

Supporting Information

Polycation-chaperoned in-stem molecular beacon system

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Preparation of ISMBs and PLL-g-Dex and conditions for the spectroscopic measurements

Phosphoramidite monomers with perylene and anthraquinone were synthesized according to the previous reports.^{1,2} Oligonucleotide syntheses (including polyacrylamide gel electrophoresis and HPLC purification) were outsourced (Nihon Techno Service Co., Ltd.). DNA sequences containing only natural nucleotides were supplied by Integrated DNA Technologies, Inc. Poly(L-lysine)-*graft*-dextran was prepared by a reductive amination reaction of poly(L-lysine) ($M_w = 8 \times 10^3$, SIGMA) with dextran ($M_n = 6 \times 10^3$, Dextran T-10, Amersham Pharmacia Biotech) as reported previously.³ The dextran content of the copolymer was 90 wt%. PLL-g-Dex was added to the solution of MB and substrate at N/P of 2, because response speed was saturated at N/P = 2 (see Fig. S7). Here, the concentrations of [amino group]_{copolymer} and [phosphate group]_{DNA} were based on positively charged amino groups derived from poly(L-lysine) and negatively charged phosphate groups, respectively.

Fluorescence spectra were measured on a JASCO model FP-6500 equipped with programmable temperature controller in a microcell (3 mm x 3 mm). Fluorescence intensity was monitored at 460 nm by exciting 425 nm. Errors in F/F_0 were estimated to be within 15%. For response-measurements in Fig. 3, MB (for a final concentration of 0.2 μM) and PLL-g-Dex (N/P = 0 or 2) were dissolved in buffered solution (100 mM NaCl, 10 mM phosphate buffer, pH 7.0) in the cell (10 mm x 10 mm) by magnetically stirring at 20 °C, followed by the addition of **Surv** (to a final concentration of 0.4 μM) to start the reaction.

- 1) H. Kashida, T. Takatsu, H. Asanuma, *Tetrahedron Lett.* **2007**, *48*, 6759.
- 2) H. Asanuma, H. Hayashi, J. Zhao, X. G. Liang, A. Yamazawa, T. Kuramochi, D. Matsunaga, Y. Aiba, H. Kashida, M. Komiyama, *Chem. Commun.* **2006**, 5062.
- 3) A. Maruyama, M. Katoh, T. Ishihara, T. Akaike, *Bioconjugate Chem.* **1997**, *8*, 3.

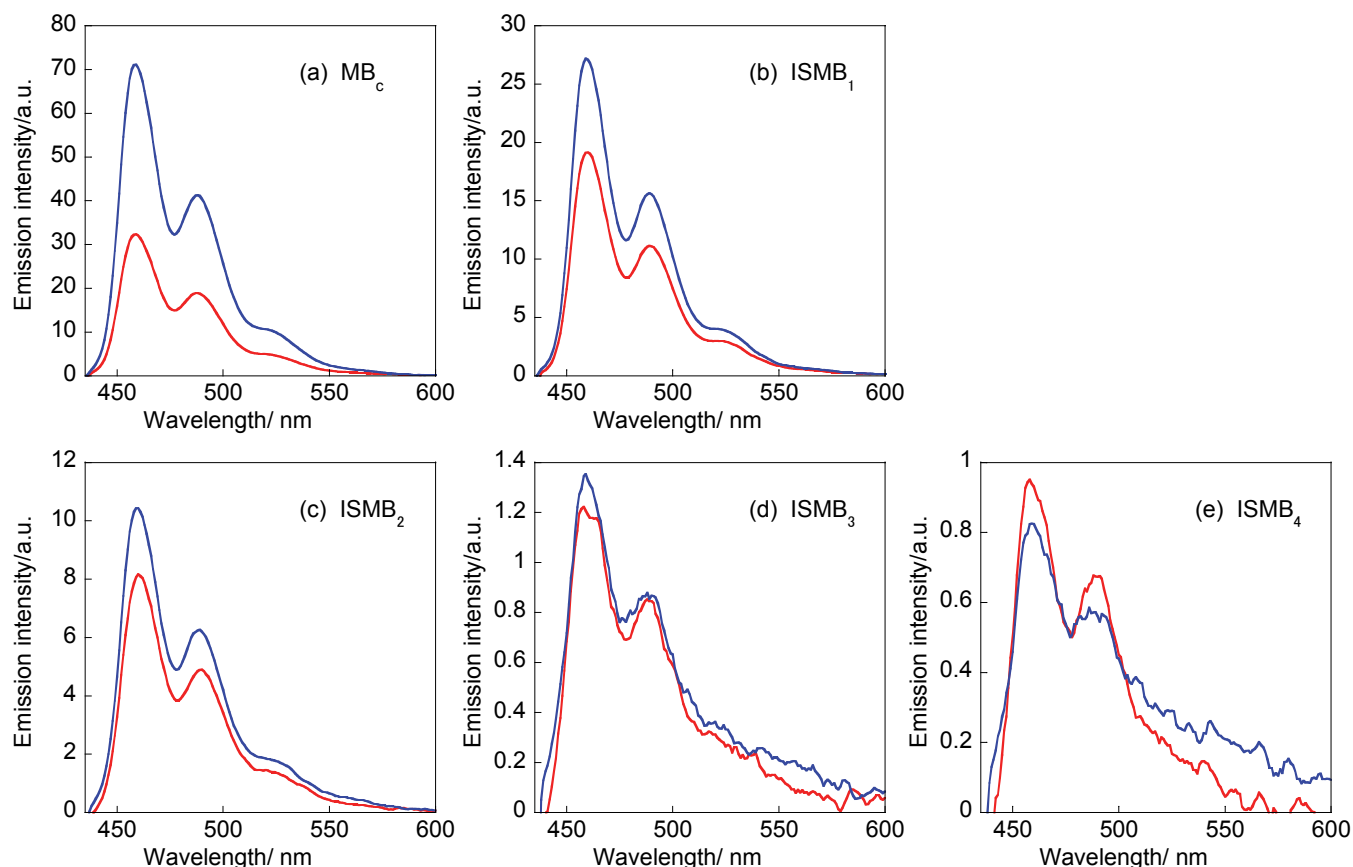
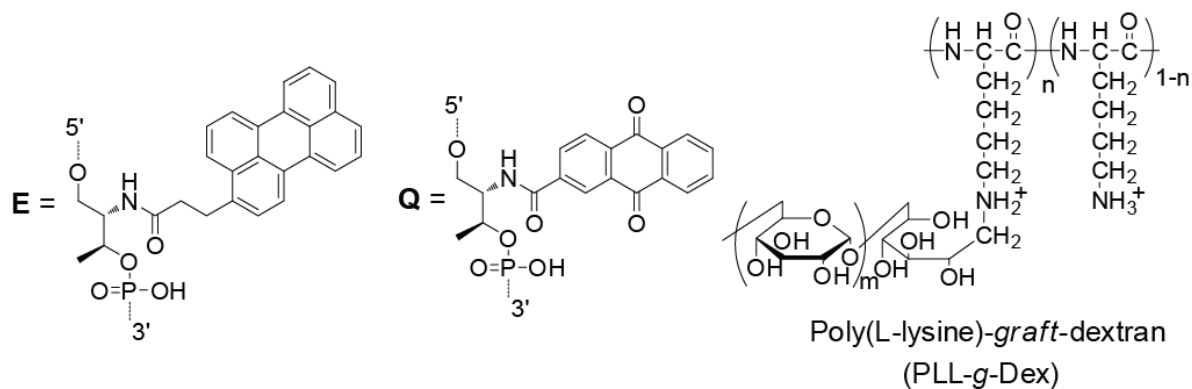


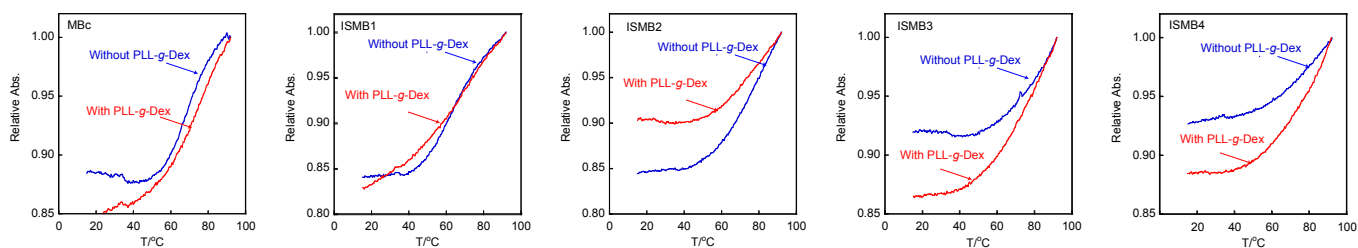
Figure S1. Fluorescence emission (Background emission) spectra of closed (a) **MB_c**, (b) **ISMB₁**, (c) **ISMB₂**, (d) **ISMB₃** and (e) **ISMB₄** in the presence (red line) and absence (blue line) of PLL-*g*-Dex.

Solution conditions; [MB] = 0.2 μM, [NaCl] = 100 mM, pH 7.0 (10 mM phosphate buffer), 20 °C, N/P = 2
 Measurement conditions of JASCO model FP-6500; bandwidth of excitation = 5nm, bandwidth of emission = 5 nm,
 Sensitivity = medium.



MB_c: 5'-ETG-GTC-CTT-GAG-AAA-GGG-C-GAC-CAQ-3'
ISMB₁: 5'-TGG-ETC-CTT-GAG-AAA-GGG-C-GAQ-CCA-3'
ISMB₂: 5'-TEG-GET-C-CTT-GAG-AAA-GGG-C-GAQ-CCQ-A-3'
ISMB₃: 5'-GEG-TEG-GET-C-CTT-GAG-AAA-GGG-C-GAQ-CCQ-ACQ-C-3'
ISMB₄: 5'-GEC-GEG-TEG-GET-C-CTT-GAG-AAA-GGG-CGA-QCC-QAC-QCC-QC-3'
Surv: 3'-ACG-CCA-CCA-GGA-ACT-CTT-TCC-CG-5'

(a) Without Surv



(b) With Surv

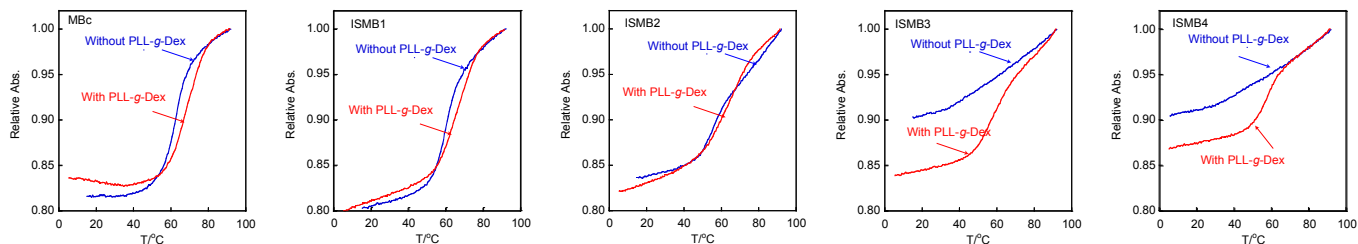


Figure S2. Melting curves of MBs (a) without and (b) with **Surv** obtained from the change of absorbance at 260 nm in the presence (red line) and absence (blue line) of PLL-g-Dex.

Solution conditions; $[\mathbf{MB}] = 1 \mu\text{M}$, $[\mathbf{Surv}] = 1 \mu\text{M}$, $[\text{NaCl}] = 100 \text{ mM}$, pH 7.0 (10 mM phosphate buffer), N/P = 0 (for blue line), 2 (for red line). Temperature ramp was $0.5 \text{ }^\circ\text{C min}^{-1}$.

Table S1. Melting temperatures of MBs determined from Figure S2.^{a)}

MB	Without Surv		With Surv	
	N/P=0	N/P=2	N/P=0	N/P=2
MB_c	67.0	74.5	62.1	68.1
ISMB₁	61.5	67.7	59.4	66.1
ISMB₂	- ^{b)}	- ^{b)}	- ^{b)}	64.0
ISMB₃	- ^{b)}	- ^{b)}	- ^{b)}	57.3
ISMB₄	- ^{b)}	- ^{b)}	- ^{b)}	56.9

a) Solution conditions; $[\mathbf{MB}] = 1 \mu\text{M}$, $[\mathbf{Surv}] = 1 \mu\text{M}$, $[\text{NaCl}] = 100 \text{ mM}$, pH 7.0 (10 mM phosphate buffer), N/P = 0 (for blue line), 2 (for red line). Temperature ramp was $0.5 \text{ }^\circ\text{C min}^{-1}$.

b) Not determined.

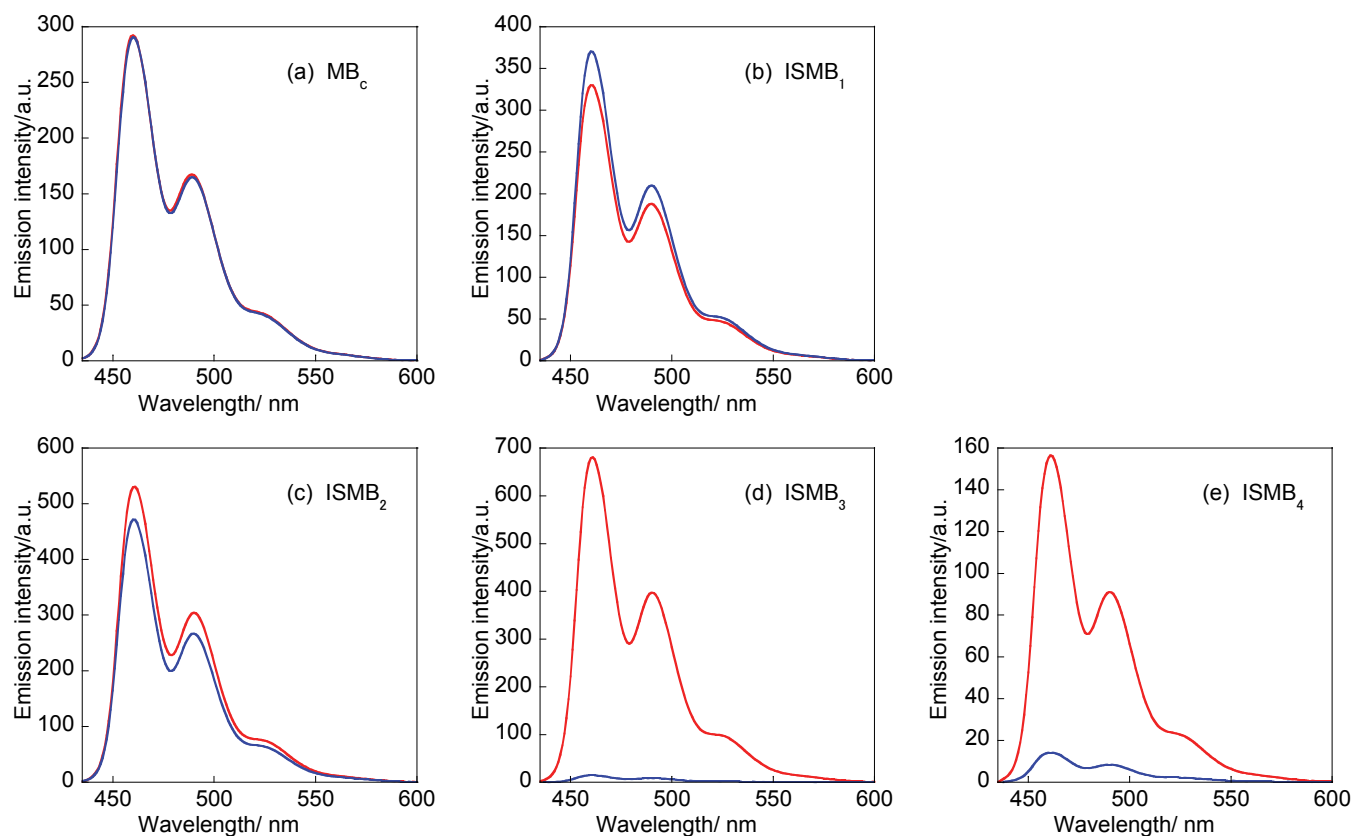


Figure S3. Fluorescence emission (signal emission) spectra of opened (a) **MB_c**, (b) **ISMB₁**, (c) **ISMB₂**, (d) **ISMB₃** and (e) **ISMB₄** in the presence (red line) and absence (blue line) of PLL-g-Dex. Solution conditions; [MB] = 0.2 μ M, [Surv] = 0.8 μ M, [NaCl] = 100 mM, pH 7.0 (10 mM phosphate buffer), 20 °C, N/P = 2. Measurement conditions of JASCO model FP-6500; bandwidth of excitation = 5nm, bandwidth of emission = 5 nm, Sensitivity = medium.

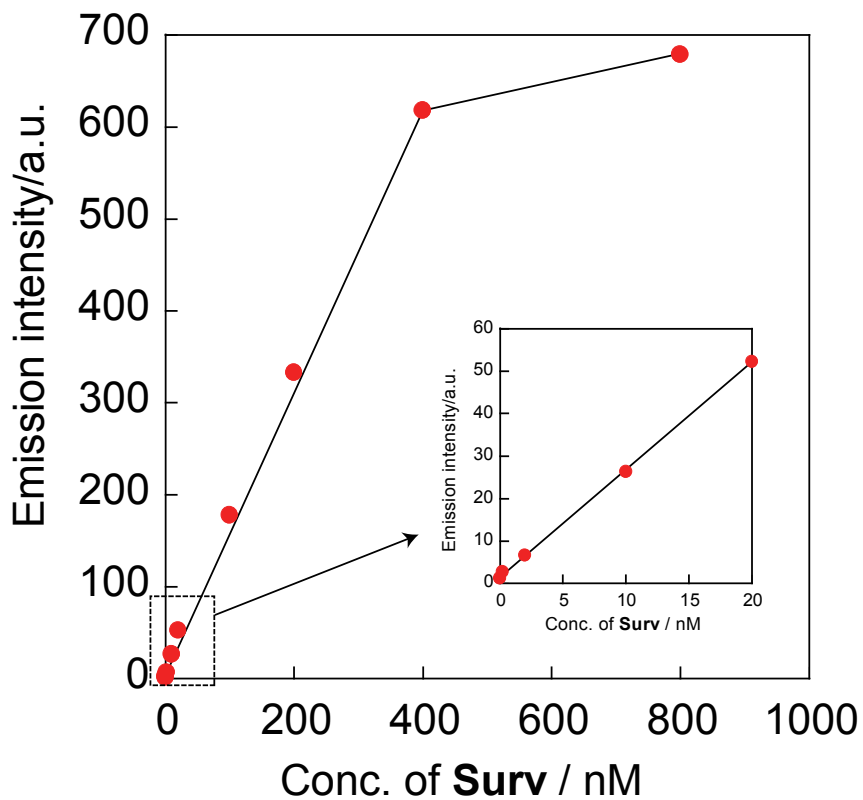


Figure S4. Linear range of **ISMB₃** for the detection of **Surv**. Plots at lower concentrations were magnified in inset. Solution conditions; [**ISMB₃**] = 200 nM, N/P = 2, [NaCl] = 100 mM, pH 7.0 (10 mM phosphate buffer), 20 °C. Measurement conditions of JASCO model FP-6500; bandwidth of excitation = 5nm, bandwidth of emission = 5 nm, Sensitivity = medium.

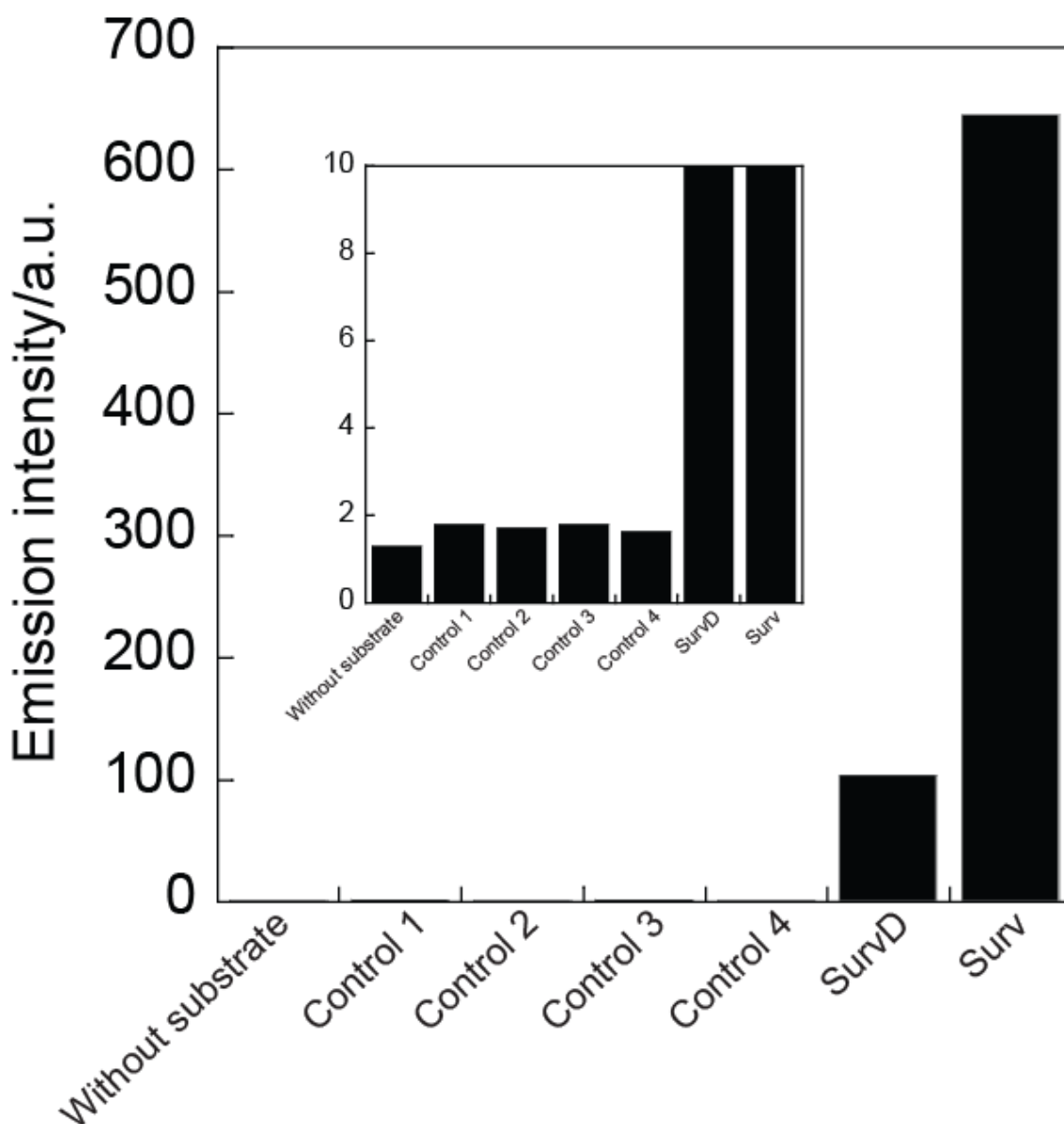


Figure S5. Change of fluorescence intensity of **ISMB₃** at 460 nm by the presence of DNA of various sequences excited at 425 nm with PLL-g-Dex. Solution conditions were as follows: [**ISMB₃**] = 0.2 μ M, [Substrate] = 0.8 μ M, [NaCl] = 100 mM, pH 7.0 (10 mM phosphate buffer), N/P = 2, 20 °C. Sequences of the substrates are listed as follows. All these oligonucleotides are a part of survivin gene.

ISMB₃ : 5'-GEGTEGGETC-CTTGAGAAAGGGC-GAQCCQACQC-3'

Control 1: 3'-AACTTACATCTCTACGCC-5'

Control 2: 3'-GAGGCCCCACGTCCGCGT-3'

Control 3: 3'-ATAAAAATCACCTCTGCC-5'

Control 4: 3'-AACTCTGCCTCAGAGTAA-5'

SurvD : 3'-ACGCCACAGGAACTCTTTCCCG-5' (One-base deletion mutant, underlined base of Surv is deleted)

Surv: 3'-ACGCCACCAGGAACTCTTTCCCG-5' (Full-match)

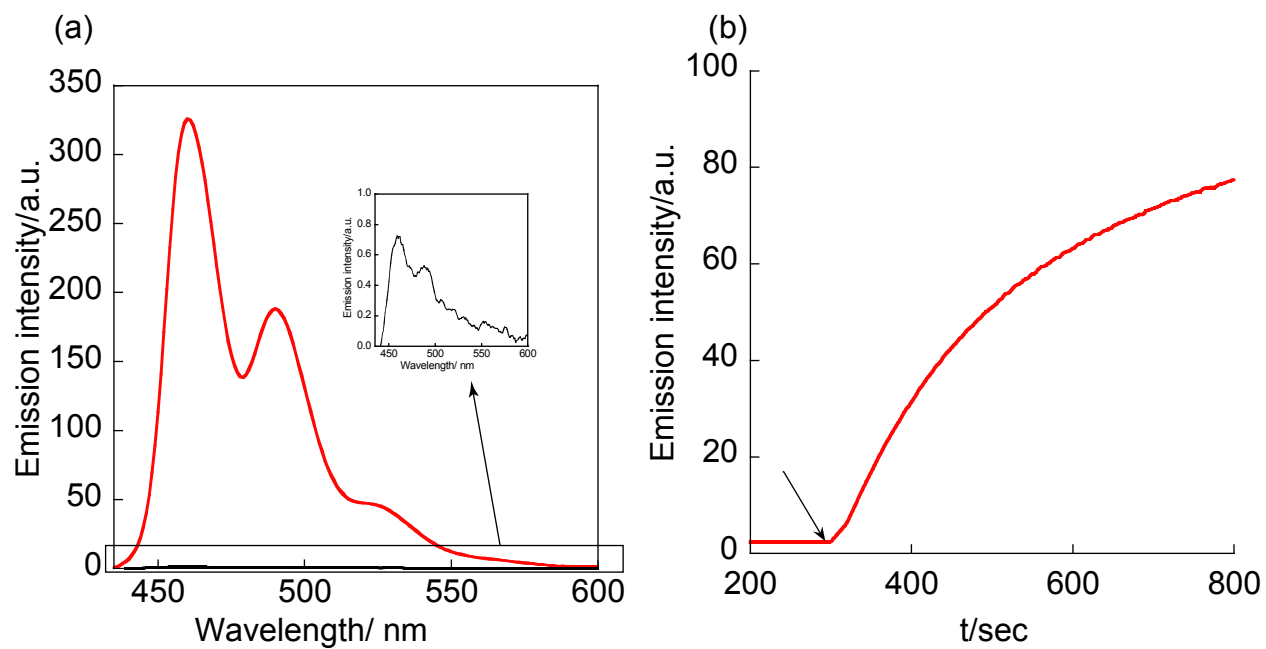


Figure S6. (a) Fluorescence emission spectra of **ISMB₃** in the absence (black line) and presence (red line) of **Surv** with 1 M NaCl at 20 °C. Solution conditions were 0.2 μM **ISMB₃**, 0.8 μM **Surv**, 1M NaCl, 10 mM phosphate buffer (pH 7.0). (b) Change in fluorescence intensity of **ISMB₃** with 1M NaCl at 20 °C. Arrow shows the point when **Surv** was added (300 sec). Solution conditions were 0.2 μM **ISMB₃**, 0.4 μM **Surv**, 1M NaCl, 10 mM phosphate buffer (pH 7.0), 20 °C.

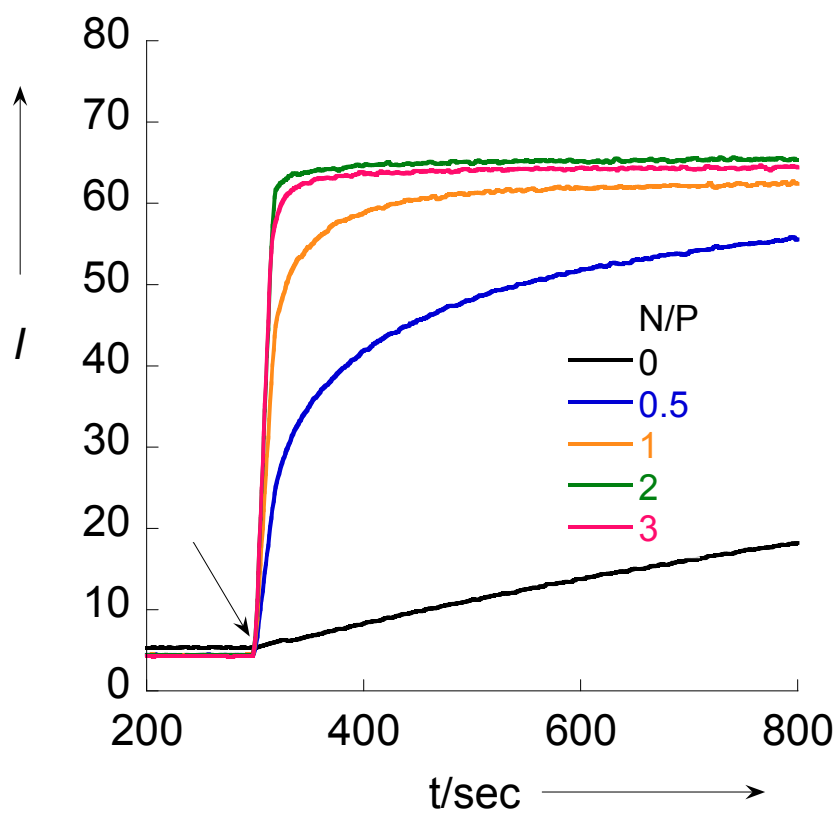


Figure S7. Response of ISMB₁ in the presence of PLL-g-Dex at various N/P ratios at 20 °C.

Black line, N/P = 0 (without PLL-g-Dex); blue line, N/P = 0.5; orange line, N/P = 1; red line, N/P = 2; green line, N/P = 3.

Arrows show the point when **Surv** was added (300 sec). Solution conditions are as follows: [ISMB₁] = 0.2 μM, [Surv] = 0.4 μM, [NaCl] = 100 mM, pH 7.0 (10 mM phosphate buffer), 20 °C.

Table S2. Signal/background (S/B) ratio of each MB in the presence and absence of PLL-g-Dex.^[a]

MB	w/o PLL-g-Dex ^[b]			with PLL-g-Dex ^[c]		
	S ^[d]	B ^[e]	S/B ^[f]	S ^[d]	B ^[e]	S/B ^[f]
MB_c	291.0	70.4	4.1	292	32.1	9.1
ISMB₁	370.0	27.1	13.7	330	19.1	17.1
ISMB₂	471.0	10.4	45.4	530	8.17	64.9
ISMB₃	14.7	1.33	11.1	679	1.19	571.0
ISMB₄	14.2	0.82	17.2	155	0.93	168.0

^[a]Solution conditions: 0.2 μM MB, 0.8 μM **Surv**, 100 mM NaCl, 10 mM phosphate buffer (pH 7), 20 °C. ^[b]N/P = 0. ^[c]N/P = 2. ^[d]Emission intensity at 460 nm in the presence of **Surv**. ^[e]Emission intensity at 460 nm in the absence of **Surv**. ^[f]The ratio of S with respect to B.