Supporting Information

for

Probe sialidase substrate specificity using chemoenzymatically synthesized sialosides containing C9-modified sialic acids

Zahra Khedri,‡^a Musleh M. Muthana,‡ Yanhong Li,‡ Saddam M. Muthana,§ Hai Yu, Hongzhi Cao¶ and Xi Chen*

Department of Chemistry, University of California, One Shields Avenue, Davis, California 95616, USA. E-mail: chen@chem.ucdavis.edu; Fax: +1 530-752-8995; Tel: +1 530-754-6037

‡These authors contributed equally

§ Current address: National Cancer Institute, Frederick, Maryland, 20702, USA

¶Current address: National Glycoengineering Research Center, Shandong University, Jinan, Shandong 250012, P. R. China

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General methods

All chemicals were obtained from commercial suppliers and used without further purification unless otherwise noted. Anhydrous solvents were used to carry out organic reactions under inert argon or nitrogen environment. ¹H NMR (600 MHz) and ¹³C NMR (151 MHz) spectra were recorded on a Varian Inova-600 spectrometer and ¹⁹F NMR (282 MHz) spectra were recorded on a Mercury-300 spectrometer. High resolution electrospray ionization (ESI) mass spectra were obtained at the Mass Spectrometry Facility in the University of California, Davis. Silica gel 60 Å (200–425 mesh, Fisher Chemical) was used for flash column chromatography. Thin-layer chromatography (TLC) was performed on silica gel plates 60 GF₂₅₄ (Sorbent technologies) using anisaldehyde sugar stain or 5% sulfuric acid in ethanol stain for detection. Gel filtration chromatography was performed using a column (100 cm × 2.5 cm) packed with Bio-gel P-2 Fine resins (Bio-Rad, Hercules, CA).



Synthesis of ManNAc, ManNGc, and mannose derivatives as precursors for sialosides

N-Acetylmannosamine (ManNAc) **1a** and mannose **11a** were purchased from Sigma. *N*-Glycolylmannosamine (ManNGc) **6a** and mannose derivatives $6-N_3$ -mannose **5a** and $6-N_3$ -ManNAc **15a** were synthesized as described previously.^{1, 2} Compounds **2a–4a**, **7a–10a**, **and 12a–14a** were synthesized as described in the following:

2-Acetamido-2,6-dideoxy-6-fluoro-D-mannopyranoside (6-F-ManNAc, 2a)



To a solution of compound 17^3 (9.04 g, 25.22 mmol) in CH₂Cl₂ (100 mL) and pyridine (20 mL), trifluoromethanesulfonic acid anhydride (Tf₂O) (4.67 mL, 27.74 mmol) was added drop wisely at -20 °C. The reaction mixture was stirred at between -20 °C to 0 °C for 3 h. The solvent was removed by rotovap, co-evaporated with toluene, and dried under vacuum for overnight. The crude product was then dissolved in anhydrous toluene (150 mL) and tetra-*n*-butylammonium azide (10.76 g, 37.82 mmol) was added. The mixture was stirred at 65 °C for 1 h and then 100 °C for 1.5 h. After cooled, the reaction mixture was diluted with toluene, washed with water, and concentrated. The residue was subjected to silica gel column chromatography (Hexanes:EtOAc = 9:1) to give **18** (8.02 g, 83%). ¹H NMR (600 MHz, CDCl₃) δ 7.51–7.50 (m, 2H), 7.42–7.35 (m, 8H), 5.57 (s, 1H), 4.87 (s, 1H), 4.72 (d, *J* = 12.0 Hz, 1H), 4.51 (d, *J* = 12.0 Hz, 1H), 4.29–4.27 (m, 1H), 4.23 (dd, *J* = 9.0, 3.6 Hz, 1H), 3.94–3.90 (m, 2H), 3.84–3.79 (m, 2H), 2.89 (s, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 137.10, 136.59, 129.43, 128.70, 128.50, 128.32, 128.17, 126.37, 102.35, 98.32, 79.10, 69.64, 68.96, 68.70, 63.83, 63.71.

A solution of **18** (5.50 g, 14.34 mmol) in dry DMF (40 mL) was stirred with NaH (0.52 g, 21.51 mmol) for 30 min at 0 °C. Tetra-*n*-butylammonium iodide (0.53 g, 1.43 mmol) and benzyl bromide (3.5 mL, 28.70 mmol) were added and the reaction was stirred at 0 °C to room temperature for

overnight. The reaction was stopped by adding MeOH (2 mL), CH_2Cl_2 was then added and the mixture was washed with water, dried over MgSO₄, and concentrated. Silica gel column chromatography of the residue (Hexanes:EtOAc = 9.5:0.5) gave **19** (6.45 g, 95%).

Compound **19** (1.85 g, 3.90 mmol) was dissolved in anhydrous CH₂Cl₂ (7 mL) with 4 Å molecular sieves (1.5 g) and stirred for 1 h at room temperature. The resulting mixture was cooled to -78 °C and Et₃SiH (0.33 mL, 2.08 mmol) and PhBCl₂ (0.24 mL, 1.88 mmol) were added successively to the stirred solution. After being stirred at -78 °C for 1.5 h, Et₃N (1 mL) and MeOH (1 mL) were added successively and the mixture was filtered over Celite and concentrated. The residue was diluted with CH₂Cl₂, washed with an aqueous solution of NaHCO₃, dried over MgSO₄, and concentrated. The crude product was purified by silica gel column (Hexanes:EtOAc = 8:2) to give **20** (1.76 g, 95%). ¹H NMR (600 MHz, CDCl₃) δ 7.47 (d, *J* = 6.6 Hz, 2H), 7.44–7.36 (m, 13H), 4.97 (d, *J* = 10.8 Hz, 1H), 4.93 (d, *J* = 1.8 Hz, 1H), 4.80 (dd, *J* = 19.2, 11.4 Hz, 2H), 4.74 (d, *J* = 3.0 Hz, 1H), 4.73 (d, *J* = 1.8 Hz, 1H), 4.53 (d, *J* = 12.0 Hz, 1H), 4.22 (dd, *J* = 6.0, 3.6 Hz, 1H), 4.03 (dd, *J* = 3.6, 1.8 Hz, 1H), 4.00 (t, *J* = 9.6 Hz, 1H), 3.90–3.84 (m, 2H), 3.79–3.76 (m, 1H), 2.44 (t, *J* = 5.4 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 138.10, 137.77, 136.68, 128.55, 128.51, 128.14, 128.09, 128.07, 128.05, 127.92, 127.90, 97.45, 79.77, 75.41, 74.24, 72.64, 72.40, 69.35, 61.81, 61.43.

To a solution of **20** (1.48 g, 3.11 mmol) in anhydrous CH₂Cl₂ (15 mL) in a Teflon flask, (diethylamino)sulfur trifluoride (DAST) (2.05 mL, 15.55 mmol) was slowly added at -40 °C. The reaction was stirred for 3 days at room temperature. After cooled to -20 °C, MeOH (5 mL) was added and the solvent was removed under reduced pressure. The residue was diluted with CH₂Cl₂, washed with water for 3 times, dried over MgSO₄, and concentrated. The crude product was purified using flash column chromatography (Hexanes:EtOAc = 10:1) to yield **21** (0.77 g, 52%). ¹H NMR (600 MHz, CDCl₃) δ 7.42–7.30 (m, 15 H), 4.94 (d, *J* = 10.8 Hz, 1H), 4.90 (s, 1H), 4.76 (dd, *J* = 24.0, 11.4 Hz, 2H), 4.70–4.59 (m, 4H), 4.51 (d, *J* = 12.0 Hz, 1H), 4.16 (dd, *J* = 9.0, 3.6 Hz, 1H), 3.98 (dd, *J* = 3.6, 1.8 Hz, 1H), 3.92 (t, *J* = 9.6 Hz, 1H), 3.83–3.76 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 137.99, 137.74, 136.70, 128.65, 128.23, 128.06, 97.58, 82.03 (d, *J* = 175.9 Hz), 79.87, 75.60, 73.58, 72.77, 71.37 (d, *J* = 18.6 Hz), 69.61, 61.39; ¹⁹F NMR (282 MHz, CDCl₃) δ -233.30 (ddd, *J* = 48.2, 48.2, 27.3 Hz, 1F).

Compound **21** (0.78 g, 1.63 mmol) was dissolved in pyridine (5 mL) and thioacetic acid (10 mL) and stirred for overnight at room temperature. The solvent was removed under diminished pressure and the crude product was purified using silica gel column chromatography (Hexanes:EtOAc = 8:2) to afford compound **22** (0.68 g, 84%). ¹H NMR (600 MHz, CDCl₃) δ 7.37–7.28 (m, 15H), 5.70 (d, *J* = 8.4 Hz, 1H), 4.98 (d, *J* = 1.8 Hz, 1H), 4.95 (d, *J* = 10.2 Hz, 1H), 4.76–4.45 (m, 8H), 4.14 (dd, *J* = 9.6, 4.8 Hz, 1H), 3.77 (dd, *J* = 10.2, 3.0 Hz, 1H), 3.65 (t, *J* = 9.9 Hz, 1H), 2.05 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 170.44, 138.22, 137.97, 136.94, 128.71, 128.69, 128.67, 128.50, 128.23, 128.18, 128.07, 99.10, 82.75 (d, *J* = 172.1 Hz), 77.75, 75.64, 73.64, 73.38 (d, *J* = 6.0 Hz), 71.52, 70.55 (d, *J* = 16.6 Hz), 70.11, 49.59, 23.73; ¹⁹F NMR (282 MHz, CDCl₃) δ -234.29 (ddd, *J* = 47.4, 47.4, 26.5 Hz, 1F).

Compound **22** (0.68 g, 1.38 mmol) was dissolved in AcOH (5 mL) and MeOH (25 mL), and Pd(OH)₂/C (80 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by flash column chromatography

(EtOAc:MeOH:H₂O = 10:1:0.1) to afford **2a** (0.26 g, 85%). ¹H NMR (600 MHz, D₂O) δ 5.13 (s, 0.7H), 5.05 (d, *J* = 1.2 Hz, 0.3H), 4.75–4.60 (m, 2H), 4.46 (dd, *J* = 4.8, 1.8 Hz, 0.3H), 4.31 (d, *J* = 1.2 Hz, 0.7H), 4.08–3.54 (m, 3H), 2.08 (s, 1H), 2.04 (s, 2H); ¹³C NMR (151 MHz, D₂O) δ 175.90, 174.99, 93.31, 93.28, 82.99 (d, *J* = 21.6 Hz), 81.85 (d, *J* = 21.6 Hz), 75.09 (d, *J* = 18.1 Hz), 71.92, 70.96 (d, *J* = 17.3 Hz), 68.65, 65.96 (d, *J* = 7.2 Hz), 65.74 (d, *J* = 4.1 Hz), 54.07, 53.37, 22.19, 22.03; ¹⁹F NMR (282 MHz, D₂O) δ -234.17 (ddd, *J* = 47.9, 47.9, 25.4 Hz, 0.4F), -235.04 (ddd, *J* = 50.7, 50.7, 19.7 Hz, 0.6F).

2-Acetamido-2-deoxy-6-O-methyl-D-mannopyranoside (6-O-Me-ManNAc, 3a)



A solution of **20** (1.70 g, 3.57 mmol) in anhydrous DMF (15 mL) was stirred with NaH (0.35 g, 14.30 mmol) at room temperature for 30 min. Methyl iodide (0.9 mL, 14.30 mmol) was added slowly at 0 °C and the mixture was stirred for overnight at room temperature. The mixture was diluted with CH₂Cl₂, washed with water, dried over MgSO₄, and concentrated. Purification of the residue by a silica gel column chromatography (Hexanes:EtOAc = 9.5:0.5) gave **23** (1.69 g, 97%). ¹H NMR (600 MHz, CDCl₃) δ 7.40–7.29 (m, 15H), 4.91-4.89 (m, 2H), 4.74 (dd, *J* = 19.8, 11.4 Hz, 2H), 4.62 (d, *J* = 10.8 Hz, 1H), 4.47 (d, *J* = 12.0 Hz, 1H), 4.12 (dd, *J* = 9.6, 3.6 Hz, 1H), 3.97 (dd, *J* = 3.6, 1.2 Hz, 1H), 3.93 (t, *J* = 9.9 Hz, 1H), 3.78–3.76 (m, 1H), 3.64 (dd, *J* = 10.8, 4.2 Hz, 1H), 3.58 (dd, *J* = 10.8, 1.8 Hz, 1H), 3.41 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 138.49, 138.09, 137.05, 128.71, 128.67, 128.27, 128.06, 97.70, 80.05, 75.59, 74.67, 72.84, 71.83, 71.43, 69.45, 61.70, 59.56.

Compound **23** (1.26 g, 2.57 mmol) was then dissolved in pyridine (5 mL) and thioacetic acid (10 mL) and stirred for overnight at room temperature. The solvent was removed under diminished pressure and the crude product was purified using silica gel column (Hexanes:EtOAc = 7:3) to afford compound **24** (1.09 g, 84%), which was dissolved in AcOH (5 mL) and MeOH (25 mL), and Pd(OH)₂/C (80 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by silica gel column chromatography (EtOAc:MeOH:H₂O = 10:1:0.2) to afford **3a** (0.42 g, 83%). ¹H NMR (600 MHz, D₂O) δ 5.11 (s, 0.6H), 5.02 (d, *J* = 1.8 Hz, 0.4H), 4.45 (dd, *J* = 1.8, 4.2 Hz, 0.4H), 4.32 (dd, *J* = 4.8, 1.2 Hz, 0.6H), 4.04 (dd, *J* = 9.6, 4.2 Hz, 0.4H), 3.96–3.94 (m, 0.6H), 3.83–3.81 (m, 0.4H), 3.76–3.69 (m, 2H), 3.62 (t, *J* = 10.2 Hz, 0.6H), 3.54–3.51 (m, 1H), 3.43 (s, 1.2H), 3.42 (s, 1.8H), 2.09 (s, 1.2H), 2.05 (s, 1.8H); ¹³C NMR (151 MHz, D₂O) δ 175.87, 174.94, 93.33, 93.24, 75.10, 72.14, 71.01, 70.90, 70.76 68.90, 67.06, 66.73, 58.69, 58.61, 54.22, 53.36, 22.22, 22.08.

2-Acetamido-2,6-dideoxy-D-mannopyranoside (6-Deoxy-ManNAc, 4a)



A stirred solution of **20** (0.72 g, 1.51 mmol) in anhydrous pyridine (10 mL) was treated with a solution of *p*-toluenesulfonyl chloride (0.87 g, 4.53 mmol) in pyridine (2 mL) at 0 °C and the mixture was stirred for overnight at room temperature. The reaction mixture was stopped by adding MeOH (2 mL) and concentrated. Purification of the residue by column chromatography on silica gel (Hexanes:EtOAc = 8.5:1.5) afforded **25** (0.89 g, 93%). ¹H NMR (600 MHz, CDCl₃) δ 7.81 (dd, *J* = 8.4, 1.8 Hz, 2H), 7.37–7.20 (m, 17H), 4.85 (dd, *J* = 10.8, 1.8 Hz, 1H), 4.77 (s, 1H), 4.74–4.67 (m, 2H), 4.59 (dd, *J* = 12.0, 1.2 Hz, 1H), 4.50 (dd, *J* = 10.8, 1.2 Hz, 1H), 4.41 (dd, *J* = 11.4, 1.2 Hz), 4.21 (s, 2H), 4.09 (m, 1H), 3.92 (d, *J* = 1.2 Hz, 1H), 3.81–3.79 (m, 1H), 3.76–3.73 (m, 1H), 2.42 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 144.88, 137.75, 137.60, 136.55, 133.03, 129.85, 128.61, 128.21, 97.22, 79.83, 75.36, 73.76, 72.67, 70.15, 69.49, 68.81, 61.15, 21.74.

To a solution of **25** (0.89 g, 1.41 mmol) in Et₂O (50 mL), LiAlH₄ (0.23 g, 6.06 mmol) was slowly added and the mixture was refluxed for 50 min at 50 °C. The resulting mixture was cooled in an ice bath, diluted with toluene (35 mL) and followed by gradual addition of EtOAc (2 mL). After removing the solvents under reduced pressure, column chromatography on silica gel (Hexanes:EtOAc = 3:7) afforded **26** (0.57 g, 96%). ¹H NMR (600 MHz, CDCl₃) δ 7.38–7.29 (m, 15H), 4.91 (d, *J* = 10.8 Hz, 1H), 4.81 (s, 1H), 4.70–4.63 (m, 4H), 4.47 (d, *J* = 12.0 Hz, 1H), 3.92 (dd, *J* = 6.0, 1.8 Hz, 1H), 3.80 (m, 1H), 3.42 (t, *J* = 9.6 Hz, 1H), 3.40 (d, *J* = 3.6 Hz, 1H), 1.52 (s, 2H), 1.32 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 138.59, 138.53, 137.55, 128.57, 128.54, 128.53, 128.16, 128.08, 127.91, 127.85, 100.11, 80.37, 80.11, 75.44, 71.82, 69.16, 67.81, 52.33, 18.25.

Compound **26** (0.57 g, 1.31 mmol) was dissolved in pyridine (4 mL) and thioacetic acid (8 mL) and stirred for overnight at room temperature. The solvent was removed under diminished pressure and the crude product was purified using silica gel column (Hexanes:EtOAc = 7:3) to afford compound **27** (0.52 g, 84%). ¹H NMR (600 MHz, CDCl₃) δ 7.38–7.34 (m, 15H), 5.86 (d, *J* = 12.0 Hz, 1H), 4.98 (d, *J* = 12.0 Hz, 1H), 4.93 (s, 1H), 4.78–4.75 (m, 2H), 4.66 (dd, *J* = 24.0, 12.0 Hz, 2H), 4.51 (dd, *J* = 12.0, 6.0 Hz, 2H), 4.13 (dd, *J* = 4.0, 1.8 Hz, 1H), 3.89–3.84 (m, 1H), 3.24 (t, *J* = 6.0 Hz, 1H), 2.07 (s, 3H), 1.35 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 170.49, 138.32, 137.94, 137.03, 128.44, 128.30, 127.96, 127.92, 127.81, 98.39, 80.39, 77.59, 75.45, 71.26, 69.41, 67.11, 49.69, 23.54, 18.16.

Compound **27** (0.52 g, 1.09 mmol) was dissolved in AcOH (4 mL) and MeOH (20 mL), and Pd(OH)₂/C (50 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by flash column chromatography (EtOAc:MeOH:H₂O = 10:1:0.1) to afford **4a** (0.18 g, 82%). ¹H NMR (600 MHz, D₂O) δ 5.07 (s,

0.6H), 5.00 (s, 0.4H), 4.45 (d, J = 3.6 Hz, 0.4H), 4.31 (d, J = 4.8 Hz, 0.6H), 4.01 (dd, J = 9.6, 4.8 Hz, 0.5H), 3.93–3.91 (m, 0.5H), 3.78 (dd, J = 10.2, 4.8 Hz, 0.5H), 3.46–3.38 (m, 1H), 3.29 (t, J = 9.6 Hz, 0.5H), 2.11 (s, 1.4H), 2.07 (s, 1.6H), 1.31 (d, J = 6.6 Hz, 1.4H), 1.29 (d, J = 6.0 Hz, 1.6H); ¹³C NMR (151 MHz, D₂O) δ 178.45, 177.53, 95.63, 95.52, 75.24, 75.20, 74.86, 74.43, 71.12, 70.81, 56.94, 56.22, 24.92, 24.73, 19.52, 19.50.





Triphenylphosphine (2.05 g, 7.80 mmol) and water (1.23 mL, 0.07 mmol) were added to a stirred solution of **19** (1.23 g, 2.60 mmol) in THF (40 mL) and the mixture was heated at 45 °C for 3 h. The solvent was evaporated. Purification of the residue by a silica gel column (Hexanes:EtOAc = 9:1) afforded product **28** (1.12 g, 97%). ¹H NMR (600 MHz, CDCl₃) δ 7.49 (d, *J* = 7.8 Hz, 2H), 7.36–7.24 (m, 13H), 5.59 (s, 1H), 4.82 (s, 1H), 4.79 (d, *J* = 11.4 Hz, 1H), 4.66 (t, *J* = 12.0 Hz, 2H), 4.46 (d, *J* = 12.0 Hz, 1H), 4.22 (dd, *J* = 10.2, 4.8 Hz, 1H), 4.05 (t, *J* = 9.0 Hz, 1H), 3.97 (dd, *J* = 10.2, 4.8 Hz, 1H), 3.43 (d, *J* = 3.6 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 138.51, 137.59, 137.03, 128.84, 128.47, 128.33, 128.17, 128.02, 127.94, 127.58, 126.05, 101.56, 100.99, 78.90, 75.75, 72.74, 69.23, 68.90, 63.86, 53.72.

Compound **28** (1.12 g, 2.50 mmol) was dissolved in CH₂Cl₂ (20 mL) and Et₃N (0.72 mL, 5.20 mmol) was added drop wisely at 0 °C. Benzyloxyacetyl chloride (0.49 mL, 3.12 mmol) was added to the reaction mixture and stirred at 0 °C for 2 h and then room temperature for 2 h. The residue was concentrated and purified by silica gel flash column chromatography (Hexanes:EtOAc = 7:3) to give **29** (1.43 g, 96%). ¹H NMR (600 MHz, CDCl₃) δ 7.53–7.52 (d, *J* = 7.8 Hz, 2H), 7.42–7.26 (m, 18H), 6.99 (d, *J* = 7.8 Hz, 1H), 5.54 (s, 1H), 5.04 (s, 1H), 4.76–4.65 (m, 4H), 4.63–4.54 (m, 4H), 4.24–3.93 (m, 4H), 3.76–3.69 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 170.25, 138.13, 137.43, 137.00, 136.83, 129.09, 128.75, 128.65, 128.43, 128.34, 128.31, 128.19, 128.14, 127.89, 127.81, 127.73, 126.20, 101.79, 99.28, 79.01, 73.58, 73.44, 71.82, 69.86, 69.63, 68.91, 63.48, 50.46.

Compound **29** (0.37 g, 0.62 mmol) was dissolved in anhydrous CH_2Cl_2 (7 mL) containing 4 Å molecular sieves (1.5 g). After the mixture was stirred for 1 h at room temperature, it was cooled to -78 °C and Et₃SiH (0.30 mL, 1.85 mmol) and PhBCl₂ (0.27 mL, 2.10 mmol) were added successively to the stirred solution. After being stirred at -78 °C for 1.5 h, Et₃N (1 mL) and MeOH (1 mL) were added successively and the mixture was filtered over Celite and concentrated. The residue was diluted with CH_2Cl_2 , washed with an aqueous solution of NaHCO₃, dried over MgSO₄, and concentrated. The crude product was purified by silica gel flash column chromatography (Hexanes:EtOAc = 1:1) to give **30**

(0.31 g, 85%). ¹H NMR (600 MHz, CDCl₃) δ 7.39–7.30 (m, 20H), 6.95 (d, J = 9.0 Hz, 1H), 5.01 (s, 1H), 4.92 (d, J = 10.8 Hz, 1H), 4.80 (d, J = 10.8 Hz, 1H), 4.76 (dd, J = 9.0, 4.2 Hz, 1H), 4.69 (d, J = 12.0 Hz, 1H), 4.62–4.60 (m, 2H), 4.56–4.53 (m, 3H), 4.20 (dd, J = 9.0, 4.0 Hz, 1H), 4.05 (dd, J = 24.6, 15.6 Hz, 2H), 3.81–3.74 (m, 3H), 3.62 (t, J = 9.6 Hz, 1H), 1.89 (s, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 170.27, 138.37, 138.20, 137.13, 137.05, 128.88, 128.75, 128.74, 128.66, 128.64, 128.41, 128.39, 128.28, 128.12, 127.99, 127.88, 98.59, 77.89, 75.47, 74.14, 73.63, 71.66, 71.46, 69.85, 69.73, 62.13, 49.21.

To a solution of **30** (0.31 g, 0.52 mmol) and Et₃N (0.14 mL, 1.04 mmol) in anhydrous CH₂Cl₂ (12 mL) in a Teflon flask, DAST (0.27 mL, 2.08 mmol) was slowly added at -40 °C. The reaction was stirred for 3 days at room temperature. After cooled to -20 °C, MeOH (1 mL) was added and the solvent was removed under reduced pressure. The residue was diluted with CH₂Cl₂, washed with water for 3 times, dried over MgSO₄, and concentrated. The crude product was purified using silica gel flash column chromatography (Hexanes:EtOAc = 7:3) to yield **31** (0.14 g, 45%). ¹H NMR (600 MHz, CDCl₃) δ 7.37–7.26 (m, 20 H), 6.91 (d, *J* = 8.4 Hz, 1H), 4.99 (s, 1H), 4.89 (d, *J* = 10.8 Hz, 1H), 4.78 (d, *J* = 11.4 Hz, 1H), 4.76–4.74 (m, 1H), 4.67–4.50 (m, 8H), 4.16 (dd, *J* = 9.6, 4.2 Hz, 1H), 4.03 (d, *J* = 4.8 Hz, 2H), 3.80 (dd, *J* = 28.2, 10.2 Hz, 1H), 3.60 (t, *J* = 9.6 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 170.12, 138.11, 138.02, 137.00, 136.87, 128.75, 128.64, 128.59, 128.53, 128.33, 128.18, 128.16, 128.03, 127.89, 127.73, 98.68, 82.03, (d, *J* = 173.5 Hz), 77.76, 75.52, 73.51, 73.28 (d, *J* = 6.8 Hz), 71.34, 70.59 (d, *J* = 18.6 Hz), 69.93, 69.59, 48.90; ¹⁹F NMR (282 MHz, CDCl₃) δ -234.76 (ddd, *J* = 48.8, 48.8, 30.4 Hz, 1F).

Compound **31** (0.14 g, 0.23 mmol) was dissolved in AcOH (4 mL) and MeOH (20 mL), and Pd(OH)₂/C (25 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by silica gel column chromatography (EtOAc:MeOH:H₂O = 10:1:0.1) to afford **7a** (47 mg, 85%). ¹H NMR (600 MHz, D₂O) δ 5.16 (s, 0.6H), 5.09 (s, 0.4H), 4.73–4.59 (m, 2H), 4.48 (dd, *J* = 4.2, 1.8 Hz, 0.3H), 4.34 (dd, *J* = 4.8, 1.8 Hz, 0.7H), 4.16 (s, 1H), 4.12 (s, 2H), 3.99 (dd, *J* = 28.8, 15.6 Hz, 0.6H), 3.87 (dd, *J* = 7.2, 4.2 Hz, 0.4H), 3.68 (t, *J* = 10.2 Hz, 0.5H), 3.57 (t, *J* = 9.6 Hz, 0.5H); ¹³C NMR (151 MHz, D₂O) δ 176.17, 175.25, 93.14 (2C), 82.87 (d, *J* = 28.0 Hz), 81.75 (d, *J* = 29.7 Hz), 75.10 (d, *J* = 8.3 Hz), 71.88, 70.99 (d, *J* = 18.6 Hz), 68.65, 65.93, 65.70, 61.04 (d, *J* = 16.9 Hz), 53.85, 53.00; ¹⁹F NMR (282 MHz, D₂O) δ -234.78 (ddd, *J* = 48.2, 48.2, 30.2 Hz, 0.4F), -235.50 (ddd, *J* = 46.8, 46.8, 28.2 Hz, 0.6F).

2-Deoxy-2-glycolamido-6-O-methyl-D-mannopyranoside (6-O-Me-ManNGc, 8a)



Triphenylphosphine (0.42 g, 1.59 mmol) and water (0.25 mL, 0.01 mmol) were added to a stirred solution of **23** (0.26 g, 0.53 mmol) in THF (10 mL) and the mixture was heated at 45 °C for 3 h. The solvent was removed and the crude product was dried under vacuum. The dried compound was

dissolved in CH₂Cl₂ (10 mL) and Et₃N (0.15 mL, 1.06 mmol) was added dropwise at 0 °C. Then, acetoxyacetyl chloride (0.07 mL, 0.64 mmol) was added to the reaction mixture and stirred at 0 °C for 2 h and then room temperature for 2 h. The residue was concentrated and purified by flash silica gel column chromatography (Hexanes:EtOAc = 1:1) to give **32** (0.28 g, 96%). ¹H NMR (600 MHz, CDCl₃) δ 7.37–7.29 (m, 15H), 6.43 (d, *J* = 9.0 Hz, 1H), 4.97 (s, 1H), 4.91 (d, *J* = 10.8 Hz, 1H), 4.77 (d, *J* = 10.8 Hz, 1H), 4.74 (dd, *J* = 9.0, 6.0 Hz, 1H), 4.68 (d, *J* = 12.0 Hz, 1H), 4.60 (dd, *J* = 24.6, 15.6 Hz, 3H), 4.51 (dd, *J* = 10.8, 6.6 Hz, 2H), 4.15 (dd, *J* = 9.6, 4.8 Hz, 1H), 3.79 (d, *J* = 9.6 Hz, 1H), 3.70 (t, *J* = 9.6 Hz, 1H), 3.66 (dd, *J* = 10.2 Hz, 1H), 3.55 (dd, *J* = 10.8, 1.8 Hz, 1H), 3.37 (s, 3H), 2.14 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 169.37, 167.26, 138.32, 137.99, 136.86, 128.57, 128.48, 128.34, 128.18, 128.09, 127.89, 127.84, 98.42, 77.69, 75.27, 73.62, 71.34, 70.97, 70.77, 69.69, 63.09, 59.38, 49.04, 20.67.

Compound **32** (0.28 g, 0.49 mmol) was dissolved in AcOH (5 mL) and MeOH (25 mL), and Pd(OH)₂/C (30 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite, washed with MeOH, dried under reduced pressure. The concentrated product was dissolved in aqueous solution of LiOH (0.5 M, 20 mL) and stirred at room temperature for overnight. The solvent was removed and the crude product was purified by silica gel column chromatography (EtOAc:MeOH:H₂O = 10:1:0.2) to afford **8a** (103 mg, 83%). ¹H NMR (600 MHz, D₂O) δ 5.12 (s, 0.7H), 5.05 (s, 0.3H), 4.47 (d, *J* = 4.2 Hz, 0.3H), 4.35 (d, *J* = 4.8 Hz, 0.7H), 4.16 (s, 0.6H), 4.13 (s, 1.4H), 4.07–3.46 (m, 5H), 3.40 (s, 1H), 3.39 (s, 2H); ¹³C NMR (151 MHz, D₂O) δ 175.32, 175.27, 93.35, 93.24, 75.45, 72.30, 71.38, 71.05, 69.12, 67.43, 67.18, 61.37, 58.99, 54.13, 53.25.

2,6-Dideoxy-2-glycolamido-D-mannopyranoside (6-Deoxy-ManNGc, 9a)



To a solution of **26** (0.25 g, 0.57 mmol) in CH₂Cl₂ (15 mL) was added Et₃N (0.16 mL, 1.15 mmol) at 0 °C. Benzyloxyacetyl chloride (0.11 mL, 0.68 mmol) was added to the reaction mixture and stirred at 0 °C for 2 h and then room temperature for 2 h. The residue was concentrated and purified by silica gel flash column chromatography (Hexanes:EtOAc = 1:1) to give **33** (0.24 g, 74%). ¹H NMR (600 MHz, CDCl₃) δ 7.38–7.27 (m, 20H), 6.90 (d, *J* = 9.0 Hz, 1H), 4.90 (d, *J* = 7.8 Hz, 0.7H), 4.88 (s, 0.3H), 4.77 (d, *J* = 10.8 Hz, 1H), 4.73 (dd, *J* = 9.0, 4.2 Hz, 1H), 4.67 (d, *J* = 12.0 Hz, 1H), 4.58 (dd, *J* = 25.8, 13.8 Hz, 3H), 4.53–4.49 (m, 3H), 4.11 (dd, *J* = 9.0, 4.2 Hz, 1H), 4.04 (dd, *J* = 27.6, 15.0 Hz, 2H), 3.85–3.81 (m, 1H), 3.13 (t, *J* = 9.6 Hz, 1H), 1.28 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 170.00, 138.38, 138.19, 137.20, 137.06, 128.72, 128.56, 128.48, 128.30, 128.23, 128.16, 128.07, 128.02, 127.87, 127.79, 127.72, 98.27, 80.42, 77.75, 75.52, 73.47, 71.28, 69.66, 69.49, 67.31, 49.17, 18.23.

Compound **33** (0.24 g, 0.41 mmol) was dissolved in AcOH (5 mL) and MeOH (25 mL), and Pd(OH)₂/C (30 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous

shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by silica gel column chromatography (EtOAc:MeOH:H₂O = 10:1:0.1) to afford **9a** (75 mg, 83%). ¹H NMR (600 MHz, D₂O) δ 5.08 (s, 0.5H), 5.03 (s, 0.5H), 4.46 (d, *J* = 4.8 Hz, 0.5H), 4.32 (d, *J* = 4.8 Hz, 0.5H), 4.16 (s, 1H), 4.13 (s, 1H), 4.02 (dd, *J* = 10.2, 4.8 Hz, 0.5H), 3.95–3.90 (m, 0.5H), 3.79 (dd, *J* = 10.2, 4.2 Hz, 0.5H), 3.47–3.44 (m, 0.5H), 3.35 (t, *J* = 9.6 Hz, 0.5H), 3.25 (t, *J* = 9.6 Hz, 0.5H), 1.30 (d, *J* = 6.0 Hz, 1.5H), 1.27 (d, *J* = 6.0 Hz, 1.5H); ¹³C NMR (151 MHz, D₂O) δ 176.17, 175.28, 92.92, 92.72, 72.60, 72.32, 71.80, 71.74, 68.47, 68.14, 61.09, 61.02, 54.05, 53.21, 53.17, 16.85, 16.80.

6-Azido-2,6-dideoxy-2-glycolamido-D-mannopyranoside (6-N₃-ManNGc, 10a)



ManNAc **1a** (1.0 g, 4.52 mmol) was dissolved in an aqueous solution of HCl (2 M, 40 mL) and stirred at 60 °C for overnight. The solvent was removed and the residue was dried under vacuum. The resulting compound was dissolved in a mixture of CH₃CN (70 mL) and water (70 mL) and followed by addition of NaHCO₃ (3.80 g, 45.20 mmol) at 0 °C. Acetoxyacetyl chloride (0.97 mL, 9.04 mmol) was added to the reaction mixture and stirred at 0 °C for 2 h and then room temperature for 2 h. MeOH was added to the mixture and the solvents were removed. The residue was purified by silica gel flash column chromatography (EtOAc:MeOH:H₂O = 8:1:0.2) to give **34** (1.20 g, 95%). ¹H NMR (600 MHz, D₂O) δ 5.11 (d, *J* = 1.2 Hz, 0.6H), 5.02 (d, *J* = 1.2 Hz, 0.4H), 4.71 (d, *J* = 1.2 Hz, 0.7H), 4.66 (m, 2.3H), 4.47 (dd, *J* = 4.8, 1.8 Hz, 0.4H), 4.33 (dd, *J* = 4.2, 1.2 Hz, 0.6H), 4.04 (dd, *J* = 9.6, 4.8 Hz, 0.6H), 3.85–3.80 (m, 2.4H), 3.59 (t, *J* = 9.6 Hz, 0.6H), 3.48 (t, *J* = 10.2 Hz, 0.4H), 2.16 (s, 1H), 2.15 (s, 2H); ¹³C NMR (151 MHz, D₂O) δ 173.62, 173.51, 172.46, 171.47, 170.69, 93.02, 92.97, 76.49, 72.12, 68.93, 66.83, 66.58, 62.73, 61.40, 60.46, 54.26, 53.31, 20.08.

A stirred solution of **34** (1.20 g, 4.30 mmol) in anhydrous pyridine (8 mL) was treated with a solution of *p*-toluenesulfonyl chloride (0.90 g, 4.73 mmol) in pyridine (3 mL) at 0 °C and the mixture was stirred for 3 h. The reaction mixture was stopped by adding MeOH (2 mL) and concentrated. Purification of the residue by column chromatography (EtOAc:MeOH = 9:1) afforded **35** (1.21 g, 65%). A mixture containing compound **35** (1.21 g, 2.79 mmol), NaN₃ (1.09 g, 16.70 mmol), and tetra*n*-butylammonium iodide (0.20 g, 0.55 mmol) in anhydrous DMF (15 mL) was stirred at 60 °C for overnight. The mixture was concentrated and purified by silica gel flash column chromatography (EtOAc:MeOH = 9.5:0.5) to afford product **36** (0.63 g, 75%). ¹H NMR (600 MHz, D₂O) δ 5.21 (d, *J* = 2.4 Hz, 0.5H), 5.16 (d, *J* = 6.6 Hz, 0.5H), 4.72 (d, *J* = 1.8 Hz, 1.5H), 4.69 (s, 1.5H), 4.49–3.51 (m, 5H), 2.22 (s, 1.5H), 2.20 (s, 1.5H); ¹³C NMR (151 MHz, D₂O) δ 173.56, 170.81, 170.52, 170.17, 93.07, 91.02, 70.62, 70.13, 69.98, 69.42, 68.67, 67.84, 67.11, 64.38, 62.83, 61.48, 53.97, 53.40, 51.10, 20.15, 20.07.

Compound **36** (0.63 g, 2.07 mmol) was dissolved in dry MeOH (15 mL) containing a catalytic amount of NaOMe. The mixture was stirred at room temperature for overnight and neutralized using Dowex (H⁺) resin. The resulted suspension was filtrated and concentrated. Silica gel column chromatography (EtOAc:MeOH:H₂O = 8:1:0.3) of the crude product afforded product **10a** (0.49 g, 90%). ¹H NMR (600 MHz, D₂O) δ 5.20 (d, *J* = 3.6 Hz, 0.7H), 5.14 (s, 0.3H), 4.47 (d, *J* = 4.2 Hz, 0.5H), 4.35 (d, *J* = 4.8 Hz, 0.5H), 4.17 (s, 0.4H), 4.13–4.12 (m, 2.6H), 4.09–3.45 (m, 4H); ¹³C NMR (151 MHz, D₂O) δ 176.00, 175.28, 94.90, 93.08, 76.65, 76.15, 73.87, 72.27, 71.80, 70.96, 70.19, 69.01, 67.03, 61.10, 60.78, 56.58, 53.81, 53.07.

6-Deoxy-6-fluoro-D-mannopyranoside (6-F-Man, 12a)



To a solution of **37**⁴ (0.36 g, 0.60 mmol) in anhydrous CH₂Cl₂ (10 mL) in a Teflon flask, DAST (0.60 mL, 4.54 mmol) was slowly added at -40 °C. The reaction was stirred for 3 days at room temperature. After cooled down to -20 °C, MeOH (2 mL) was added and the solvent was removed under reduced pressure. The residue was diluted with CH₂Cl₂, washed with water for 3 times, dried over MgSO₄, and concentrated. The crude product was purified using silica gel flash column chromatography (Hexanes:EtOAc = 4:1) to yield **38** (0.21 g, 60%). ¹H NMR (300 MHz, CDCl₃) δ 8.23–7.27 (m, 20 H), 6.63 (d, *J* = 2.1 Hz, 1H), 6.14–6.02 (m, 2H), 5.88 (dd, *J* = 2.7, 2.1 Hz, 1H), 4.72 (t, *J* = 2.1 Hz, 1H), 4.56 (t, *J* = 2.4 Hz, 1H), 4.49–4.34 (m, 1H); ¹³C NMR (75.5 MHz, CDCl₃) δ 165.87, 165.50, 165.48, 164.01, 134.32, 133.99, 133.89, 133.67, 130.39, 130.27, 130.14, 130.05, 129.99, 129.83, 128.93, 128.76, 128.64, 91.49, 73.12, 72.12, 70.04, 69.53, 65.81; ¹⁹F NMR (282 MHz, CDCl₃) δ -233.40 (ddd, *J* = 49.5, 49.5, 25.5 Hz, 1F).

To a suspension of **38** (0.21 g, 0.35 mmol) in dry MeOH (10 mL) was added NaOMe (40 mg) and the mixture was stirred overnight at room temperature. The reaction was neutralized with Dowex (H⁺) resin, filtrated, concentrated and purified by silica gel flash column chromatography (EtOAc:MeOH:H₂O = 10:1:0.3) to afford **12a** (62 mg, 98%). ¹H NMR (600 MHz, D₂O) δ 5.18 (d, *J* = 1.8 Hz, 0.6H), 4.92 (s, 0.4H), 4.76–4.59 (m, 2H), 3.94–3.93 (m, 1H), 3.85 (dd, *J* = 9.6, 3.0 Hz, 1H), 3.76 (t, *J* = 10.2 Hz, 1H), 3.67–3.66 (m, 1H); ¹³C NMR (151 MHz, D₂O) δ 94.37, 93.9, 83.05 (d, *J* = 18.6 Hz), 81.92 (d, *J* = 18.6 Hz), 74.74 (d, *J* = 17.7 Hz), 72.99, 71.27 (d, *J* = 17.7 Hz), 70.67, 70.21, 65.72 (d, *J* = 7.2 Hz), 65.47 (d, *J* = 6.8 Hz); ¹⁹F NMR (282 MHz, D₂O) δ -234.16 (ddd, *J* = 49.5, 49.5, 27.3 Hz, 0.6F), -235.03 (ddd, *J* = 49.5, 49.5, 29.3 Hz, 0.4F).

6-O-Methyl-D-mannopyranoside (6-O-Me-Man, 13a)



A solution of **39**² (2.23 g, 4.12 mmol) in anhydrous DMF (20 mL) was stirred with NaH (0.39 g, 16.48 mmol) at room temperature for 30 min. Methyl iodide (1.02 mL, 16.48 mmol) was added slowly at 0 °C and the mixture was stirred for 6 h. The mixture was diluted with CH₂Cl₂, washed with water, dried over MgSO₄, and concentrated. Purification of the residue by silica gel column chromatography (Hexanes:EtOAc = 10:0.5) gave **40** (2.26 g, 99%). ¹H NMR (600 MHz, CDCl₃) δ 7.39–7.30 (m, 20H), 5.00 (s, 1H), 4.97 (d, *J* = 10.8 Hz, 1H), 4.75–4.73 (m, 3H), 4.66–4.64 (m, 3H), 4.48 (d, *J* = 12.0 Hz, 1H), 4.05–3.98 (m, 2H), 3.85–3.84 (m, 2H), 3.71 (dd, *J* = 10.2, 4.8 Hz, 1H), 3.65 (dd, *J* = 10.8, 1.8 Hz, 1H), 3.44 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 138.82, 138.51, 137.60, 128.65, 128.59, 128.57, 128.30, 128.18, 128.06, 128.00, 127.93, 127.88, 127.77, 97.60, 80.44, 75.50, 75.22, 74.78, 72.83, 72.41, 71.97 (2C), 69.23, 59.54.

Compound **40** (2.26 g, 4.07 mmol) was dissolved in AcOH (8 mL) and MeOH (40 mL), and Pd(OH)₂/C (400 mg) was added. The resulted mixture was hydrogenated for 2 days with vigorous shaking under H₂ atmosphere (80 mbar), and then the catalyst was removed by filtration over Celite and washed with MeOH. The concentrated product was purified by silica gel column chromatography (EtOAc:MeOH:H₂O = 10:1:0.1) to afford **13a** (0.63 g, 80%). ¹H NMR (600 MHz, D₂O) δ 5.13 (s, 0.7H), 4.86 (d, *J* = 1.8 Hz, 0.3H), 3.91–3.71 (m, 3H), 3.66–3.45 (m, 3H), 3.39 (s, 1H) 3.38 (s, 2H); ¹³C NMR (151 MHz, D₂O) δ 94.12, 93.78, 74.70, 73.10, 71.60, 71.29, 70.90, 70.75, 70.27, 67.11, 66.80, 58.62, 58.53.

6-Deoxy-D-mannopyranoside (6-Deoxy-Man, 14a)



To a solution of **41**⁵ (1.44 g, 6.15 mmol) in CH₂Cl₂ (15 mL) was added Et₃N (1.04 mL, 7.40 mmol) and *p*-toluenesulfonyl chloride (1.46 g, 7.40 mmol). The reaction was stirred at room temperature for overnight. The mixture was concentrated under reduced pressure and the crude product was purified using silica gel flash column chromatography (Hexanes:EtOAc = 10:1) to give **42** (2.10 g, 88%). ¹H NMR (400 MHz, CDCl₃) δ 7.80 (d, *J* = 8.4 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 2H), 4.83 (s, 1H), 4.29 (d, *J* = 3.2 Hz, 2H), 4.09 (d, *J* = 3.2 Hz, 2H), 3.75–3.70 (m, 1H), 3.62–3.59 (m, 1H), 3.34 (s, 3H), 2.75 (d, *J* = 5.2 Hz, 1H), 2.44 (s, 3H), 1.47 (s, 3H), 1.33 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.14, 133.08, 130.06, 128.19, 109.99, 98.56, 78.13, 75.56, 69.35, 68.77, 68.49, 55.41, 28.02, 26.18, 21.86.

Sodium borohydride (0.96 g, 25.46 mmol) was added slowly to a stirred solution of 42 (1.97 g, 5.07 mmol) in DMSO (20 mL). The mixture was stirred at 80 °C for 2 h. The reaction solution was

poured into cold water (70 mL), extracted with Et₂O, filtrated, and dried over MgSO₄. Solvent was removed under reduced pressure and the crude product was purified using column chromatography on silica gel (Hexanes:EtOAc = 3:7) to afford **43** (1.03 g, 93%). ¹H NMR (400 MHz, CDCl₃) δ 4.84 (s, 1H), 4.13–4.03 (m, 2H), 3.66–3.61 (m, 1H), 3.38 (s, 3H), 2.60 (d, *J* = 5.6 Hz, 1H), 1.52 (s, 3H), 1.35 (m, 3H), 1.30 (d, *J* = 8.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 109.74, 98.40, 78.80, 75.85, 74.66, 65.97, 55.18, 28.21, 26.35, 17.71.

To a solution of compound **43** (0.96 g, 4.40 mmol) in water (50 mL) was added Dowex (H⁺) resin (5.2 g). The mixture was stirred vigorously and refluxed for 16 h. The suspension was filtered and the solvent was removed under diminished pressure. Silica gel column chromatography purification (EtOAc:MeOH:H₂O = 10:0.8:0.1) gave compound **14a** (0.68 g, 94%). ¹H NMR (600 MHz, D₂O) δ 5.06 (s, 0.6H), 4.82 (s, 0.4H), 3.88–3.79 (m, 1.6H), 3.74 (dd, *J* = 9.6, 3.0 Hz, 0.4H), 3.54 (dd, *J* = 9.0, 3.0 Hz, 0.4H), 3.40–3.29 (m, 1.6H), 1.24 (d, *J* = 5.4 Hz, 1H), 1.22 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (151 MHz, D₂O) δ 94.11, 93.62, 72.86, 72.30, 72.14, 71.94, 71.45, 70.93, 70.06, 68.39, 16.93, 16.88.

Enzymatic synthesis of sialosides 1b-15b and 1c-15c

General procedure for one-pot three-enzyme preparative synthesis of $\alpha 2$ –3- (1b–15b) and $\alpha 2$ –6-linked (1c–15c) sialosides using Gal βp NP as the sialyltransferase acceptor. Gal βp NP (1.0 eq, 10 mM, 0.1 mmol), a sialic acid precursor (1.2–1.5 eq, 12 mM, 0.12 mmol), sodium pyruvate (6.0 eq, 60 mM, 0.6 mmol), and CTP (2.0 eq, 20 mM, 0.2 mmol) were dissolved in water in a 50 mL centrifuge tube containing Tris-HCl buffer (100 mM, pH 8.5) and MgCl₂ (20 mM). After addition of appropriate amount of an Pm aldolase, an *N. meningitides* CMP-sialic acid synthetase, and a sialyltransferase (PmST1 or Pd2,6ST), water was added to bring the volume of the reaction mixture to 10 mL. The reaction was incubated in an isotherm incubator with agitating in 120 rpm for 1–16 h at 37 °C or for 12–48 h at room temperature. The product formation was monitored by TLC using EtOAc:MeOH:H₂O:AcOH = 7:2:0.5:0.1 (by volume) as the developing solvent and stained with *p*-anisaldehyde sugar stain solution. The reaction was terminated by adding the same volume of ice-cold EtOH and incubating at 4 °C for 30 min. The mixture was centrifuged to remove precipitates. The supernatant was concentrated and passed through a Bio-gel P-2 gel filtration column with water to obtain the desired product. Silica gel column purification (EtOAc:MeOH:H₂O = 7:2:0.5) was used for further purification.

4-Nitrophenyl *O*-(5-acetamido-9-fluoro-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-β-D-galactopyranoside (Neu5Ac9Fα2–3GalβpNP, 2b). Yield, 98%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.26 (d, *J* = 9.0 Hz, 2H), 7.24 (d, *J* = 9.6 Hz, 2H), 5.28 (d, *J* = 7.8 Hz, 1H), 4.71–4.56 (m, 2H), 4.22 (dd, *J* = 10.2, 3.0 Hz, 1H), 4.05–3.97 (m, 2H), 3.91–3.85 (m, 3H), 3.74 (d, *J* = 6.0 Hz, 2H), 3.71–3.64 (m, 3H), 2.77 (dd, *J* = 12.6, 4.8 Hz, 1H), 2.01 (s, 3H), 1.82 (t, *J* = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.10, 173.97, 161.84, 142.66, 126.24, 116.56, 100.05, 99.80, 84.47 (d, *J* = 165.6 Hz), 75.61, 72.84 (d, *J* = 11.0 Hz), 70.58, 70.45, 68.93, 68.55, 67.39, 67.00, 60.81, 51.79, 39.85, 22.22; ¹⁹F NMR (282 MHz, D₂O) δ -234.76 (ddd, *J* = 46.5, 46.5, 27.9 Hz, 1F); HRMS (ESI) calculated for $C_{23}H_{31}FN_2O_{15}Na$ (M+H) 617.1606, found 617.1579.

4-Nitrophenyl *O*-(5-acetamido-3,5-dideoxy-9-*O*-methyl-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-β-D-galactopyranoside (Neu5Ac9OMeα2–3Galβ*p*NP, 3b). Yield, 94%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.25 (d, *J* = 9.0 Hz, 2H), 7.24 (d, *J* = 8.4 Hz, 2H), 5.28 (d, *J* = 8.4 Hz, 1H), 4.22 (dd, *J* = 9.6, 2.4 Hz, 1H), 4.03 (d, *J* = 3.0 Hz, 1H), 3.98–3.96 (m, 1H), 3.92–3.83 (m, 3H), 3.75 (d, *J* = 6.0 Hz, 2H), 3.70–3.62 (m, 3H), 3.57 (dd, *J* = 9.0, 1.8 Hz, 1H), 3.52 (dd, *J* = 10.2, 6.6 Hz, 1H), 3.34 (s, 3H), 2.78 (dd, *J* = 12.0, 4.8 Hz, 1H), 2.01 (s, 3H), 1.81 (t, *J* = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.16, 173.98, 161.92, 142.68, 126.30, 116.62, 99.92, 99.88, 75.66 (2C), 73.40, 72.97, 70.42, 68.97, 68.58, 68.32, 67.47, 60.87, 58.61, 51.83, 39.88, 22.25; HRMS (ESI) calculated for C₂₄H₃₄N₂O₁₆Na (M+H) 629.1806, found 629.1781.

4-Nitrophenyl *O*-(5-acetamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Neu5Ac9Deoxyα2–3GalβpNP, 4b). Yield, 90%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.22 (d, J = 7.8 Hz, 2H), 7.20 (d, J = 8.4 Hz, 2H), 5.24 (d, J = 7.8 Hz, 1H), 4.18 (dd, J = 9.6, 1.8 Hz, 1H), 3.99 (d, J = 2.4 Hz, 1H), 3.92–3.79 (m, 4H), 3.71 (d, J = 6.0 Hz, 2H), 3.65–3.61 (m, 2H), 3.33–3.29 (m, 1H), 2.73 (dd, J = 12.0, 4.2 Hz, 1H), 1.98 (s, 3H), 1.77 (t, J = 12.0 Hz, 1H), 1.19 (d, J = 6.0 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 177.63, 176.61, 164.42, 145.20, 128.82, 119.14, 102.61, 102.41, 78.18, 78.16, 75.77, 75.48, 71.48, 71.16, 70.27, 69.99, 63.40, 54.49, 42.34, 24.78, 21.14; HRMS (ESI) calculated for C₂₃H₃₂N₂O₁₅Na (M+H) 599.1700, found 599.1691.

4-Nitrophenyl *O*-(5-acetamido-9-azido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-β-D-galactopyranoside (Neu5Ac9N₃α2–3GalβpNP, 5b). Yield, 87%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.21 (d, *J* = 9.0 Hz, 2H), 7.21 (d, *J* = 9.0 Hz, 2H), 5.26 (d, *J* = 7.8 Hz, 1H), 4.22 (dd, *J* = 9.6, 3.0 Hz, 1H), 4.03–3.99 (m, 2H), 3.91–3.83 (m, 2H), 3.75 (d, *J* = 6.0 Hz, 2H), 3.71 (s, 1H), 3.69–3.42 (m, 5H), 2.77 (dd, *J* = 12.0, 4.2 Hz, 1H), 2.02 (s, 3H), 1.81 (t, *J* = 12.6 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.17, 173.98, 161.88, 142.64, 126.25, 116.59, 100.01, 99.88, 75.61, 72.88, 70.54, 68.92, 68.50, 67.40, 62.64, 60.83, 59.48, 53.18, 51.84, 39.89, 22.21; HRMS (ESI) calculated for C₂₃H₃₁N₅O₁₅Na (M+H) 640.1714, found 640.1684.

4-Nitrophenyl *O*-(9-fluoro-5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Neu5Gc9Fα2–3GalβpNP, 7b). Yield, 98%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.26 (d, J = 9.6 Hz, 2H), 7.24 (d, J = 9.0 Hz, 2H), 5.28 (d, J = 7.8 Hz, 1H), 4.72–4.56 (m, 2H), 4.23 (dd, J = 10.2, 3.0 Hz, 1H), 4.10 (s, 2H), 4.03 (d, J = 3.0 Hz, 1H), 3.96 (t, J = 10.2 Hz, 1H), 3.92–3.88 (m, 2H), 3.81–3.70 (m, 6H), 2.80 (dd, J = 12.6, 4.8 Hz, 1H), 1.84 (t, J = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.89, 174.01, 161.86, 142.64, 126.24, 116.57, 100.06, 99.81, 84.45 (d, J = 170.8 Hz), 75.66, 75.59, 72.57, 70.58 (d, J = 14.6 Hz), 68.94, 68.28, 67.37, 66.93, 61.14, 60.80, 51.48, 39.91; ¹⁹F NMR (282 MHz, D₂O) δ -234.83 (ddd, J = 45.7, 45.7, 27.6 Hz, 1F); HRMS (ESI) calculated for C₂₃H₃₁FN₂O₁₆Na (M+H) 633.1555, found 633.1525.

4-Nitrophenyl O-(5-glycolylamido-3,5-dideoxy-9-O-methyl-D-glycero- α -D-galacto-2nonulopyranosylonic acid)-(2 \rightarrow 3)-O- β -D-galactopyranoside (Neu5Gc9OMe α 2–3Gal βp NP, 8b). Yield, 40%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.26 (d, *J* = 9.6 Hz, 2H), 7.24 (d, *J* = 9.6 Hz, 2H), 5.28 (d, *J* = 7.8 Hz, 1H), 4.23 (dd, *J* = 10.2, 3.0 Hz, 1H), 4.10 (s, 2H), 4.03 (d, *J* = 3.6 Hz, 1H), 3.99–3.88 (m, 4H), 3.80–3.74 (m, 4H), 3.67 (dd, *J* = 10.8, 1.8 Hz, 1H), 3.57 (dd, *J* = 9.6, 1.8 Hz, 1H), 3.52 (dd, *J* = 10.8, 6.6 Hz, 1H), 3.34 (s, 3H), 2.79 (dd, *J* = 12.6, 4.2 Hz, 1H), 1.83 (t, *J* = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.89, 174.00, 161.87, 142.67, 126.24, 116.57, 100.00, 99.85, 75.67, 75.63, 73.34, 72.66, 70.43, 68.93, 68.28, 68.22, 67.41, 61.13, 60.83, 58.53, 51.48, 39.90; HRMS (ESI) calculated for C₂₄H₃₄N₂O₁₇Na (M+H) 645.1755, found 645.1727.

4-Nitrophenyl *O*-(5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Neu5Gc9Deoxyα2–3GalβpNP, 9b). Yield, 90%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.29 (d, *J* = 9.0 Hz, 2H), 7.28 (d, *J* = 9.0 Hz, 2H), 5.31 (d, *J* = 7.8 Hz, 1H), 4.26 (dd, *J* = 9.6, 3.0 Hz, 1H), 4.14 (s, 2H), 4.07 (d, *J* = 3.0 Hz, 1H), 3.98–3.91 (m, 3H), 3.82–3.78 (m, 4H), 3.75 (s, 1H), 3.39 (dd, *J* = 9.0, 1.8 Hz, 1H), 2.82 (dd, *J* = 12.0, 4.8 Hz, 1H), 1.86 (t, *J* = 12.6 Hz, 1H), 1.26 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 175.85, 174.10, 161.88, 142.67, 126.27, 116.59, 100.08, 99.83, 75.60 (2C), 73.15, 72.66, 68.90, 68.33, 67.76, 67.42, 61.10, 60.84, 51.63, 39.84, 18.53; HRMS (ESI) calculated for C₂₃H₃₂N₂O₁₆Na (M+H) 615.1650, found 615.1624.

4-Nitrophenyl *O*-(9-azido-5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-β-D-galactopyranoside (Neu5Gc9N₃α2–3Galβ*p*NP, 10b). Yield, 85%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.22 (d, *J* = 9.0 Hz, 2H), 7.22 (d, *J* = 9.6 Hz, 2H), 5.28 (d, *J* = 7.8 Hz, 1H), 4.25 (dd, *J* = 10.2, 3.0 Hz, 1H), 4.11 (s, 2H), 4.04 (d, *J* = 3.0 Hz, 1H), 3.96–3.75 (m, 9H), 3.62–3.57 (m, 2H), 2.80 (dd, *J* = 12.6, 4.8 Hz, 1H), 1.84 (t, *J* = 12.0 Hz, 1H); ¹³CNMR (151 MHz, D₂O) δ 176.04, 174.07, 162.02, 142.78, 126.37, 116.75, 100.18, 100.02, 75.73, 72.91, 72.10, 69.08, 68.34 (2C), 67.58, 62.86, 61.30, 60.98, 59.65, 51.69, 40.05; HRMS (ESI) calculated for C₂₃H₂₉N₅O₁₆Na (M-H) 631.1609, found 631.1574.

4-Nitrophenyl *O*-(9-fluoro-3,9-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Kdn9Fα2–3Galβ*p*NP, 12b). Yield, 94%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.22 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.4 Hz, 2H), 5.24 (d, *J* = 7.8 Hz, 1H), 4.73– 4.57 (m, 2H), 4.17 (dd, *J* = 9.6, 3.0 Hz, 1H), 4.02–3.94 (m, 3H), 3.89–3.84 (m, 2H), 3.72 (d, *J* = 6.0 Hz, 2H), 3.62–3.51 (m, 3H), 2.70 (dd, *J* = 12.6, 4.8 Hz, 1H), 1.75 (t, *J* = 12.0, 1H); ¹³C NMR (151 MHz, D₂O) δ 176.90, 164.80, 145.61, 129.13, 119.64, 103.05, 102.95, 87.81 (d, *J* = 164.6 Hz), 78.57, 76.84, 73.65 (d, *J* = 16.6 Hz), 73.31, 72.84, 71.87, 70.37, 69.80, 69.77 (d, *J* = 15.1 Hz), 63.80, 42.43; ¹⁹F NMR (282 MHz, D₂O) δ -234.43 (ddd, *J* = 47.9, 47.9, 25.4 Hz, 1F); (ESI) calculated for C₂₁H₂₈NO₁₅FNa (M+H) 576.1341, found 576.1372.

4-Nitrophenyl *O*-(9-*O*-methyl-3-deoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Kdn9OMeα2–3GalβpNP, 13b). Yield, 78%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.22 (d, J = 8.4 Hz, 2H), 7.20 (d, J = 8.4 Hz, 2H), 5.24 (d, J = 7.2 Hz, 1H), 4.16 (dd, J = 9.0, 3.0 Hz, 1H), 3.98 (d, J = 3.0 Hz, 1H), 4.00 (t, J = 7.2, 1H), 3.88–3.48 (m, 10H), 3.32 (s, 3H), 2.68 (dd, J = 12.0, 4.2 Hz, 1H), 1.72 (t, J = 12.6 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 174.16, 161.90, 142.71, 126.29, 116.63, 100.02, 99.93, 75.67 (2C), 74.01, 73.57, 70.60, 70.45, 69.90, 68.96, 68.03, 67.40, 60.86, 58.60, 39.54; HRMS (ESI) calculated for $C_{22}H_{31}NO_{16}Na$ (M+H) 588.1541, found 588.1533.

4-Nitrophenyl *O*-(3,9-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Kdn9Deoxyα2–3GalβpNP, 14b). Yield, 94%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.22 (d, J = 9.0 Hz, 2H), 7.20 (d, J = 9.6 Hz, 2H), 5.23 (d, J = 7.8 Hz, 1H), 4.15 (dd, J = 9.6, 2.4 Hz, 1H), 3.98 (d, J = 3.0 Hz, 1H), 3.91–3.82 (m, 3H), 3.70 (d, J = 6.0 Hz, 2H), 3.59–3.47 (m, 4H), 2.68 (dd, J = 12.6, 4.8 Hz, 1H), 1.72 (t, J = 12.6 Hz, 1H), 1.22 (d, J = 6.0 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 174.19, 161.83, 142.65, 126.23, 116.52, 99.99, 99.77, 75.54 (2C), 73.91, 72.69, 70.40, 69.93, 68.87, 67.94, 67.29, 60.76, 39.37, 18.46; HRMS (ESI) calculated for C₂₁H₂₉NO₁₅Na (M+H) 558.1435, found 558.1429.

4-Nitrophenyl *O*-(9-azido-3,9-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-β-D-galactopyranoside (Kdn9N₃α2–3Galβ*p*NP, 15b). Yield, 91%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.28 (d, *J* = 9.0 Hz, 2H), 7.28 (d, *J* = 9.6 Hz, 2H), 5.32 (d, *J* = 7.8 Hz, 1H), 4.27 (dd, *J* = 10.2, 3.0 Hz, 1H), 4.10–4.05 (m, 2H), 3.98–3.59 (m, 9H), 3.54 (dd, *J* = 13.2, 6.6 Hz, 1H), 2.78 (dd, *J* = 12.0, 4.2 Hz, 1H), 1.84 (t, *J* = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 174.31, 161.92, 142.74, 126.33, 116.69, 100.22, 100.13, 75.74, 74.02, 70.92, 70.44, 69.98, 69.01, 68.64, 67.44, 62.73, 60.89, 53.36, 39.58. HRMS (ESI) calculated for C₂₁H₂₈N₄O₁₅Na (M+H) 599.1449, found 599.1476.

4-Nitrophenyl *O*-(5-acetamido-9-fluoro-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Ac9Fα2–6Galβ*p*NP, 2c). Yield, 95%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.27 (d, *J* = 9.0 Hz, 2H), 7.25 (d, *J* = 9.0 Hz, 2H), 5.17 (d, *J* = 7.2 Hz, 1H), 4.71–4.56 (m, 2H), 4.00–3.92 (m, 4H), 3.83 (t, *J* = 7.8 Hz, 1H), 3.78–3.64 (m, 6H), 2.74 (dd, *J* = 12.6, 3.6 Hz, 1H), 1.99 (s, 3H), 1.64 (t, *J* = 12.0 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 175.11, 173.67, 162.04, 142.69, 126.29, 116.57, 100.40, 100.00, 84.52 (d, *J* = 166.8 Hz), 74.18, 72.48 (2C), 70.42 (2C), 68.57, 68.38, 67.10, 63.20, 51.94, 40.42, 22.17; ¹⁹F NMR (282 MHz, D₂O) δ -234.70 (ddd, *J* = 46.5, 46.5, 27.9 Hz, 1F); HRMS (ESI) calculated for C₂₃H₃₁FN₂O₁₅Na (M+H) 617.1606 , found 617.1571.

4-Nitrophenyl *O*-(5-acetamido-3,5-dideoxy-9-*O*-methyl-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Ac9OMeα2–6Galβ*p*NP, 3c). Yield, 92%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.27 (d, J = 9.0 Hz, 2H), 7.25 (d, J = 9.6 Hz, 2H), 5.17 (d, J = 7.2 Hz, 1H), 4.01–3.99 (m, 2H), 3.95–3.91 (m, 2H), 3.84 (dd, J = 9.6, 7.8 Hz, 1H), 3.77 (dd, J = 10.2, 3.6 Hz, 1H), 3.72 (dd, J = 15.6, 5.4 Hz, 1H), 3.69–3.63 (m, 4H), 3.52–3.49 (m, 2H), 3.35 (s, 3H), 2.74 (dd, J = 12.0, 4.2 Hz, 1H), 1.99 (s, 3H), 1.63 (t, J = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.13, 173.70, 162.04, 142.67, 126.28, 116.56, 100.39, 100.00, 74.16, 73.37, 72.62, 72.49, 70.42, 70.31, 68.53 (2C), 68.36, 63.14, 58.60, 51.95, 40.40, 22.15; HRMS (ESI) calculated for C₂₄H₃₄N₂O₁₆ (M+H) 629.1806, found 629.1773.

4-Nitrophenyl *O*-(5-acetamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Ac9Deoxyα2–6Galβ*p*NP, 4c). Yield, 97%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.25 (d, J = 9.0 Hz, 2H), 7.22 (d, J = 9.6 Hz, 2H), 5.14 (d, J = 7.8 Hz, 1H), 3.98–3.96 (m, 2H), 3.89 (dd, J = 15.0, 6.6 Hz, 2H), 3.81 (dd, J = 10.2, 7.8 Hz, 1H), 3.74 (dd,

J = 10.2, 3.6 Hz, 1H), 3.72–3.68 (m, 1H), 3.66–3.59 (m, 3H), 3.27 (dd, J = 9.0, 1.8 Hz, 1H), 2.71 (dd, J = 12.6, 4.8 Hz, 1H), 1.96 (s, 3H), 1.60 (t, J = 12.6 Hz, 1H), 1.19 (d, J = 6.6 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 174.96, 173.67, 161.95, 142.67, 126.25, 116.51, 100.34, 99.92, 74.14, 73.27, 72.60, 72.45, 70.39, 68.51, 68.41, 67.61, 63.13, 52.03, 40.36, 22.11, 18.52; HRMS (ESI) calculated for C₂₃H₃₂N₂O₁₅Na (M+H) 599.1700, found 599.1692.

4-Nitrophenyl *O*-(5-acetamido-9-azido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Ac9N₃α2–6GalβpNP, 5c). Yield, 81%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.28 (d, J = 9.6 Hz, 2H), 7.26 (d, J = 9.6 Hz, 2H), 5.17 (d, J = 7.8 Hz, 1H), 4.01–3.91 (m, 4H), 3.84 (t, J = 7.8 Hz, 1H), 3.79–3.62 (m, 6H), 3.52 (d, J = 9.0 Hz, 1H), 3.43 (dd, J = 13.2, 6.6 Hz, 1H), 2.73 (dd, J = 12.6, 4.2 Hz, 1H), 2.00 (s, 3H), 1.63 (t, J = 12.6 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.12, 173.67, 162.01, 142.67, 126.27, 116.56, 100.45, 100.01, 74.17, 72.54, 72.48, 70.47, 70.41, 68.97, 68.54, 68.31, 63.18, 53.21, 51.98, 40.38, 22.14; HRMS (ESI) calculated for C₂₃H₃₁N₅O₁₅Na (M+H) 640.1714, found 640.1682.

4-Nitrophenyl *O*-(9-fluoro-5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Gc9Fα2–6Galβ*p*NP, 7c). Yield, 95%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.28 (d, J = 9.6 Hz, 2H), 7.25 (d, J = 9.0 Hz, 2H), 5.18 (d, J = 7.2 Hz, 1H), 4.71–4.55 (m, 2H), 4.08 (s, 2H), 4.01–3.93 (m, 4H), 3.86–3.75 (m, 5H), 3.68–3.64 (m, 2H), 2.76 (dd, J = 12.6, 4.2 Hz, 1H), 1.66 (t, J = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.89, 173.73, 162.05, 142.71, 126.30, 116.58, 100.46, 100.04, 84.52 (d, J = 167.3 Hz), 74.21, 72.51, 72.28, 70.57, 70.45, 68.59, 68.12, 67.06, 63.24, 61.12, 51.67, 40.47; ¹⁹F NMR (282 MHz, D₂O) δ -234.80 (ddd, J = 48.8, 48.8, 27.3 Hz, 1F); HRMS (ESI) calculated for C₂₃H₃₁FN₂O₁₆Na (M+H) 633.1555, found 633.1522.

4-Nitrophenyl *O*-(5-glycolylamido-3,5-dideoxy-9-*O*-methyl-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Gc9OMeα2–6Galβ*p*NP, 8c). Yield, 30%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.27 (d, J = 8.4 Hz, 2H), 7.25 (d, J = 9.0 Hz, 2H), 5.17 (d, J = 7.2 Hz, 1H), 4.08 (s, 2H), 4.01 (d, J = 2.4 Hz, 1H), 4.00 (d, J = 4.2 Hz, 1H), 3.93 (dd, J = 15.6, 7.2 Hz, 2H), 3.86–3.76 (m, 5H), 3.53–3.50 (m, 2H), 3.53–3.50 (m, 2H), 3.35 (s, 3H), 2.76 (dd, J = 12.0, 3.6 Hz, 1H), 1.65 (t, J = 12.0 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 175.89, 173.69, 162.05, 142.71, 126.31, 116.59, 100.42, 100.04, 74.20, 73.39, 72.52, 72.37, 70.46, 70.40, 68.57, 68.33, 68.12, 63.19, 61.12, 58.61, 51.67, 40.47; HRMS (ESI) calculated for C₂₄H₃₄N₂O₁₇Na (M+H) 645.1755, found 645.1720.

4-Nitrophenyl *O*-(5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Gc9Deoxyα2–6GalβpNP, 9c). Yield, 84%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.28 (d, J = 9.6 Hz, 2H), 7.26 (d, J = 9.0 Hz, 2H), 5.18 (d, J = 7.8 Hz, 1H), 4.08 (s, 2H), 4.01–3.99 (m, 2H), 3.95–3.91 (m, 2H), 3.85–3.74 (m, 5H), 3.66 (dd, J = 10.8, 4.2 Hz, 1H), 3.30 (dd, J = 8.4, 1.2 Hz, 1H), 2.75 (dd, J = 12.6, 4.8 Hz, 1H), 1.64 (t, J = 12.0 Hz, 1H), 1.22 (d, J = 6.0 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 175.85, 173.75, 162.05, 142.71, 126.31, 116.60, 100.44, 100.06, 74.20, 73.26, 72.52, 72.38, 70.46, 68.56, 68.19, 67.71, 63.18, 61.12, 51.80, 40.47, 18.56; HRMS (ESI) calculated for C₂₃H₃₂N₂O₁₆Na (M+H) 615.1650, found 615.1616.

4-Nitrophenyl *O*-(9-azido-5-glycolylamido-3,5,9-trideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Neu5Gc9N₃α2–6Galβ*p*NP, 10c). Yield, 82%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.30 (d, J = 9.0 Hz, 2H), 7.28 (d, J = 9.0 Hz, 2H), 5.21 (d, J = 7.8 Hz, 1H), 4.12 (s, 2H), 4.06–4.03 (m, 2H), 3.99 (t, J = 8.4 Hz, 1H), 3.83-3.72 (m, 7H), 3.63 (dd, J = 10.2, 3.6 Hz, 1H), 3.56 (dd, J = 12.0, 6.6 Hz, 1H), 3.49 (d, J = 9.0 Hz, 1H), 2.80 (dd, J = 12.0, 3.6 Hz, 1H), 1.69 (t, J = 12.0 Hz, 1H); ¹³CNMR (151 MHz, D₂O) δ 175.96, 173.66, 161.98, 142.62, 126.25, 116.53, 100.37, 100.01, 74.14, 72.46, 71.94, 70.42, 68.53, 68.24, 68.03, 63.16, 62.71, 61.08, 59.57, 51.68, 40.42; HRMS (ESI) calculated for C₂₃H₂₉N₅O₁₆ (M-Na-H) 631.1609, found 631.1565.

4-Nitrophenyl *O*-(3,9-dideoxy-9-fluoro-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Kdn9Fα2–6Galβ*p*NP, 12c). Yield, 98%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.25 (d, *J* = 9.0 Hz, 2H), 7.23 (d, *J* = 9.6 Hz, 2H), 5.15 (d, *J* = 7.8 Hz, 1H), 4.73– 4.56 (m, 2H), 4.02–3.88 (m, 5H), 3.81 (t, *J* = 8.4 Hz, 1H), 3.75 (dd, *J* = 9.6, 3.0 Hz, 1H), 3.61–3.55 (m, 3H), 3.41 (t, *J* = 9.6 Hz, 1H), 2.67 (dd, *J* = 12.6, 4.8 Hz, 1H), 1.57 (t, *J* = 12.6 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 173.85, 161.99, 142.63, 126.25, 116.50, 100.35, 99.98, 84.55 (d, *J* = 166.8 Hz), 74.17, 73.50, 72.45, 70.62, 70.38, 70.24, 69.97, 68.55, 66.75, 66.70, 63.22, 40.02; ¹⁹F NMR (282 MHz, D₂O) δ -234.42 (ddd, *J* = 47.9, 47.9, 28.2 Hz, 1F); HRMS (ESI) calculated for C₂₁H₂₈NO₁₅FNa (M+H) 576.1341, found 576.1330.

4-Nitrophenyl *O*-(9-*O*-methyl-3-deoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Kdn9OMeα2–6GalβpNP, 13c). Yield, 62%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.24 (d, *J* = 7.2 Hz, 2H), 7.21 (d, *J* = 7.8 Hz, 2H), 5.13 (d, *J* = 7.2 Hz, 1H), 3.96– 3.95 (m, 2H), 3.89 (t, *J* = 8.4 Hz, 2H), 3.80 (t, *J* = 8.4 Hz, 1H), 3.74–3.67 (m, 3H), 3.60–3.50 (m, 4H), 3.83 (t, *J* = 9.0 Hz, 1H), 3.34 (s, 3H), 2.65 (dd, *J* = 12.0, 3.6 Hz, 1H), 1.55 (t, *J* = 12.0 Hz, 1H). ¹³C NMR (151 MHz, D₂O) δ 174.01, 162.36, 143.06, 126.56, 117.05, 100.82, 100.46, 74.55, 73.97, 73.93, 72.91, 70.98, 70.82, 70.66, 70.40, 68.92, 68.60, 63.48, 58.90, 40.28; HRMS (ESI) calculated for $C_{22}H_{31}NO_{16}Na$ (M+H) 588.1541, found 588.1531.

4-Nitrophenyl *O*-(3,9-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Kdn9Deoxyα2–6GalβpNP, 14c). Yield, 70%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.25 (d, *J* = 9.6 Hz, 2H), 7.22 (d, *J* = 9.6 Hz, 2H), 5.14 (d, *J* = 7.2 Hz, 1H), 3.97–3.95 (m, 2H), 3.89 (t, *J* = 8.4 Hz, 2H), 3.82–3.79 (m, 1H), 3.74 (dd, *J* = 10.2, 3.6 Hz, 1H), 3.60–3.51 (m, 4H), 3.39 (t, *J* = 9.6 Hz, 1H), 2.66 (dd, *J* = 12.6, 4.8 Hz, 1H), 1.55 (t, *J* = 12.6 Hz, 1H), 1.22 (d, *J* = 6.0 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 173.90, 161.99, 142.69, 126.26, 116.52, 100.35, 99.99, 74.17, 73.59, 72.80, 72.46, 70.40, 70.31, 70.11, 68.50, 67.92, 63.18, 40.05, 18.54; HRMS (ESI) calculated for $C_{21}H_{29}NO_{15}Na$ (M+H) 558.1435, found 558.1424.

4-Nitrophenyl *O*-(9-azido-3,9-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→6)-*O*-β-D-galactopyranoside (Kdn9N₃α2–6Galβ*p*NP, 15c). Yield, 59%; white solid. ¹H NMR (600 MHz, D₂O) δ 8.28 (d, J = 9.0 Hz, 2H), 7.25 (d, J = 9.0 Hz, 2H), 5.17 (d, J = 8.4 Hz, 1H) 4.00– 3.90 (m, 4H), 3.85–3.76 (m, 3H), 3.68–3.61 (m, 4H), 3.46 (dd, J = 13.2, 6.6 Hz, 1H), 3.41 (t, J = 9.6 Hz, 1H), 2.69 (dd, J = 12.0, 4.2 Hz, 1H), 1.59 (t, J = 12.0, 1H); ¹³C NMR (151 MHz, D₂O) δ 171.69, 159.89, 140.57, 124.14, 114.44, 98.34, 97.69, 72.07, 71.42, 70.37, 68.70, 68.29, 68.11, 67.91, 66.53, 66.43, 61.10, 51.19, 37.87; HRMS (ESI) calculated for $C_{21}H_{28}N_4O_{15}Na$ (M+H) 599.1449, found 599.1442.

Sialidase substrate specificity assays

Materials: *Clostridium perfringens* sialidase (type VI), *Vibrio cholerae* sialidase (type III), and β -galactosidase from *Aspergillus oryzae* were purchased from Sigma. *Salmonella typhimurium* sialidase and *Streptococcus pneumoniae* sialidase were bought from Prozyme. All of these enzymes were used without further purification. PmST1 was expressed in *E. coli* and purified as described previously.⁶ Sodium pyruvate (Fisher Biotech), *N*-acetylmannosamine (ManNAc, Sigma), mannose (Acros Organics), *para*-nitrophenyl β -D-galactopyranoside (Gal β pNP, Sigma), and cytidine 5'-triphosphate disodium salt (CTP, Sigma) were from commercially available sources. 384-Well plates for sialidase assays were from Fisher Biotech.

All sialidase assays were carried out at 37 °C in duplicate for 30 min in 384-well plates in a final volume of 20 μ L containing a sialoside substrate (0.3 mM) and the β-galactosidase (12 μ g, 126 mU). The amount of the β-galactosidase required to completely hydrolyze Galβ*p*NP (0.3 mM) within the time frame of the assay was predetermined. The assay conditions for various sialidases were as follows: *S. typhimurium* sialidase (1.5 mU) in sodium acetate buffer (100 mM, pH 5.5) containing NaCl (100 mM); *C. perfringens* sialidase (2 mU) in MES buffer (100 mM, pH 5.0); *V. cholerae* sialidase (1 mU) in sodium acetate buffer (100 mM, pH 5.5) containing NaCl (100 mM); *S. pneumoniae* sialidase (0.2 mU) in sodium acetate buffer (100 mM, pH 6.0); PmST1 (7.5 μ g) in sodium acetate buffer (100 mM, pH 5.5); NEU2 (6 μ g) in MES buffer (100 mM, pH 5.0). The reactions were stopped by adding 40 μ L of CAPS buffer (*N*-cyclohexyl-3-aminopropane sulfonic acid, 0.5 M, pH 10.5). The amount of *para*-nitrophenolate formed was determined by measuring the A_{405nm} of the reaction mixtures using a microplate reader. Reactions of Galβ*p*NP and an excess amount of β-galactosidase were used as controls.

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