris*[3-carboxylethoxy]methyl]3'-{*tris*[2'-ethoxy-β-D-

galactopyranoside-ethoxy]methyl]methylamide}-3- β -alanine 6. General procedure B using 2,3-bis(benzyolxy)-2-carbonyl-3-{tris[3-carboxyl ethoxy]methyl]3'-{tris[2'-ethoxy-2,3,4,6-tetra-O-acetyl- β -D-galactopyranoside-ethoxy]methyl]methylamide}-3- β -alanine (15 mg, 2.8 μ mol), sodium methoxide (10 mg, 2.4 μ mol) and 10% Pd/C (12 mg) yielded 2,3-bis(hydroxy)-2carbonyl-3-{*tris*[3-carboxylethoxy]methyl]3'-{*tris*[2'-ethoxy- β -D-galactopyranoside-

ethoxy]methyl]methylamide}-3-β-alanine **6** (10 mg, 69%). [α]_D^{r,t} = -24.3 (c = 1.0, H₂O); ¹H NMR (400 MHz, MeOD): δ 7.26 (d, 1H, J = 9.2 Hz), 6.92 (d, 1H, J = 8.0 Hz), 6.73 (t, 1H, J = 8.0 Hz), 4.35 (d, 9H, J = 8.0 Hz), 3.88 (m, 18H), 3.72-3.55 (m, 118H), 3.55-3.36 (m, 37H), 2.45 (t, 18H, J = 8.0 Hz), 2.39 (t, 8H, J = 9.2 Hz), 2.23 (t, 4H, J = 8.0 Hz); ¹³C NMR (100 MHz, MeOD): δ 176.7, 173.6, 162.3, 149.5, 145.6, 118.6, 118.1, 115.5, 75.0, 72.6, 70.6, 68.5, 67.2, 60.9, 39.3, 35.8; (MALDI-ToF) (*m*/*z*); C₁₄₃H₂₄₅N₁₇O₈₅Na 3583.5269; found : 3583.5234 [M+Na]⁺.

6. Ligand titration with Ferric Ions. Aliquots of stock solutions of the ligands 10, 19, 25 and increasing amounts of $FeCl_3$ (both in spectroscopic grade methanol) were mixed, and after 15 min diluted with methanol/0.1 N aqueous tris-buffer pH 8.5 (4/1) to a constant ligand concentration of 0.3 mM for UV/vis titrations. Measurements were performed after equilibration

for 24 h at room temperature. UV/vis absorption spectra were recorded on a Hewlett-Packard diode array 8450A spectrophotometer.

Spectral evidence for the formation of the 3:1 complexes for catechol from iron(III) in airsaturated buffers is shown in figure 1 for complex 2 at 490 nm.



Figure 1. Uv-visible spectroscopic titrations of ligands: Variation of $O.D_{490}$ intensity as a function of the molar Iron(III)/ligand fraction. Solvent: methanol/water(80:20); ligand concentration = 0.3 mM; 0.05 M (tris-base, pH 8.0); $T (25.0 \pm 0.2)$ °C, **10 (•)**, **19(**), **25 (•)**.

7. Tubidimetric analysis :

The solution of lectin (1.0 mg/ml) in HEPES buffer (10 mM Hepes pH 6.5, 1mM MgCl₂, 1mM CaCl₂, 1% BSA) and complex 6-12 (1.0 mM) in water was added. The time dependent turbidity kinetics was recorded by measuring the absorption coefficient at 700 nm after every one minutes. After 20 minutes the turbidity was disturbed by adding 100 mM of mannose to the solution.

8. Imaging Experiment :

4

The snapshot experiment was performed with 1 mM of 4 and 7 complex and 5 mg/mL (0.1 ml) of ConA lectin in HEPES buffer solution (10 mM Hepes pH 6.5, 1mM MgCl₂, 1mM CaCl₂, 1% BSA). Complex 4 showed precipitation with ConA after 1 hr.

7



Figure 2. Visible images of the complex 4 & 7 in presence and absence of ConA lectin.

9. Bacterial Strains and 96-well growth promotion assay. The mutants *E.coli ORN178 and ORN208* were provided by Prof. Orndorff (College of Veterinary Medicine, Raleigh, NC United States). Such mutants are ideal for the study of mannose mediate interactions. Bacteria were grown in LB medium overnight at 37 °C on a rotary shaker at 180 RPM. An aliquots of bacteria were removed from the batch culture to monitor growth until they reached an approximate OD_{600} of 0.6. Approximately 100 µL of bacterial cells were aliquot in to previously autoclaved 2.2 ml 96-well polypropylene blocks and the mixed with 1ml of freshly prepared LB broth medium containing 0.1 mM of 2,2'-dipyridyl, followed by 0.1µM to 0.1 mM concentration of iron complexes (1-7). After covering the arrays with a sterile mat cap, the cultures were placed on a rotating platform shaker at 250 rpm and incubated at 37°C for at least 6 h. Figure 4 displayed the optimum concentration required for bacterial growth.

Once we optimum concentration of the ligand (50 μ M) for different rate of growth. 96-plate assay was repeated at different time interval to monitor the kinetics of growth. The inhibition assay was performed by adding NaN₃ (100 μ M) or compound 4 (50 μ M) before adding 50 μ M of iron complexes (1-7).



Figure 3. Fe(III)-glycodendrimer dependent growth promotion under iron limitation. Mannose positive mutants of *E. coli* (ORN178) was grown in a medium containing the iron chelator 2,2'- dipyridyl with different concentration of complex 1-7. A_{600} value represent the means \pm standard deviations from there parallel cultures.



Figure 4. Fe(III)-glycodendrimer dependent growth promotion under iron limitation. Mannose negative mutants of *E. coli* (ORN208) was grown in a medium containing the iron chelator 2,2'-dipyridyl (×) siderophore $1(\blacksquare)$, complex $2(\blacklozenge)$, $3(\bullet)$, $4(\Box)$, $5(\diamondsuit)$ and $7(\circ)$. A₆₀₀ value represent the means \pm standard deviations from their parallel cultures.



Figure 5. Fe(III)-glycodendrimer dependent growth promotion under iron limitation. Mannose positive mutants of *E. Coli* (ORN178) was grown in medium containing the iron chelator 2,2'-dipyridyl and compound $1(\blacksquare),2(\bullet)3(\bullet)$ and with complex 4, (50 µM) for 30 mins and complex $1(\Box)$, 2 (\diamond) and 3 (\circ). A₆₀₀ value represent the means \pm standard deviations from their parallel cultures.

10. LB Plate assay. LB plate assay for performed according to earlier protocols.² All stains were tested on plates containing bipyridine (100 mM). Sterile aqueous solution of siderophore were prepared in the concentration of 0.1 mM and added on autoclaved circular filter papers. Plates were incubated for 24h. The radiuses of bacterial growth around the filter papers indicate specific update and delivery of iron into the system (fig 6).



Figure 6. LB plates assay with compound 1-7 and E. coli ORN 178 stain.

11. References

- 1. R. Kikkeri, L. H. Hossain, P. H. Seeberger, Chem Comm. 2008, 2127
- 2. W. Rabsch, G. Winkelmann, BioMetals, 1991, 4, 244.