

Supplementary Information

Regioselectivity among six secondary hydroxyl groups: Selective acylation of the least reactive hydroxyl groups of inositol

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Materials and General Procedures

Chromatograms were visualized under UV light and by dipping plates into either phosphomolybdic acid in MeOH or anisaldehyde in ethanol, followed by heating. The ^1H NMR, COSY, and HMQC spectra were recorded on a Bruker (500 MHz) NMR spectrometer. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.0 ppm) or with the solvent reference relative to TMS employed as the internal standard (CDCl_3 , δ 7.26 ppm; D_2O , δ 4.79 ppm). Data are reported as follows: chemical shift (multiplicity [singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m)], coupling constants [Hz], integration and peak identification). All NMR signals were assigned on the basis of ^1H NMR, ^{13}C NMR, COSY and HMQC experiments. ^{13}C spectra were recorded with complete proton decoupling. Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard. All NMR data were collected at 25 °C. Melting points were determined using Stuart SMP30 melting point apparatus and are uncorrected. Flash column chromatography was performed using Silica Gel (200-400 mesh). All reactions were carried out under argon or nitrogen atmosphere employing oven dried glassware.

Preparation of H_2SO_4 -silica

A slurry of 20 g of silica gel (200-400 mesh, FINAR make) with 150 mL of diethyl ether was made in a 250 mL RB flask. To this slurry, 1 mL of conc. H_2SO_4 was added very slowly with vigorous shaking over a period of 30 min. The ether was evaporated under reduced pressure and the free flowing silica was heated at 120 °C in an oven for 1h. The resulting solid was kept under high vacuum at room temperature for 2h and was stored in a desiccator.

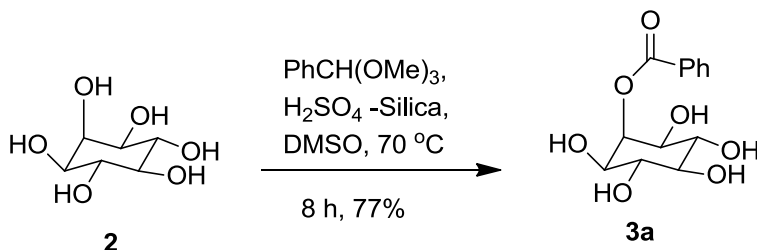
General procedure for the synthesis of 2-*O*-acyl-*myo*-inositol

A mixture of *myo*-inositol (2 mmol) and trialkylorthoester (2.4 mmol) in dry DMF / DMSO (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 6-12 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed successively with reaction solvent. The combined filtrate was evaporated under reduced pressure (freeze drying in case of DMSO). The residue obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield respective 2-*O*-acyl-*myo*-inositol.

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Synthetic Procedures and Characterization

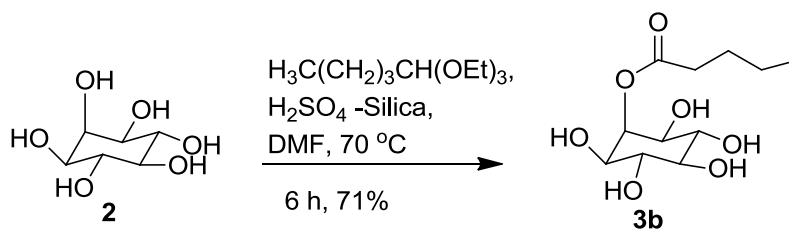
2-O-benzoyl-*myo*-inositol (3a)



A mixture of *myo*-inositol **2** (0.36 g, 2 mmol) and trimethylorthobenzoate (0.42 mL, 2.4 mmol) in dry DMSO (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 8 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed with DMSO. The DMSO was removed by freeze drying. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield compound **3a** (0.44 g, 77%) as a white solid.

mp: 232-234 °C. ^1H NMR (500 MHz, DMSO-d_6) δ 7.96-7.94 (m, 2H, Ar-H), 7.66 (t, J = 7.3 Hz, 1H, Ar-H), 7.55 (t, J = 7.7 Hz, 2H, Ar-H), 5.44 (s, 1H,), 4.91-4.79 (m, -OH), 3.49-3.15 (m, 4H), 3.06 (t, J = 6.4 Hz, H-5); ^{13}C NMR (125 MHz, DMSO-d_6) δ 165.10, 132.93, 130.60, 129.14, 12.53, 75.75, 74.83, 73.21, 69.89. Elemental analysis: calcd for $\text{C}_{13}\text{H}_{16}\text{O}_7$: C, 54.93; H, 5.67%. Found: C, 54.76; H, 5.39%.

2-O-pentanoyl-*myo*-inositol (3b)

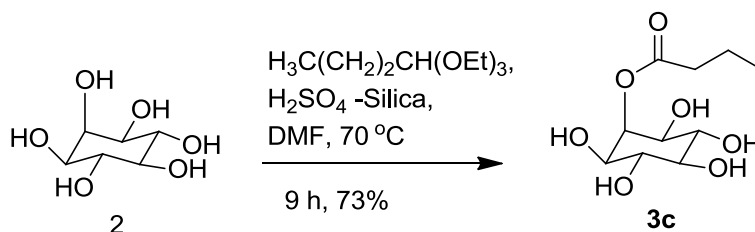


A mixture of *myo*-inositol (0.36 g, 2 mmol) and triethylorthovalerate (0.57 mL, 2.4 mmol) in dry DMF (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 6 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed with DMF. The DMF was evaporated under reduced pressure. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield compound **3b** (0.37 g, 71%) as a white solid.

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mp: 155-157 °C. ^1H NMR (500 MHz, DMSO- d_6) δ 5.20 (t, J = 2.6 Hz, H-2), 4.73 (bs, -OH), 3.34-3.26 (m, 4H, Ins-H), 2.97 (t, J = 8.6 Hz, H-5), 2.28 (t, J = 7.3 Hz, 2H), 1.54-1.50 (m, 2H), 1.49-1.31 (m, 2H), 0.88 (t, J = 7.3 Hz, 3H); ^{13}C NMR (125 MHz, DMSO- d_6) δ 172.13, 74.80, 74.45, 73.00, 69.73, 33.65, 26.65, 21.54, 13.67. Elemental analysis: calcd for $\text{C}_{11}\text{H}_{20}\text{O}_7$: C, 49.99; H, 7.63%. Found: C, 49.83; H, 7.51%.

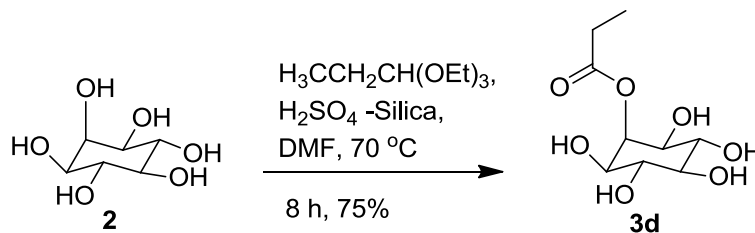
2-O-butanoyl-*myo*-inositol (3c)



A mixture of *myo*-inositol (0.36 g, 2 mmol) and triethylorthobutyrate (0.52 mL, 2.4 mmol) in dry DMF (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 9 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed with DMF. The DMF was evaporated under reduced pressure. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield compound **3c** (0.363 g, 73%) as a white solid.

mp: 158-160 °C. ^1H NMR (500 MHz, DMSO- d_6) δ 5.19 (t, J = 2.5 Hz, H-2), 4.70 (bs, -OH), 3.33-3.25 (m, 4H, Ins-H), 2.96 (t, J = 8.6 Hz, H-5), 2.24 (t, J = 7.2 Hz, 2H), 1.56-1.52 (m, 2H), 0.90 (t, J = 7.4 Hz, 3H); ^{13}C NMR (125 MHz, DMSO- d_6) δ 171.99, 74.83, 74.43, 73.03, 69.77, 35.89, 18.05, 13.47. Elemental analysis: calcd for $\text{C}_{10}\text{H}_{18}\text{O}_7$: C, 48.00; H, 7.25%. Found: C, 48.24; H, 7.43%.

2-O-propanoyl-*myo*-inositol (3d)



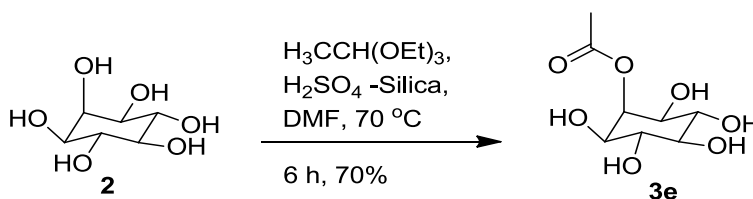
A mixture of *myo*-inositol (0.36 g, 2 mmol) and triethylorthopropionate (0.48 mL, 2.4 mmol) in dry DMF (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 8 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed with DMF. The DMF was evaporated under reduced

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pressure. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield compound **3d** (0.349 g, 75%) as a white solid.

mp: 164-166 °C. ^1H NMR (500 MHz, DMSO- d_6) δ 5.19 (t, J = 2.6 Hz, H-2), 3.54 (bs, -OH), 3.33-3.27 (m, 4H, Ins-H), 2.97 (t, J = 8.5 Hz, H-5), 2.30 (q, J = 7.5 Hz, 2H), 1.03 (t, J = 7.5 Hz, 3H); ^{13}C NMR (125 MHz, DMSO- d_6) δ 173.00, 74.61, 74.46, 72.81, 69.61, 27.14, 9.06. Elemental analysis: calcd for $\text{C}_9\text{H}_{16}\text{O}_7$: C, 45.76; H, 6.83%. Found: C, 45.48; H, 6.91%.

2-O-acetyl-*myo*-inositol (3e)



A mixture of *myo*-inositol (0.36 g, 2 mmol) and triethylorthoacetate (0.37 mL, 2.4 mmol) in dry DMF (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 6 h under argon atmosphere. The resulting solution was neutralized by adding solid NaHCO_3 , filtered and washed with DMF. The DMF was evaporated under reduced pressure. The residue obtained was chromatographed using isopropanol and ethyl acetate (1:9 v/v) to yield compound **3e** (0.31 g, 70%) as a white solid.

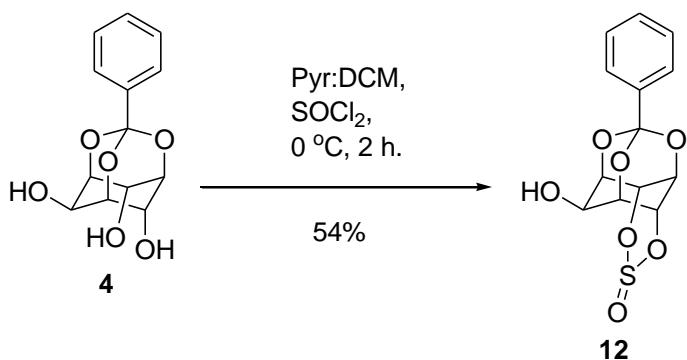
mp: 170-172 °C. ^1H NMR (500 MHz, DMSO- d_6) δ 5.18 (t, J = 2.5 Hz, H-2), 4.70 (bs, -OH), 3.33-3.27 (m, 4H, Ins-H), 2.97 (t, J = 8.5 Hz, H-5), 1.99 (s, 3H); ^{13}C NMR (125 MHz, DMSO- d_6) δ 169.69, 74.83, 74.68, 72.99, 69.76, 21.05. Elemental analysis: calcd for $\text{C}_8\text{H}_{14}\text{O}_7$: C, 43.24; H, 6.35%. Found: C, 43.11; H, 6.17%.

2-O-formyl-*myo*-inositol (3f)

A mixture of *myo*-inositol (0.36 g, 2 mmol) and triethylorthoformate (0.42 mL, 2.4 mmol) in dry DMF (10 mL) was stirred in presence of H_2SO_4 -silica (50 mg) at 70 °C for 6 h under argon atmosphere. A major spot (R_f 0.15, isopronaol: ethylacetate 1:9 v/v) was observed just above the *myo*-inositol. The resulting solution was neutralized by adding solid NaHCO_3 and the filtrate was evaporated. However, during the filtration and chromatography the labile formate ester got hydrolyzed to the inositol.^[1]

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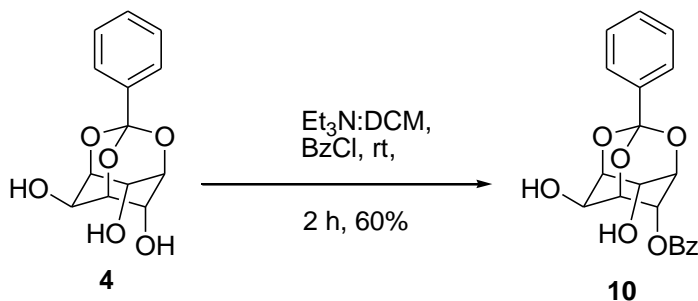
Myo-inositol-4,6-O-sulfite-1,3,5-orthobenzoate (12)



To a solution of *myo*-inositol-1,3,5-orthobenzoate^[2] (0.11 g, 0.41 mmol) in a mixture of pyridine and dichloromethane (1:4, v/v, 10 mL) was added thionylchloride (0.04 mL, 0.49 mmol) drop wise at 0 °C. The reaction was stirred at 0 °C for 2 h. After the completion of starting material (checked by TLC), the reaction mixture was concentrated under reduced pressure. The residue was co-evaporated with toluene to dryness. The resulting residue was adsorbed on silica by dissolving it into dichloromethane. The crude product was purified by column chromatography using ethyl acetate and petroleum ether (1:4, v/v) to yield **12** (0.08 g, 54 %) as a white solid.

mp: 115-117 °C. ¹H NMR (500 MHz, CDCl₃) δ 7.56-7.54 (m, 2H, Ar-*H*), 7.36-7.31 (m, 3H, Ar-*H*), 5.84 (t, *J* = 4.2 Hz, 1H, H-5), 5.22 (t, *J* = 4.5 Hz, 2H, H-4 & H-6), 4.46 (dd, *J* = 2.85 and 1.60 Hz, 2H, H-1 & H-3), 4.14 (bs, 1H, H-2); ¹³C NMR (125 MHz, CDCl₃) δ 135.51, 130.22, 128.25, 125.22, 107.31, 72.52, 68.55, 60.70, 60.27. Elemental analysis: calcd for C₁₃H₁₂O₇S: C, 50.00; H, 3.87; S, 10.27%. Found: C, 50.18; H, 3.79; S, 10.42%.

4-O-benzoyl-myio-inositol 1,3,5-orthobenzoate (10)

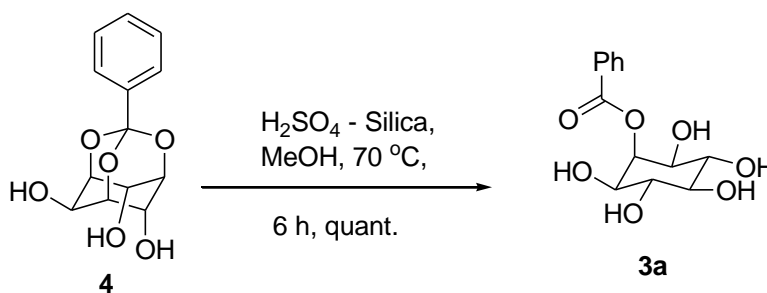


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To a solution of *myo*-inositol-1,3,5-orthobenzoate (0.180 g, 0.67 mmol) in a mixture of triethylamine and dichloromethane (1:4, v/v, 10 mL) benzoyl chloride (0.09 mL, 0.74 mmol) was added drop wise at 0 °C. The reaction was allowed to stir at rt for 2 h. After the completion of starting material (checked by TLC), the reaction mixture was concentrated under reduced pressure. The residue was dissolved in dichloromethane (20 mL) and washed successively with water (3 x 10 mL) and brine (10 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude product was purified by column chromatography using ethyl acetate and petroleum ether (1:4, v/v) to yield **10** (0.145 g, 60 %) as a white solid.

mp: 135-137 °C. ¹H NMR (500 MHz, CDCl₃) δ 7.94-7.92 (m, 2H, Ar-*H*), 7.61-7.59 (m, 2H, Ar-*H*), 7.54 (t, *J* = 7.5 Hz, 1H, Ar-*H*), 7.39 (t, *J* = 7.5 Hz, 2H, Ar-*H*), 7.33-7.31 (m, 3H, Ar-*H*), 5.87-5.86 (m, 1H, H-4), 4.73 (bs, 1H, H-6), 4.58-4.57 (m, 1H, H-5), 4.50 (dd, *J* = 3.90 and 1.80 Hz, 1H, H-3), 4.39 (dd, *J* = 3.90 and 1.90 Hz, 1H, H-1), 4.22 (bs, 1H, H-2); ¹³C NMR (125 MHz, CDCl₃) δ 165.03, 136.40, 133.83, 129.88, 129.81, 128.91, 128.74, 128.18, 125.30, 107.80, 75.62, 73.31, 69.05, 68.59, 67.35, 60.29. Elemental analysis: calcd for C₂₀H₁₈O₇: C, 64.86; H, 4.90%. Found: C, 64.63; H, 4.87%.

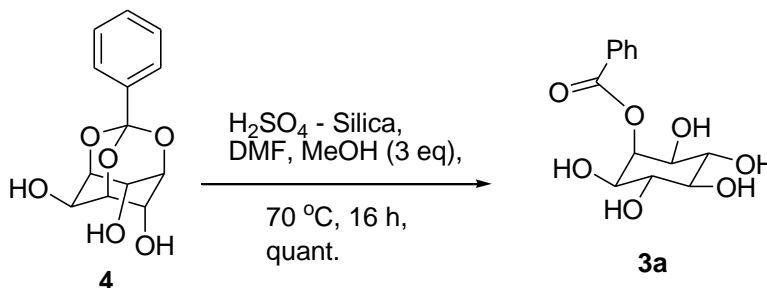
Hydrolysis of *myo*-inositol 1,3,5-orthobenzoate (**4**) in wet methanol



A mixture of *myo*-inositol-1,3,5-orthobenzoate (0.05 g, 0.18 mmol) and H₂SO₄-silica (20 mg) in methanol (5 mL) was stirred at 70 °C. The reaction was monitored by TLC. When the reaction was complete (6 h), the reaction mixture was neutralized by adding solid NaHCO₃, filtered and washed with MeOH (10 mL). The MeOH was evaporated under reduced pressure. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9, v/v) to yield compound **3a** (0.045 g, quant) as a white solid.

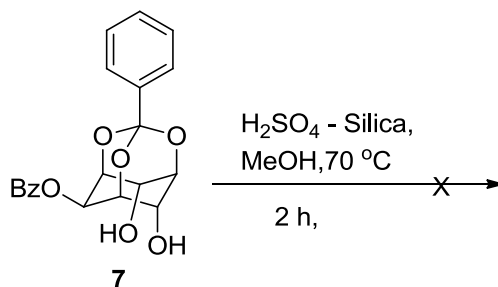
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Hydrolysis of *myo*-inositol 1,3,5-orthobenzoate (4) in dry DMF and 3 equiv of methanol



A mixture of *myo*-inositol-1,3,5-orthobenzoate (0.05 g, 0.18 mmol), dry methanol (22 μL , 0.54 mmol, 3 eq) and H_2SO_4 -silica (20 mg) in dry DMF (5 mL) was stirred at $70\text{ }^\circ\text{C}$ under argon atmosphere. The reaction was monitored by TLC. When the reaction was complete (16 h), the reaction mixture was neutralized by adding solid NaHCO_3 , filtered and washed with MeOH (10 mL). The solvents were evaporated under reduced pressure. The residue thus obtained was chromatographed using isopropanol and ethyl acetate (1:9, v/v) to yield compound 3a (0.045 g, quant) as a white solid.

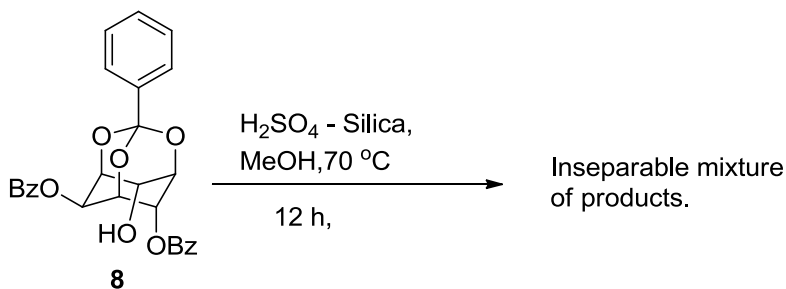
Hydrolysis of 2-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate (7)



To a solution of 2-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate^[3] (0.05 g, 0.13 mmol) in methanol (5 mL) was added H_2SO_4 -silica (20 mg) and stirred the reaction at $70\text{ }^\circ\text{C}$ for 2 h. There was no change in starting material as judged by TLC. The reaction was further heated to $80\text{ }^\circ\text{C}$ for 12 h. There was no change in reaction mixture and the starting material could be recovered.

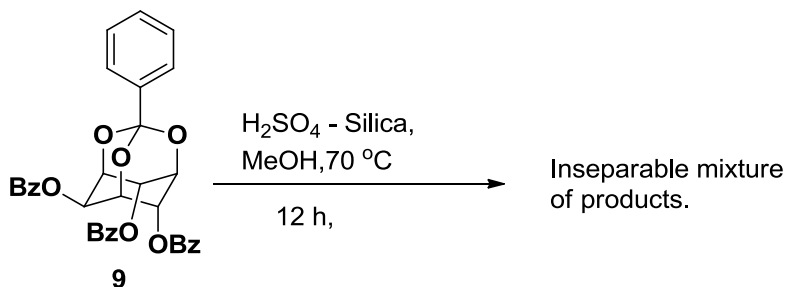
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Hydrolysis of 2,4-di-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate (8)



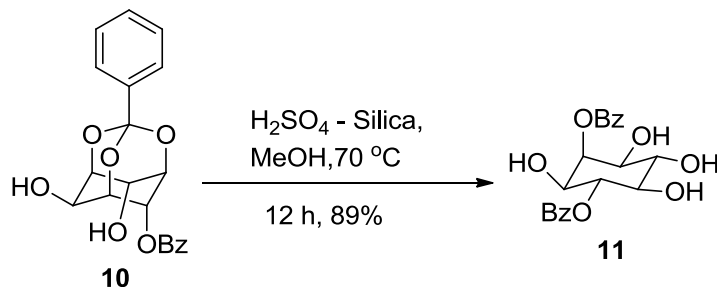
To a solution of 2,4-di-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate^[3] (0.05 g, 0.10 mmol) in methanol (5 mL), H₂SO₄-silica (20 mg) was added and the mixture was stirred at 70 °C for 12 h. There was no reaction as judged by TLC. However, an inseparable mixture of high polar products was obtained when the reaction was heated to 80 °C for 60 h.

Hydrolysis of 2,4,6-tri-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate (9)



To a solution of 2,4,6-tri-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate^[3] (0.05 g, 0.086 mmol) in methanol (5 mL), H₂SO₄-silica (20 mg) was added and the mixture was stirred at 70 °C for 12 h. There was no change in starting material as judged by TLC. As in the case of dibenzoate, an inseparable mixture of products was obtained when the reaction was done at higher temperature (80 °C) for several four days.

Hydrolysis of 4-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate (10)



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To a solution of 4-*O*-benzoyl-*myo*-inositol 1,3,5-orthobenzoate (0.15 g, 0.4 mmol) in methanol (10 mL), H₂SO₄-silica (50 mg) was added and the mixture was stirred at 70 °C. The reaction was monitored by TLC. When the reaction was complete (12 h), it was quenched by adding solid NaHCO₃. The resulting mixture was filtered and washed with methanol. The solvent was evaporated under reduced pressure. The crude product was purified by flash chromatography using ethyl acetate and petroleum ether (9:1, v/v) as the eluent to get the known⁴ compound **11** (0.14 g, 89%) as a white solid. The ¹H NMR of **11** was reported in CD₃OD. We have compared the NMR data of our sample in CD₃OD with the reported data and found that they are identical. However, we observed that dibenzoate **11** underwent some transesterification in CD₃OD to give minor amounts of impurities (See ¹H NMR spectrum on page No. S62). But dibenzoate **11** was stable in DMSO-d₆ solution. So we have recorded the ¹H NMR spectrum of **11** in DMSO-d₆ to establish its purity. Also a COSY NMR was done to assign the peaks.

Reported data⁴

mp: 193-195 °C, ¹H NMR (500 MHz, CD₃OD) δ 8.14-7.43 (m, 10H, 2Ph), 5.75 (app. t, *J* = 2.8 Hz, 1H, H-2), 5.53 (app. t, *J* = 9.9 Hz, 1H, H-4), 4.01 (dd, *J* = 10.1 Hz & 2.8 Hz, 1H, H-3), 3.85 (app. t, *J* = 9.5 Hz, 1H, H-6), 3.73 (dd, *J* = 9.8 Hz & 2.7 Hz, 1H, H-1), 3.60 (app. t, *J* = 9.4 Hz, 1H, H-5).

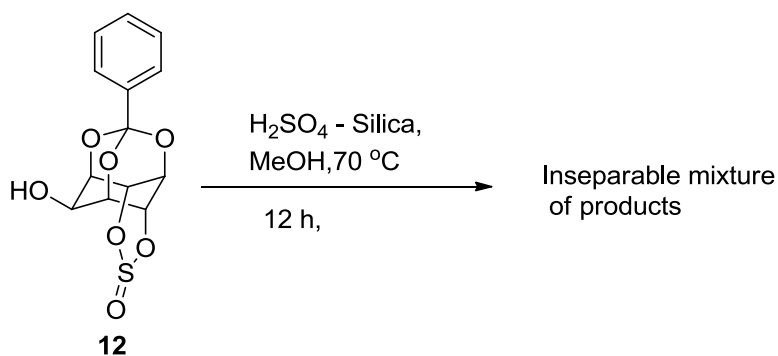
Data obtained

mp: 194 °C, ¹H NMR (500 MHz, CD₃OD) δ 8.03 (d, *J* = 7.3 Hz, 2H, Ar-H), 7.98 (d, *J* = 7.3 Hz, 2H, Ar-H), 7.54-7.47 (m, 2H, Ar-H), 7.43-7.35 (m, 4H, Ar-H), 5.65 (t, *J* = 2.8 Hz, 1H, H-2), 5.44 (t, *J* = 9.9 Hz, 1H, H-4), 3.92 (dd, *J* = 10.1 Hz & 2.8 Hz, 1H, H-3), 3.76 (t, *J* = 9.6 Hz, 1H, H-6), 3.64 (dd, *J* = 9.8 Hz & 2.8 Hz, 1H, H-1), 3.51 (t, *J* = 9.4 Hz, 1H, H-5).

¹H NMR (500 MHz, DMSO-d₆) δ 8.03-7.98 (m, 4H, Ar-H), 7.71-7.52 (m, 6H, Ar-H), 5.53 (t, *J* = 2.5 Hz, 1H, H-2), 5.32-5.28 (m, 2H, H-4, 3-OH), 5.16 (d, *J* = 5.6 Hz, 5-OH), 5.09-5.04 (m, 2H, 1-OH, 6-OH), 3.92-3.88 (m, 1H, H-3), 3.63-3.58 (m, 2H, H-1, H-6), 3.46-3.43 (m, 1H, H-5).

Hydrolysis of *myo*-inositol-4,6-*O*-sulfite-1,3,5-orthobenzoate (**12**)

Supplementary Information

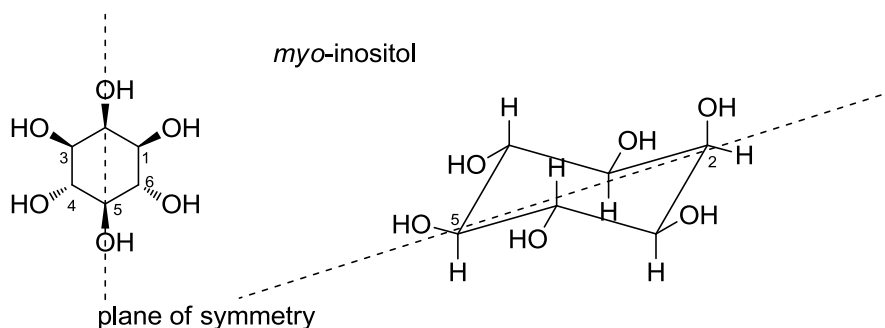


A mixture of *myo*-inositol-4,6-*O*-sulfite-1,3,5-orthobenzoate (0.05 g, 0.16 mmol) and H_2SO_4 -silica (20 mg) in methanol (5 mL) was stirred at $70\text{ }^\circ\text{C}$ for 12 h. There was no change in starting material as judged by TLC. But prolonged (4 days) heating at $80\text{ }^\circ\text{C}$ resulted in the formation of an inseparable mixture of products.

Structural Assignment of 2-*O*-esters

Myo-inositol being a meso compound with a plane of symmetry through C2 and C5 (see figure below), esterification of C2-OH or C5-OH gives a meso product and hence a symmetrical NMR spectrum with four signals in the ratio 1:2:2:1 and monoesterification at any other hydroxyl will result in an unsymmetrical product (hence unsymmetrical spectra). ^{13}C NMR of all the monoesters obtained from H_2SO_4 -silica catalyzed reaction between *myo*-inositol and trialkylorthoester revealed that they are symmetrical compounds. Since H-2 is an equatorially oriented hydrogen flanked by two axially oriented hydrogens on neighboring carbons (dihedral angle ϕ around 60°), its $^3J_{\text{HH}}$ coupling constants will be small. As H-5 is an axial hydrogen flanked by two axial hydrogens (dihedral angle ϕ around 180°), its $^3J_{\text{HH}}$ coupling constants will be higher. H of $-\text{CH}(\text{OH})-$ motif will shift downfield on esterification of the OH and hence it is easy to determine the position of esterification. The most downfield shifted hydrogen ($\delta > 5.2$ ppm; as a result of esterification) in all these monoesters (**3a-e**), showed a smaller coupling constant of ≈ 2.6 Hz suggesting that 2-OH is acylated and the other lone proton (upfield) showed a large coupling constant of ≈ 8.6 Hz (H-5) confirming that 5-OH is not acylated. These signal assignments were further confirmed by 2D NMR.

Supplementary Information



Reactivity comparison of H₂SO₄-silica with other acid catalysts

Treatment of *myo*-inositol with trialkylorthoesters in presence of acid catalysts such as PTSA,⁵ Camphorsulfonic acid,⁶ Amberlyst⁷ and triflic acid⁸ have been reported to yield *myo*-inositol 1,3 5-orthoesters in good yields. We have reproduced some of these results in our own lab several times and in all the cases, we got only the corresponding orthoesters but not the 2-O-ester. Thus H₂SO₄-silica is a special case wherein the corresponding 2-O-ester is formed regioselectively instead of the orthoesters.

Establishment of regioselectivity

The ¹³C NMR of the crude reaction mixture showed minor amounts of inositol along with the major 2-O-esters. In order to prove that the minor impurity was not any other isomeric ester but just inositol, we have added *myo*-inositol to a pure sample of **3a** (approximately 1:1 ratio) and recorded its ¹³C NMR. A comparison of this mixed NMR spectrum with ¹³C NMR spectrum of the crude reaction mixture established that the minor impurity in the crude mixture is just inositol.

References

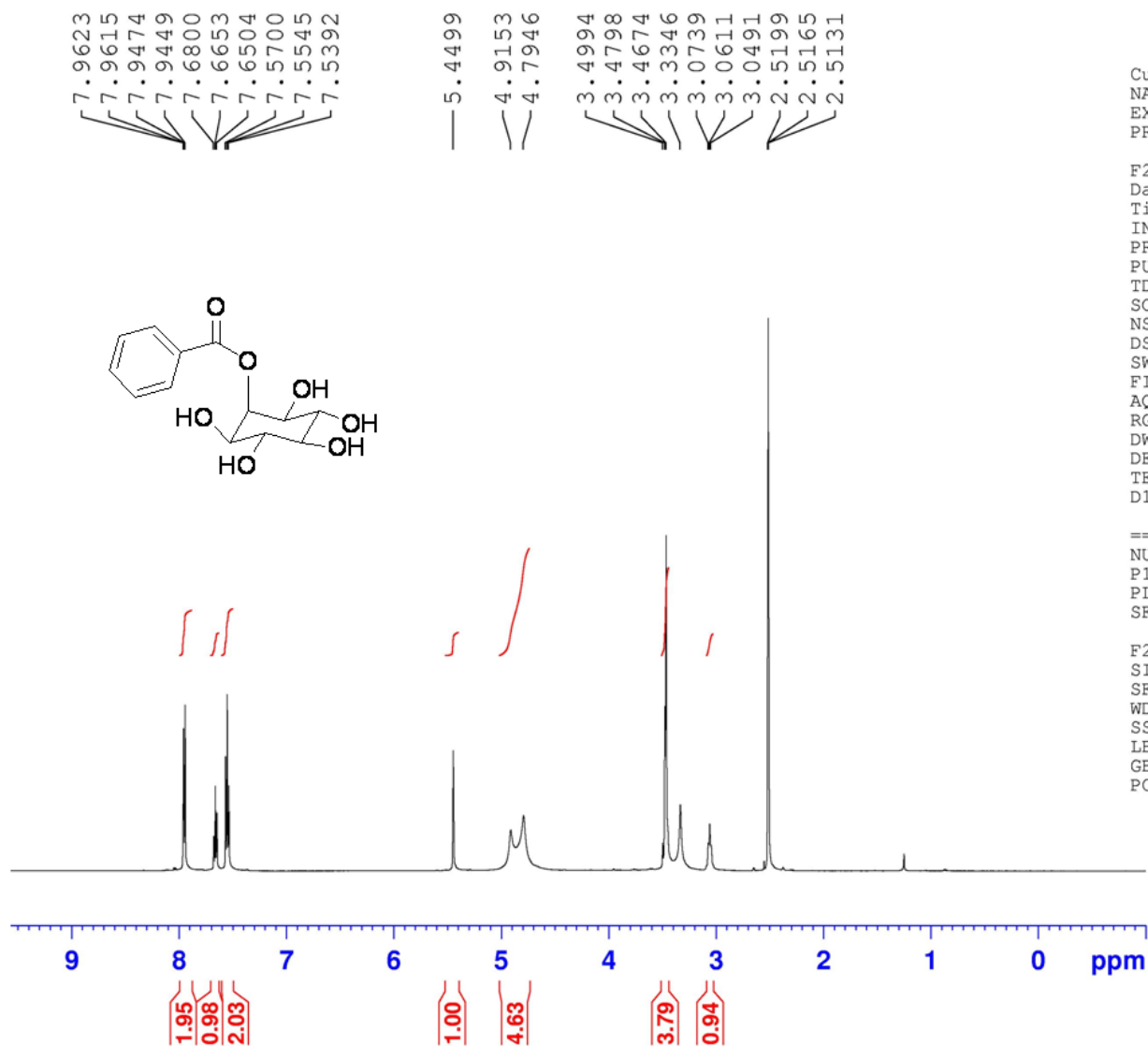
- [1] S. W. Garret, C. Liu, A. M. Riley, B. V. L. Potter, *J. Chem. Soc. Perkin Trans. 1*, **1998**, 1367-1368.
- [2] G. Bhosekar, C. Murali, R. G. Gonnade, M. S. Shashidhar, M. M. Bhadbhade, *Crystal Growth & Design* **2005**, 5, 1977-1982.
- [3] C. Murali, R. G. Gonnade, M. S. Shashidhar, M. M. Bhadbhade, *Eur. J. Org. Chem.* **2007**, 1153–1159.
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- [5] H. W. Lee and Y. Kishi. *J. Org. Chem.* **1985**, 50, 4402-4404.
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Supplementary Information

- [7] A. Krief, W. Dumont, D. Billen, J. J. Leteson, P. Lestrade, P. J. Murphy and D. Lacroix *Tetrahedron Lett.* **2004**, 45, 1461-1463.
- [8] P. Andersch and M. P. Schneider, *Tetrahedron Asym.* **1993**, 4, 2135-2138.

Supplementary Information

¹H NMR of **3a** in DMSO- d₆



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EXPNO 249
PROCNO 1

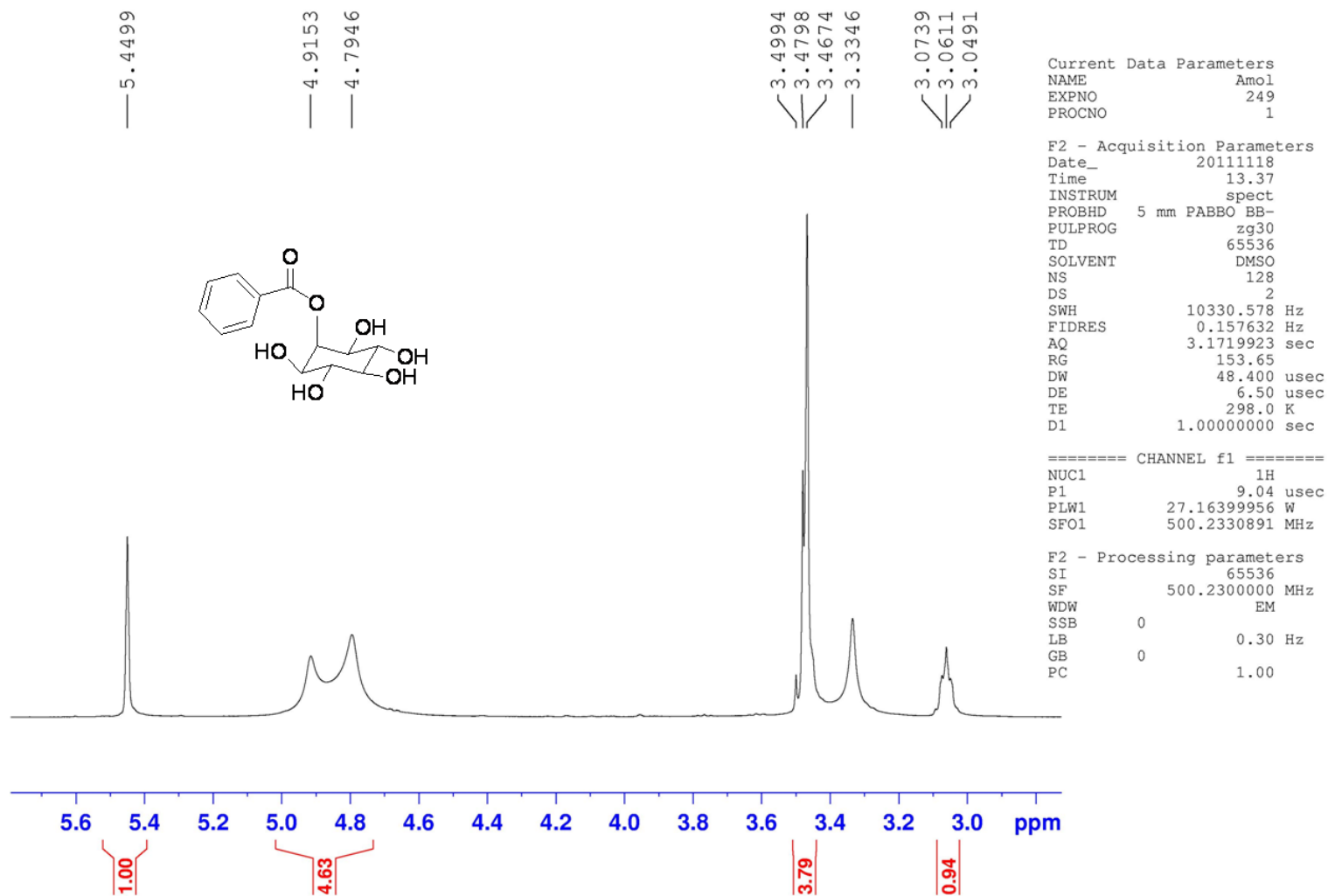
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Time 13.37
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 128
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 153.65
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

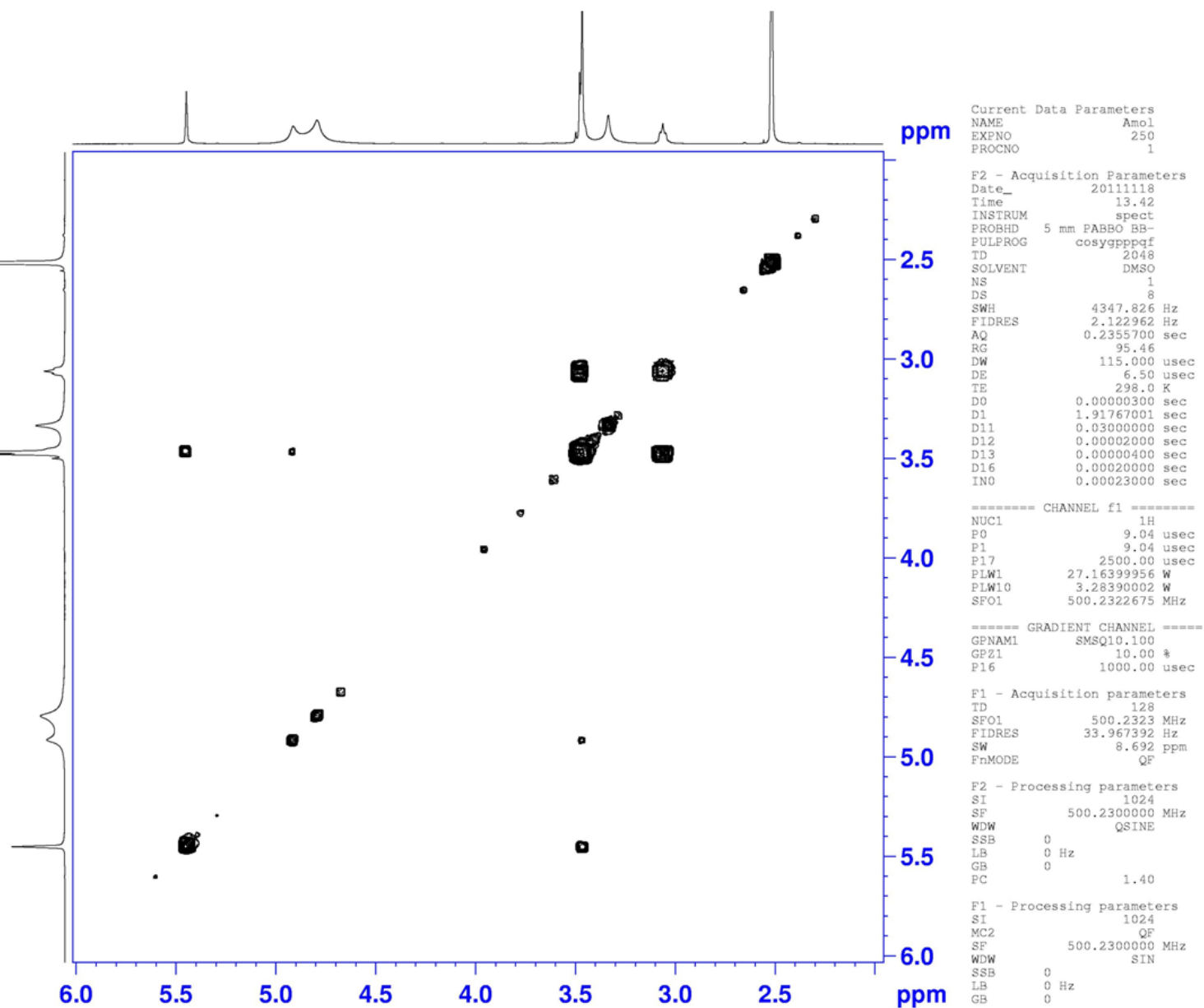
Supplementary Information

¹H NMR of **3a** (zoom)



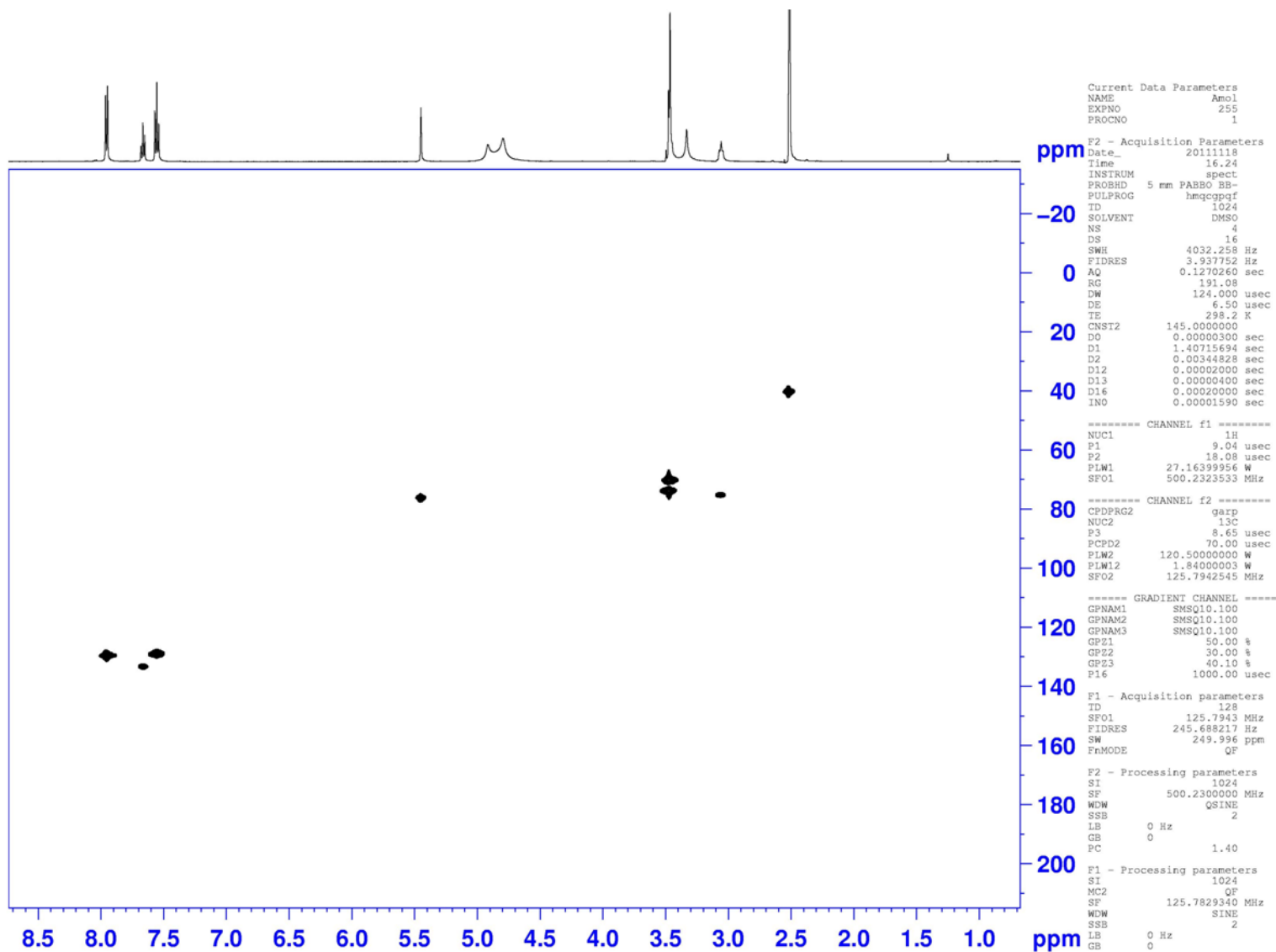
Supplementary Information

COSY of 3a



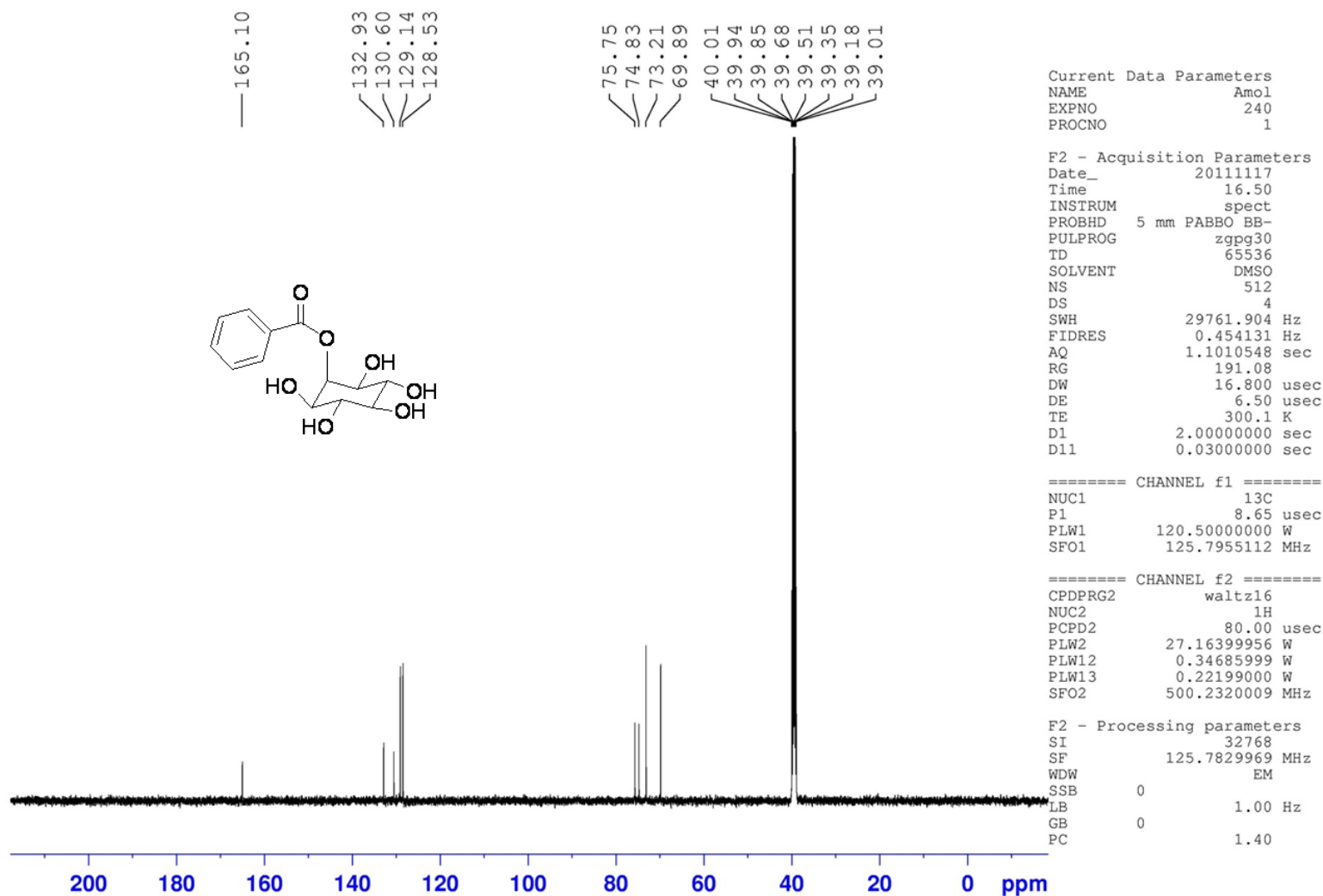
Supplementary Information

HMQC of 3a



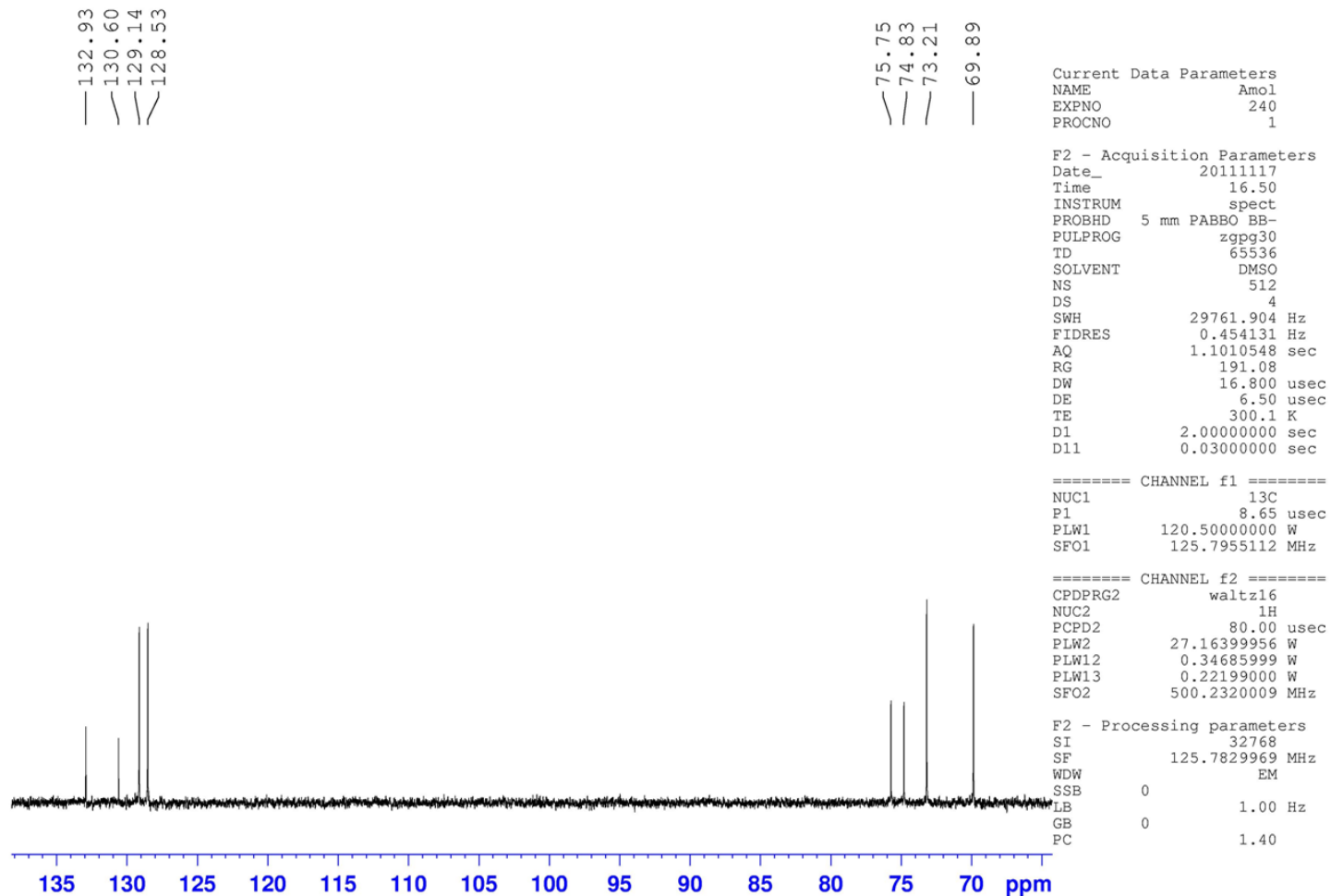
Supplementary Information

¹³C NMR of **3a** in DMSO-d₆



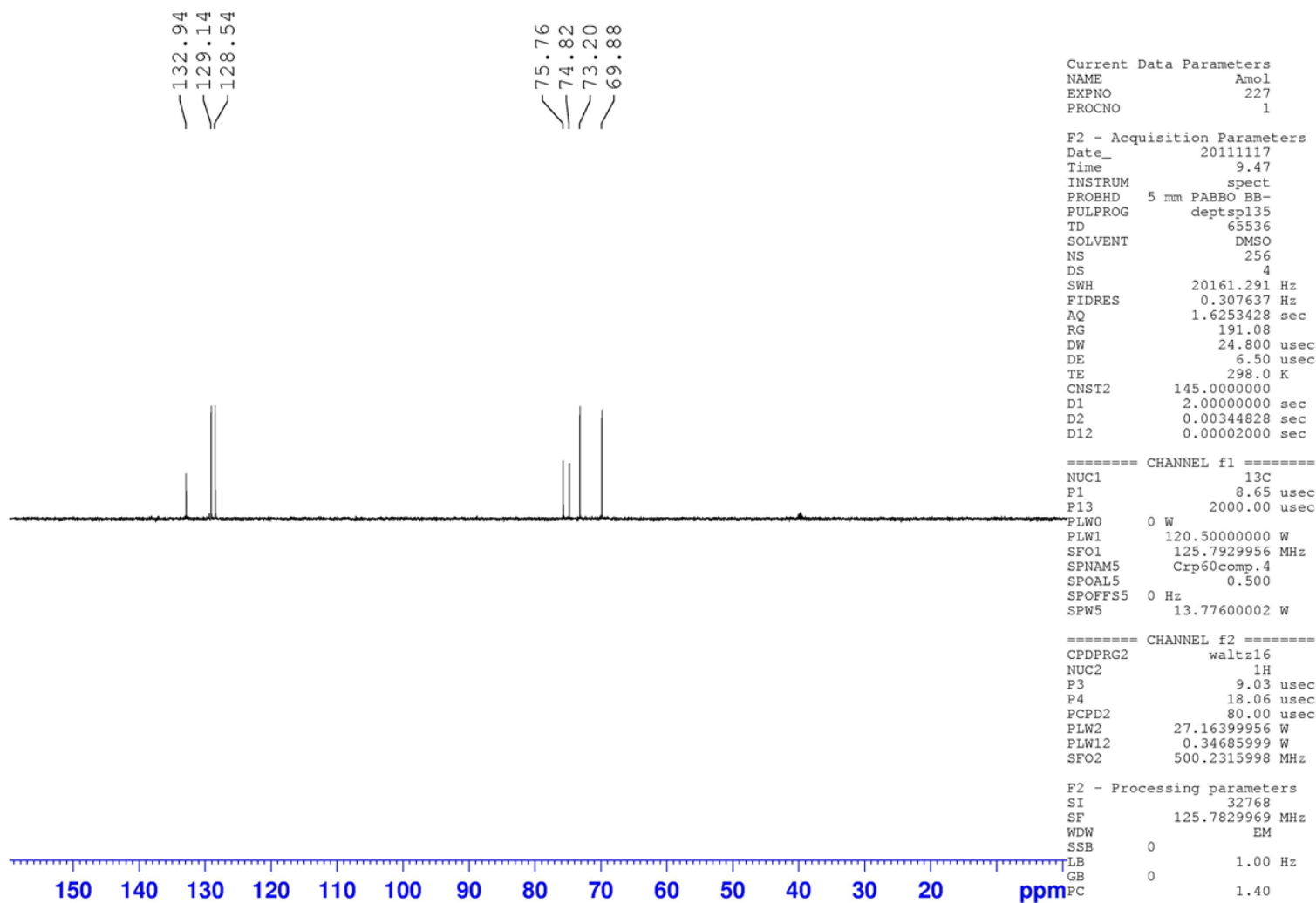
Supplementary Information

¹³C NMR of **3a** (zoom)



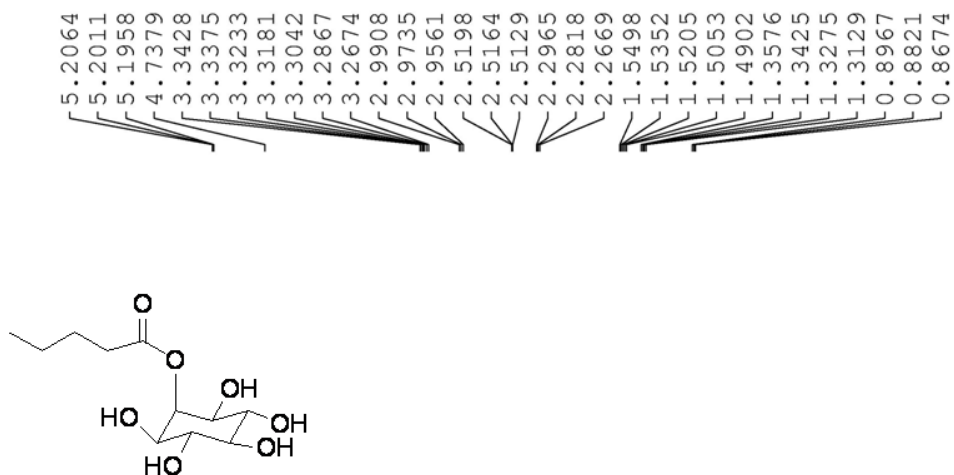
Supplementary Information

DEPT of 3a



Supplementary Information

^1H NMR of **3b** in DMSO-d_6

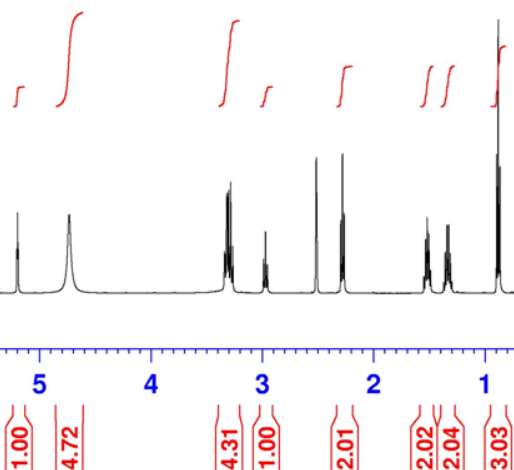


Current Data Parameters
NAME Amol
EXPNO 198
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111114
Time 0.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 64
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 107.19
DW 48.400 usec
DE 6.50 usec
TE 295.5 K
D1 1.00000000 sec

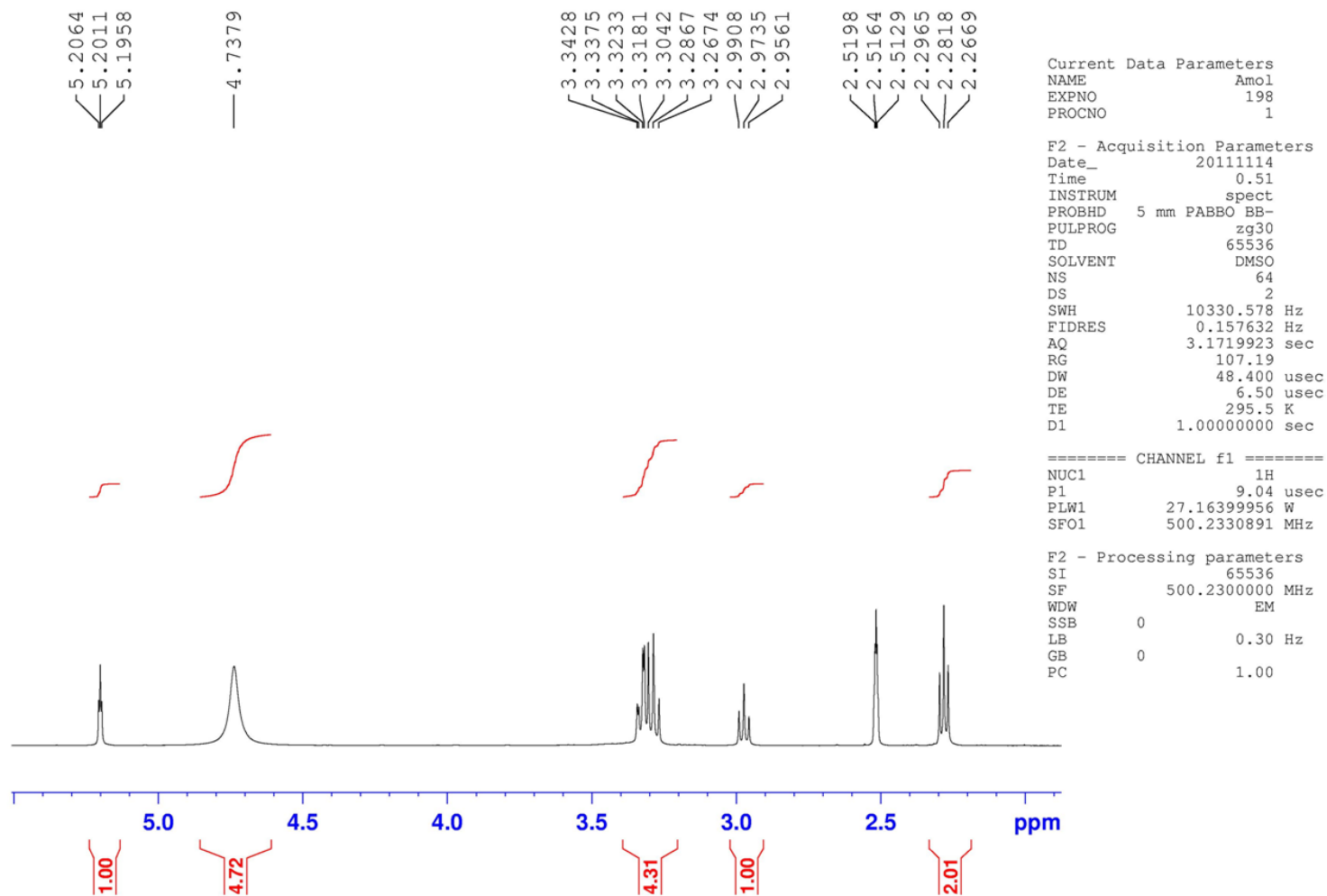
===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Supplementary Information

¹H NMR of **3b** (zoom)



Current Data Parameters

NAME	Amol
EXPNO	199
PROCNO	1

F2 - Acquisition Parameters

Date_	20111114
Time	0.52
INSTRUM	spect
PROBHD	5 mm FABBO BB-
PULPROG	cosygpppqf
TD	2048
SOLVENT	DMSO
NS	1
DS	8
SWH	3597.122 Hz
FIDRES	1.756407 Hz
AQ	0.2847220 sec
RG	64.12
DW	139.000 usec
DE	6.50 usec
TE	295.5 K
D0	0.00000300 sec
D1	1.86851799 sec
D11	0.03000000 sec
D12	0.00002000 sec
D13	0.00000400 sec
D16	0.00020000 sec
IN0	0.00027800 sec

===== CHANNEL f1 =====

NUC1	1H
P0	9.04 usec
P1	9.04 usec
P17	2500.00 usec
PLW1	27.16399956 W
PLW10	3.28390002 W
SFO1	500.2313902 MHz

===== GRADIENT CHANNEL =====

GPNAME1	SMSQ10.100
GPZ1	10.00 %
P16	1000.00 usec

F1 - Acquisition parameters

TD	128
SFO1	500.2314 MHz
FIDRES	28.102518 Hz
SW	7.191 ppm
FnMODE	QF

F2 - Processing parameters

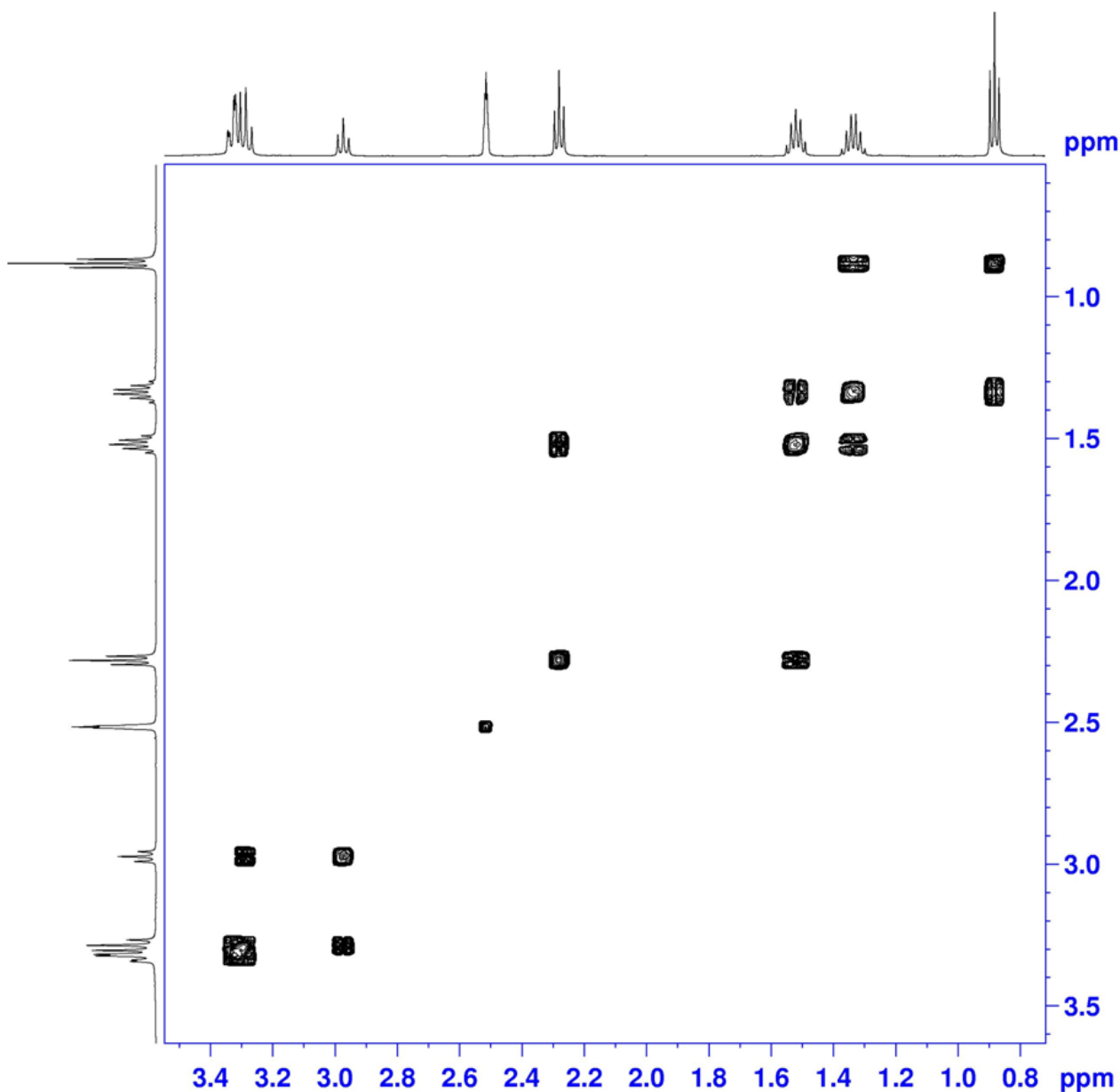
SI	1024
SF	500.2300000 MHz
WDW	QSINE
SSB	0
LB	0 Hz
GB	0
PC	1.40

F1 - Processing parameters

SI	1024
MC2	QF
SF	500.2300000 MHz
WDW	SIN
SSB	0
LB	0 Hz
GB	0

Supplementary Information

COSY of **3b** (zoom)



```

Current Data Parameters
NAME          Amol
EXPNO         199
PROCNO        1

F2 - Acquisition Parameters
Date_         20111114
Time          0.52
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       cosygpppgf
TD            2048
SOLVENT       DMSO
NS            1
DS            8
SWH           3597.122 Hz
FIDRES        1.756407 Hz
AQ            0.2847220 sec
RG            64.12
DW            139.000 usec
DE            6.50 usec
TE            295.5 K
D0            0.00000300 sec
D1            1.86851799 sec
D11           0.03000000 sec
D12           0.00002000 sec
D13           0.00000400 sec
D16           0.00020000 sec
IN0           0.00027800 sec

===== CHANNEL f1 =====
NUC1          1H
P0            9.04 usec
P1            9.04 usec
P17           2500.00 usec
PLW1          27.16399956 W
PLW10         3.28390002 W
SFO1          500.2313902 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SMSQ10.100
GPZ1          10.00 %
P16           1000.00 usec

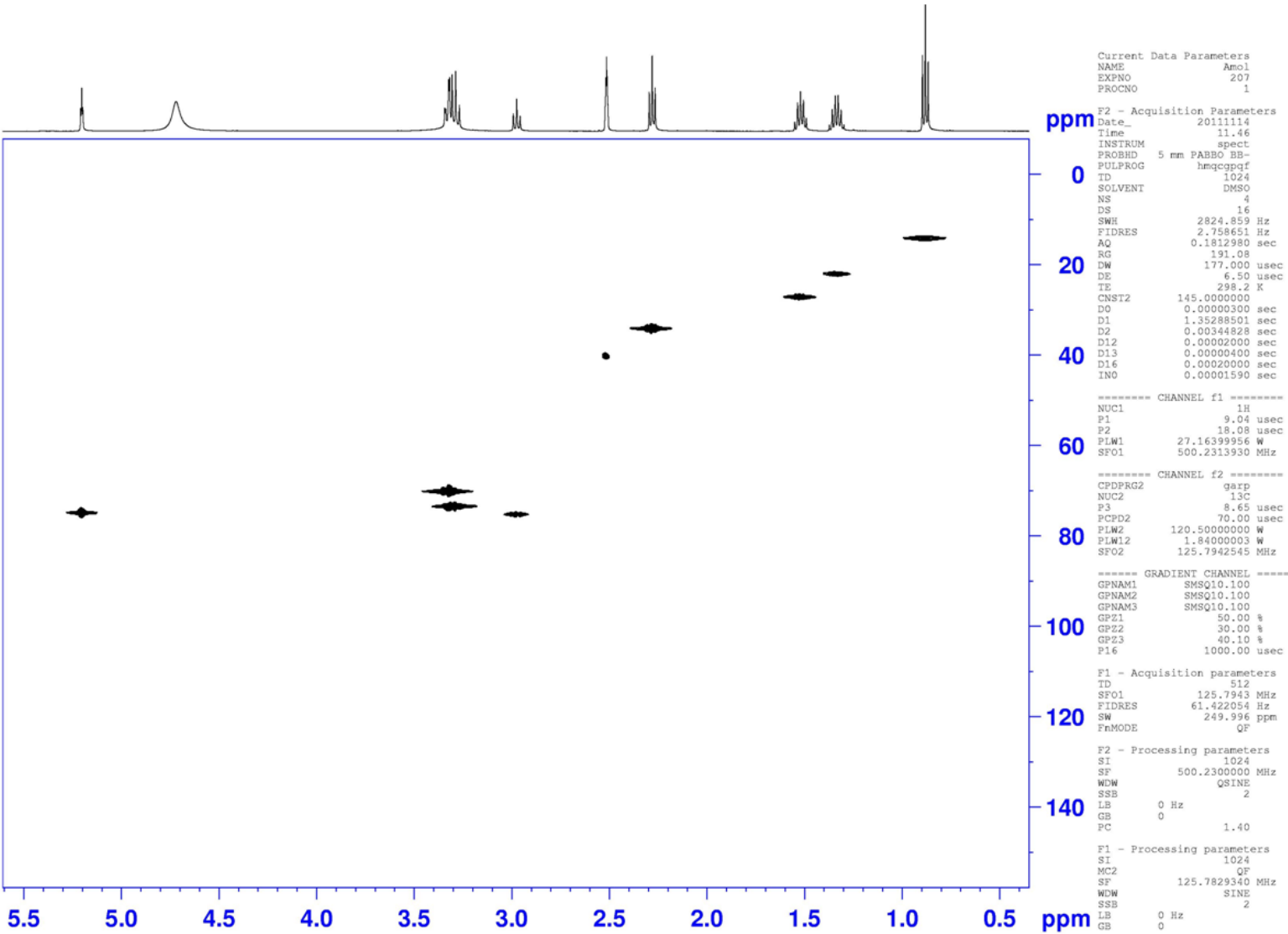
F1 - Acquisition parameters
TD            128
SFO1          500.2314 MHz
FIDRES        28.102518 Hz
SW            7.191 ppm
FnMODE        QF

F2 - Processing parameters
SI            1024
SF            500.2300000 MHz
WDW           QSINE
SSB           0
LB            0 Hz
GB            0
PC            1.40

F1 - Processing parameters
SI            1024
MC2           QF
SF            500.2300000 MHz
WDW           SIN
SSB           0
LB            0 Hz
GB            0
    
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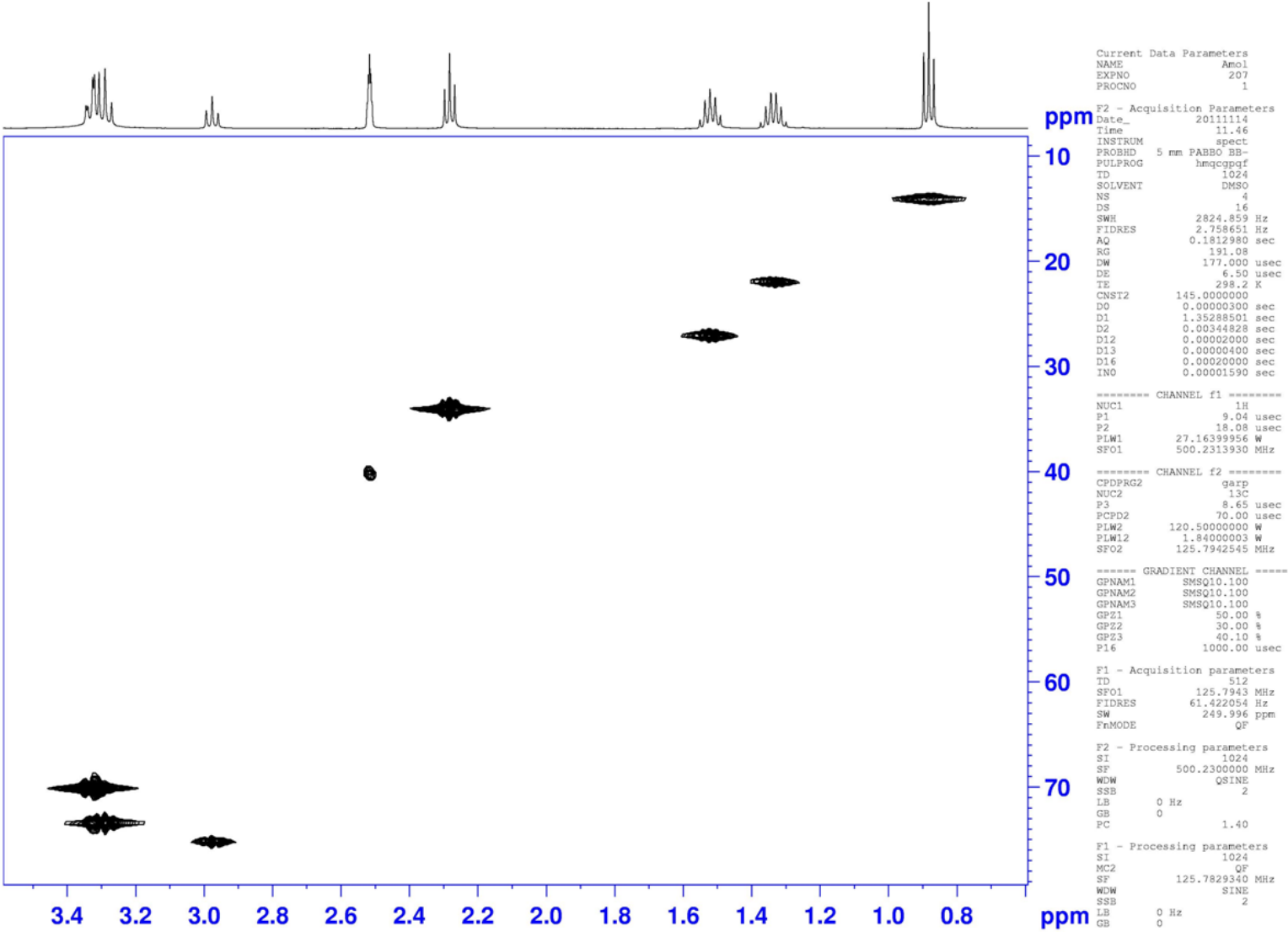
Supplementary Information

HMQC of 3b



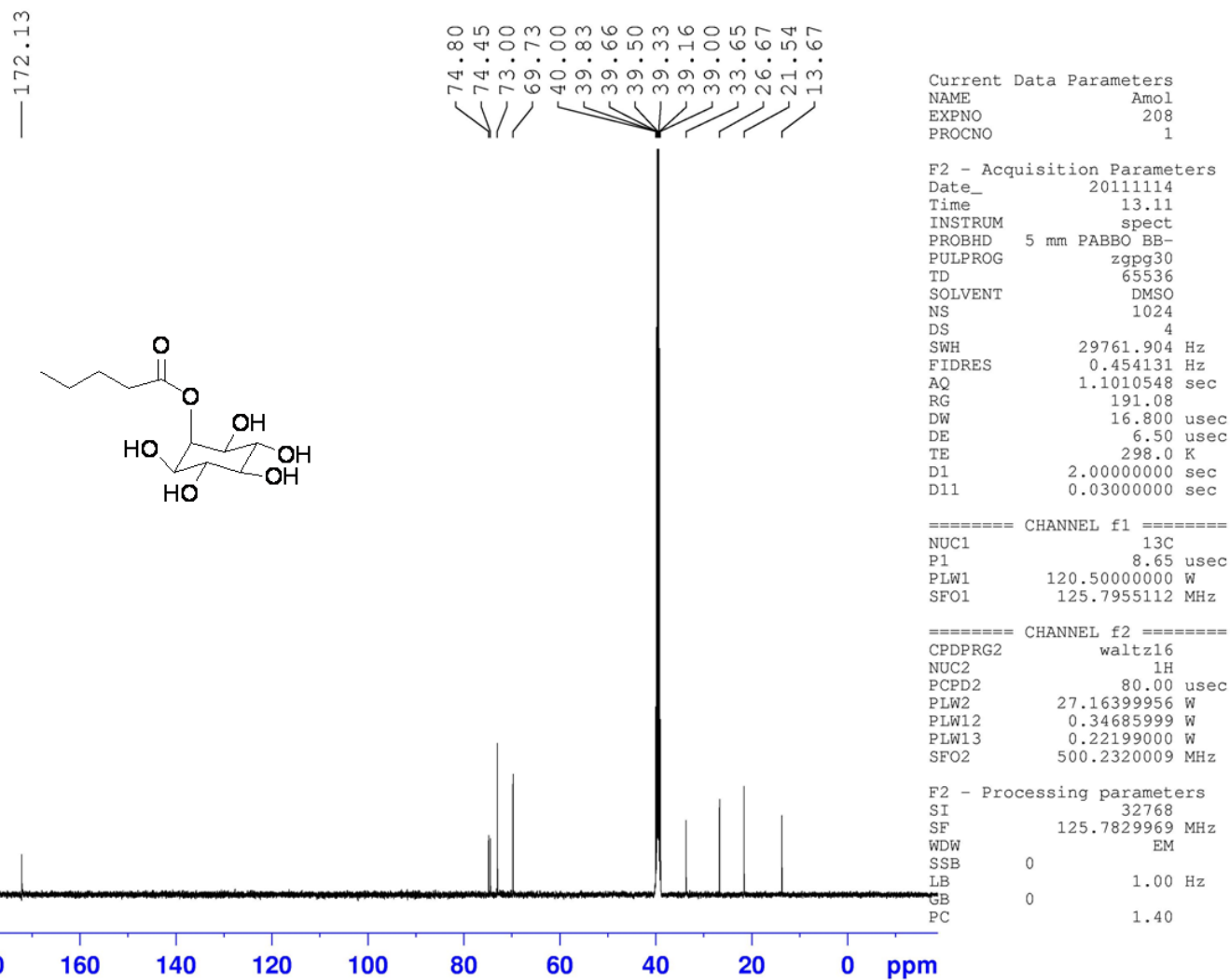
Supplementary Information

HMQC of **3b** (zoom)



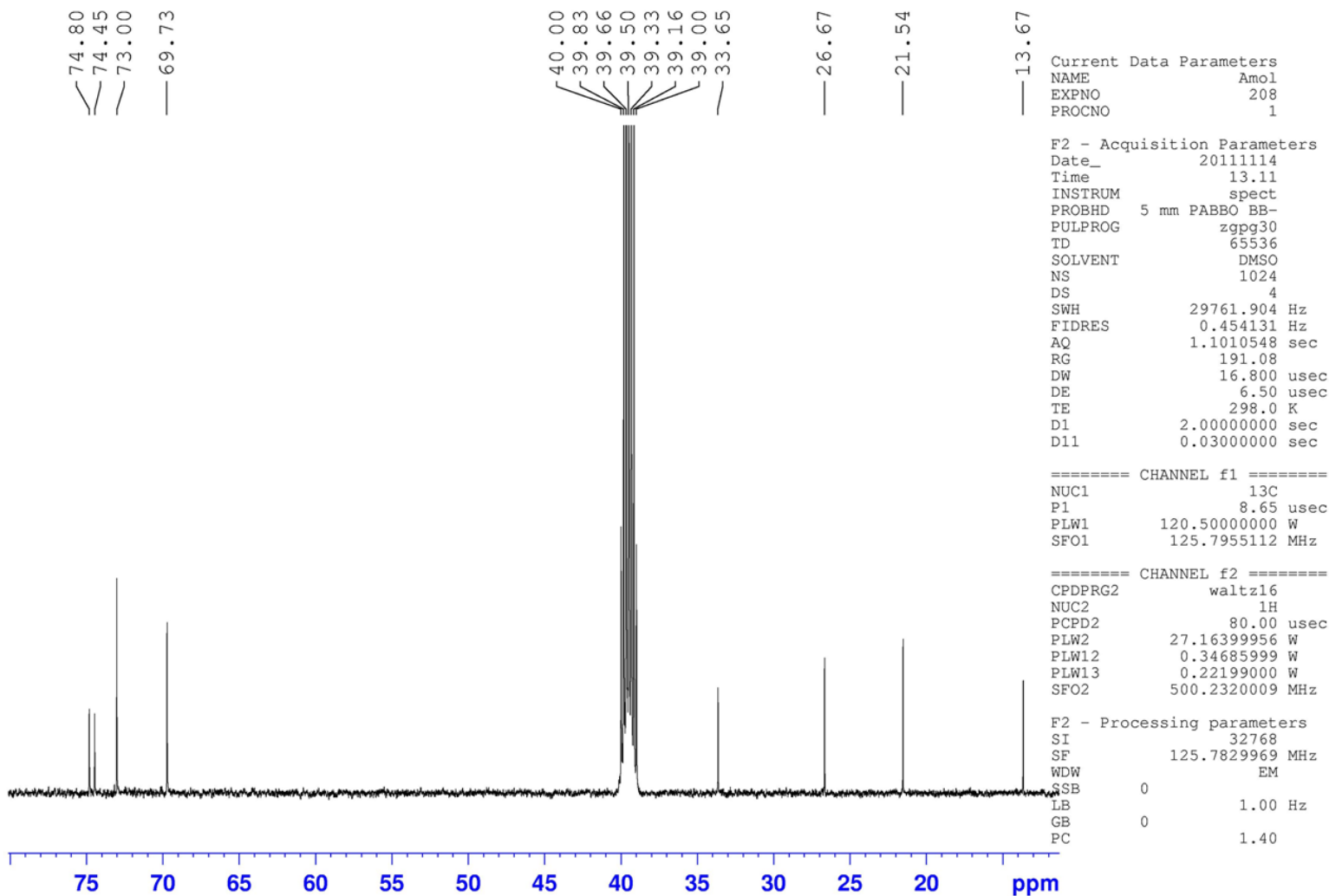
Supplementary Information

¹³C NMR of **3b** in DMSO-d₆



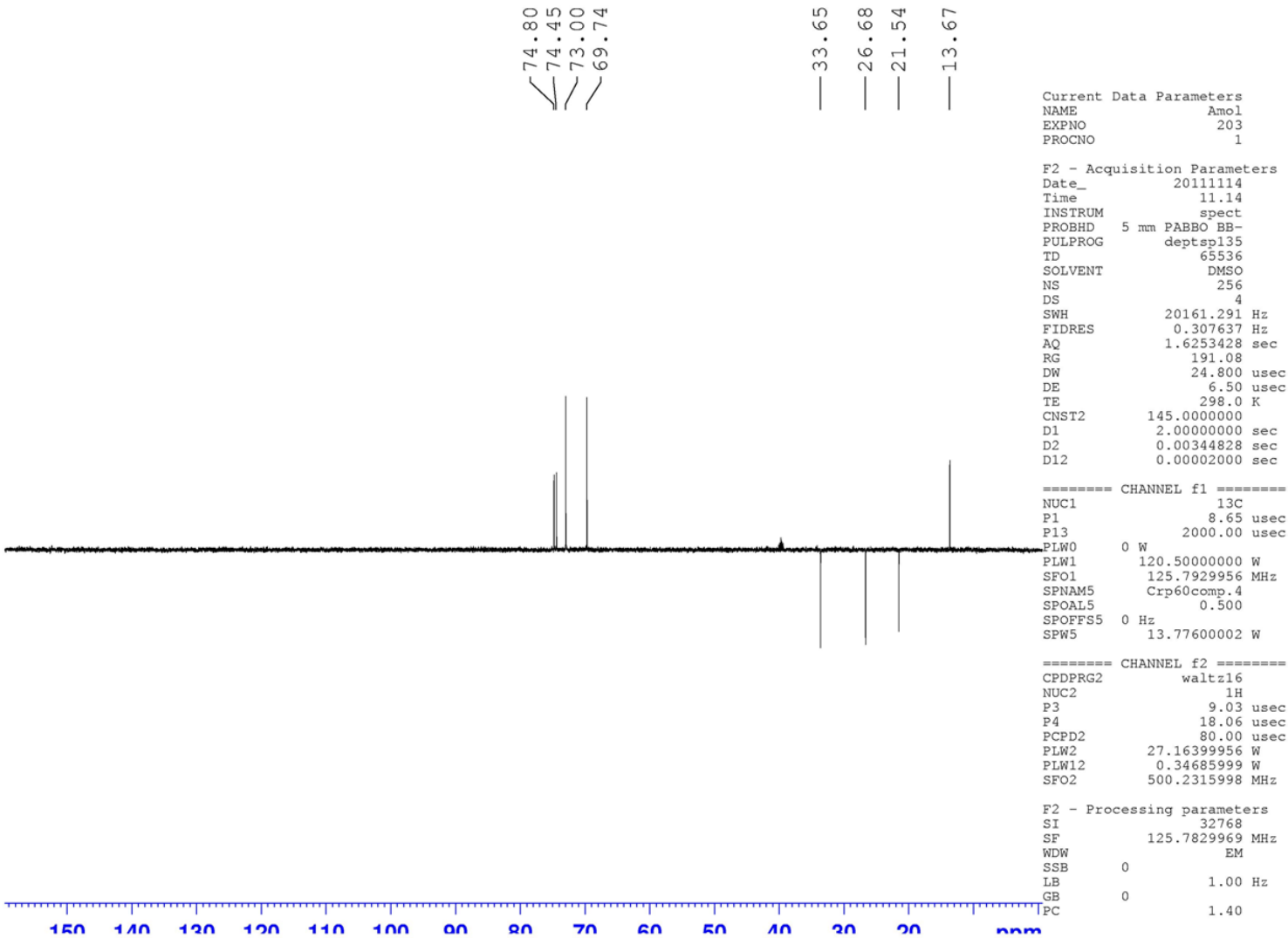
Supplementary Information

¹³C NMR of **3b** (zoom)



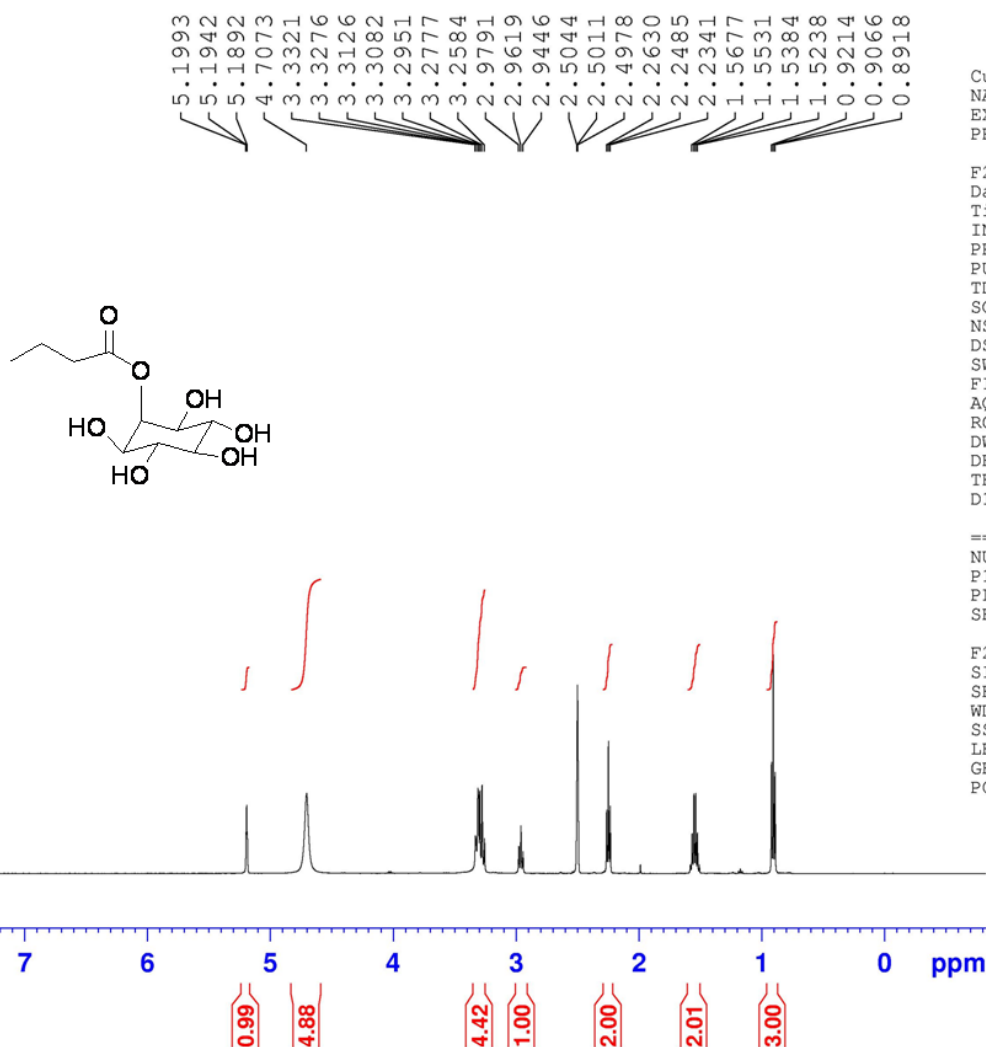
Supplementary Information

DEPT of **3b**



Supplementary Information

¹H NMR of **3c** in DMSO-d₆



Current Data Parameters
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EXPNO 201
PROCNO 1

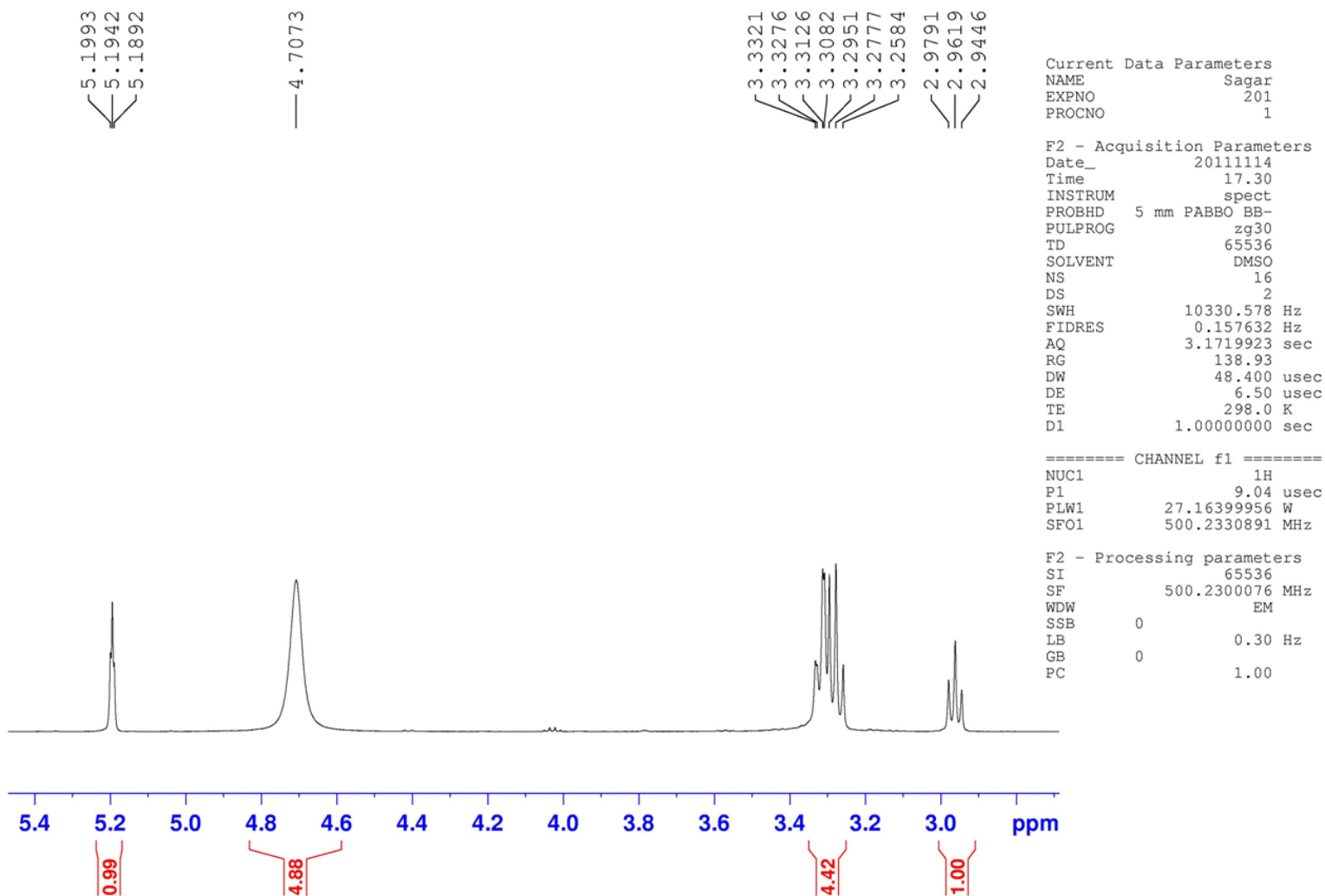
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Date_ 20111114
Time 17.30
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 138.93
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
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SF 500.2300076 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

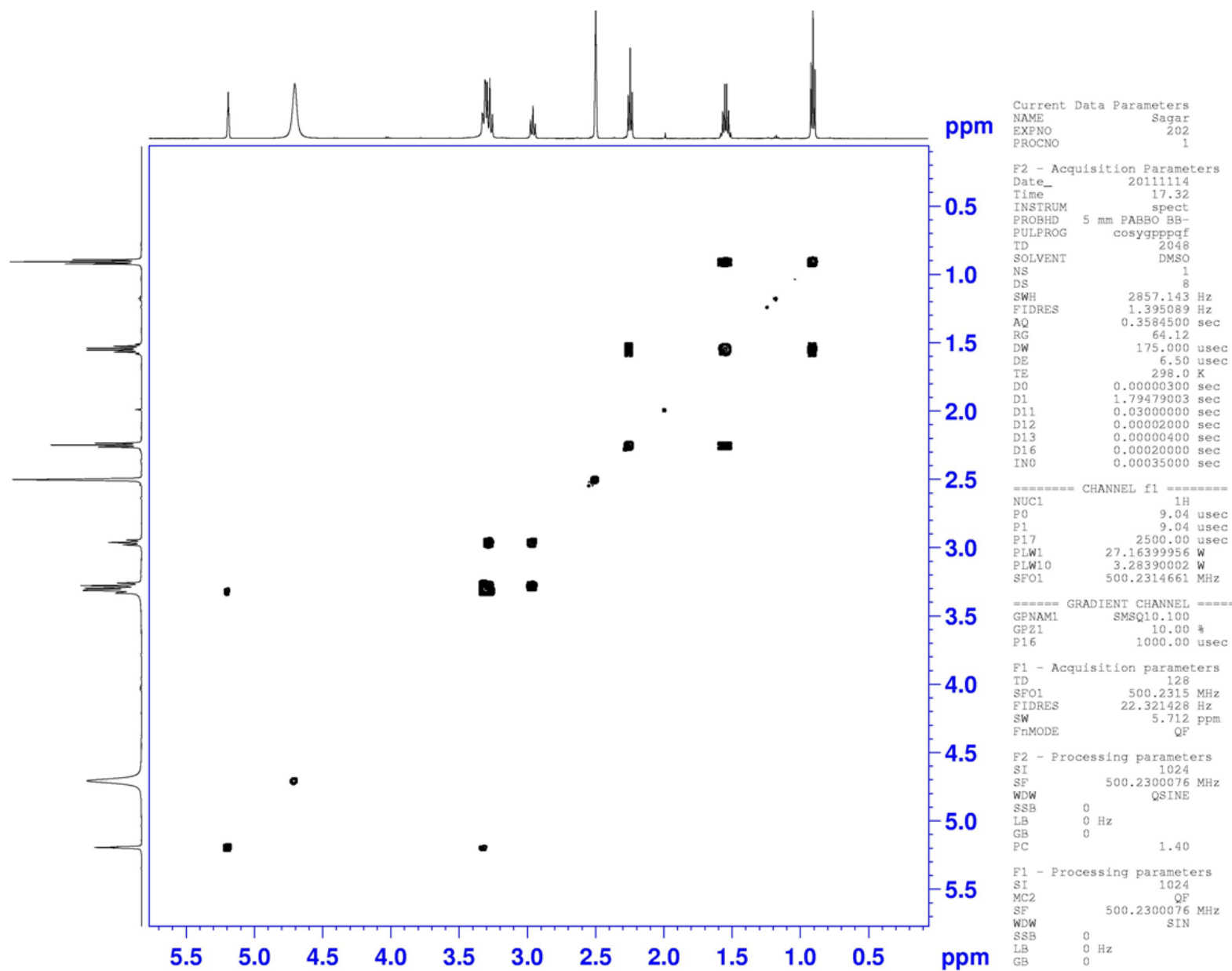
Supplementary Information

¹H NMR of **3c** (zoom)



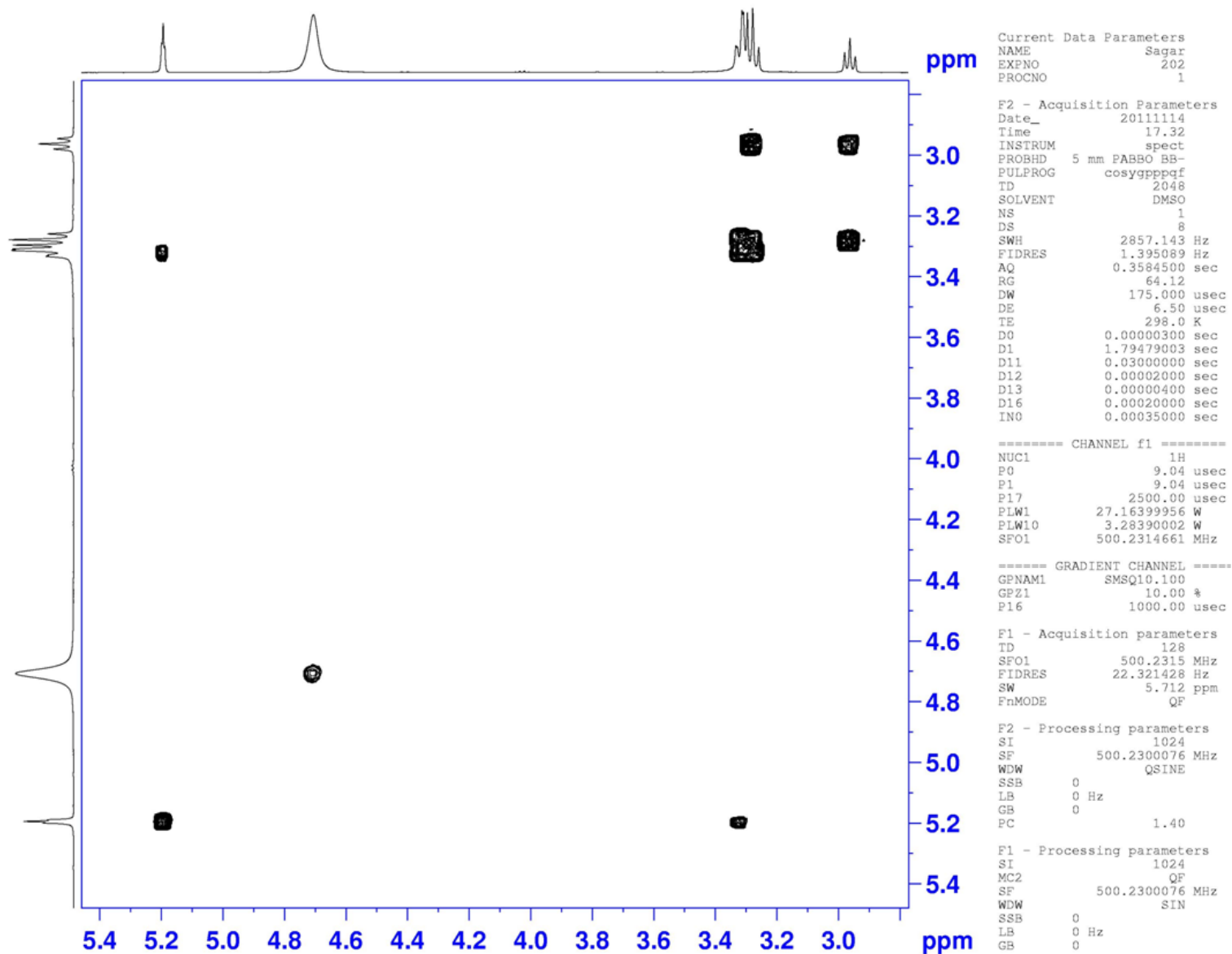
Supplementary Information

COSY of 3c



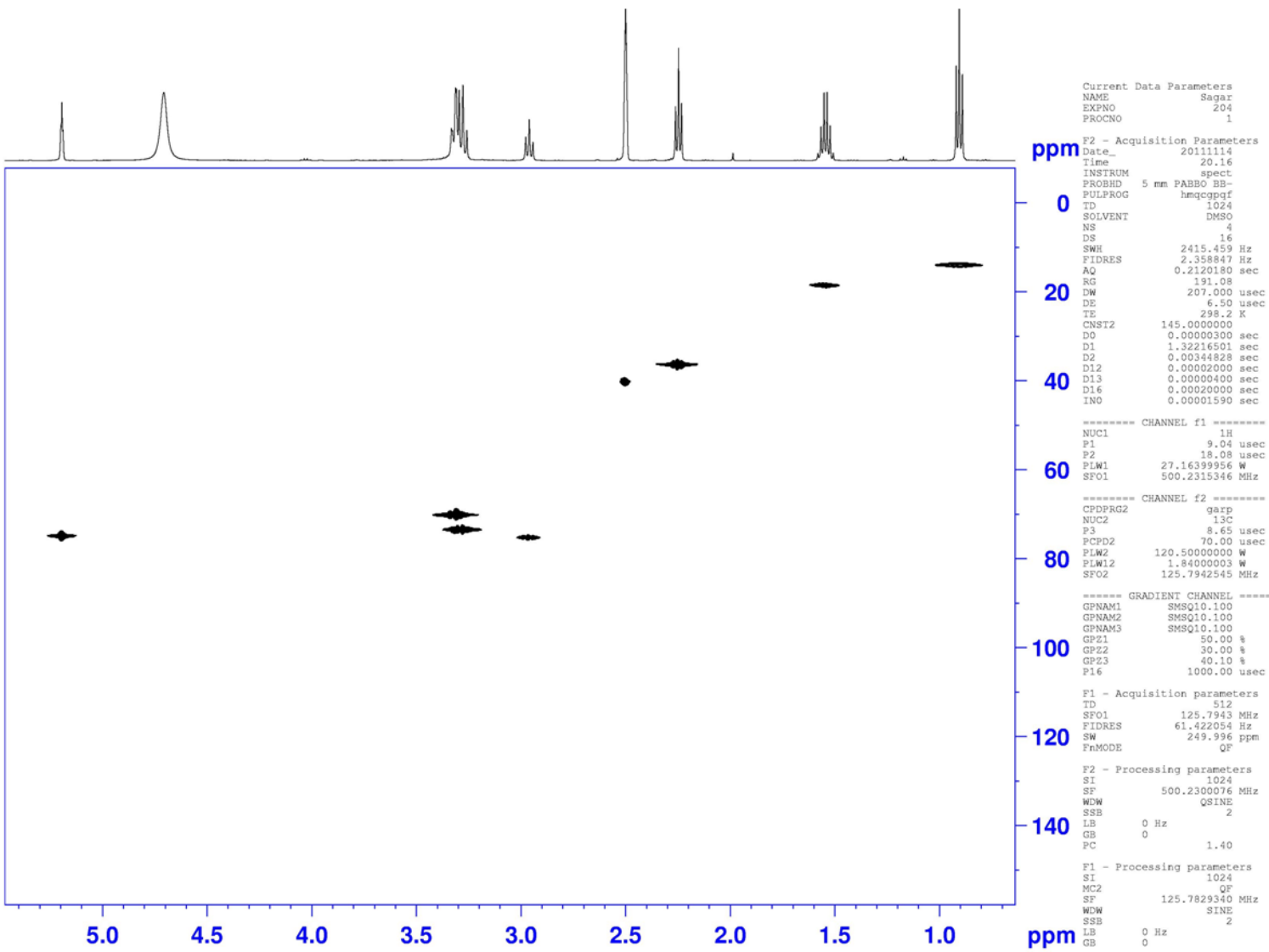
Supplementary Information

COSY of **3c** (zoom)



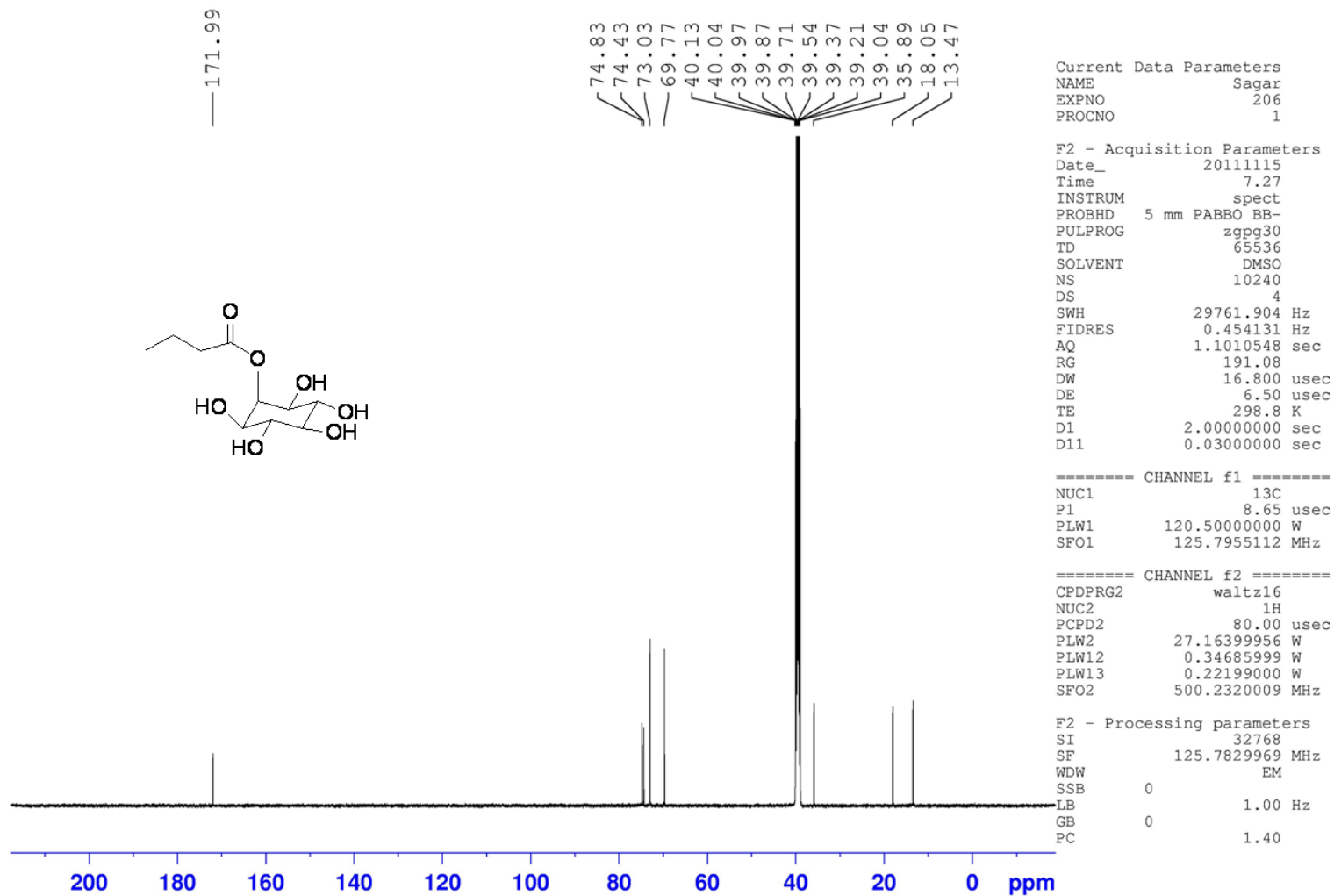
Supplementary Information

HMQC of 3c



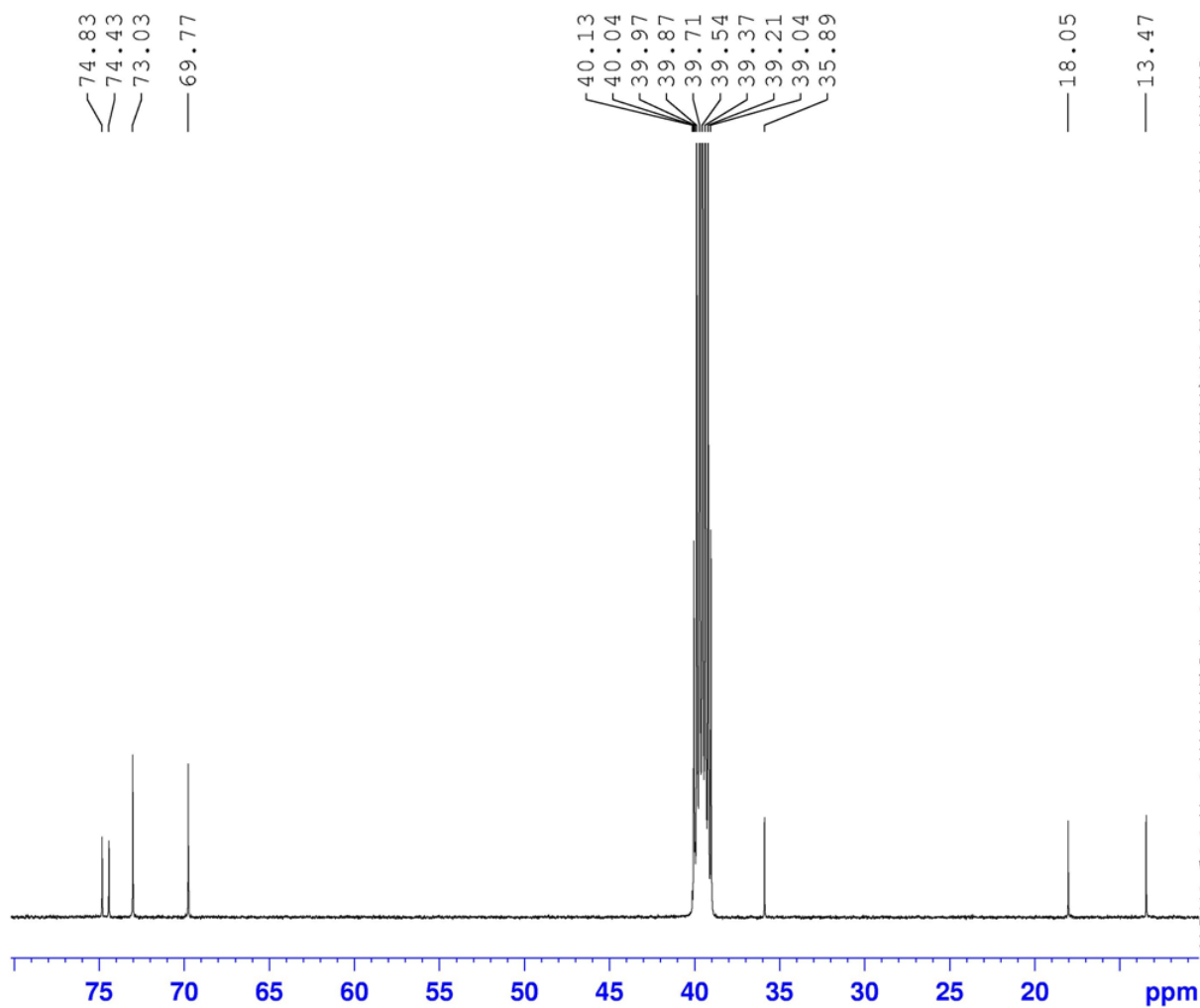
Supplementary Information

¹³C NMR of **3c** in DMSO-d₆



Supplementary Information

¹³C NMR of **3c** (zoom)



Current Data Parameters
NAME Sagar
EXPNO 206
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111115
Time 7.27
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 10240
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 191.08
DW 16.800 usec
DE 6.50 usec
TE 298.8 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
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SFO1 125.7955112 MHz

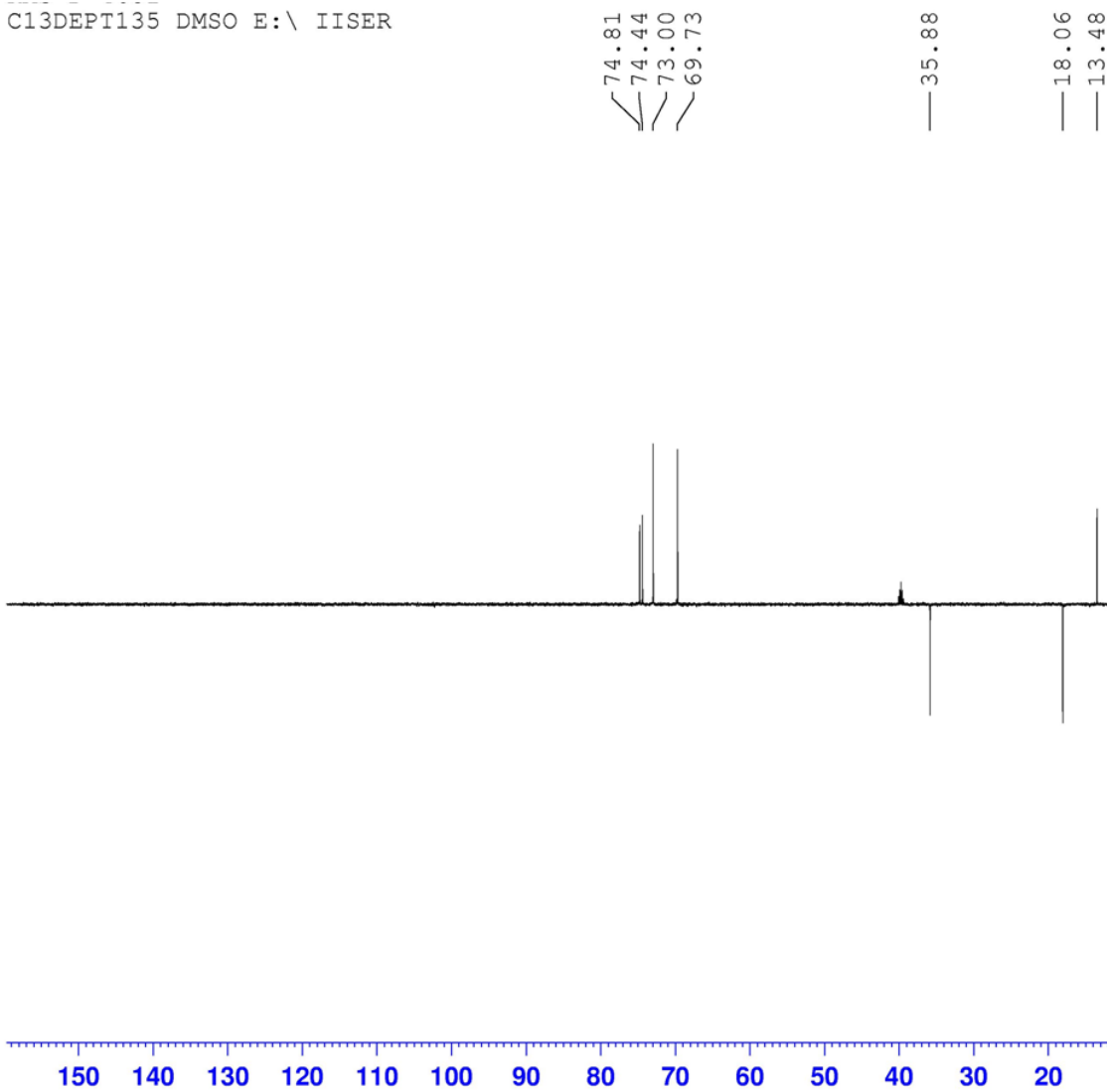
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 27.16399956 W
PLW12 0.34685999 W
PLW13 0.22199000 W
SFO2 500.2320009 MHz

F2 - Processing parameters
SI 32768
SF 125.7829969 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Supplementary Information

DEPT of 3c

C13DEPT135 DMSO E:\ IISER



Current Data Parameters
NAME Sagar
EXPNO 203
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111114
Time 20.15
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG deptsp135
TD 65536
SOLVENT DMSO
NS 2560
DS 4
SWH 20161.291 Hz
FIDRES 0.307637 Hz
AQ 1.6253428 sec
RG 191.08
DW 24.800 usec
DE 6.50 usec
TE 298.0 K
CNST2 145.0000000
D1 2.00000000 sec
D2 0.00344828 sec
D12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.65 usec
P13 2000.00 usec
PLW0 0 W
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SFO1 125.7929956 MHz
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SPW5 13.77600002 W

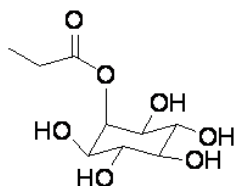
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P4 18.06 usec
PCPD2 80.00 usec
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PLW12 0.34685999 W
SFO2 500.2315998 MHz

F2 - Processing parameters
SI 32768
SF 125.7829969 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Supplementary Information

¹H NMR of **3d** in DMSO-d₆

5.2032
5.1979
5.1927
3.5408
3.3394
3.3338
3.3200
3.3147
3.3073
3.2903
3.2708
2.9938
2.9768
2.5236
2.5201
2.5165
2.5129
2.5094
2.3222
2.3072
2.2921
2.2771
1.0477
1.0327
1.0176

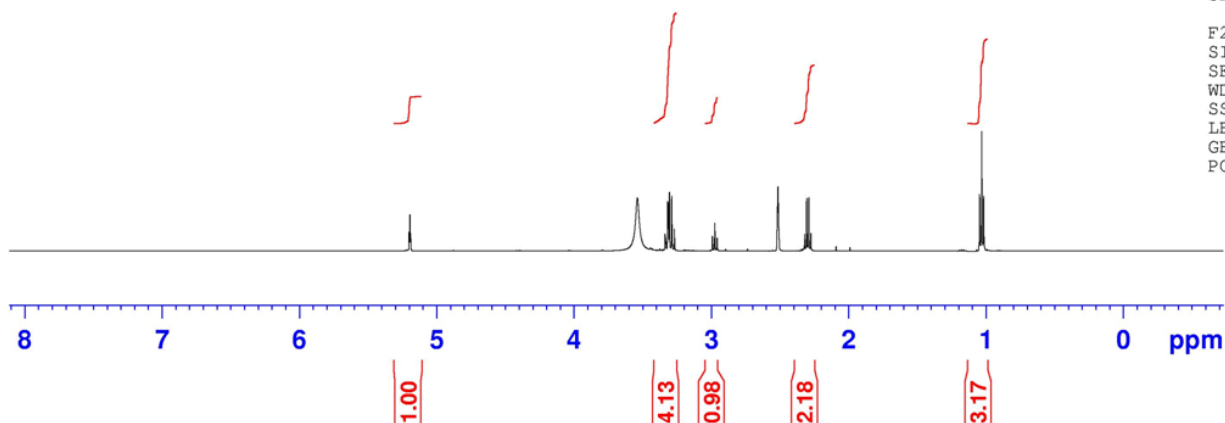


Current Data Parameters
NAME Sagar
EXPNO 220
PROCNO 1

F2 - Acquisition Parameters
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Time 14.35
INSTRUM spect
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PULPROG zg30
TD 65536
SOLVENT DMSO
NS 64
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 125.62
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.00000000 sec

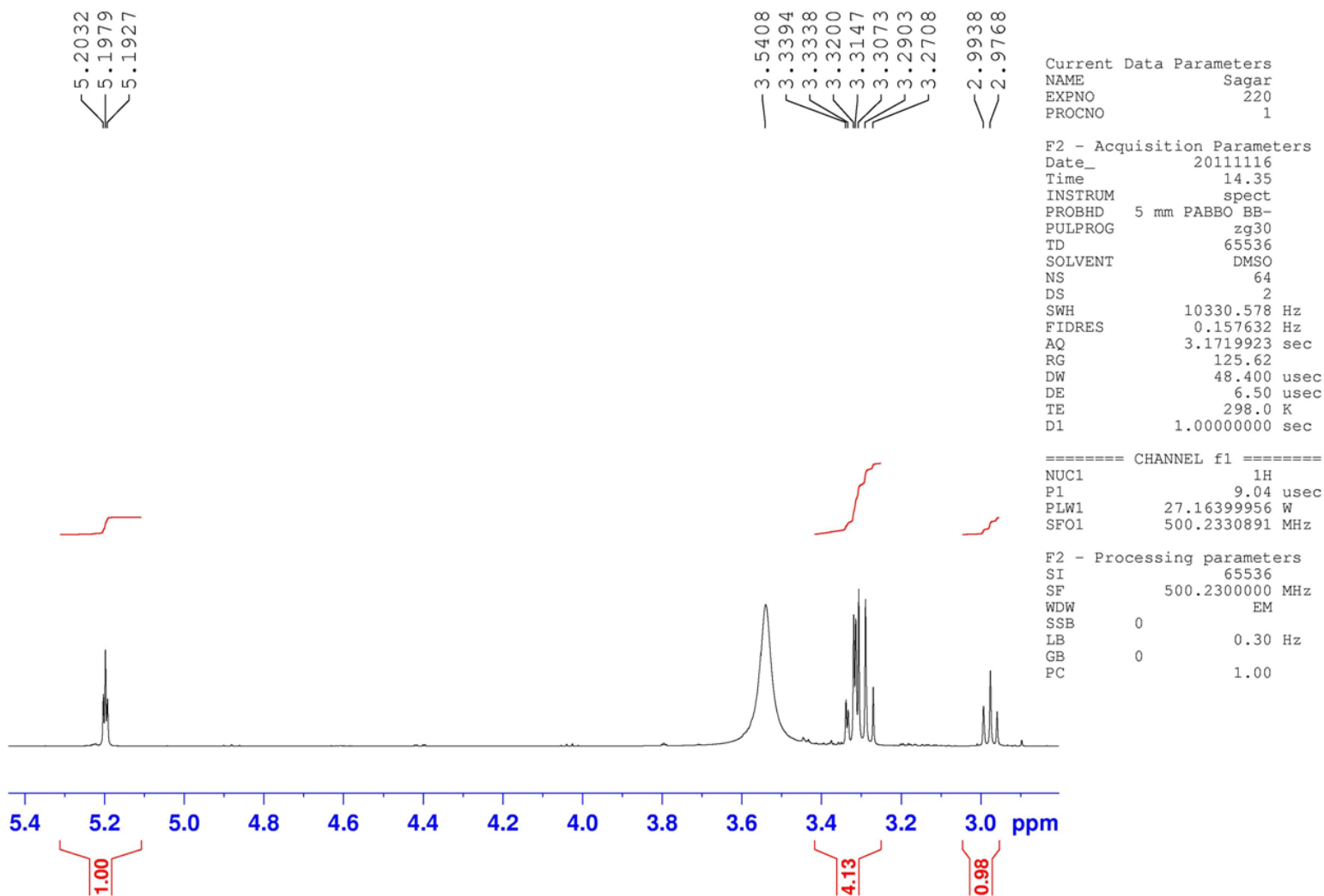
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NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



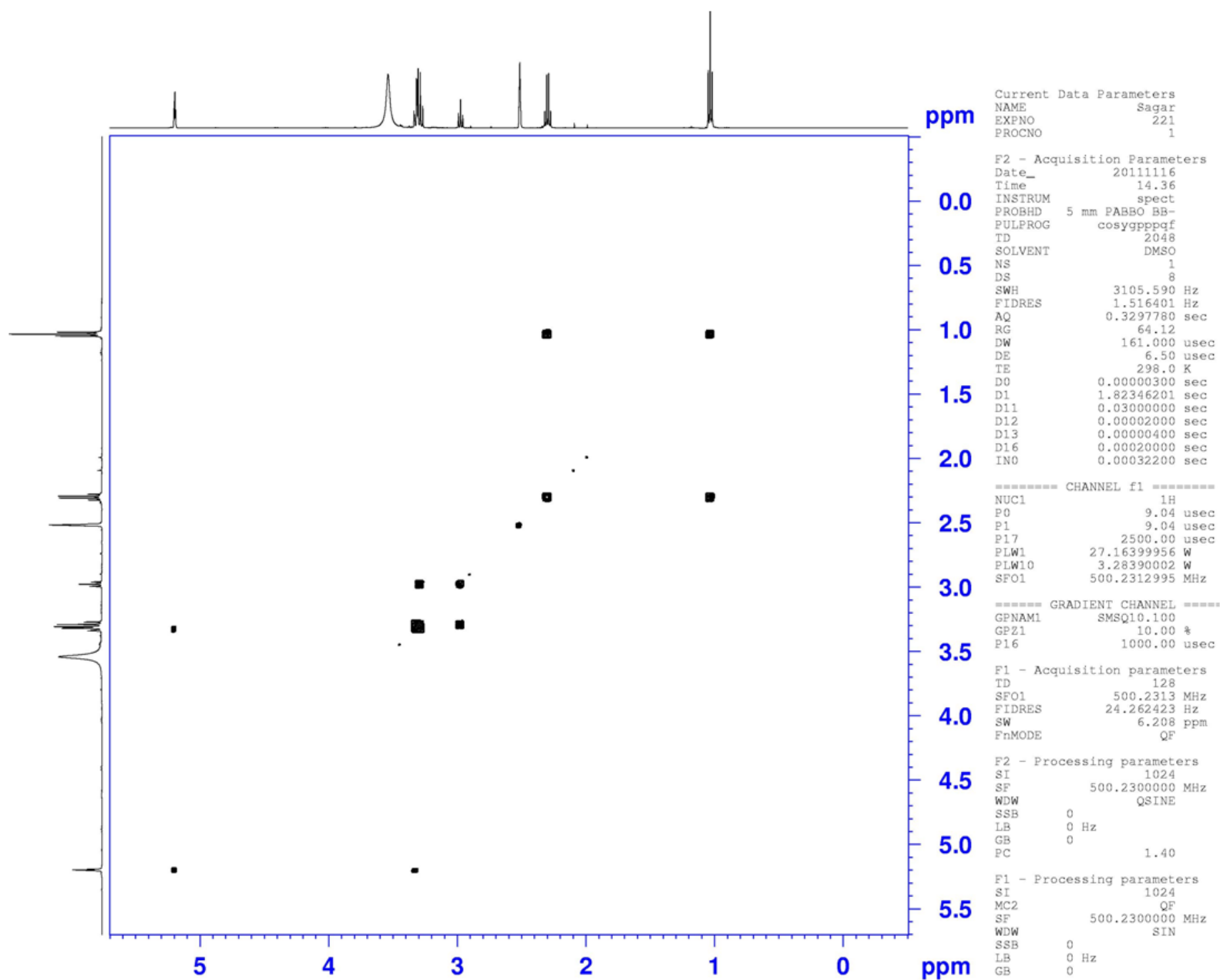
Supplementary Information

¹H NMR of **3d** (zoom)



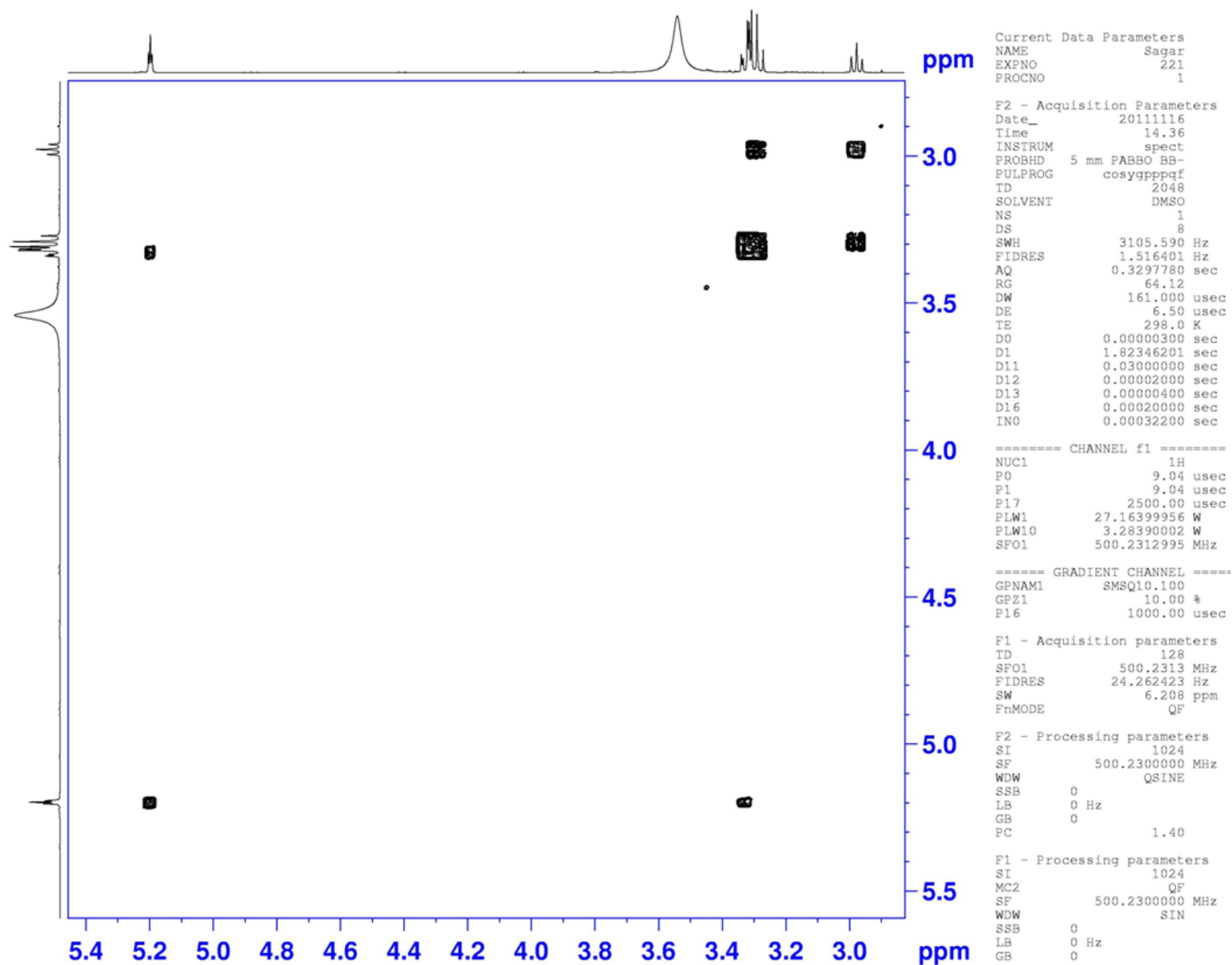
Supplementary Information

COSY of **3d**



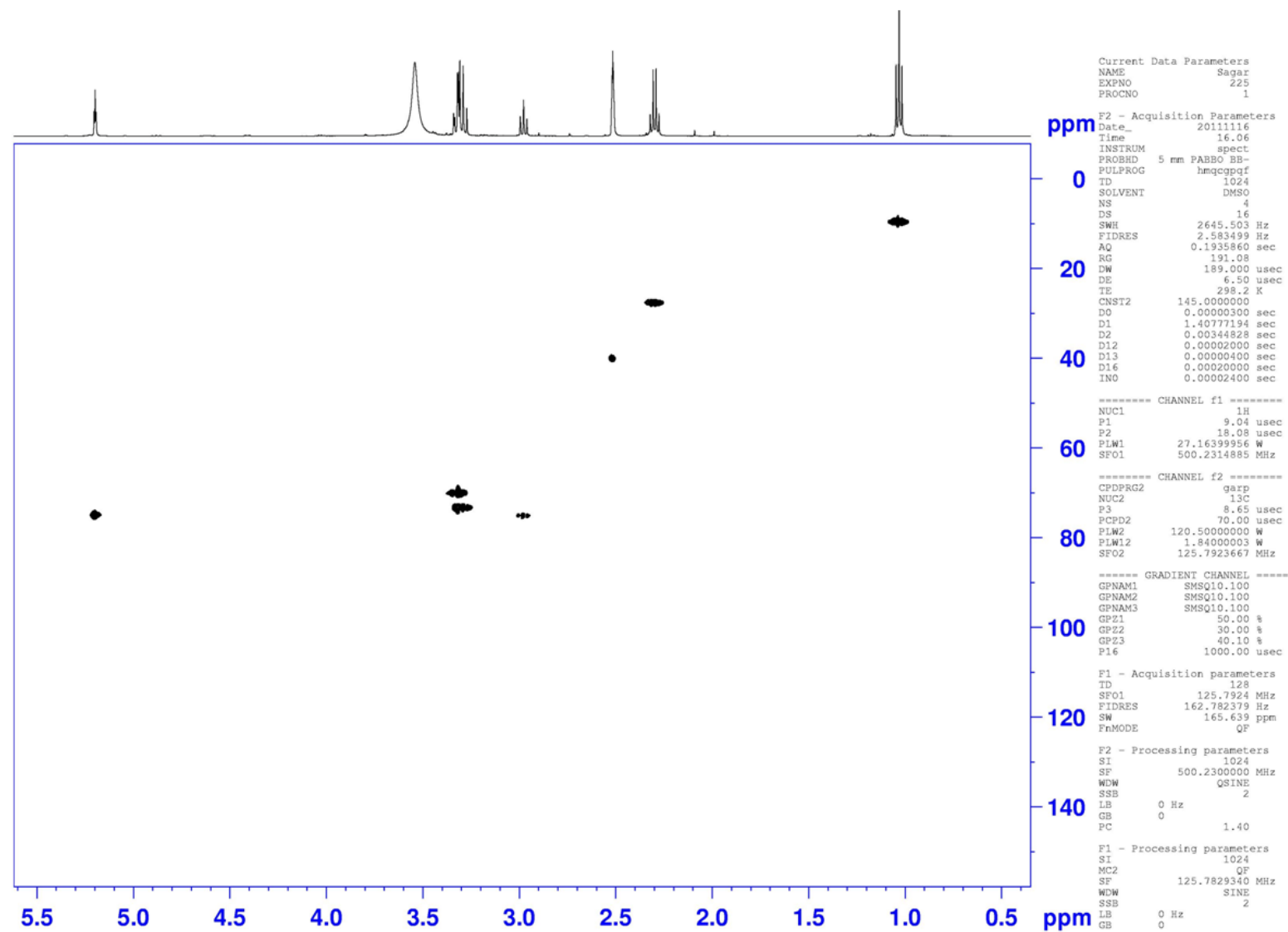
Supplementary Information

COSY of **3d** (zoom)



Supplementary Information

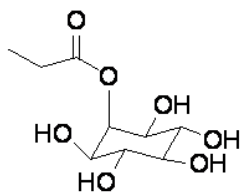
HMQC of 3d



Supplementary Information

¹³C NMR of **3d** in DMSO-d₆

— 173.00



74.61
74.46
72.81
69.61
39.95
39.89
39.80
39.72
39.63
39.55
39.46
39.30
39.13
38.96
38.80
27.14
9.06

Current Data Parameters
NAME Sagar
EXPNO 223
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111116
Time 16.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 191.08
DW 16.800 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.65 usec
PLW1 120.50000000 W
SFO1 125.7955112 MHz

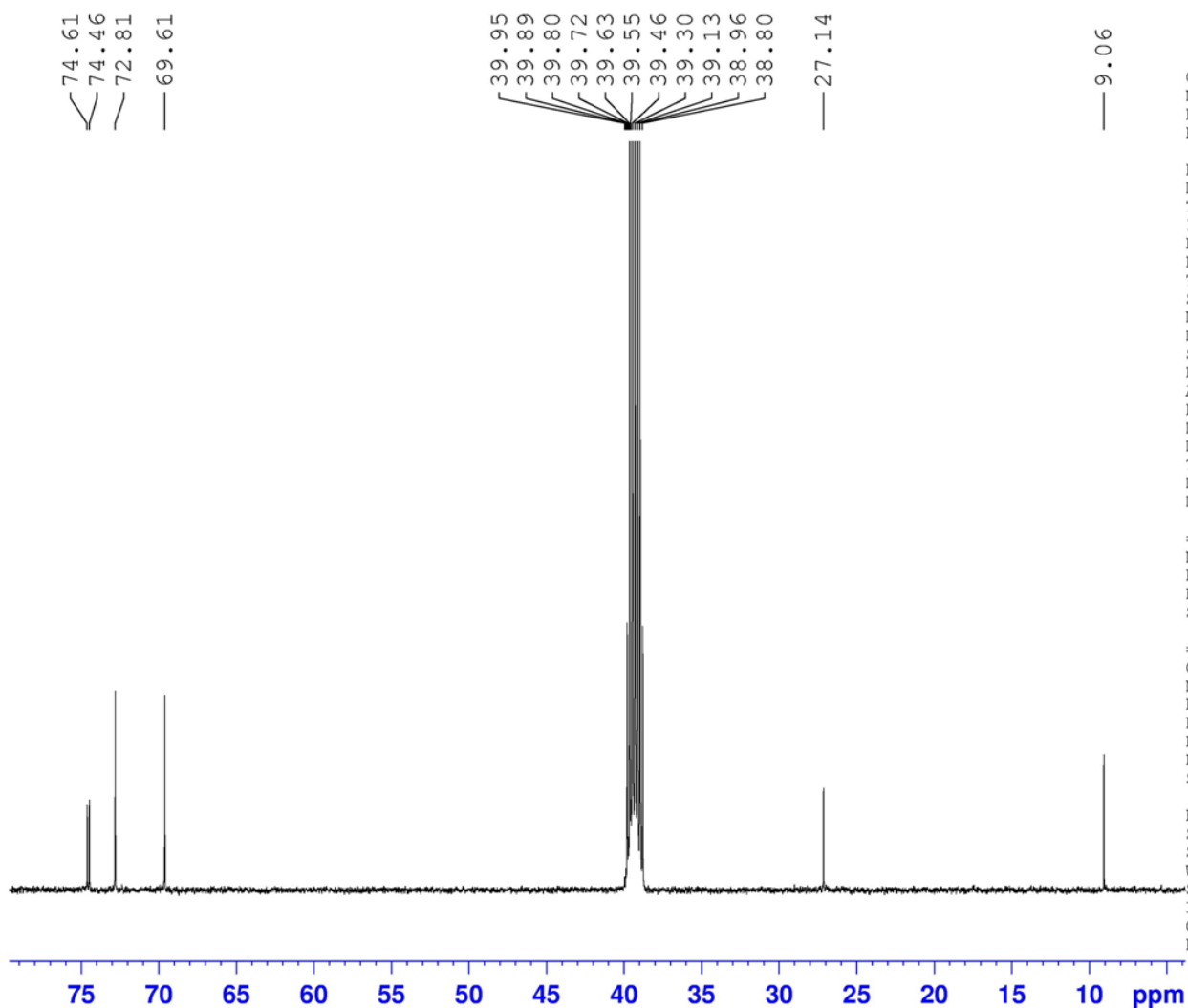
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 27.16399956 W
PLW12 0.34685999 W
PLW13 0.22199000 W
SFO2 500.2320009 MHz

F2 - Processing parameters
SI 32768
SF 125.7829969 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

200 180 160 140 120 100 80 60 40 20 0 ppm

Supplementary Information

¹³C NMR of **3d** (zoom)



```

Current Data Parameters
NAME          Sagar
EXPNO         223
PROCNO        1

F2 - Acquisition Parameters
Date_         20111116
Time          16.03
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       zgpg30
TD            65536
SOLVENT       DMSO
NS            1024
DS            4
SWH           29761.904 Hz
FIDRES        0.454131 Hz
AQ            1.1010548 sec
RG            191.08
DW            16.800 usec
DE            6.50 usec
TE            298.0 K
D1            2.00000000 sec
D11           0.03000000 sec

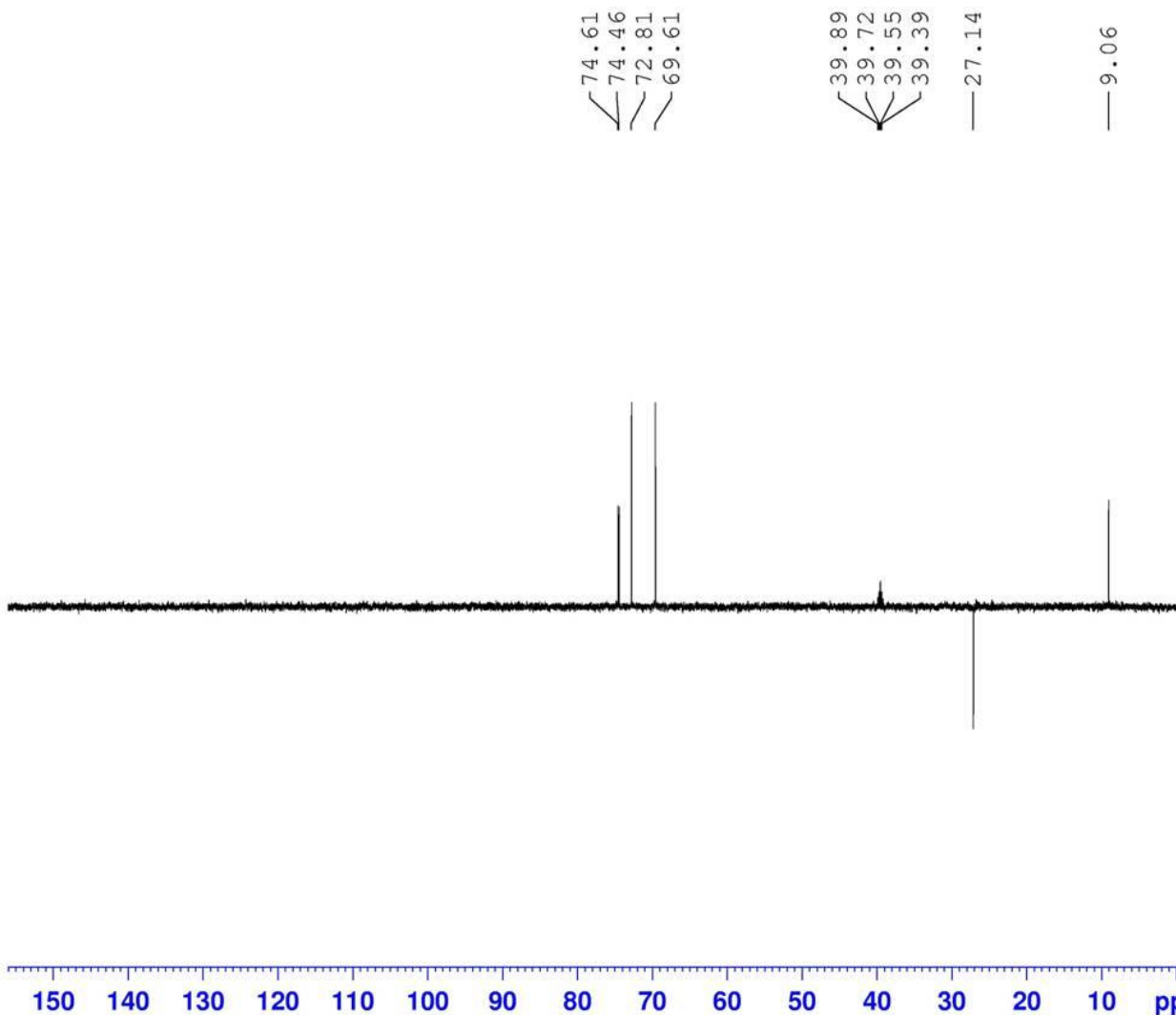
===== CHANNEL f1 =====
NUC1           13C
P1             8.65 usec
PLW1          120.50000000 W
SFO1          125.7955112 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         80.00 usec
PLW2          27.16399956 W
PLW12         0.34685999 W
PLW13         0.22199000 W
SFO2          500.2320009 MHz

F2 - Processing parameters
SI            32768
SF           125.7829969 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40
    
```

Supplementary Information

DEPT of 3d



Current Data Parameters
NAME Sagar
EXPNO 222
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111116
Time 14.58
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG deptsp135
TD 65536
SOLVENT DMSO
NS 256
DS 4
SWH 20161.291 Hz
FIDRES 0.307637 Hz
AQ 1.6253428 sec
RG 191.08
DW 24.800 usec
DE 6.50 usec
TE 298.0 K
CNST2 145.0000000
D1 2.00000000 sec
D2 0.00344828 sec
D12 0.00002000 sec

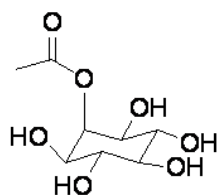
===== CHANNEL f1 =====
NUC1 13C
P1 8.65 usec
P13 2000.00 usec
PLW0 0 W
PLW1 120.50000000 W
SFO1 125.7929956 MHz
SPNAM5 Crp60comp.4
SPOAL5 0.500
SPOFFS5 0 Hz
SPW5 13.77600002 W

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
P3 9.03 usec
P4 18.06 usec
PCPD2 80.00 usec
PLW2 27.16399956 W
PLW12 0.34685999 W
SFO2 500.2315998 MHz

F2 - Processing parameters
SI 32768
SF 125.7829969 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Supplementary Information

¹H NMR of **3e** in DMSO-d₆



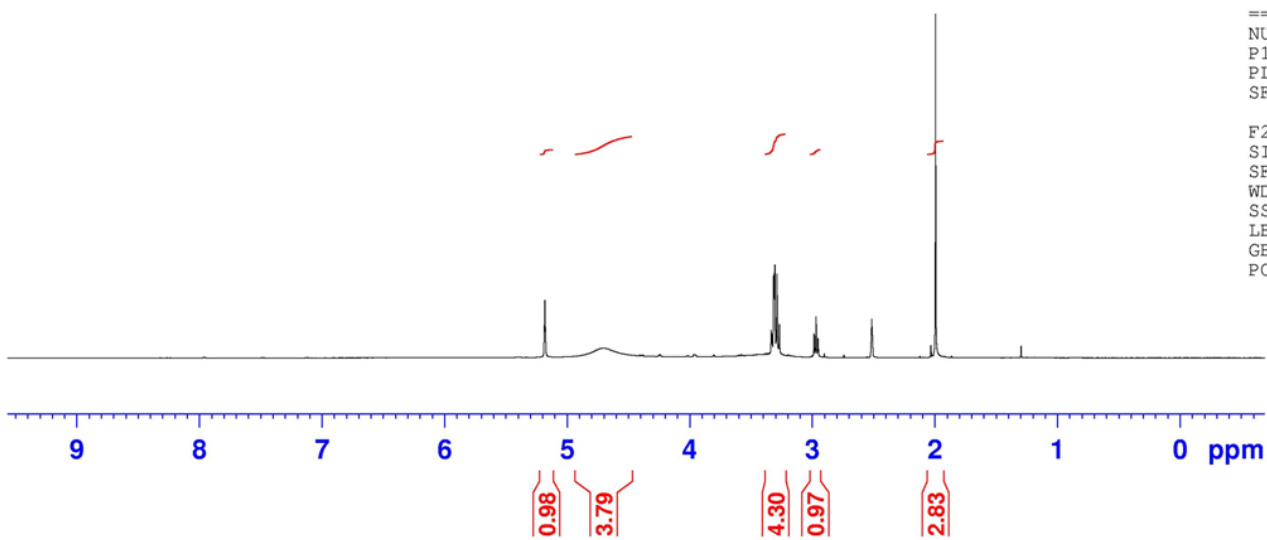
5.1894
5.1843
5.1793
4.7098
3.3375
3.3322
3.3180
3.3129
3.3071
3.2903
3.2708
2.9882
2.9712
2.9544
2.5199
2.5165
2.5130
1.9960
1.2977

Current Data Parameters
NAME Amol
EXPNO 239
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111117
Time 21.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 64
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 85.95
DW 48.400 usec
DE 6.50 usec
TE 298.1 K
D1 1.00000000 sec

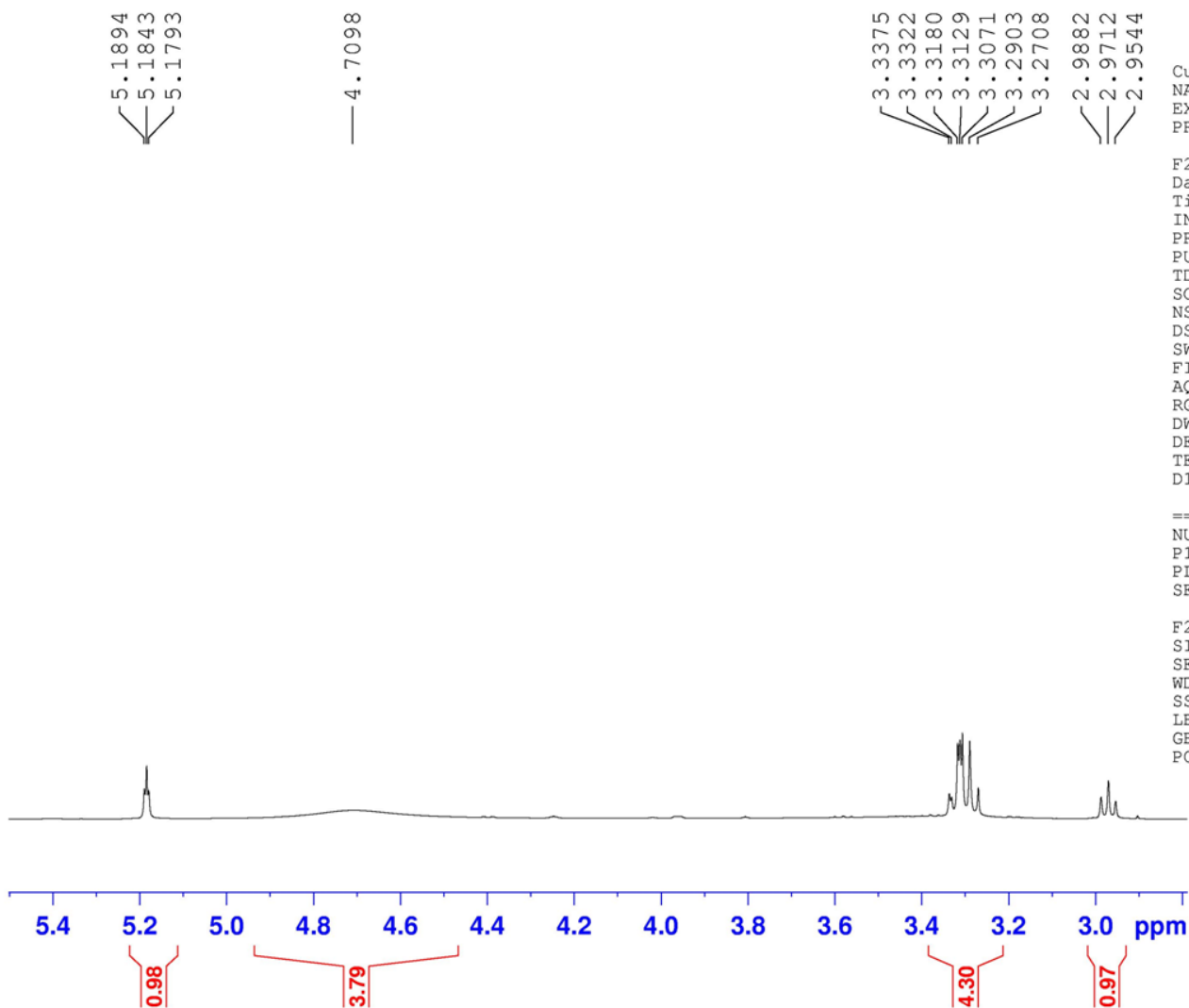
===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Supplementary Information

¹H NMR of **3e** (zoom)



Current Data Parameters
NAME Amol
EXPNO 239
PROCNO 1

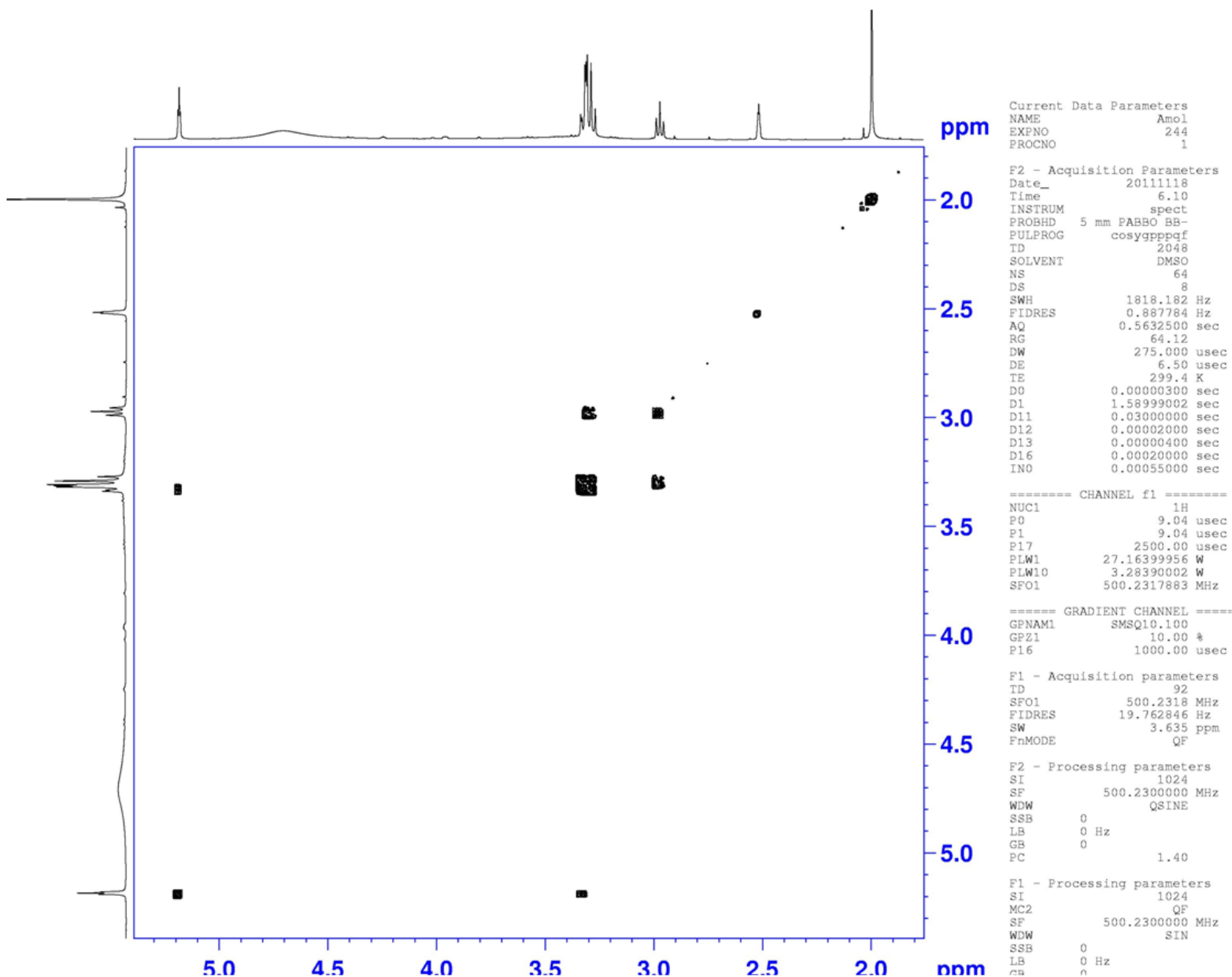
F2 - Acquisition Parameters
Date_ 20111117
Time 21.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 64
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 85.95
DW 48.400 usec
DE 6.50 usec
TE 298.1 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

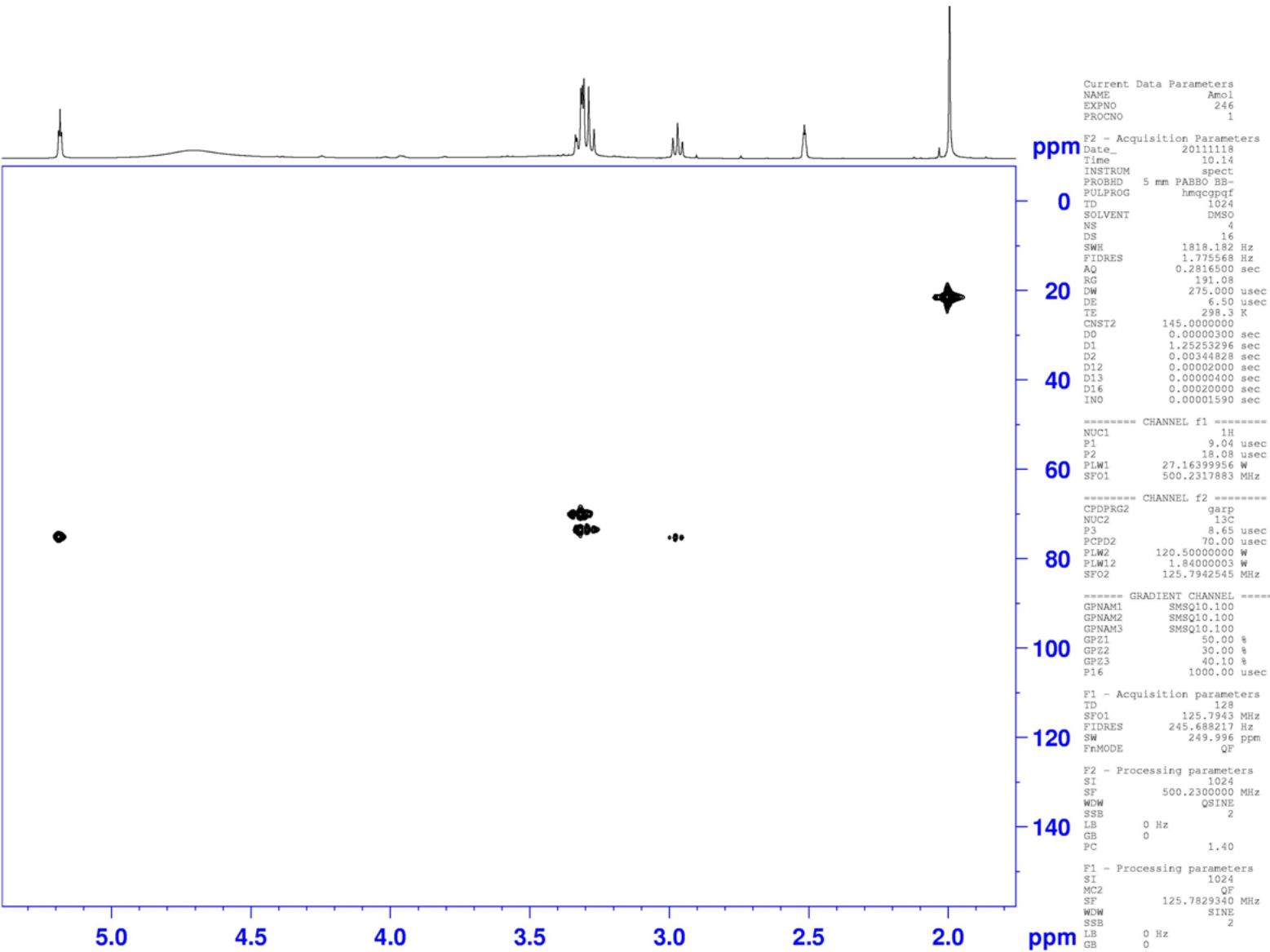
Supplementary Information

COSY of 3e



Supplementary Information

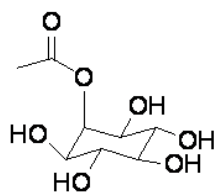
HMQC of 3e



Supplementary Information

¹³C NMR of **3e** in DMSO-d₆

— 169.69



74.83
74.68
72.99
69.76
40.02
39.86
39.69
39.52
39.36
39.19
39.02
21.05

Current Data Parameters
NAME Amol
EXPNO 243
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111118
Time 6.07
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 10240
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 191.08
DW 16.800 usec
DE 6.50 usec
TE 299.8 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.65 usec
PLW1 120.50000000 W
SFO1 125.7955112 MHz

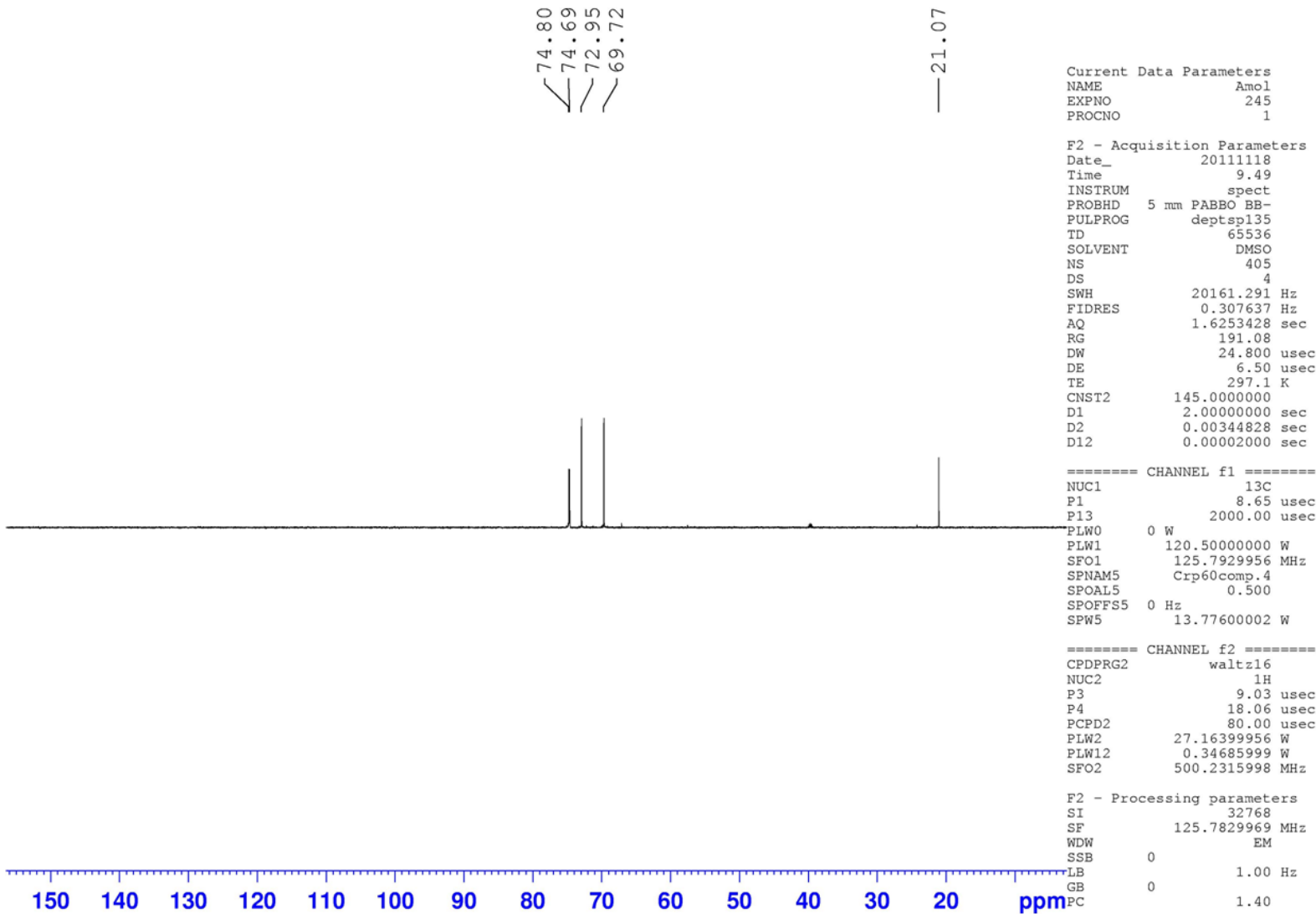
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 27.16399956 W
PLW12 0.34685999 W
PLW13 0.22199000 W
SFO2 500.2320009 MHz

F2 - Processing parameters
SI 32768
SF 125.7829969 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

200 180 160 140 120 100 80 60 40 20 0 ppm

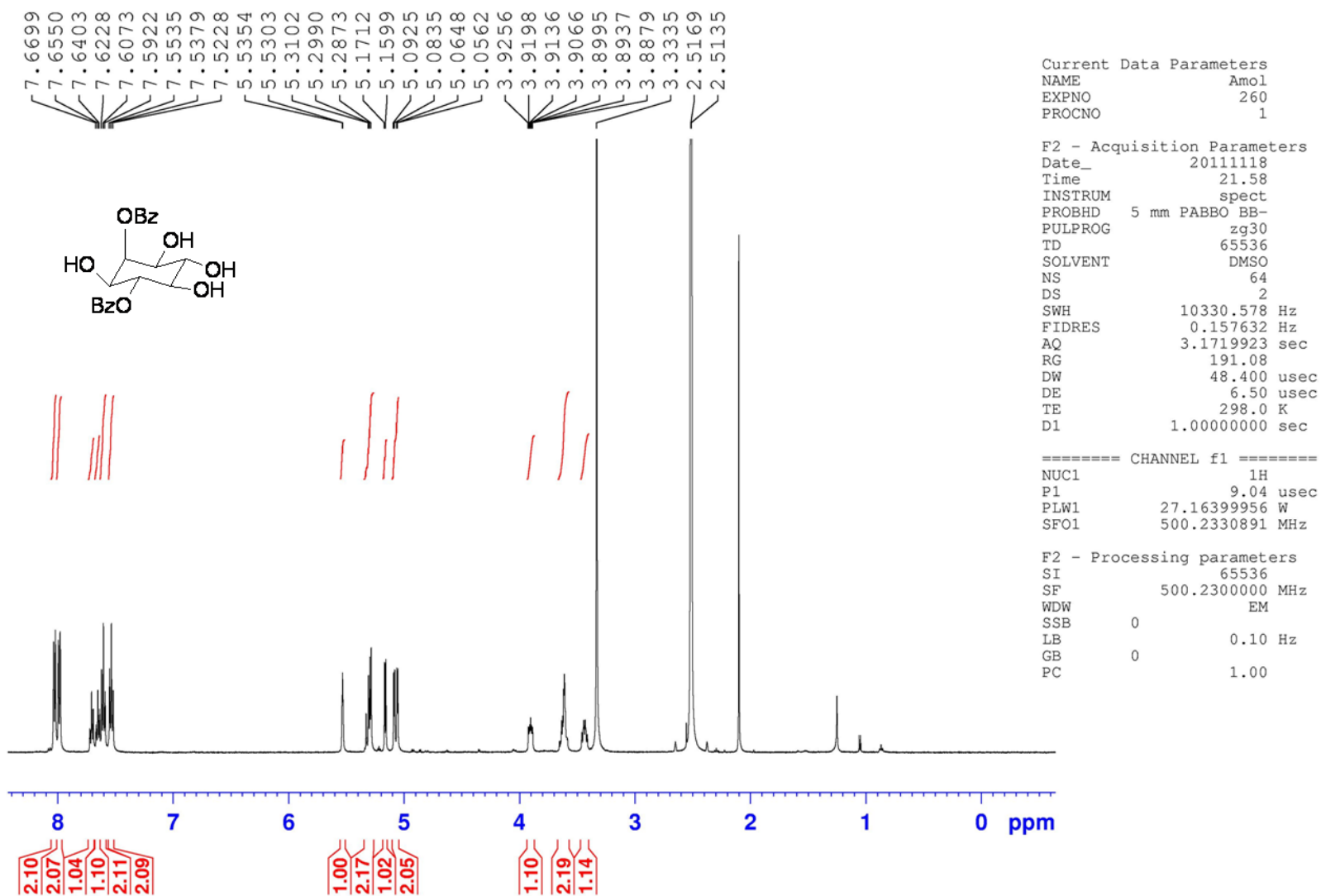
Supplementary Information

DEPT of **3e**



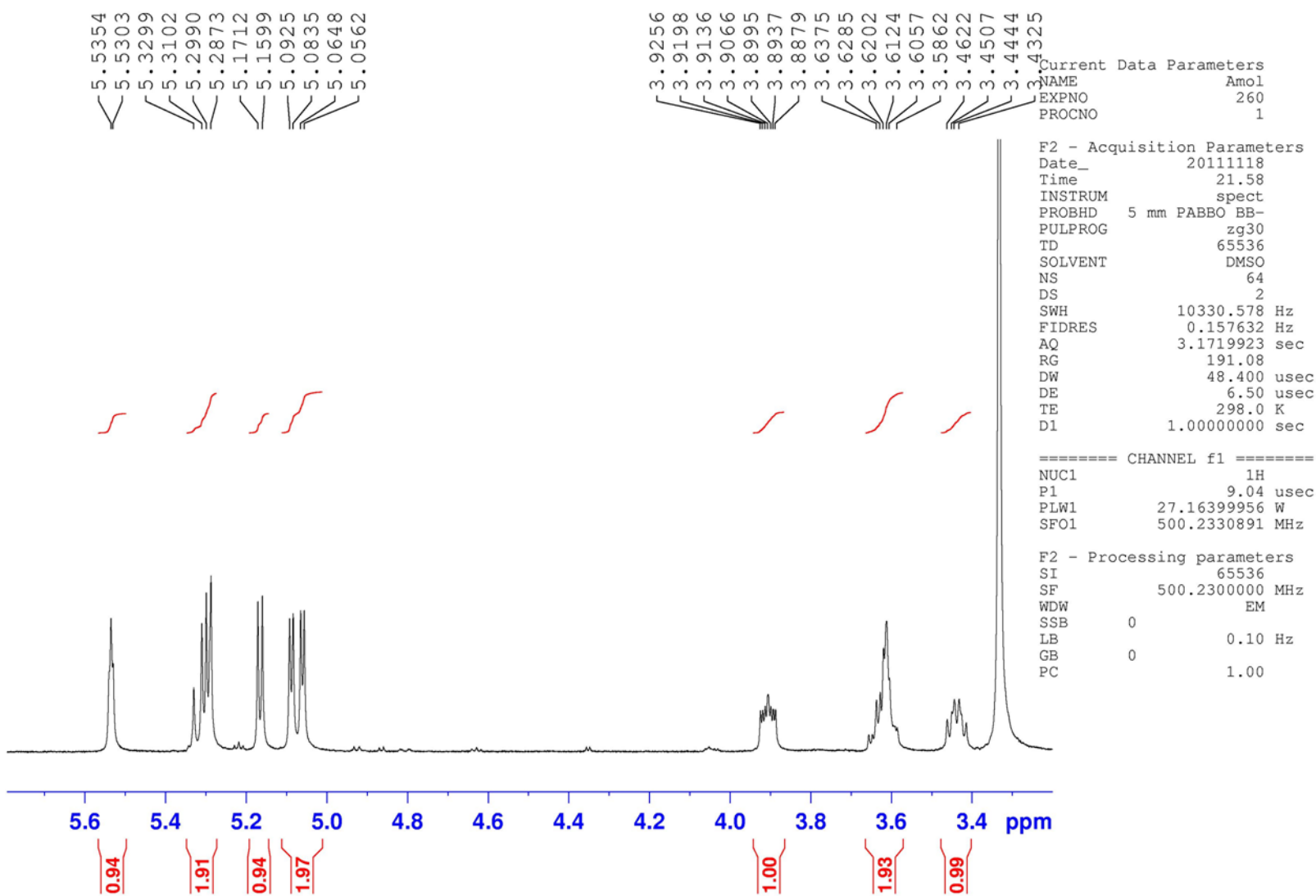
Supplementary Information

¹H NMR of **11** in DMSO-d₆



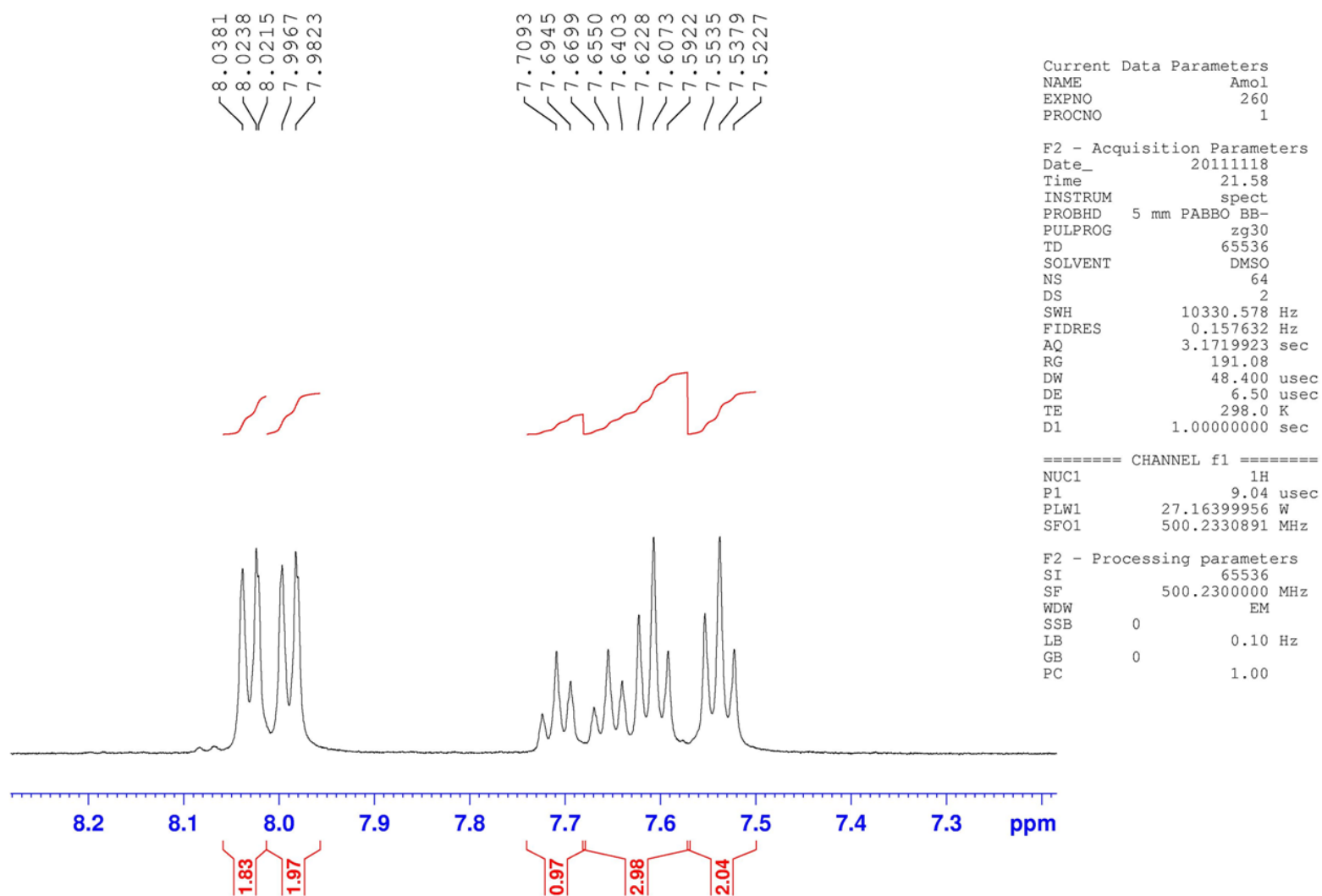
Supplementary Information

¹H NMR of **11** in DMSO-d₆ (zoom)



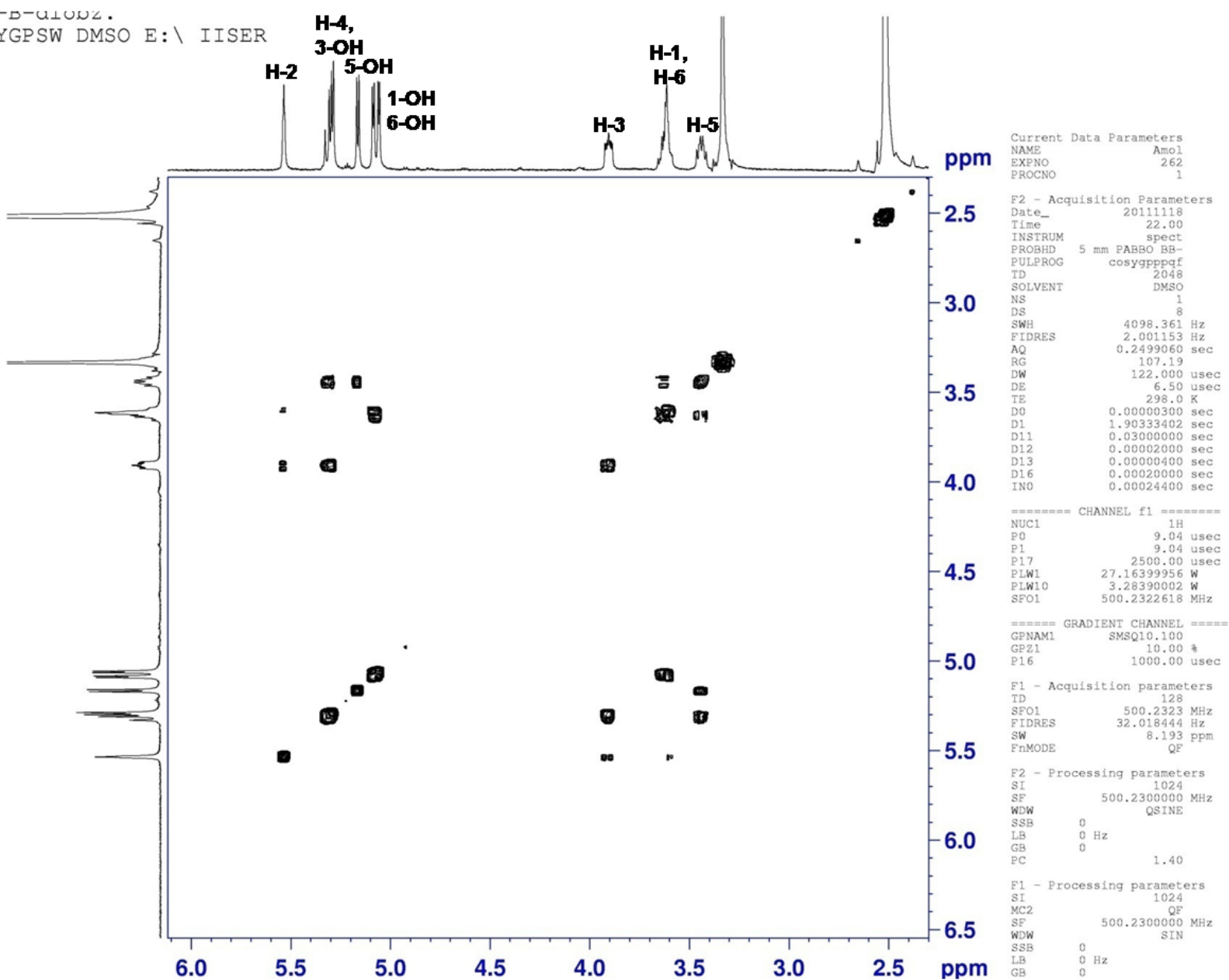
Supplementary Information

¹H NMR of **11** in DMSO-d₆ (zoom)



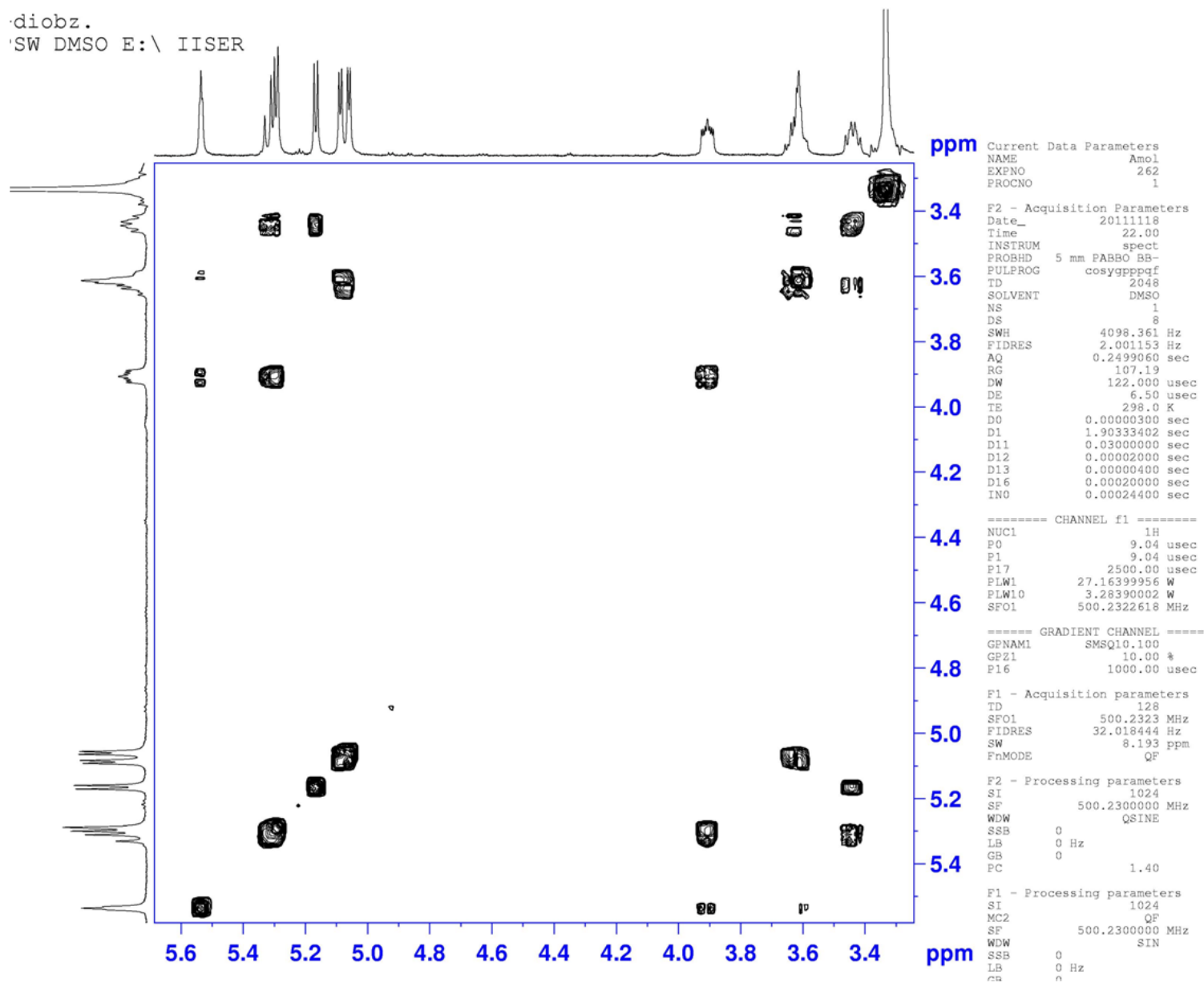
Supplementary Information

COSY of **11** in DMSO-d₆



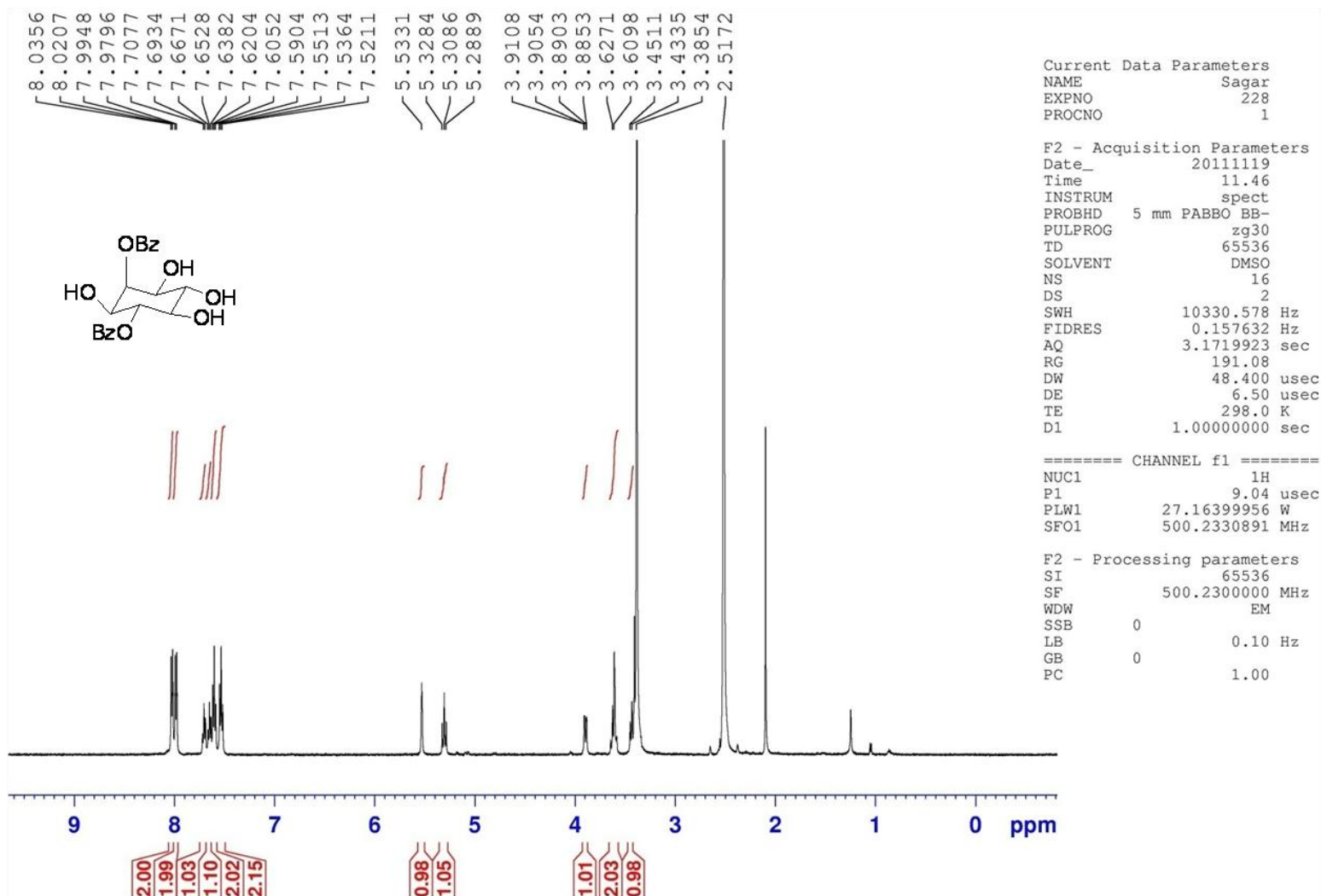
Supplementary Information

COSY of **11** in DMSO-d₆ (zoom)



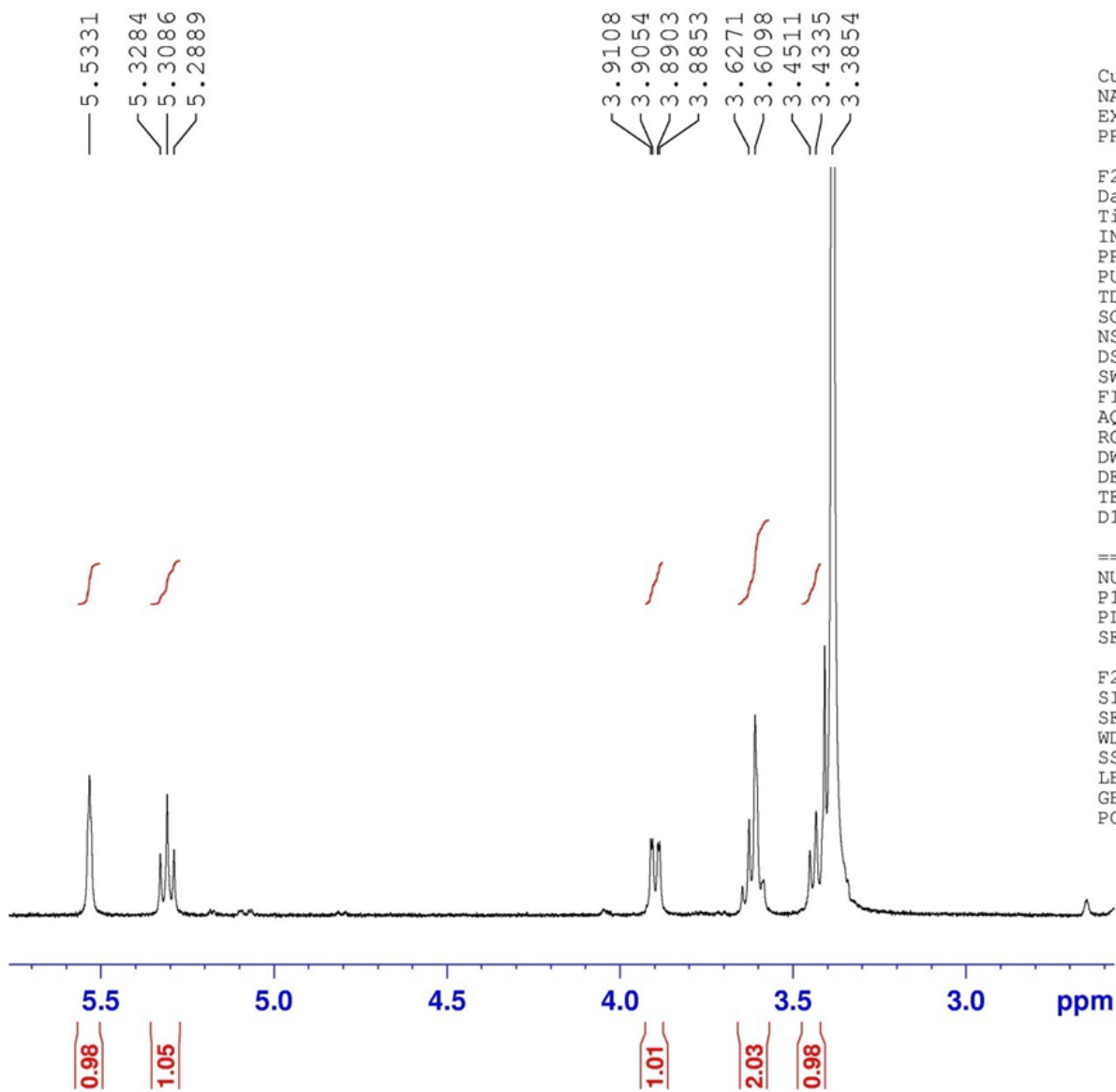
Supplementary Information

¹H NMR of **11** in DMSO-d₆ (D₂O Exchange)



Supplementary Information

¹H NMR of **11** in DMSO-d₆ (D₂O Exchange) (zoom)



Current Data Parameters
NAME Sagar
EXPNO 228
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111119
Time 11.46
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 191.08
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.00000000 sec

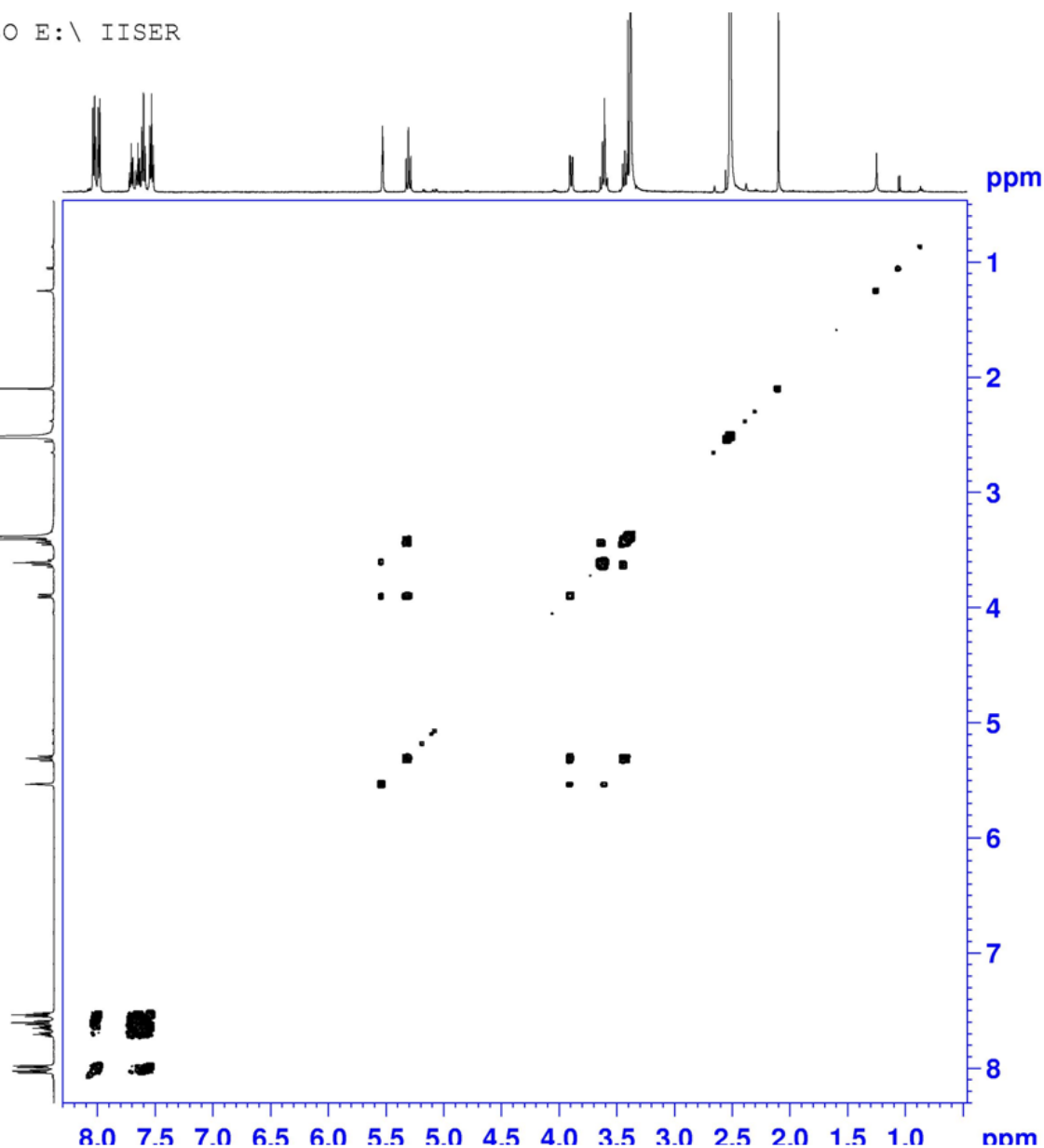
===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.230000 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

Supplementary Information

COSY of **11** in DMSO-d₆ (D₂O Exchange)

GPSW DMSO E:\ IISER



Current Data Parameters
NAME Sagar
EXPNO 236
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111119
Time 19.01
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG cosygpppqf
TD 2048
SOLVENT DMSO
NS 1
DS 8
SWH 4065.041 Hz
FIDRES 1.984883 Hz
AQ 0.2519540 sec
RG 107.19
DW 123.000 use
DE 6.50 use
TE 298.3 K
D0 0.0000300 sec
D1 1.90128601 sec
D11 0.03000000 sec
D12 0.00002000 sec
D13 0.00000400 sec
D16 0.00020000 sec
IN0 0.00024600 sec

===== CHANNEL f1 =====
NUC1 1H
P0 9.04 use
P1 9.04 use
P17 2500.00 use
PLW1 27.16399956 W
PLW10 3.28390002 W
SFO1 500.2322653 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SMSQ10.100
GPZ1 10.00 %
P16 1000.00 use

F1 - Acquisition parameters
TD 128
SFO1 500.2323 MHz
FIDRES 31.758131 Hz
SW 8.126 ppm
FnMODE QF

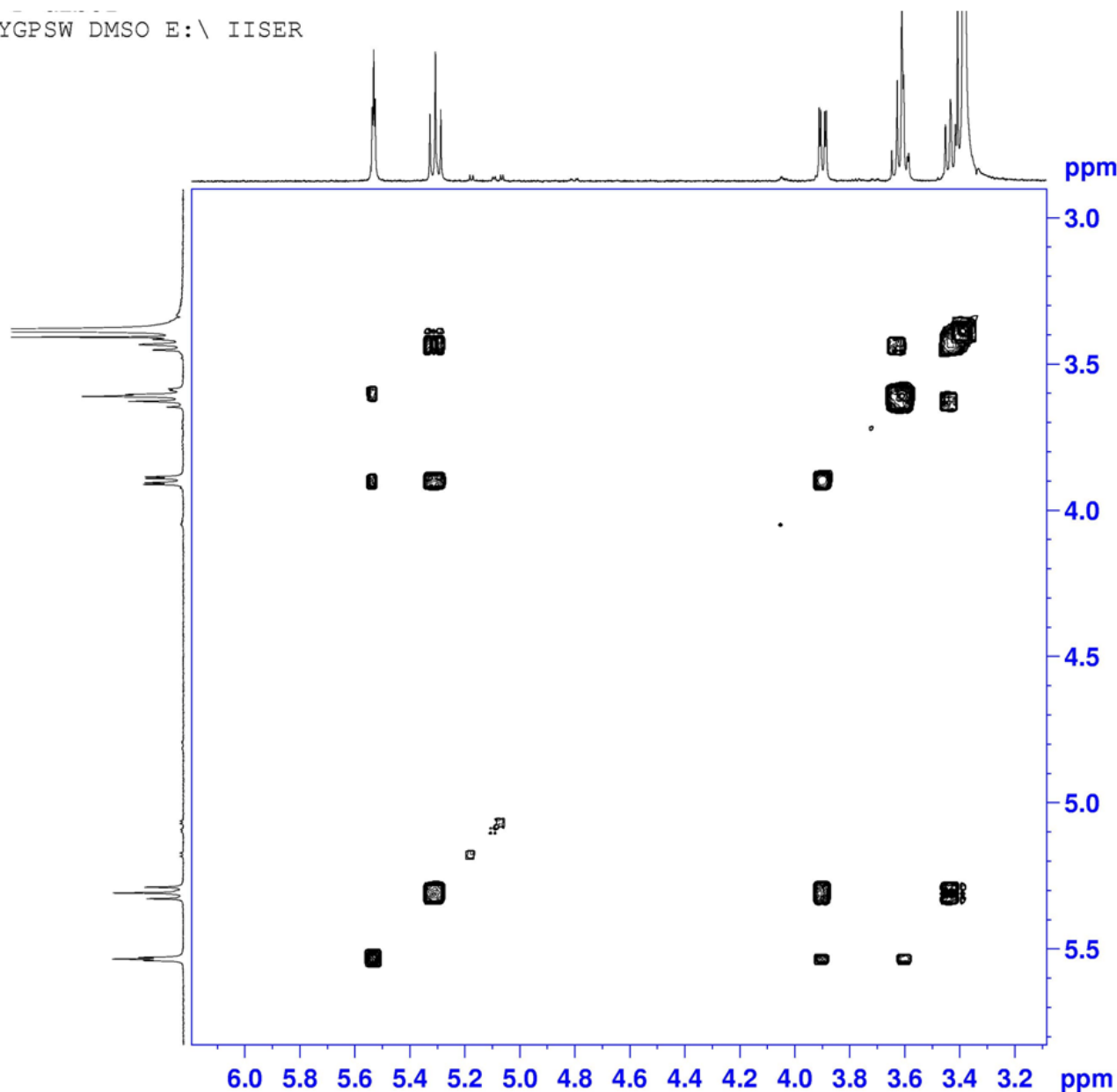
F2 - Processing parameters
SI 1024
SF 500.2300000 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 500.2300000 MHz
WDW SIN
SSB 0
LB 0 Hz
GB 0

Supplementary Information

COSY of **11** in DMSO-d₆ (D₂O Exchange) (zoom)

YGPSW DMSO E:\ IISER



Current Data Parameters
NAME Sagar
EXPNO 236
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111119
Time 19.01
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG cosygpppqf
TD 2048
SOLVENT DMSO
NS 1
DS 8
SWH 4065.041 Hz
FIDRES 1.984883 Hz
AQ 0.2519540 sec
RG 107.19
DW 123.000 usec
DE 6.50 usec
TE 298.3 K
D0 0.00000300 sec
D1 1.90128601 sec
D11 0.03000000 sec
D12 0.00002000 sec
D13 0.00000400 sec
D16 0.00020000 sec
IN0 0.00024600 sec

===== CHANNEL f1 =====
NUC1 1H
P0 9.04 usec
P1 9.04 usec
P17 2500.00 usec
PLW1 27.16399956 W
PLW10 3.28390002 W
SFO1 500.2322653 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SMSQ10.100
GPZ1 10.00 %
P16 1000.00 usec

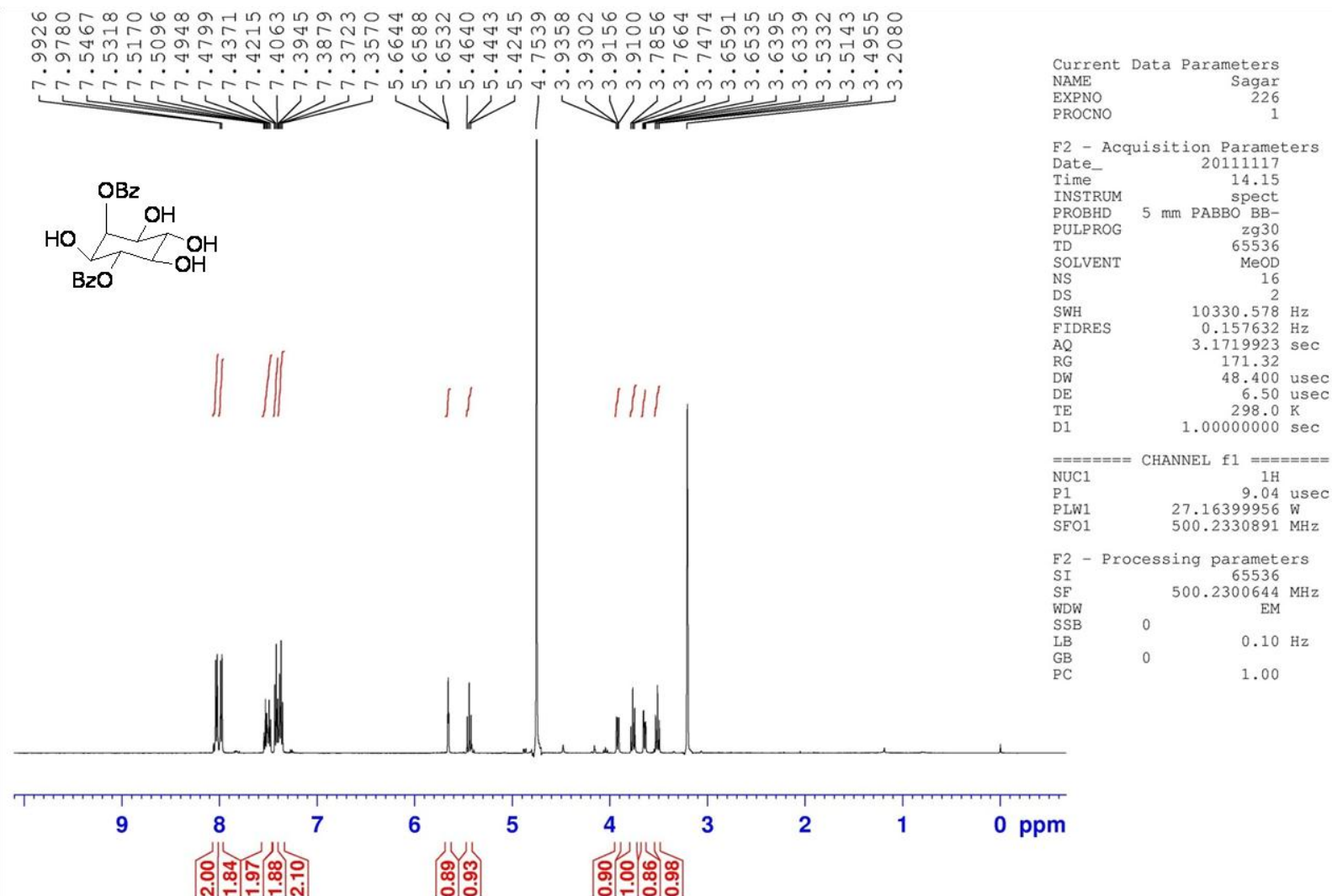
F1 - Acquisition parameters
TD 128
SFO1 500.2323 MHz
FIDRES 31.758131 Hz
SW 8.126 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 500.2300000 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 500.2300000 MHz
WDW SIN
SSB 0
LB 0 Hz
GB 0

Supplementary Information

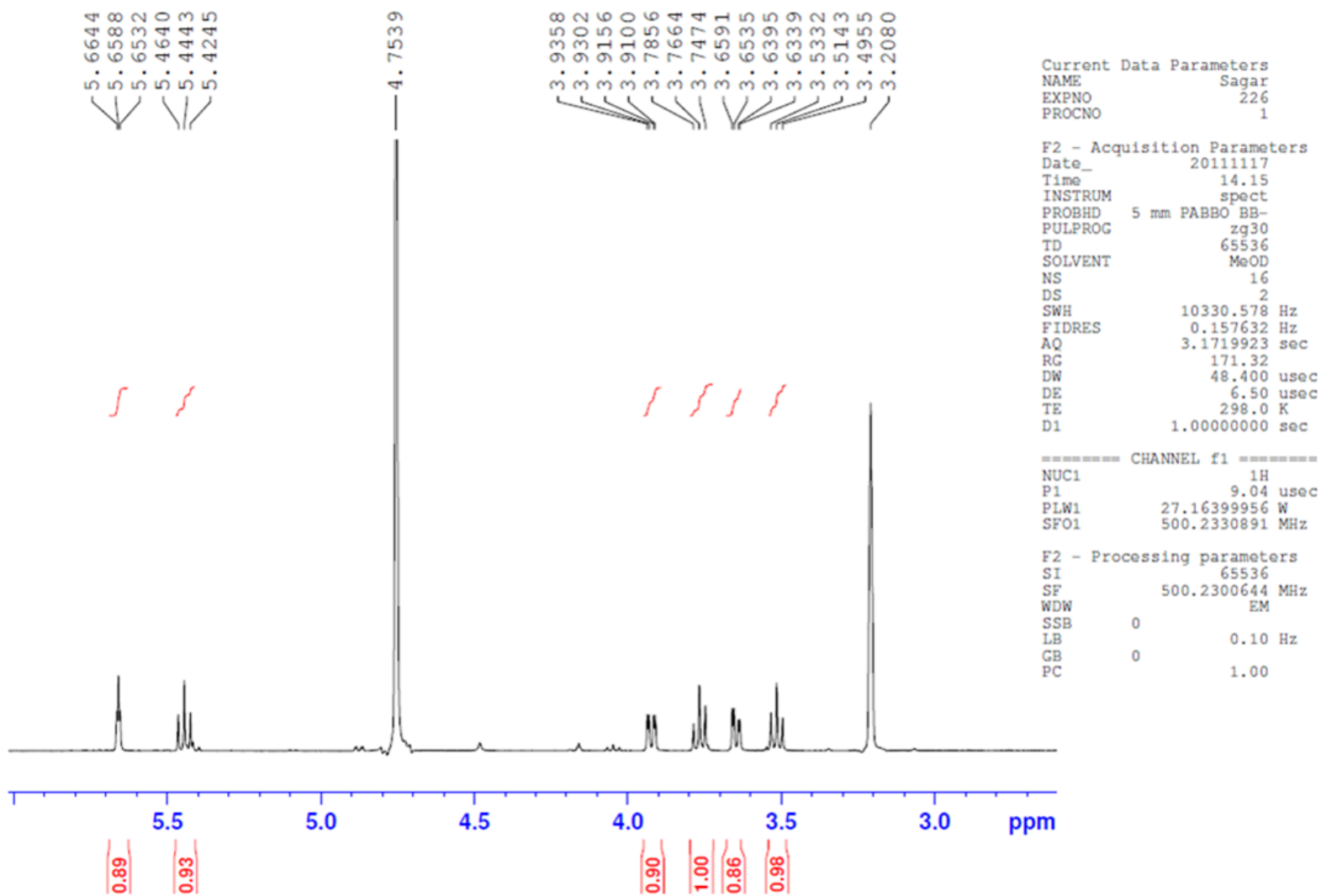
¹H NMR of **11** in CD₃OD



The minor peaks in the spectrum are due to the transesterification of dibenzoate in the presence of methanol.

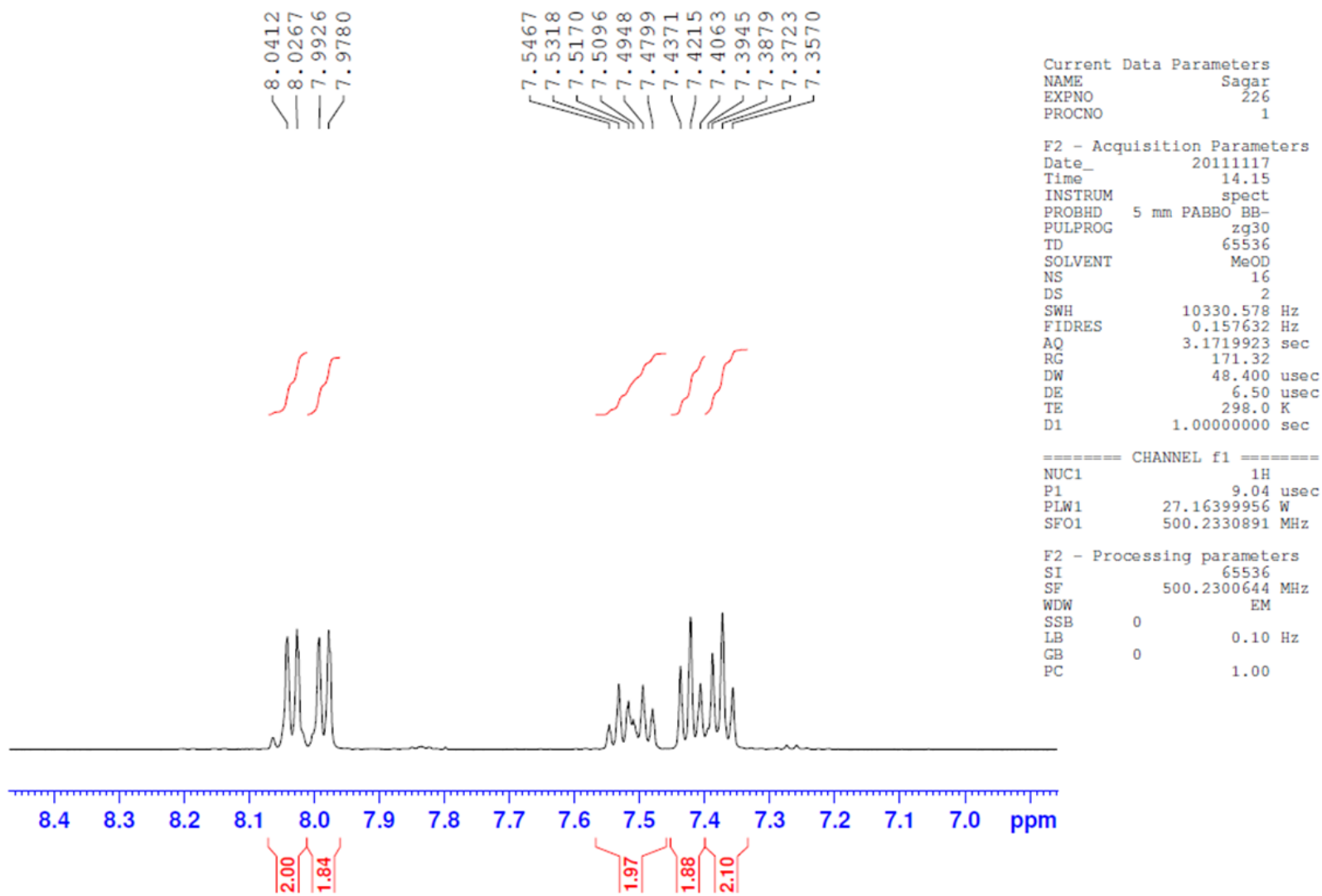
Supplementary Information

¹H NMR of **11** in in CD₃OD (zoom)



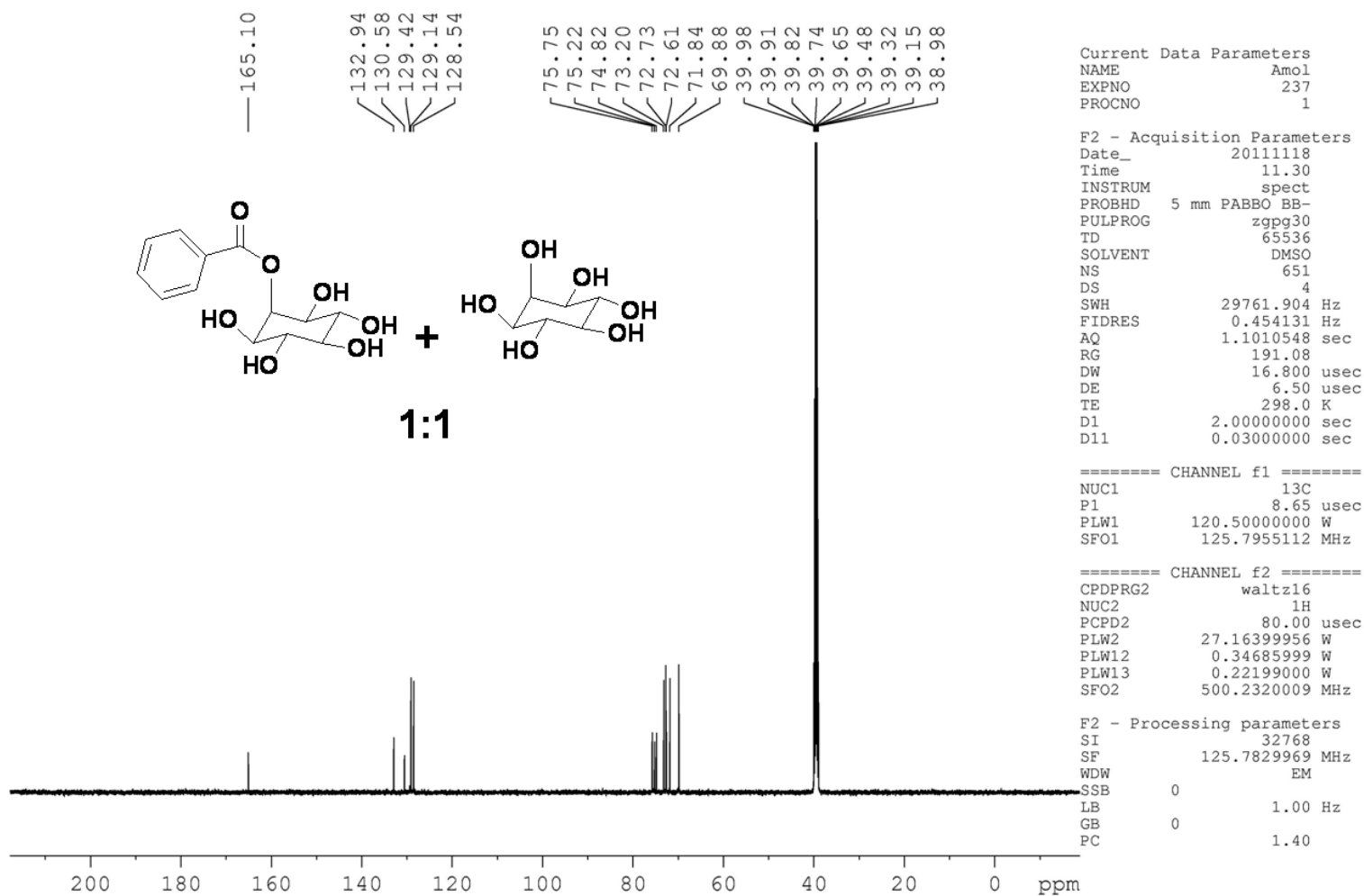
Supplementary Information

¹H NMR of **11** in in CD₃OD (zoom)



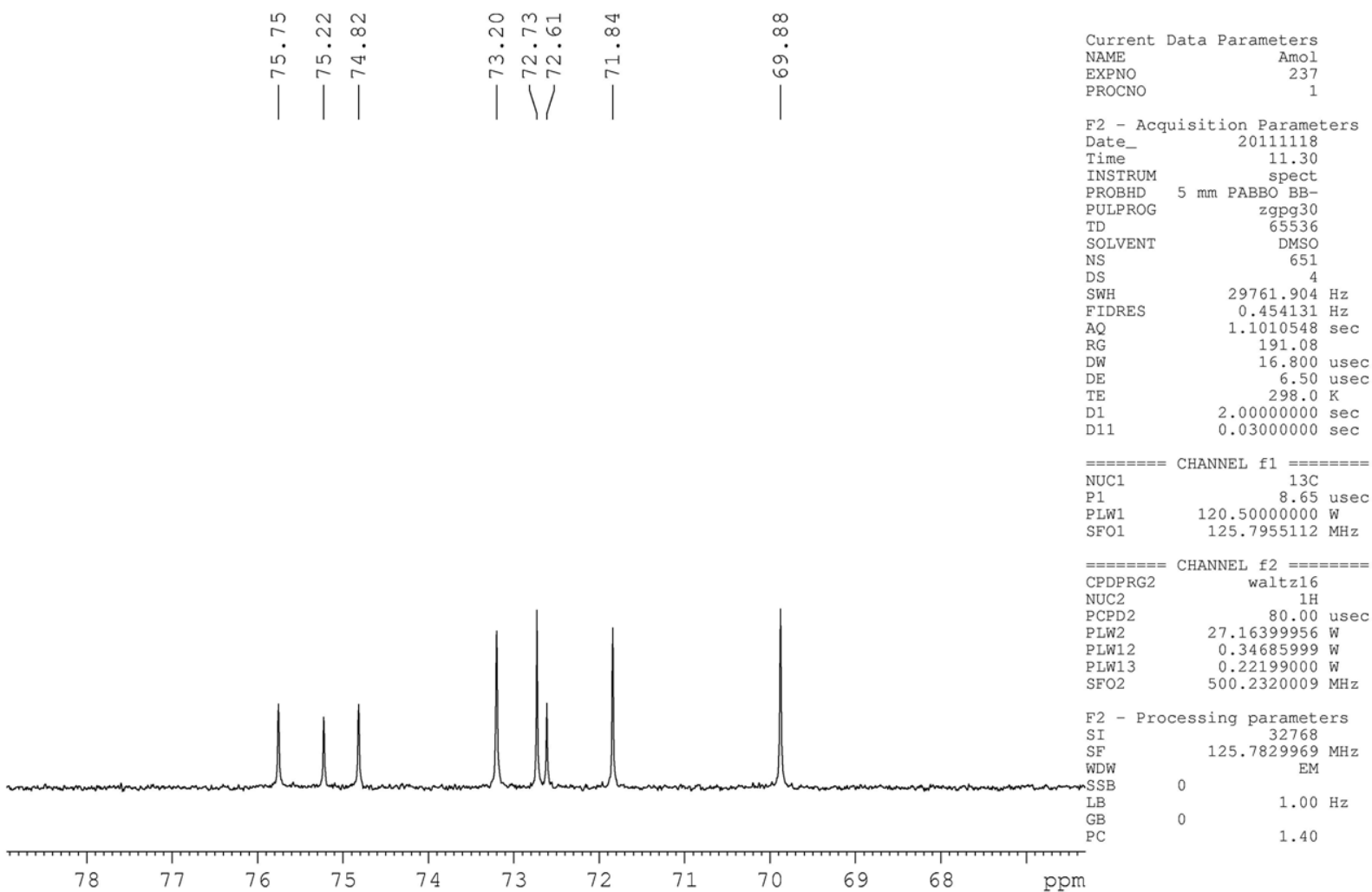
Supplementary Information

¹³C NMR of **3a** + *myo*-inositol (1:1)



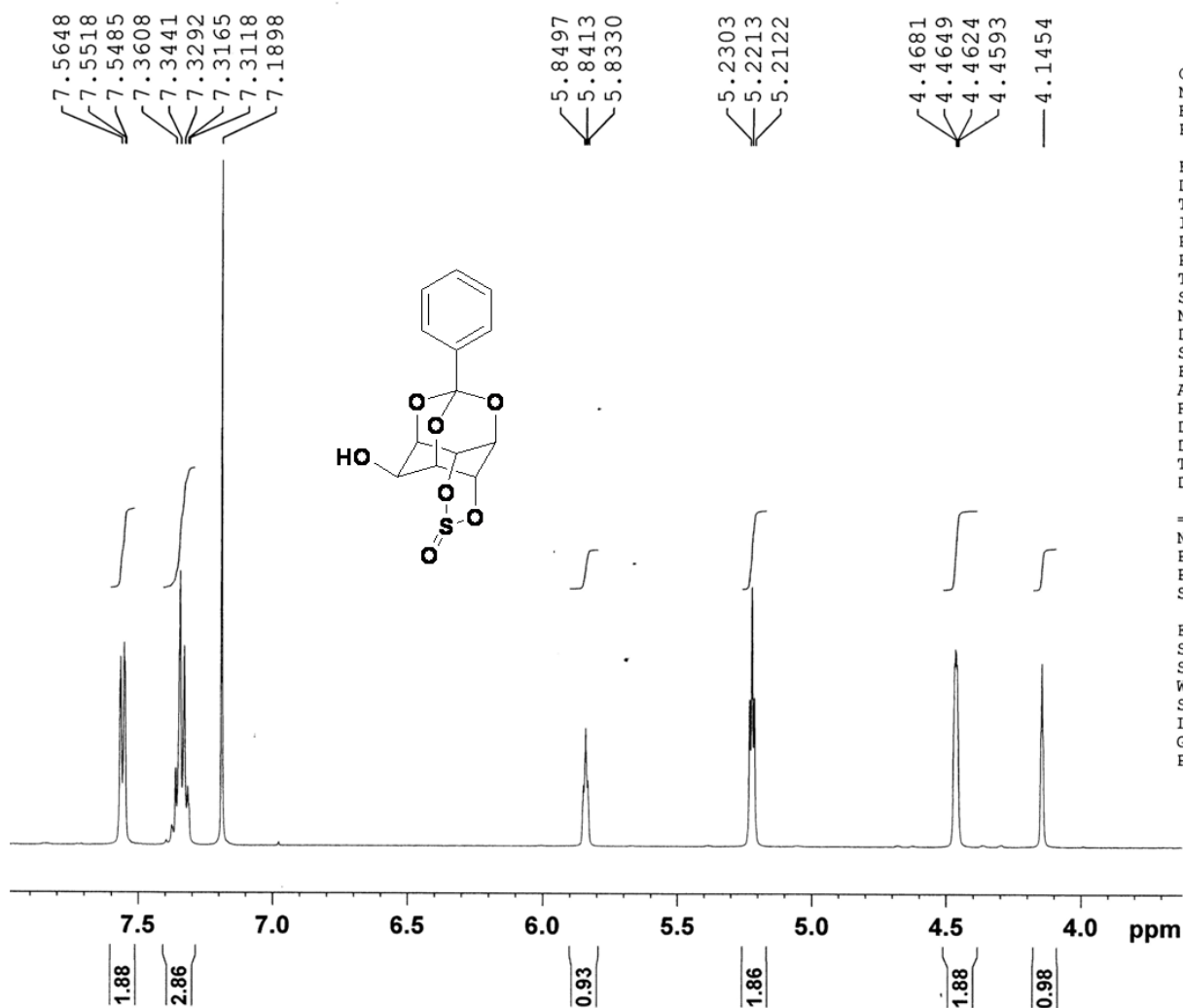
Supplementary Information

¹³C NMR of **3a** + *myo*-inositol (1:1) (zoom)



Supplementary Information

¹H NMR of **12** (zoom)



Current Data Parameters
NAME Amol
EXPNO 53
PROCNO 1

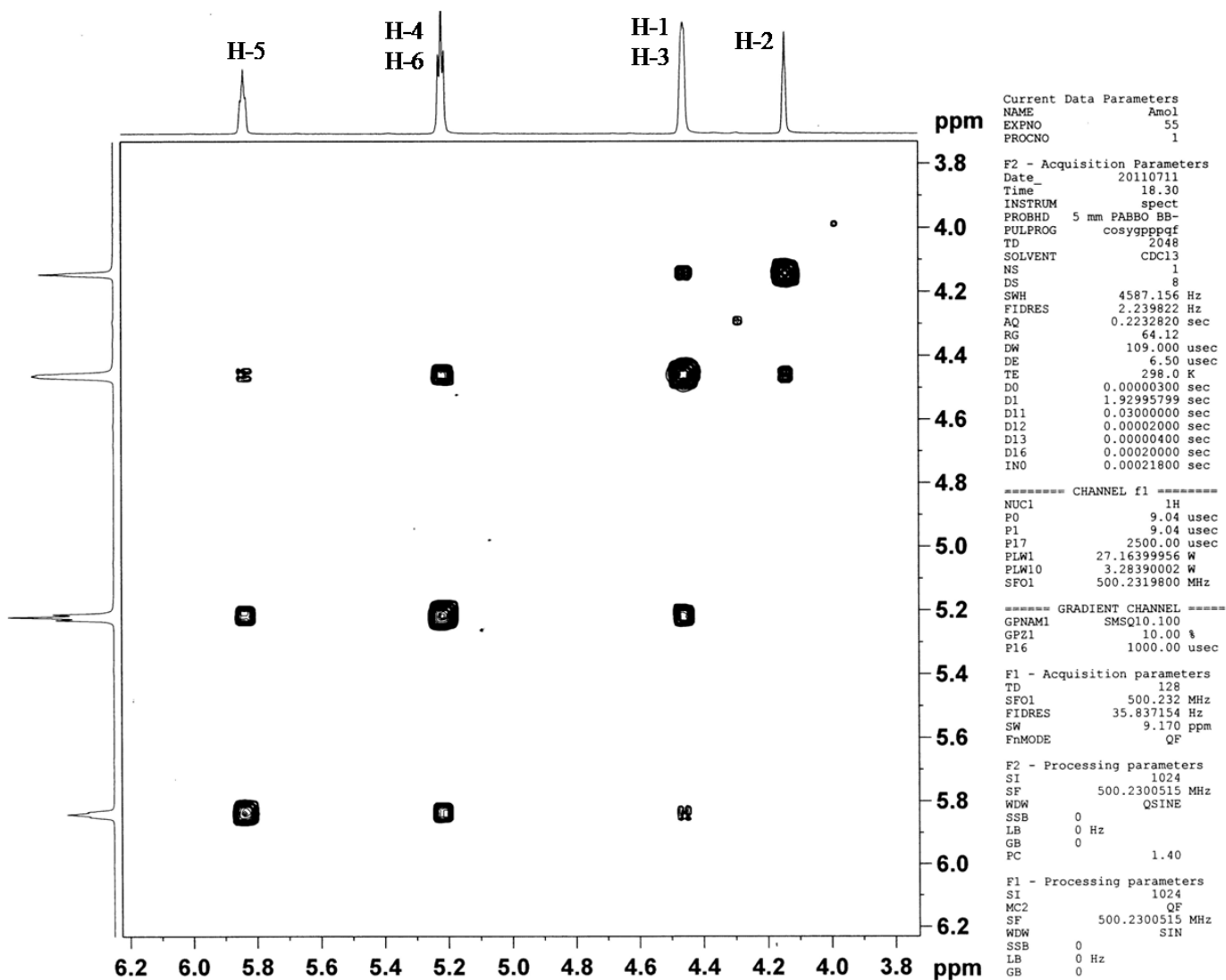
F2 - Acquisition Parameters
Date 20110711
Time 17.58
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 191.08
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300514 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

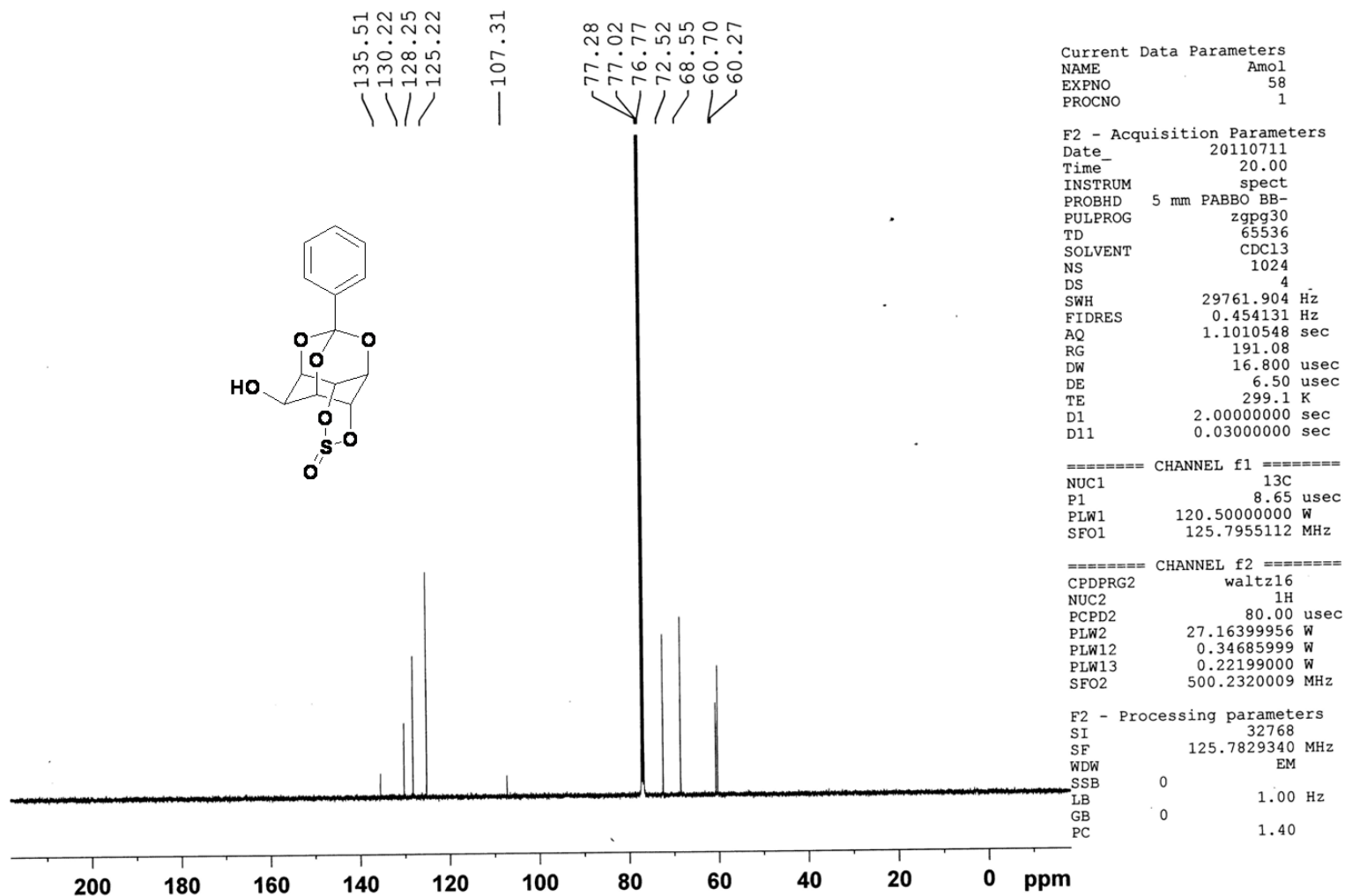
Supplementary Information

COSY of 12



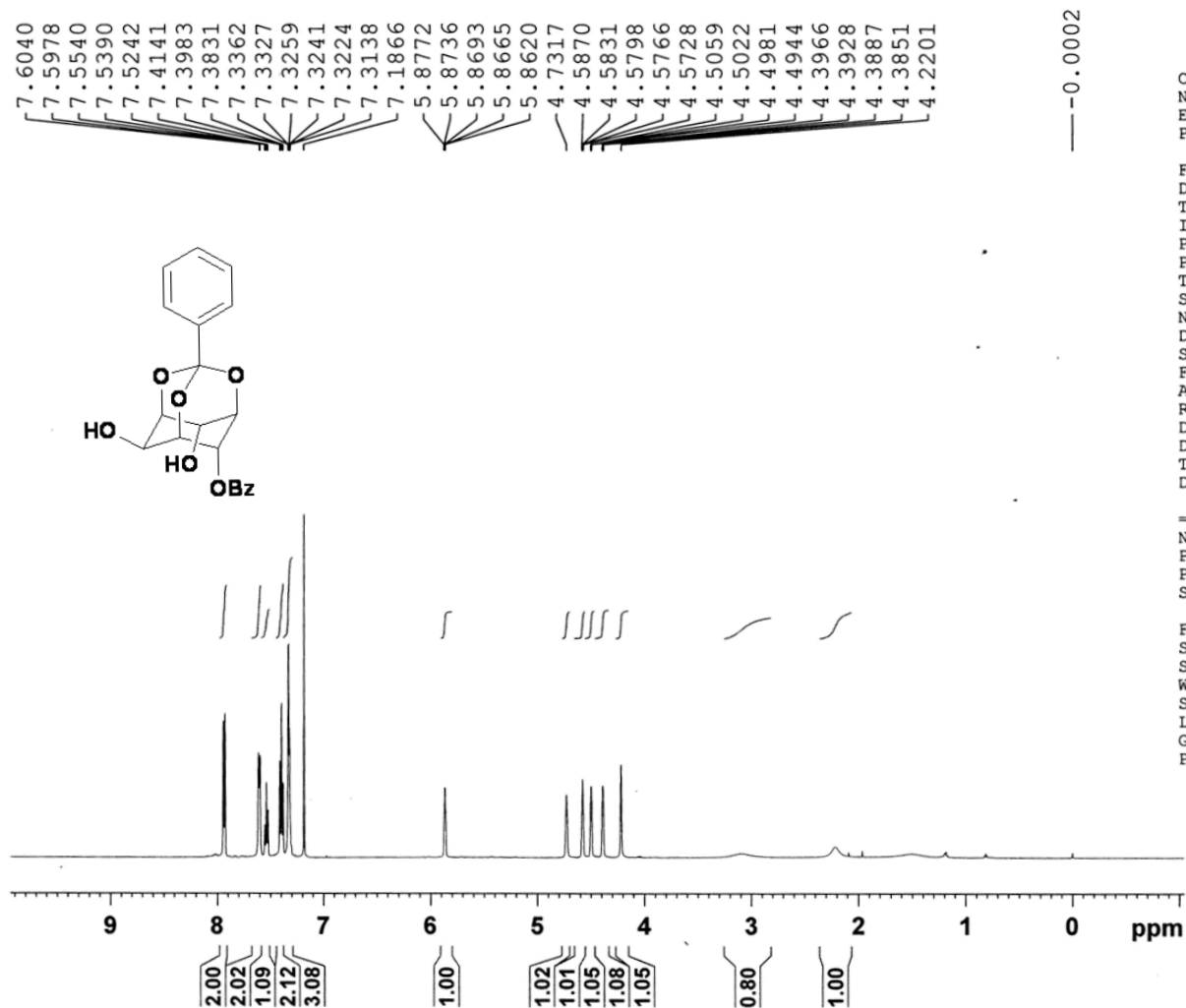
Supplementary Information

¹³C NMR of **12**



Supplementary Information

¹H NMR of **10**



Current Data Parameters
NAME Sagar
EXPNO 30
PROCNO 1

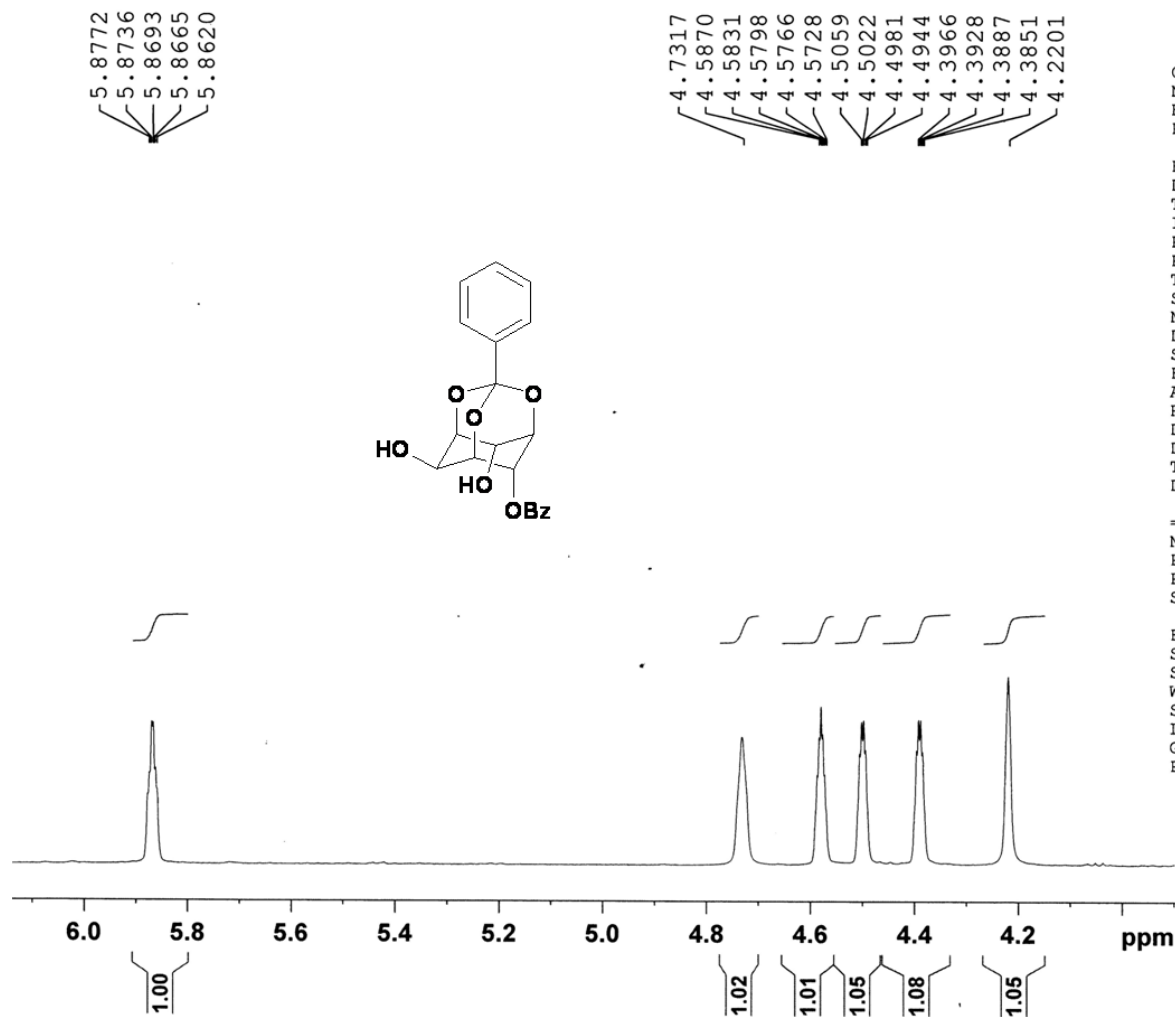
F2 - Acquisition Parameters
Date_ 20110712
Time_ 16.28
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 191.08
DW 48.400 usec
DE 6.50 usec
TE 302.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300533 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

Supplementary Information

¹H NMR of **10** (zoom)



Current Data Parameters
NAME Sagar
EXPNO 30
PROCNO 1

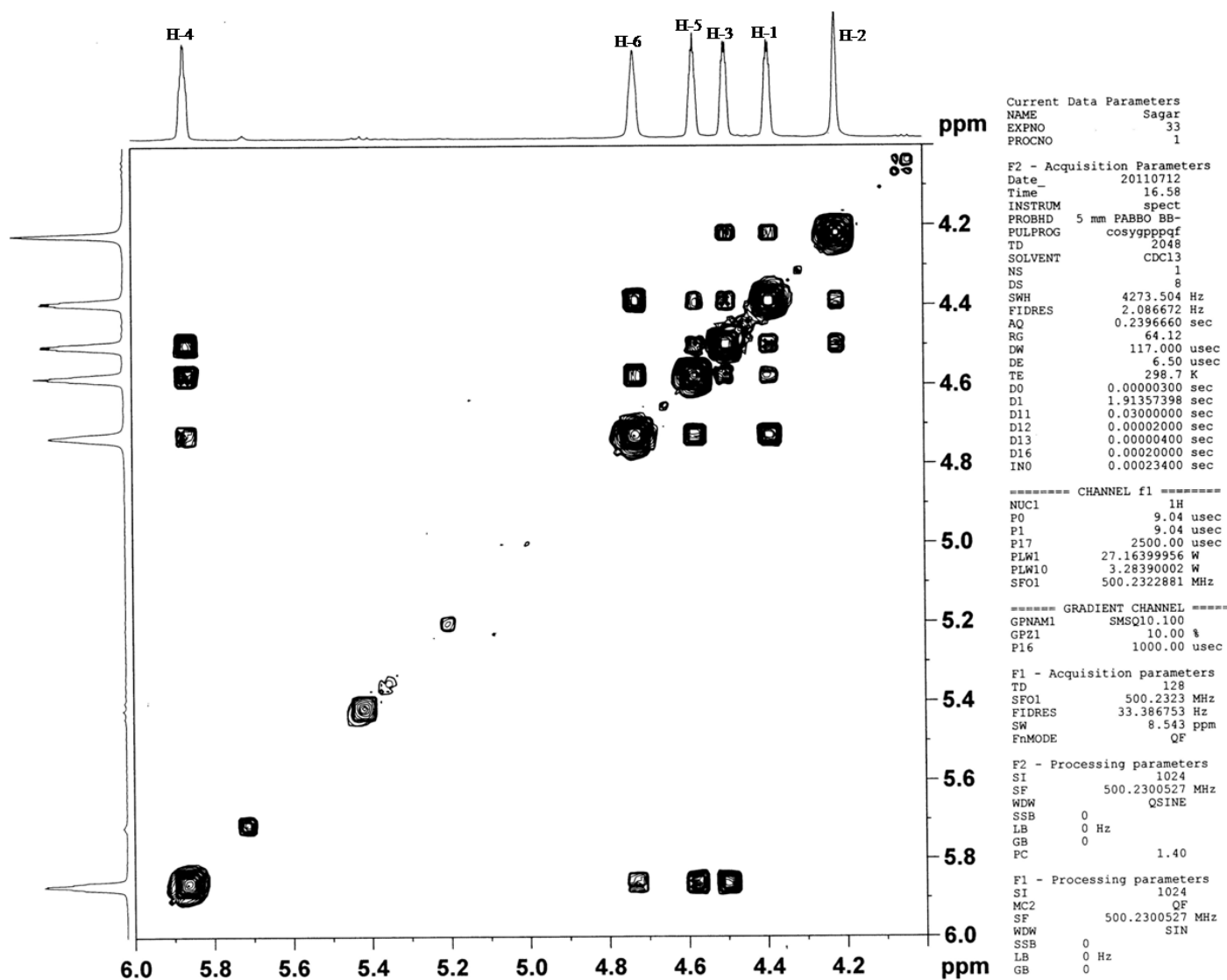
F2 - Acquisition Parameters
Date_ 20110712
Time 16.28
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 191.08
DW 48.400 usec
DE 6.50 usec
TE 302.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.04 usec
PLW1 27.16399956 W
SFO1 500.2330891 MHz

F2 - Processing parameters
SI 65536
SF 500.2300533 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

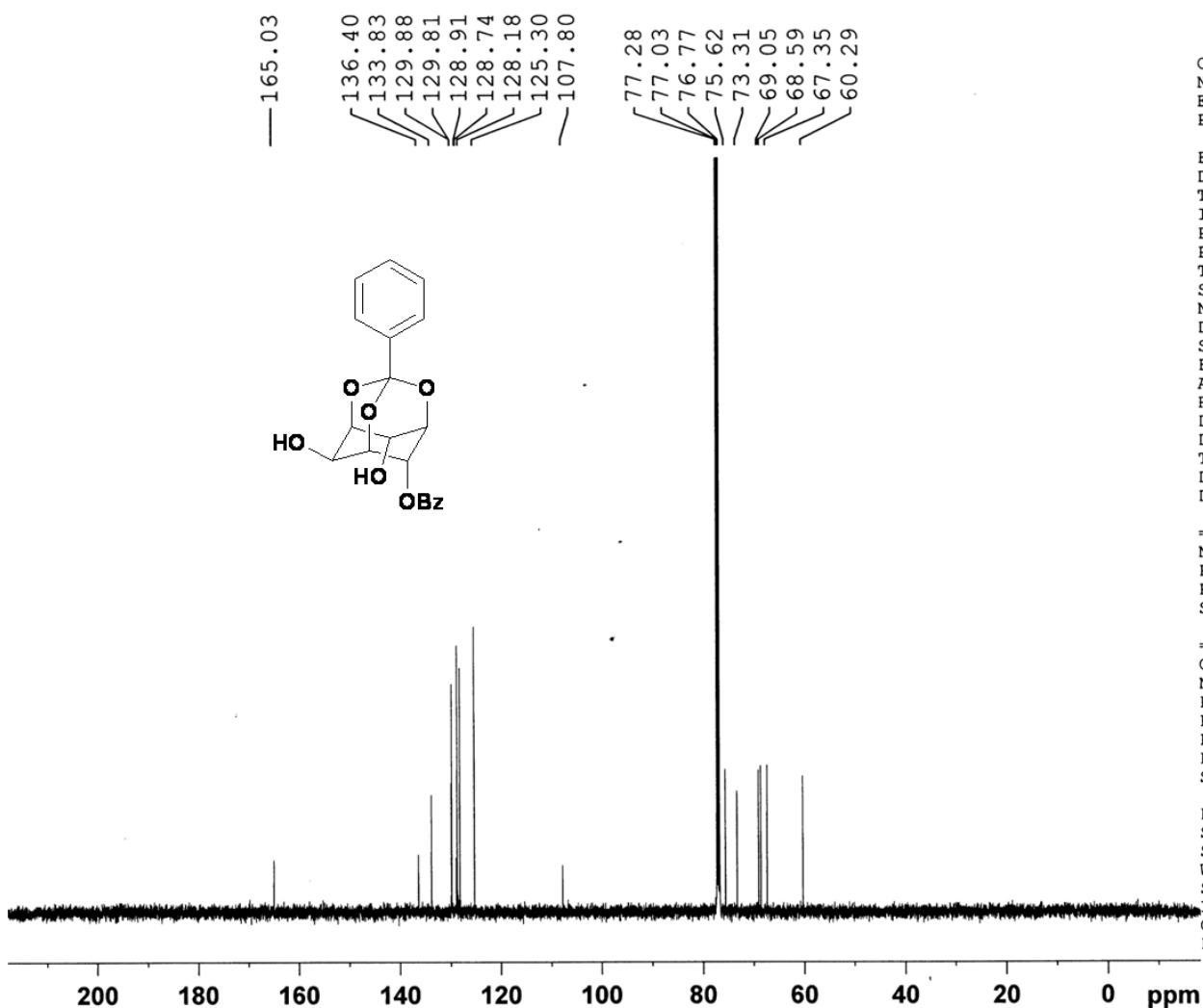
Supplementary Information

COSY of 10



Supplementary Information

¹³C NMR of **10**



```

Current Data Parameters
NAME          Sagar
EXPNO         35
PROCNO        1

F2 - Acquisition Parameters
Date_         20110712
Time          17.08
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            502
DS            4
SWH           29761.904 Hz
FIDRES        0.454131 Hz
AQ            1.1010548 sec
RG            191.08
DW            16.800 usec
DE            6.50 usec
TE            298.7 K
D1            2.00000000 sec
D11           0.03000000 sec

===== CHANNEL f1 =====
NUC1           13C
P1             8.65 usec
PLW1          120.50000000 W
SFO1          125.7955112 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         80.00 usec
PLW2          27.16399956 W
PLW12         0.34685999 W
PLW13         0.22199000 W
SFO2          500.2320009 MHz

F2 - Processing parameters
SI            32768
SF           125.7829340 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40
    
```