Rhodium-catalyzed 1,3-Acyloxy Migration and Subsequent Intramolecular [4+2] Cycloaddition of Vinylallene and Unactivated Alkyne

Suyu Huang,^{*a,b*} Xiaoxun Li,^{*b*} Claire L. Lin,^{*b*} Ilia A. Guzei,^{*c*} and Weiping Tang*^{*b*}

^a Department of Chemistry and Key Laboratory for Chemical Biology of Fujian Province, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen, Fujian, 361005 PR China.

^b School of Pharmacy, University of Wisconsin, Madison, WI, 53705 USA. E-mail: wtang@pharmacy.wisc.edu

^c Department of Chemistry, University of Wisconsin, Madison, WI,53705 USA.

General remarks

All reactions in non-aqueous media were conducted under a positive pressure of dry argon in glassware that had been oven dried prior to use unless noted otherwise. Anhydrous solutions of reaction mixtures were transferred via an oven dried syringe or cannula. All solvents were dried prior to use unless noted otherwise. Reagents were purchased from Aldrich, Acros, TCI, or VWR unless otherwise noted. Thin layer chromatography was performed using precoated silica gel plates (EMD Chemical Inc. 60, F254). Flash column chromatography was performed with silica gel (Sillicycle, 40-63µm). Infrared spectra (IR) were obtained as neat oils on a Bruker Equinox 55 Spectrophotometer. ¹H and ¹³C Nuclear magnetic resonance spectra (NMR) were obtained on a Varian Unity-Inova 400 MHz or 500 MHz recorded in ppm (δ) downfield of TMS ($\delta = 0$) in CDCl₃, CD₃OD. Signal splitting patterns were described as singlet (s), doublet (d), triplet (t), quartet (q), quintet (quint), or multiplet (m), with coupling constants (*J*) in hertz. High resolution mass spectra (HRMS) were performed by Analytical Instrument Center at the School of Pharmacy or Department of Chemistry on an Electron Spray Injection (ESI) mass spectrometer.



General strategies for the preparation of substrates:

Representative experimental procedures: Preparation of substrates **4a**, **4b**, **4k**, **4l**, **4m**, and **4n**:



To a stirred suspension of NaH (670 mg, 16.7 mmol, 55-65% in mineral oil) in anhydrous THF (10 mL) was added commercially available alcohol **S2** (2.50 g, 30.4 mmol) in THF (15 mL) at 0°C. The reaction mixture was allowed to warm to room temperature and stirred for 30 min. The solution of bromide **S1** (2.00 g, 15.2 mmol) in THF (10 mL) was added to the above reaction. The reaction mixture was heated under reflux for overnight. It was then quenched with water (30 mL) and extracted with Et_2O (3 × 20 mL). The combined organic layers were washed with brine, dried over anhydrous NaSO₄, filtered and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 20) to give the enyne **S3** (1.90 g, 95%) as a yellow oil.

To a stirred solution of enyne **S3** (465 mg, 3.47 mmol) in anhydrous THF (17 mL) was added n-BuLi in hexane (1.6 M, 2.39 mL, 3.82 mmol) at -78 °C. After stirring for 30 min at -78 °C, acetone (302 mg, 5.20 mmol) was added to the reaction mixture dropwise. After the addition was completed, the reaction was allowed to warm to room temperature and stirred for 30 min. After the reaction mixture was cooled to 0 °C, acetic anhydride (390 mg, 3.82 mmol) was added. When the reaction was completed as determined by TLC, the reaction mixture was quenched by the addition of saturated aqueous ammonium chloride (15 mL) and extracted with ethyl ether (3 × 20 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 10) to yield **4a** (380 mg, 70% over 2 steps) as a colorless oil.

Substrates 4b, 4k, 4l, 4m, and 4n was obtained following procedures similar to 4a.

Preparation of **S5** and **S6**:



To a stirred suspension of imidazole (5.00g, 73.2 mmol) in anhydrous CH_2Cl_2 (100 mL) was added commercially available alcohol **S2** (3.00 g, 36.6 mmol) in CH_2Cl_2 (10 mL) at 0°C. TBSCl (6.62 g, 43.9 mmol) in CH_2Cl_2 (10 mL) was then added to the above solution. The reaction mixture was allowed to warm to room temperature and stirred overnight. After the reaction was completed, the reaction mixture was quenched with water (50 mL) and extracted with CH_2Cl_2 (3 × 50 mL). The combined organic layers were washed with brine, dried over anhydrous NaSO₄, filtered and concentrated under vacuum to give crude TBS-protected alcohol for the next step without purification.

Product S4 was obtained from crude TBS-protected alcohol according to procedures described for 4a. The yield was 88% over 3 steps.

To a stirred solution of **S4** (4.00 g, 13.5 mmol) in anhydrous THF (120 mL) was added TBAF in THF (1.0 M, 14.8 mL, 14.8 mmol) at 0 °C. The reaction mixture was stirred for 10 min at 0°C. It was then quenched with saturated aqueous ammonium chloride (60 mL) and was extracted with Et_2O (3 × 50 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 2) to yield **S5** (2.36 g, 97%) as a colorless oil.

To a stirred solution of alcohol **S5** (529 mg, 2.9 mmol) in anhydrous CH_2Cl_2 (20 mL) was added CBr_4 (1.25 g, 3.8 mmol) and PPh₃ (1.14 g, 4.4 mmol) at 0°C. The reaction mixture was stirred for 15 min at 0°C and then concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 10) to yield **S6** (650 mg, 92%) as a colorless oil. Preparation of **S10**:



To a stirred solution of vinyl iodide S8^{1,2} (2.60 g, 13.1 mmol) and propargyl ester S9 (1.58 g,

12.5 mmol) in anhydrous Et₃N (15 mL) was added (Ph₃P)₂PdCl₂ (175 mg, 0.25 mmol) and CuI (95 mg, 0.50 mmol) at rt. The reaction mixture was stirred overnight. It was then quenched with saturated aqueous ammonium chloride (20 mL) and extracted with EtOAc (3×20 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 2) to yield **S10** (2.00 g, 83%) as a colorless oil.

Preparation of substrates 4c, 4d and 4g:



To a stirred solution of alcohol **S5** (958 mg, 5.26 mmol) and Ph₃P (1.38 g, 5.26 mmol) in anhydrous THF (25 mL) was added amide **S11**³ (1.00 g, 4.78 mmol) in THF (5 mL) at 0 °C and stirred for 10 min. To the above solution was added diisopropyl azodicarboxylate (1.11 mL, 5.74 mmol) dropwise at 0 °C. After the addition was completed, the reaction was allowed to warm to room temperature and stirred for 3 h. The reaction mixture was quenched by addition of saturated aqueous sodium bicarbonate (20 mL) and extracted with ethyl ether (3 × 20 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 5) to yield **4c** (1.70 g, 95%) as a colorless oil.

Following procedures similar to 4c, substrate 4d was prepared from $S12^4$ and substrate 4g was prepared from S11 and S10.

Preparation of substrate 4e and 4f:



To a stirred solution of propargyl ester 4c (208 mg, 0.56 mmol) and iodobenzene (147 mg, 0.72 mmol) in anhydrous Et₃N (6 mL) was added (Ph₃P)₂PdCl₂ (7.8 mg, 0.011 mmol) and CuI

(4.2 mg, 0.022 mmol) at rt. The reaction mixture was stirred overnight. It was then quenched with saturated aqueous ammonium chloride (10 mL) and extracted with EtOAc (3×10 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 2) to yield **4e** (230 g, 93%) as a yellow oil. Substrate **4f** was prepared from **4d** following similar procedure.

Preparation of **4h** and **4i**:



To a stirred suspension of NaH (37 mg, 0.925 mmol, 55-65% in mineral oil) in anhydrous THF (5 mL) was added $S13^5$ (169 mg, 0.918 mmol) in THF (5 mL) at 0°C. The reaction mixture was allowed to warm to room temperature and stirred for 30 min. Then the mixture was cooled to 0°C, the bromide S6 (152 mg, 0.623 mmol) in THF (10 mL) was added dropwise. The reaction mixture was stired 1 h at room temperature, then was quenched with saturated aqueous ammonium chloride (20 mL), and extracted with Et₂O (3 × 20 mL). The combined organic layers were washed with brine, dried over anhydrous NaSO₄, filtered and concentrated in vacuo. The residue was purified by flash column chromatography (Ethyl acetate/ Hexane = 1 : 5) to give substrate 4h (170 mg, 79%) as a colorless oil. Substrate 4i was prepared from S14⁶ following similar procedure.



To a stirred suspension of commercially available phosphonium bromide **S16** (1.47 g, 3.24 mmol) in anhydrous THF (30 mL) was added NaHMDS in THF (1.0 M, 3.24 mL, 3.24 mmol) at 0° C.⁸ The reaction mixture was allowed to warm to room temperature and stirred for 40 min. The mixture was cooled to -78 °C. A solution of aldehyde **S15**⁷ (493 mg, 2.90 mmol) in THF (5 mL)

was added to the above reaction mixture dropwise. After the addition was completed, the reaction was allowed to warm to room temperature and stirred for 12 h. The reaction mixture was poured into H₂O (30 mL) and extracted with Et₂O (3×30 mL). The combined organic layers were washed with brine, dried over anhydrous NaSO₄, filtered and concentrated under vacuum. The residue was purified by flash column chromatography (Hexane/Ethyl acetate = 99 : 1) to yield a TMS-protected enyne (390 mg, 51%) as a yellow oil.

To a stirred solution of TMS-protected enyne (390 mg, 1.47 mmol) in anhydrous THF (8 mL) was added TBAF in THF (1.0 M, 1.47 mL, 1.47 mmol) at 0 °C. The reaction mixture was stirred for 10 min at 0°C and was then quenched with H₂O (10 mL). The mixture was extracted with Et₂O (3 × 10 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to yield crude enyne **S17** for next step without purification.

Substrate **4j** was prepared from crude enyne **S17** according to procedures described for **4a**. The yield was 32% over 4 steps.

Characterization data for substrates:

4a

(E)-7-(but-2-ynyloxy)-2-methylhept-5-en-3-yn-2-yl acetate (4a)

¹H NMR (400MHz, CDCl₃) δ 1.67 (s, 6H), 1.85 (t, *J*= 2.4 Hz, 3H), 2.02 (s, 3H), 4.07 (dd, *J*= 1.6, 5.6 Hz, 2H), 4.09 (q, *J*= 2.4 Hz, 2H), 5.76 (td, *J*= 1.6, 16.0 Hz, 1H), 6.15 (td, *J*= 5.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 22.2, 29.2, 58.0, 69.2, 72.6, 75.0, 82.0, 83.0, 91.1, 112.0, 139.4, 169.5. IR (film): 2988, 2851, 2360, 1742, 1365, 1241, 1192, 1136, 1112, 1016, 957, 913, 832, 732. HRMS (ESI) *m/z* calcd. For C₁₄H₁₈O₃ (M+Na)⁺ 257.1148, found 257.1157.



(E)-7-(but-2-ynyloxy)-2-methylhept-5-en-3-yn-2-yl pivalate (4b)

¹H NMR (400MHz, CDCl₃) δ 1.16 (s, 9H), 1.64 (s, 6H), 1.84 (t, *J*= 2.4 Hz, 3H), 4.05 (dd, *J*= 1.6, 5.6 Hz, 2H), 4.08 (q, *J*= 2.4 Hz, 2H), 5.75 (td, *J*= 1.6, 16.0 Hz, 1H), 6.12 (td, *J*= 5.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 27.3, 29.1, 39.3, 58.1, 69.3, 72.1, 75.0, 81.7, 83.0, 91.4, 112.2, 139.2, 176.8. IR (film): 2980, 1733, 1479, 1362, 1285, 1171, 1124, 1029, 953, 846, 770, 733. HRMS (ESI) *m/z* calcd. For C₁₇H₂₄O₃ (M+Na)⁺ 299.1618, found 299.1616.



(E)-2-methyl-7-(4-methyl-N-(prop-2-ynyl)phenylsulfonamido)hept-5-en-3-yn-2-yl acetate (4c)

¹H NMR (400MHz, CDCl₃) δ 1.66 (s, 6H), 2.02 (s, 3H), 2.02 (t, *J*= 2.8 Hz, 1H), 2.43 (s, 3H), 3.85 (dd, *J*= 1.2, 6.4 Hz, 2H), 4.08 (d, *J*= 2.4 Hz, 2H), 5.74 (td, *J*= 1.6, 16.0 Hz, 1H), 5.99 (td, *J*= 6.8, 15.6 Hz, 1H), 7.30 (dd, *J*= 0.8, 8.4 Hz, 2H), 7.72 (td, *J*= 2.0, 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.8, 22.2, 29.1, 36.3 48.1, 72.4, 74.3, 76.4, 81.5, 91.8, 114.4, 128.0, 129.8, 136.0, 136.9, 143.9, 169.6. IR (film): 1740, 1600, 1435, 1350, 1244, 1193, 1162, 1137, 1093, 1052, 1018, 961, 897, 817, 738, 704, 660. HRMS (ESI) *m*/*z* calcd. For C₂₀H₂₃NO₄S (M+Na)⁺ 396.1240, found 396.1230.



(E)-7-(N-(but-2-ynyl)-4-methylphenylsulfonamido)-2-methylhept-5-en-3-yn-2-yl acetate (4d)

¹H NMR (400MHz, CDCl₃) δ 1.54 (t, *J*= 2.4 Hz, 3H), 1.66 (s, 6H), 2.02 (s, 3H), 2.42 (s, 3H), 3.81 (dd, *J*= 1.6, 6.4 Hz, 2H), 4.00 (q, *J*= 2.4 Hz, 2H), 5.72 (td, *J*= 1.6, 16.0 Hz, 1H), 5.99 (td, *J*= 6.4, 16.0 Hz, 1H), 7.30 (d, *J*= 8.0 Hz, 2H), 7.72 (td, *J*= 1.6, 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 3.4, 21.7, 22.2, 29.1, 36.9, 48.1, 71.6, 72.4, 81.6, 82.2, 91.6, 114.0, 128.1, 129.5, 136.2, 137.3, 143.6, 169.6. IR (film): 1740, 1598, 1434, 1348, 1241, 1160, 1136, 1091, 1047, 1016, 958, 905, 815, 731, 656. HRMS (ESI) *m*/*z* calcd. For C₂₁H₂₅NO₄S (M+Na)⁺ 410.1397, found 410.1394.



(E) - 2 - methyl - 7 - (4 - methyl - N - (3 - phenyl prop - 2 - ynyl) phenyl sulfon amido) hept - 5 - en - 3 - yn - 2 - yl acetate (4e)

¹H NMR (400MHz, CDCl₃) δ 1.67 (s, 6H), 2.02 (s, 3H), 2.34 (s, 3H), 3.91 (dd, *J*= 1.2, 6.4 Hz, 2H), 4.29 (s, 2H), 5.77 (td, *J*= 1.2, 16.0 Hz, 1H), 6.06 (td, *J*= 6.8, 15.6 Hz, 1H), 7.03-7.09 (m, 2H), 7.20-7.31 (m, 5H), 7.76 (d, *J*= 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.6, 22.2, 29.1, 37.3, 48.5, 72.4, 81.6, 81.6, 86.2, 91.8, 114.3, 122.3, 128.0, 128.3, 128.7, 129.8, 131.7, 136.0, 137.1, 143.9, 169.6. IR (film): 2360, 2341, 1739, 1598, 1491, 1442, 1349, 1265, 1246, 1162, 1136, 1049, 1017, 959, 900, 815, 735, 703, 659. HRMS (ESI) *m/z* calcd. For C₂₆H₂₇NO₄S (M+Na)⁺ 472.1553, found 472.1551.



(E)-7-(N-(3-(4-bromophenyl)prop-2-ynyl)-4-methylphenylsulfonamido)-2-methylhept-5-en-3-yn-2-yl acetate (4f)

¹H NMR (400MHz, CDCl₃) δ 1.66 (s, 6H), 2.02 (s, 3H), 2.35 (s, 3H), 3.90 (dd, *J*= 1.2, 6.4 Hz, 2H), 4.26 (s, 2H), 5.76 (td, *J*= 1.2, 16.0 Hz, 1H), 6.05 (td, *J*= 6.4, 16.0 Hz, 1H), 6.93 (td, *J*= 2.4, 8.8 Hz, 2H), 7.26 (d, *J*= 8.8 Hz, 2H), 7.38 (td, *J*= 2.0, 8.4 Hz, 2H), 7.74 (td, *J*= 2.0, 8.4 Hz, 2H);

¹³C NMR (100MHz, CDCl₃) δ 21.7, 22.2, 29.1, 37.2, 48.6, 72.4, 81.5, 83.0, 85.1, 91.9, 114.3, 121.2, 123.0, 128.0, 129.8, 131.6, 133.2, 136.1, 137.0, 143.9, 169.6. IR (film): 1738, 1485, 1348, 1243, 1161, 1135, 1011, 958, 898, 824, 733, 702, 660. HRMS (ESI) *m*/*z* calcd. For C₂₆H₂₆BrNO₄S (M+Na)⁺ 550.0658, found 550.0667.



4g

(E)-2,6-dimethyl-7-(4-methyl-N-(prop-2-ynyl)phenylsulfonamido)hept-5-en-3-yn-2-yl acetate (4g)

¹H NMR (400MHz, CDCl₃) δ 1.69 (s, 6H), 1.90 (s, 3H), 1.97 (t, *J*= 2.4 Hz, 1H), 2.02 (s, 3H), 2.43 (s, 3H), 3.77 (s, 2H), 4.02 (d, *J*= 2.4 Hz, 2H), 5.51-5.54 (m, 1H), 7.27-7.32 (m, 2H), 7.70-7.75 (m, 2H); ¹³C NMR (100MHz, CDCl₃) δ 17.2, 21.8, 22.2, 29.3, 36.0, 52.6, 72.6, 74.4, 76.2, 81.0, 95.0, 109.4, 128.0, 129.7, 136.1, 143.9, 145.2, 169.6. IR (film): 3296, 2987, 1738, 1598, 1432, 1349, 1268, 1243, 1160, 1136, 1112, 1092, 1014, 958, 899, 816, 736, 703, 660. HRMS (ESI) *m/z* calcd. For C₂₁H₂₅NO₄S (M+Na)⁺ 410.1397, found 410.1402.



(E)-dimethyl 2-(6-acetoxy-6-methylhept-2-en-4-ynyl)-2-(but-2-ynyl)malonate (4h)

¹H NMR (400MHz, CDCl₃) δ 1.66 (s, 6H), 1.75 (t, *J*= 2.4 Hz, 3H), 2.02 (s, 3H), 2.73 (q, *J*= 2.4 Hz, 2H), 2.83 (d, *J*= 7.6 Hz, 2H), 3.74 (s, 6H), 5.63 (d, *J*=16.0 Hz, 1H), 5.91 (td, *J*= 7.6, 15.6 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.7, 22.2, 23.4, 29.2, 36.0, 53.1, 57.3, 72.6, 73.2, 79.5, 82.2, 90.1, 114.0, 137.7, 169.6, 170.4. IR (film): 2955, 1736, 1437, 1377, 1265, 1244, 1209, 1136, 1099, 1066, 1017, 958, 736, 703. HRMS (ESI) *m*/*z* calcd. For C₁₉H₂₄O₆ (M+Na)⁺ 371.1465, found 371.1472.



(E)-dimethyl 2-(6-acetoxy-6-methylhept-2-en-4-ynyl)-2-(but-3-ynyl)malonate (4i) ¹H NMR (400MHz, CDCl₃) δ 1.66 (s, 6H), 1.97 (t, *J*= 2.0 Hz, 1H), 2.02 (s, 3H), 2.11-2.22 (m,

4H), 2.71 (dd, J= 1.2, 7.6 Hz, 2H), 3.74 (s, 6H), 5.58 (d, J=15.6 Hz, 1H), 5.91 (td, J= 7.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 14.2, 22.2, 29.2, 31.8, 36.9, 52.9, 57.1, 69.2, 72.6, 82.1, 83.1, 90.3, 113.8, 137.6, 169.6, 171.0. IR (film): 2954, 1731, 1435, 1366, 1241, 1197, 1135, 1079, 1016, 959, 834, 737, 702. HRMS (ESI) m/z calcd. For C₁₉H₂₄O₆ (M+Na)⁺ 371.1465, found 371.1480.



(E)-2-methyl-11-phenylundeca-5-en-3,10-diyn-2-yl acetate (4j)

¹H NMR (400MHz, CDCl₃) δ 1.64-1.73 (m, 2H), 1.67 (s, 6H), 2.01 (s, 3H), 2.27 (dt, *J*= 7.2, 7.2 Hz, 2H), 2.41 (t, *J*= 7.2 Hz, 2H), 5.56 (d, *J*= 15.6 Hz, 1H), 6.15 (td, *J*= 7.2, 16.0 Hz, 1H), 7.24-7.30 (m, 3H), 7.35-7.41 (m, 2H); ¹³C NMR (100MHz, CDCl₃) δ 19.0, 22.2, 27.8, 29.3, 32.2, 72.7, 81.4, 82.8, 89.2, 89.7, 110.1, 124.1, 127.8, 128.4, 131.8, 144.2, 169.6. IR (film): 1740, 1489, 1440, 1365, 1240, 1192, 1134, 1014, 956, 829, 756, 736, 692. HRMS (ESI) *m/z* calcd. For C₂₀H₂₂O₂ (M+Na)⁺ 317.1512, found 317.1515.



4k

(E)-1-(5-(but-2-ynyloxy)pent-3-en-1-ynyl)cyclohexyl acetate (4k)

¹H NMR (400MHz, CDCl₃) δ 1.26-1.40 (m, 1H), 1.46-1.56 (m, 1H), 1.56-1.66 (m, 4H), 1.79-1.89 (m, 2H), 1.85 (t, *J*= 2.4 Hz, 3H), 2.03 (s, 3H), 2.07-2.18 (m, 2H), 4.08 (dd, *J*= 1.6, 5.6 Hz, 2H), 4.10 (q, *J*= 2.4 Hz, 2H), 5.80 (td, *J*= 1.6, 16.0 Hz, 1H), 6.16 (td, *J*= 5.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 22.2, 22.8, 25.4, 37.3, 58.1, 69.2, 75.0, 76.0, 83.0, 84.2, 90.1, 112.2, 139.3, 169.4. IR (film): 2937, 1737, 1446, 1367, 1302, 1266, 1231, 1184, 1134, 1074, 1019, 962, 913, 735, 703. HRMS (ESI) *m*/*z* calcd. For C₁₇H₂₂O₃ (M+Na)⁺ 297.1461, found 297.1458.



(E)-8-(but-2-ynyloxy)-2,2-dimethyloct-6-en-4-yn-3-yl acetate (4l)

¹H NMR (400MHz, CDCl₃) δ0.99 (s, 9H), 1.84 (t, *J*= 2.4 Hz, 3H), 2.09 (s, 3H), 4.07 (dd, *J*= 1.6, 5.6 Hz, 2H), 4.09 (q, *J*= 2.4 Hz, 2H), 5.20 (d, *J*= 1.6 Hz, 1H), 5.75 (dtd, *J*= 1.6, 1.6, 16.0 Hz, 1H),

6.17 (td, J= 5.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 21.1, 25.8, 35.5, 58.2, 69.2, 72.5, 75.0, 83.1, 83.7, 86.5, 111.7, 139.9, 170.4. IR (film): 2967, 1738, 1369, 1234, 1165, 1139, 1076, 1017, 976, 909, 736, 702. HRMS (ESI) m/z calcd. For C₁₆H₂₂O₃ (M+Na)⁺ 285.1461, found 285.1464.



(E)-8-(but-2-ynyloxy)-2-methyloct-6-en-4-yn-3-yl acetate (4m)

¹H NMR (400MHz, CDCl₃) δ 0.99 (d, *J*= 6.8 Hz, 3H), 1.02 (d, *J*= 6.8 Hz, 3H), 1.86 (t, *J*= 2.4 Hz, 3H), 1.94-2.05 (m, 1H), 2.09 (s, 3H), 4.08 (dd, *J*= 1.6, 5.6 Hz, 2H), 4.10 (q, *J*= 2.4 Hz, 2H), 5.33 (dd, *J*= 2.0, 5.6 Hz, 1H), 5.77 (dtd, *J*= 2.0, 2.0, 16.0 Hz, 1H), 6.19 (td, *J*= 5.6, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 17.8, 18.4, 21.2, 32.7, 58.2, 69.2, 69.6, 75.0, 83.1, 83.9, 86.2, 111.6, 140.0, 170.3. IR (film): 2967, 1738, 1446, 1371, 1355, 1229, 1168, 1111, 1077, 1018, 981, 954, 904, 737. HRMS (ESI) *m/z* calcd. For C₁₅H₂₀O₃ (M+Na)⁺ 271.1305, found 271.1313.





(E)-7-(but-2-ynyloxy)-1-(tert-butyldimethylsilyloxy)hept-5-en-3-yn-2-yl acetate (4n)

¹H NMR (400MHz, CDCl₃) δ 0.08 (d, *J*=1.6Hz, 6H), 0.89 (s, 9H), 1.86 (t, *J*= 2.0 Hz, 3H), 2.10 (s, 3H), 3.80 (d, *J*= 1.6 Hz, 1H), 3.82 (s, 1H), 4.06-4.12 (m, 4H), 5.56 (dt, *J*= 2.0, 6.8 Hz, 1H), 5.75 (qd, *J*= 1.6, 16.0 Hz, 1H), 6.19 (td, *J*= 5.2, 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ -5.1, 3.8, 18.5, 21.2, 25.9, 58.2, 65.0, 65.5, 69.1, 74.9, 83.1, 84.1, 85.0, 111.2, 140.5, 170.1. IR (film): 2952, 2929, 2857, 1747, 1472, 1370, 1252, 1227, 1132, 1079, 1040, 954, 836, 778. HRMS (ESI) *m*/*z* calcd. For C₁₉H₃₀O₄Si (M+Na)⁺ 373.1806, found 373.1801.



(E)-7-(but-2-ynyloxy)-2-methylhepta-2,3,5-trien-4-yl pivalate (8b)

Allene **8b** was obtained as an unseparable mixture with isomer enyne **4b** (**4b/8b**=1:2). ¹H NMR (400MHz, CDCl₃) δ 1.29 (s, 9H), 1.83 (s, 6H), 1.84-1.87 (m, 3H), 4.04-4.13 (m, 4H), 5.69 (td, *J*= 6.0, 16.0 Hz, 1H), 6.07 (d, *J*= 16.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 3.8, 21.4, 27.5, 39.4, 58.0, 69.7, 75.3, 82.7, 108.3, 118.6, 123.8, 127.2, 175.9, 196.1.

General procedures for the Rh-catalyzed tandem acyloxy migration cycloaddition:

Condition A: To a solution of $[Rh(CO)_2Cl]_2$ (1.9 mg, 5 mol %) in DCE (0.05 M) was added substrate (0.10 mmol). The solution was stirred at 80°C until the reaction was complete as determined by TLC analysis. Then the reaction mixture was added to the solution of K₂CO₃ (28 mg, 0.20 mmol) in methanol (2 mL) at 0 °C. After the reaction was completed, the mixture was quenched with saturated aqueous ammonium chloride (5 mL) and was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by chromatography on silica gel to afford the corresponding cyclohexenone.

Condition B: To a solution of $[Rh(CO)_2Cl]_2$ (1.9 mg, 5 mol %) and $[(CF_3)_2CHO]_3P$ (10.6 mg, 20 mol %) in DCE (0.05 M) was added propargylic ester (0.10 mmol). The solution was stirred at 80°C until the reaction was complete as determined by TLC analysis. Then the reaction mixture was added to the solution of K₂CO₃ (28 mg, 0.20 mmol) in methanol (2 mL) at 0 °C. After the reaction was completed, the mixture was quenched with saturated aqueous ammonium chloride (5 mL) and extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by chromatography on silica gel to afford the corresponding cyclohexenone.



7-methyl-6-(propan-2-ylidene)-1,3,3a,6-tetrahydroisobenzofuran-5-yl acetate (5a)

¹H NMR (400MHz, CDCl₃) δ 1.88 (s, 3H), 1.89 (s, 3H), 1.91-1.94 (m, 3H), 2.16 (s, 3H), 3.20-3.31 (m, 1H), 3.34 (dd, *J*= 7.6, 11.6 Hz, 1H), 4.26 (t, *J*= 7.6 Hz, 1H), 4.37 (d, *J*= 14.0 Hz, 1H), 4.44 (d, *J*= 13.6 Hz, 1H), 5.47 (d, *J*= 1.6 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 19.7, 21.1, 23.3, 24.4, 42.0, 69.1, 72.2, 113.4, 125.6, 128.4, 130.9, 139.3, 150.8, 168.7. IR (film): 2851, 1759, 1683, 1438, 1368, 1270, 1203, 1157, 1073, 1038, 1017, 958, 918, 820, 736. HRMS (ESI) *m/z* calcd. For C₁₄H₁₈O₃ (M+Na)⁺ 257.1148, found 257.1158.



7-methyl-6-(propan-2-ylidene)-1,3,3a,6-tetrahydroisobenzofuran-5-yl pivalate (5b)

¹H NMR (400MHz, CDCl₃) δ 1.27 (s, 9H), 1.87 (s, 3H), 1.88 (s, 3H), 1.92-1.96 (m, 3H), 3.20-3.30 (m, 1H), 3.34 (dd, *J*= 7.6, 11.2 Hz, 1H), 4.25 (t, *J*= 8.0 Hz, 1H), 4.36 (d, *J*= 13.6 Hz, 1H), 4.44 (d, *J*= 13.6 Hz, 1H), 5.41 (d, *J*= 1.6 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 19.6, 23.7, 24.0, 27.5, 39.2, 42.0, 69.0, 72.1, 113.4, 125.8, 128.8, 130.7, 139.3, 151.5, 176.4. IR (film): 2976, 2855, 1748, 1687, 1640, 1482, 1463, 1398, 1368, 1274, 1157, 1128, 1112, 1074, 1039, 924, 835, 764, 738. HRMS (ESI) *m/z* calcd. For C₁₇H₂₄O₃ (M+Na)⁺ 299.1618, found 299.1631.



7-methyl-6-(propan-2-ylidene)-3,3a,4,6-tetrahydroisobenzofuran-5(1H)-one (7a)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.84-1.87 (m, 3H), 1.90 (s, 3H), 2.07 (dd, *J*= 10.8, 18.0 Hz, 1H), 2.11 (s, 3H), 2.60 (dd, *J*= 6.4, 18.0 Hz, 1H), 2.92-3.06 (m, 1H), 3.38 (t, *J*= 8.4 Hz, 1H), 4.30 (t, *J*= 8.4 Hz, 1H), 4.37 (d, *J*= 14.0 Hz, 1H), 4.47 (d, *J*= 14.4 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 19.4, 23.2, 25.0, 38.1, 41.5, 69.5, 74.8, 123.8, 134.6, 141.5, 143.1, 203.5. IR (film): 2936, 2844, 2360, 1683, 1596, 1433, 1365, 1331, 1302, 1225, 1193, 1167, 1107, 1074, 1037, 1000, 979, 922, 876, 802, 700. HRMS (ESI) *m*/*z* calcd. For C₁₂H₁₆O₂ (M+Na)⁺ 215.1043, found 215.1041.



6-(propan-2-ylidene)-2-tosyl-2,3,3a,4-tetrahydro-1H-isoindol-5(6H)-one (7c)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.89 (s, 3H), 2.16 (s, 3H), 2.19 (dd, *J*= 12.4, 13.6 Hz, 1H), 2.44 (s, 3H), 2.62 (dd, *J*= 5.6, 13.6 Hz, 1H), 2.70 (t, *J*= 9.6 Hz, 1H), 2.88-3.04 (m, 1H), 3.84 (d, *J*= 15.2 Hz, 1H), 3.89 (dd, *J*= 7.6, 9.2 Hz, 1H), 4.19 (d, *J*= 15.2 Hz, 1H), 6.44 (q, *J*= 2.4 Hz, 1H), 7.34 (d, *J*= 8.0 Hz, 2H), 7.72 (d, *J*= 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.8, 23.1, 23.7, 39.1, 44.3, 51.2, 54.7, 119.6, 127.0, 127.9, 130.1, 133.3, 137.1, 144.1, 146.9, 199.1. IR (film): 3055, 2935, 1681, 1585, 1451, 1342, 1289, 1265, 1214, 1161, 1093, 1056, 1036, 985,

923, 868, 824, 810, 736, 705, 667. HRMS (ESI) m/z calcd. For C₁₈H₂₁NO₃S (M+Na)⁺ 354.1134, found 354.1125.



7-methyl-6-(propan-2-ylidene)-2-tosyl-2,3,3a,4-tetrahydro-1H-isoindol-5 (6H)-one (7d)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.79-1.83 (m, 3H), 1.85 (s, 3H), 1.93 (dd, *J*= 10.8, 18.0 Hz, 1H), 2.07 (s, 3H), 2.45 (s, 3H), 2.55 (dd, *J*= 6.4, 17.6 Hz, 1H), 2.62 (t, *J*= 8.8 Hz, 1H), 2.92-3.08 (m, 1H), 3.68 (d, *J*= 14.8 Hz, 1H), 3.81 (dd, *J*= 8.4, 9.2 Hz, 1H), 4.04 (d, *J*= 14.4 Hz, 1H), 7.36 (dd, *J*= 0.8, 8.0 Hz, 2H), 7.73 (d, *J*= 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 19.4, 21.8, 23.2, 25.0, 36.8, 42.0, 50.4, 54.2, 126.5, 128.2, 130.0, 132.3, 134.2, 137.2, 143.8, 144.2, 202.3. IR (film): 2923, 1683, 1598, 1345, 1266, 1163, 1094, 1033, 816, 736, 704, 666. HRMS (ESI) *m*/*z* calcd. For C₁₉H₂₃NO₃S (M+Na)⁺ 368.1291, found 368.1292.



7-phenyl-6-(propan-2-ylidene)-2-tosyl-2,3,3a,4-tetrahydro-1H-isoindol-5(6H)-one (7e)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.33 (s, 3H), 2.10 (s, 3H), 2.12 (dd, *J*= 11.6, 17.6 Hz, 1H), 2.43 (s, 3H), 2.61 (dd, *J*= 5.6, 17.6 Hz, 1H), 2.99 (dd, *J*= 5.6, 10.0 Hz, 1H), 3.08-3.20 (m, 1H), 3.57 (dd, *J*= 8.4, 10.0 Hz, 1H), 3.89 (dd, *J*= 2.0, 14.4 Hz, 1H), 4.18 (dd, *J*= 1.2, 14.8 Hz, 1H), 7.07-7.14 (m, 2H), 7.22-7.28 (m, 1H), 7.29-7.37 (m, 4H), 7.65-7.71 (m, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.8, 23.5, 25.5, 37.5, 42.7, 51.4, 52.6, 127.5, 127.7, 128.2, 128.9, 130.0, 132.3, 132.8, 133.0, 138.0, 140.1, 144.2, 147.8, 202.1. IR (film): 1682, 1597, 1440, 1344, 1160, 1092, 1034, 815, 764, 734, 701, 665. HRMS (ESI) *m*/*z* calcd. For C₂₄H₂₅NO₃S (M+Na)⁺ 430.1447, found 430.1459.



7-(4-bromophenyl)-6-(propan-2-ylidene)-2-tosyl-2,3,3a,4-tetrahydro-1H-isoindol-5(6H)-one (7f)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.35 (s, 3H), 2.10 (s, 3H), 2.13 (dd, *J*= 11.6, 17.6 Hz, 1H), 2.44 (s, 3H), 2.62 (dd, *J*= 5.6, 17.6 Hz, 1H), 2.98 (dd, *J*= 5.2, 10.0 Hz, 1H), 3.07-3.20 (m, 1H), 3.57 (dd, *J*= 8.4, 9.6 Hz, 1H), 3.83 (dd, *J*= 2.0, 14.8 Hz, 1H), 4.14 (d, *J*= 15.2 Hz, 1H), 6.99 (td, *J*= 2.4, 8.4 Hz, 2H), 7.34 (d, *J*= 8.0 Hz, 2H), 7.46 (td, *J*= 2.4, 8.4 Hz, 2H), 7.68 (d, *J*= 8.0 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.8, 23.4, 25.7, 37.6, 42.5, 51.3, 52.6, 121.4, 128.2, 129.4, 130.1, 132.0, 132.1, 132.2, 132.4, 138.8, 139.0, 144.3, 148.1, 201.7. IR (film): 2923, 1683, 1597, 1488, 1344, 1266, 1161, 1092, 1035, 1009, 814, 735, 707, 665. HRMS (ESI) *m/z* calcd. For C₂₄H₂₄BrNO₃S (M+Na)⁺ 508.0552, found 508.0558.



3a-methyl-6-(propan-2-ylidene)-2-tosyl-2,3,3a,4-tetrahydro-1H-isoindol-5(6H)-one (7g) Condition A¹H NMR (400MHz, CDCl₃) δ 1.04 (s, 3H), 1.89 (s, 3H), 2.18 (s, 3H), 2.36 (d, *J*= 13.6 Hz, 1H), 2.44 (s, 3H), 2.47 (d, *J*= 13.6 Hz, 1H), 2.85 (d, *J*= 8.8 Hz, 1H), 3.50 (d, *J*= 9.2 Hz, 1H), 3.83 (d, *J*= 14.8 Hz, 1H), 4.24 (d, *J*= 15.2 Hz, 1H), 6.34 (s, 1H), 7.34 (d, *J*= 8.4 Hz, 2H), 7.73 (d, *J*= 8.4 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 21.8, 23.2, 23.8, 24.3, 44.0, 50.5, 51.5, 61.2, 118.3, 126.6, 127.9, 130.0, 133.4, 140.8, 144.0, 147.3, 199.0. IR (film): 2924, 1687, 1597, 1345, 1290, 1267, 1163, 1093, 1047, 815, 736, 705, 666. HRMS (ESI) *m/z* calcd. For C₁₉H₂₃NO₃S (M+Na)⁺ 368.1291, found 368.1296.



Dimethyl-7-methyl-5-oxo-6-(propan-2-ylidene)-3a,4,5,6-tetrahydro-1H-indene-2,2(3H)-dica rboxylate (7h)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.81 (dd, *J*= 10.0, 12.8 Hz, 1H), 1.86 (s, 6H), 2.00 (dd, *J*= 11.2, 17.6 Hz, 1H), 2.07 (s, 3H), 2.63 (dd, *J*= 5.6, 17.6 Hz, 1H), 2.72 (ddd, *J*= 0.8, 8.0, 12.8 Hz, 1H), 2.78-2.92 (m, 1H), 2.96 (d, *J*= 18.4 Hz, 1H), 3.06 (d, *J*= 18.4 Hz, 1H), 3.75 (s, 3H), 3.76 (s, 3H); ¹³C NMR (100MHz, CDCl₃) δ 19.5, 23.1, 25.1, 36.8, 37.9, 40.9, 44.3, 53.1, 53.1, 60.2, 125.9, 134.9, 141.2, 142.0, 172.1, 172.2, 204.1. IR (film): 2954, 1732, 1681, 1598, 1434,

1254, 1200, 1172, 1105, 1066, 1001, 957, 868, 800, 734, 702. HRMS (ESI) m/z calcd. For $C_{17}H_{22}O_5 (M+Na)^+$ 329.1359, found 329.1363.



Dimethyl-7-oxo-6-(propan-2-ylidene)-3,4,6,7,8,8a-hexahydronapht rhoxylate (7i)

halene-2,2(1H)-dicarboxylate (7i)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.56 (t, *J*= 12.8 Hz, 1H), 1.80 (dt, *J*= 4.4, 13.2 Hz, 1H), 1.88 (s, 3H), 2.09 (s, 3H), 2.20-2.33 (m, 1H), 2.31 (dd, *J*= 9.6, 14.0 Hz, 1H), 2.35-2.43 (m, 1H), 2.43-2.56 (m, 2H), 2.65 (dd, *J*= 6.0, 13.6 Hz, 1H), 2.69-2.80 (m, 1H), 3.72 (s, 3H), 3.77 (s, 3H), 6.25 (s, 1H); ¹³C NMR (100MHz, CDCl₃) δ 22.0, 23.1, 30.7, 32.0, 35.5, 39.2, 47.5, 52.9, 53.1, 54.9, 120.9, 129.3, 137.3, 141.5, 171.2, 172.3, 201.1. IR (film): 2954, 1730, 1688, 1434, 1295, 1248, 1157, 1087, 1024, 864, 735. HRMS (ESI) *m*/*z* calcd. For C₁₇H₂₂O₅ (M+Na)⁺ 329.1359, found 329.1357.



7-phenyl-6-(propan-2-ylidene)-2,3,3a,4-tetrahydro-1H-inden-5(6H)-one (7j)

Condition A ¹H NMR (400MHz, CDCl₃) δ 1.36 (s, 3H), 1.42-1.53 (m, 1H), 1.57-1.70 (m, 1H), 1.76-1.88 (m, 1H), 1.98-2.09 (m, 1H), 2.05-2.15 (m, 4H), 2.38-2.49 (m, 1H), 2.59 (dd, *J*= 5.2, 17.6 Hz, 1H), 2.62-2.73 (m, 1H), 2.84-2.97 (m, 1H), 7.14-7.25 (m, 3H), 7.25-7.32 (m, 2H); ¹³C NMR (100MHz, CDCl₃) δ 23.1, 25.2, 25.8, 31.4, 32.5, 38.3, 44.2, 126.3, 128.3, 128.4, 130.6, 134.8, 141.7, 143.9, 148.4, 205.6. IR (film): 2952, 1682, 1598, 1492, 1443, 1266, 1236, 1207, 1171, 1037, 762, 736, 701. HRMS (ESI) *m*/*z* calcd. For C₁₈H₂₀O (M+Na)⁺ 275.1406, found 275.1416.



6-cyclohexylidene-7-methyl-3,3a,4,6-tetrahydroisobenzofuran-5(1H)-one (7k) Condition A¹H NMR (400MHz, CDCl₃) δ 1.50-1.80 (m, 6H), 1.82-1.88 (m, 3H), 2.05 (dd, *J*= 10.0, 17.6 Hz, 1H), 2.12-2.22 (m, 1H), 2.24-2.35 (m, 1H), 2.54-2.72 (m, 3H), 2.96-3.09 (m, 1H), 3.40 (t, J= 8.8 Hz, 1H), 4.29 (t, J= 8.4 Hz, 1H), 4.35 (d, J= 14.4 Hz, 1H), 4.44 (d, J= 14.0 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 19.5, 26.5, 28.8, 28.9, 31.0, 33.6, 38.2, 41.4, 69.4, 74.8, 123.4, 132.6, 141.7, 149.3, 204.3. IR (film): 2926, 2835, 1680, 1598, 1446, 1177, 1088, 1070, 1039, 987, 920, 731. HRMS (ESI) m/z calcd. For C₁₅H₂₀O₂ (M+Na)⁺ 255.1356, found 255.1352.



(Z)-6-(2,2-dimethylpropylidene)-7-methyl-3,3a,4,6-tetrahydroisobenzofuran-5(1H)-one (7l) Condition B ¹H NMR (400MHz, CDCl₃) δ 1.19 (s, 9H), 1.73 (d, *J*= 1.2 Hz, 3H), 2.40 (t, *J*= 12.4, 1H), 2.67 (dd, *J*= 5.2, 12.4 Hz, 1H), 2.94-3.10 (m, 1H), 3.35 (dd, *J*= 8.4, 10.4 Hz, 1H), 4.24 (t, *J*= 8.0 Hz, 1H), 4.44 (d, *J*= 14.0 Hz, 1H), 4.52 (d, *J*= 14.4 Hz, 1H), 5.80 (s, 1H); ¹³C NMR (100MHz, CDCl₃) δ 15.7, 30.6, 33.6, 41.5, 43.8, 69.7, 74.5, 126.2, 134.1, 138.5, 146.3, 202.4. IR (film): 2957, 2867, 1701, 1463, 1387, 1363, 1334, 1266, 1165, 1041, 919, 737, 704. HRMS (ESI) *m*/*z* calcd. For C₁₄H₂₀O₂ (M+Na)⁺ 243.1356, found 243.1361.



7-methyl-6-neopentyl-1,3-dihydroisobenzofuran-5-ol (7l')

¹H NMR (400MHz, CDCl₃) δ 0.98 (s, 9H), 2.16 (s, 3H), 2.62 (s, 2H), 4.83 (s, 1H), 5.03 (s, 2H), 5.08 (s, 2H), 6.54 (s, 1H); ¹³C NMR (100MHz, CDCl₃) δ 17.6, 30.4, 34.5, 38.5, 73.8, 74.2, 105.6, 124.1, 131.0, 132.3, 137.0, 154.7. IR (film): 3242, 2952, 2864, 1604, 1449, 1363, 1327, 1306, 1260, 1159, 1061, 1024, 986, 907, 891, 829, 731. HRMS (ESI) *m/z* calcd. For C₁₄H₂₀O₂ (M+Na)⁺ 243.1356, found 243.1367.



(**Z**)-7-methyl-6-(2-methylpropylidene)-3,3a,4,6-tetrahydroisobenzofuran-5(1H)-one (7m) Condition **B** ¹H NMR (400MHz, CDCl₃) δ 1.00 (d, *J*= 6.4 Hz, 3H), 1.07 (d, *J*= 6.8 Hz, 3H), 1.76 (s, 3H), 2.31 (dd, *J*= 12.0, 13.2 Hz, 1H), 2.68 (dd, *J*= 5.6, 13.2 Hz, 1H), 2.93-3.07 (m, 1H),

3.35 (dd, J= 8.0, 10.4 Hz, 1H), 3.42-3.56 (m, 1H), 4.27 (t, J= 8.0 Hz, 1H), 4.47 (d, J= 14.0 Hz, 1H), 4.55 (d, J= 14.4 Hz, 1H), 5.72 (d, J= 9.6 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ 15.4, 23.1, 23.3, 27.8, 40.1, 43.5, 69.8, 74.7, 124.8, 131.3, 138.4, 145.9, 200.6. IR (film): 2963, 2866, 1694, 1596, 1466, 1388, 1361, 1332, 1218, 1138, 1040, 953, 920, 735. HRMS (ESI) *m/z* calcd. For C₁₃H₁₈O₂ (M+Na)⁺ 229.1199, found 229.1203.



(Z)-6-(2-(tert-butyldimethylsilyloxy)ethylidene)-7-methyl-3,3a,4,6-tetrahydroisobenzofuran -5(1H)-one (7n)

Condition B¹H NMR (400MHz, CDCl₃) δ 0.09 (s, 6H), 0.92 (s, 9H), 1.80 (d, *J*= 1.2 Hz, 3H), 2.27 (dd, *J*= 12.4, 14.4 Hz, 1H), 2.71 (dd, *J*= 5.2, 14.4 Hz, 1H), 2.94-3.10 (m, 1H), 3.37 (dd, *J*= 8.4, 10.4 Hz, 1H), 4.30 (t, *J*= 8.0 Hz, 1H), 4.49 (d, *J*= 14.4 Hz, 1H), 4.85 (d, *J*= 14.0 Hz, 1H), 4.70-4.84 (m, 2H), 6.08 (t, *J*= 4.8 Hz, 1H); ¹³C NMR (100MHz, CDCl₃) δ -4.98, -4.97, 15.3, 18.5, 26.2, 39.7, 42.4, 63.7, 69.7, 74.7, 124.1, 131.4, 139.5, 141.8, 200.1. IR (film): 2929, 2856, 1693, 1387, 1360, 1331, 1263, 1089, 1040, 1012, 921, 837, 778, 736, 704. HRMS (ESI) *m/z* calcd. For C₁₇H₂₈O₃Si (M+Na)⁺ 331.1700, found 331.1700.

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UNIVERSITY OF WISCONSIN-MADISON

MOLECULAR STRUCTURE LABORATORY

ILIA A. GUZEI, PH.D.

University of Wisconsin-Madison 2124 Chemistry Department 1101 University Ave Madison, WI 53706 ***** 608-263-4694 Fax 608-262-0381

E-mail: iguzei@chem.wisc.edu



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Crystallographic Experimental Section

Data Collection

A colorless crystal with approximate dimensions $0.51 \times 0.48 \times 0.42 \text{ mm}^3$ was selected under oil under ambient conditions and attached to the tip of a MiTeGen MicroMount©. The crystal was mounted in a stream of cold nitrogen at 100(1) K and centered in the X-ray beam by using a video camera.

The crystal evaluation and data collection were performed on a Bruker Quazar SMART APEXII diffractometer with Mo K_{α} ($\lambda = 0.71073$ Å) radiation and the diffractometer to crystal distance of 4.95 cm.

The initial cell constants were obtained from three series of ω scans at different starting angles. Each series consisted of 12 frames collected at intervals of 0.5° in a 6° range about ω with the exposure time of 1 second per frame. The reflections were successfully indexed by an automated indexing routine built in the APEXII program suite. The final cell constants were calculated from a set of 7099 strong reflections from the actual data collection.

The data were collected by using the full sphere data collection routine to survey the reciprocal space to the extent of a full sphere to a resolution of 0.75 Å. A total of 18915 data were harvested by collecting 5 sets of frames with 0.5° scans in ω and φ with exposure times of 1 sec per frame. These highly redundant datasets were corrected for Lorentz and polarization effects. The absorption correction was based on fitting a function to the empirical transmission surface as sampled by multiple equivalent measurements. [1]

Structure Solution and Refinement

The systematic absences in the diffraction data were consistent for the space groups P_1 and P_1 . The *E*-statistics suggested the centrosymmetric space group P_1 that yielded chemically reasonable and computationally stable results of refinement [2-4].

A successful solution by the direct methods provided most non-hydrogen atoms from the *E*-map. The remaining non-hydrogen atoms were located in an alternating series of least-squares cycles and difference Fourier maps. All non-hydrogen atoms were refined with anisotropic displacement coefficients. All hydrogen atoms were included in the structure factor calculation at idealized positions and were allowed to ride on the neighboring atoms with relative isotropic displacement coefficients.

The final least-squares refinement of 274 parameters against 5277 data resulted in residuals R (based on F^2 for $I \ge 2\sigma$) and wR (based on F^2 for all data) of 0.0274 and 0.0646, respectively. The final difference Fourier map was featureless.

The molecular diagram is drawn with 50% probability ellipsoids.

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Figure 1. A molecular drawing of **7f**. All H atoms are omitted.

Table 1. Crystal data and structure refinement for	7 f .		
Identification code	7f		
Empirical formula	C ₂₄ H ₂₄ Br N O ₃ S		
Formula weight	486.41		
Temperature	100(2) K		
Wavelength	0.71073 Å		
Crystal system	Triclinic		
Space group	PI		
Unit cell dimensions	a = 7.652(2) Å	$\alpha = 81.281(11)^{\circ}$.	
	b = 8.0993(19) Å	$\beta = 84.362(11)^{\circ}$.	
	c = 18.213(5) Å	$\gamma = 72.965(17)^{\circ}$	
Volume	1065.0(5) Å ³	•	
Ζ	2		
Density (calculated)	1.517 Mg/m ³		
Absorption coefficient	2.054 mm ⁻¹		
F(000)	500		
Crystal size	0.51 x 0.48 x 0.42 mm ³		
Theta range for data collection	2.27 to 28.32°.		
Index ranges	-9<=h<=10, -10<=k<=10, -24<	=l<=24	
Reflections collected	18915		
Independent reflections	5277 [R(int) = 0.0335]		
Completeness to theta = 25.00°	99.8 %		
Absorption correction	Numerical with SADABS		
Max. and min. transmission	0.4791 and 0.4205		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	5277 / 0 / 274		
Goodness-of-fit on F ²	1.009		
Final R indices [I>2sigma(I)]	R1 = 0.0274, $wR2 = 0.0625$		
R indices (all data)	R1 = 0.0336, $wR2 = 0.0646$		
Largest diff. peak and hole	0.501 and -0.363 e.Å ⁻³		

	Х	У	Z	U(eq)
Br(1)	-1372(1)	-718(1)	7173(1)	19(1)
S(1)	5517(1)	6424(1)	6383(1)	14(1)
O(1)	5868(2)	8037(2)	6449(1)	18(1)
O(2)	4074(2)	6401(2)	5943(1)	20(1)
O(3)	1896(2)	5773(2)	10061(1)	21(1)
N(1)	5005(2)	5643(2)	7235(1)	14(1)
C(1)	7550(2)	4970(2)	6070(1)	15(1)
C(2)	7455(3)	3603(2)	5710(1)	21(1)
C(3)	9052(3)	2428(2)	5488(1)	24(1)
C(4)	10761(3)	2583(2)	5613(1)	21(1)
C(5)	10819(2)	3986(2)	5959(1)	20(1)
C(6)	9240(2)	5168(2)	6192(1)	18(1)
C(7)	12487(3)	1275(3)	5374(1)	31(1)
C(8)	6093(2)	5752(2)	7841(1)	16(1)
C(9)	5249(2)	4824(2)	8520(1)	14(1)
C(10)	3755(2)	6043(2)	8961(1)	17(1)
C(11)	2718(2)	5061(2)	9534(1)	14(1)
C(12)	2790(2)	3255(2)	9410(1)	13(1)
C(13)	3187(2)	2927(2)	8615(1)	13(1)
C(14)	4363(2)	3681(2)	8197(1)	13(1)
C(15)	4671(2)	3912(2)	7362(1)	15(1)
C(16)	2557(2)	2022(2)	9975(1)	15(1)
C(17)	2077(3)	2314(2)	10775(1)	20(1)
C(18)	2797(2)	167(2)	9860(1)	18(1)
C(19)	2125(2)	1987(2)	8283(1)	13(1)
C(20)	277(2)	2204(2)	8499(1)	14(1)
C(21)	-751(2)	1391(2)	8173(1)	15(1)
C(22)	70(2)	342(2)	7628(1)	15(1)
C(23)	1905(2)	72(2)	7411(1)	16(1)
C(24)	2921(2)	891(2)	7742(1)	15(1)

Table 2. Atomic coordinates $(x \ 10^4)$ and equivalent isotropic displacement parameters (Å²x 10^3) for **7f**. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

Table 3.	Bond lengths [Å] and angles [°] for	7f.

Br(1)-C(22)	1.8949(17)	C(10)-H(10A)	0.9900
S(1)-O(2)	1.4331(13)	C(10)-H(10B)	0.9900
S(1)-O(1)	1.4332(13)	C(11)-C(12)	1.498(2)
S(1)-N(1)	1.6389(14)	C(12)-C(16)	1.359(2)
S(1)-C(1)	1.7568(18)	C(12)-C(13)	1.497(2)
O(3)-C(11)	1.219(2)	C(13)-C(14)	1.345(2)
N(1)-C(8)	1.475(2)	C(13)-C(19)	1.485(2)
N(1)-C(15)	1.478(2)	C(14)-C(15)	1.507(2)
C(1)-C(6)	1.391(2)	C(15)-H(15A)	0.9900
C(1)-C(2)	1.393(2)	C(15)-H(15B)	0.9900
C(2)-C(3)	1.378(3)	C(16)-C(17)	1.504(2)
C(2)-H(2)	0.9500	C(16)-C(18)	1.504(2)
C(3)-C(4)	1.393(3)	C(17)-H(17A)	0.9800
C(3)-H(3)	0.9500	C(17)-H(17B)	0.9800
C(4)-C(5)	1 393(3)	C(17)-H(17C)	0 9800
C(4)-C(7)	1 504(3)	C(18)-H(18A)	0.9800
C(5)-C(6)	1 379(3)	C(18)-H(18B)	0.9800
C(5)-H(5)	0.9500	C(18)-H(18C)	0.9800
C(6)-H(6)	0.9500	C(19)-C(20)	1.397(2)
C(7)-H(7A)	0.9800	C(19) - C(24)	1.397(2) 1 400(2)
C(7)-H(7B)	0.9800	C(20)-C(21)	1.385(2)
C(7)-H(7C)	0.9800	C(20) - H(20)	0.9500
C(8)-C(9)	1 535(2)	$C(20)$ - $\Pi(20)$ C(21)- $C(22)$	1.387(2)
C(8) - H(8A)	0.9900	C(21)-C(22) C(21)-H(21)	0.9500
C(8) - H(8R)	0.9900	$C(21)-\Pi(21)$ C(22)-C(23)	1.383(2)
$C(0) - \Gamma(0D)$	1.508(2)	C(22)-C(23) C(23)-C(24)	1.303(2) 1.387(2)
C(9) - C(14)	1.508(2) 1.524(2)	C(23) - C(24) C(23) - H(23)	0.0500
C(9) - C(10)	1.0000	$C(23)$ - $\Pi(23)$ $C(24)$ $\Pi(24)$	0.9500
$C(9)$ - $\Pi(9)$ C(10) $C(11)$	1.0000 1.510(2)	C(24)- $H(24)$	0.9300
O(2) S(1) O(1)	1.319(2) 120.23(8)	C(5) C(6) H(6)	120.3
O(2) - S(1) - O(1) O(2) - S(1) - N(1)	120.23(8)	C(3)-C(0)-H(0) C(1) $C(6)$ $H(6)$	120.3
O(2)-S(1)-N(1) O(1) S(1) N(1)	100.23(7) 105.17(7)	C(1)-C(0)-H(0) C(4) C(7) H(7A)	120.5
O(1)-S(1)-N(1) O(2) S(1) C(1)	105.17(7) 108.10(8)	C(4) - C(7) - H(7R)	109.5
O(2)-S(1)-C(1)	108.10(8) 108.04(8)	U(7A) C(7) U(7D)	109.5
V(1) - S(1) - C(1)	108.94(8)	$\Pi(/A) - C(/) - \Pi(/B)$	109.5
N(1)-S(1)-C(1)	107.46(8) 107.00(12)	U(7A) C(7) U(7C)	109.5
C(8) N(1) C(13)	107.90(12)	$\Pi(7A) - C(7) - \Pi(7C)$	109.5
C(0)-N(1)-S(1) C(15) N(1) S(1)	118.83(11) 118.48(11)	$\Pi(7D) - C(7) - \Pi(7C)$	109.3 102.24(12)
C(13)-N(1)-S(1)	118.46(11)	N(1) - C(0) - C(9)	102.34(13)
C(0)-C(1)-C(2)	120.30(10) 120.26(12)	$N(1)-C(0)-\Pi(0A)$	111.5
C(0)-C(1)-S(1)	120.20(13) 110.42(14)	V(9)-V(8)-H(8A)	111.3
C(2)-C(1)-S(1)	119.43(14)	N(1)-C(8)-H(8B)	111.3
C(3)-C(2)-C(1)	119.32(17)	C(9)-C(8)-H(8B)	111.3
C(3)-C(2)-H(2)	120.3	H(8A)-C(8)-H(8B)	109.2
C(1)-C(2)-H(2)	120.3	C(14) - C(9) - C(10)	107.34(14)
C(2)-C(3)-C(4)	121.46(17)	C(14) - C(9) - C(8)	104.45(13)
C(2)-C(3)-H(3)	119.3	C(10)-C(9)-C(8)	114.12(14)
C(4)-C(3)-H(3)	119.5	C(14)-C(9)-H(9)	110.2
C(3)-C(4)-C(5)	118.11(17)	C(10)-C(9)-H(9)	110.2
C(3)-C(4)-C(7)	120.58(18)	C(8)-C(9)-H(9)	110.2
C(5)-C(4)-C(7)	121.31(18)	C(11)-C(10)-C(9)	112.22(14)
C(6)-C(5)-C(4)	121.46(17)	C(11)-C(10)-H(10A)	109.2
C(6)-C(5)-H(5)	119.3	C(9)-C(10)-H(10A)	109.2
C(4)-C(5)-H(5)	119.3	С(11)-С(10)-Н(10В)	109.2
C(5)-C(6)-C(1)	119.32(16)	C(9)-C(10)-H(10B)	109.2

H(10A)-C(10)-H(10B)	107.9	H(17A)-C(17)-H(17C)	109.5
O(3)-C(11)-C(12)	124.35(15)	H(17B)-C(17)-H(17C)	109.5
O(3)-C(11)-C(10)	118.78(15)	C(16)-C(18)-H(18A)	109.5
C(12)-C(11)-C(10)	116.87(14)	C(16)-C(18)-H(18B)	109.5
C(16)-C(12)-C(13)	123.42(15)	H(18A)-C(18)-H(18B)	109.5
C(16)-C(12)-C(11)	122.49(14)	C(16)-C(18)-H(18C)	109.5
C(13)-C(12)-C(11)	114.07(13)	H(18A)-C(18)-H(18C)	109.5
C(14)-C(13)-C(19)	121.44(14)	H(18B)-C(18)-H(18C)	109.5
C(14)-C(13)-C(12)	117.94(14)	C(20)-C(19)-C(24)	118.25(15)
C(19)-C(13)-C(12)	120.16(14)	C(20)-C(19)-C(13)	120.14(14)
C(13)-C(14)-C(15)	128.48(15)	C(24)-C(19)-C(13)	121.60(15)
C(13)-C(14)-C(9)	120.87(14)	C(21)-C(20)-C(19)	120.85(15)
C(15)-C(14)-C(9)	109.16(13)	C(21)-C(20)-H(20)	119.6
N(1)-C(15)-C(14)	100.67(12)	C(19)-C(20)-H(20)	119.6
N(1)-C(15)-H(15A)	111.6	C(20)-C(21)-C(22)	119.47(16)
C(14)-C(15)-H(15A)	111.6	C(20)-C(21)-H(21)	120.3
N(1)-C(15)-H(15B)	111.6	C(22)-C(21)-H(21)	120.3
C(14)-C(15)-H(15B)	111.6	C(23)-C(22)-C(21)	121.14(15)
H(15A)-C(15)-H(15B)	109.4	C(23)-C(22)-Br(1)	120.03(12)
C(12)-C(16)-C(17)	125.21(15)	C(21)-C(22)-Br(1)	118.83(13)
C(12)-C(16)-C(18)	122.45(15)	C(22)-C(23)-C(24)	118.88(15)
C(17)-C(16)-C(18)	112.33(14)	C(22)-C(23)-H(23)	120.6
C(16)-C(17)-H(17A)	109.5	C(24)-C(23)-H(23)	120.6
C(16)-C(17)-H(17B)	109.5	C(23)-C(24)-C(19)	121.39(16)
H(17A)-C(17)-H(17B)	109.5	C(23)-C(24)-H(24)	119.3
C(16)-C(17)-H(17C)	109.5	C(19)-C(24)-H(24)	119.3

Symmetry transformations used to generate equivalent atoms:

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²
$\overline{\text{Br}(1)}$	24(1)	20(1)	18(1)	-6(1)	-1(1)	-13(1)
S(1)	15(1)	15(1)	11(1)	0(1)	0(1)	-6(1)
O(1)	22(1)	14(1)	17(1)	-1(1)	2(1)	-7(1)
O(2)	19(1)	26(1)	16(1)	0(1)	-4(1)	-8(1)
O(3)	25(1)	20(1)	18(1)	-8(1)	4(1)	-6(1)
N(1)	17(1)	15(1)	11(1)	-1(1)	0(1)	-8(1)
C(1)	18(1)	16(1)	11(1)	0(1)	1(1)	-5(1)
C(2)	24(1)	23(1)	19(1)	-7(1)	3(1)	-13(1)
C(3)	32(1)	20(1)	24(1)	-8(1)	7(1)	-12(1)
C(4)	26(1)	19(1)	14(1)	2(1)	2(1)	-3(1)
C(5)	18(1)	26(1)	14(1)	0(1)	-3(1)	-5(1)
C(6)	20(1)	19(1)	15(1)	-3(1)	-2(1)	-7(1)
C(7)	30(1)	27(1)	27(1)	-2(1)	5(1)	1(1)
C(8)	17(1)	19(1)	13(1)	-3(1)	0(1)	-9(1)
C(9)	15(1)	15(1)	11(1)	-2(1)	0(1)	-6(1)
C(10)	22(1)	14(1)	17(1)	-3(1)	2(1)	-7(1)
C(11)	15(1)	15(1)	14(1)	-1(1)	-2(1)	-4(1)
C(12)	12(1)	15(1)	13(1)	-3(1)	-1(1)	-5(1)
C(13)	12(1)	14(1)	13(1)	-2(1)	-1(1)	-3(1)
C(14)	14(1)	14(1)	12(1)	-3(1)	-1(1)	-4(1)
C(15)	18(1)	15(1)	12(1)	-2(1)	1(1)	-9(1)
C(16)	14(1)	18(1)	15(1)	-2(1)	-1(1)	-6(1)
C(17)	26(1)	23(1)	13(1)	0(1)	1(1)	-10(1)
C(18)	20(1)	16(1)	18(1)	1(1)	-1(1)	-7(1)
C(19)	15(1)	12(1)	13(1)	0(1)	-1(1)	-5(1)
C(20)	16(1)	15(1)	13(1)	-3(1)	0(1)	-5(1)
C(21)	15(1)	16(1)	15(1)	-2(1)	0(1)	-6(1)
C(22)	21(1)	13(1)	14(1)	-2(1)	-2(1)	-9(1)
C(23)	21(1)	14(1)	13(1)	-3(1)	2(1)	-6(1)
C(24)	16(1)	15(1)	14(1)	-1(1)	2(1)	-5(1)

Table 4.Anisotropic displacement parameters $(Å^2x \ 10^3)$ for **7f**.The anisotropicdisplacement factor exponent takes the form: $-2\pi^2$ [$h^2 \ a^{*2}U^{11} + \dots + 2 \ h \ k \ a^* \ b^* \ U^{12}$]

	X	у	Z	U(eq)
H(2)	6301	3481	5617	25
H(3)	8986	1491	5245	29
H(5)	11973	4131	6035	24
H(6)	9305	6108	6433	22
H(7A)	12526	121	5641	46
H(7B)	13552	1611	5487	46
H(7C)	12507	1242	4837	46
H(8A)	5957	6978	7903	19
H(8B)	7406	5147	7752	19
H(9)	6227	4095	8854	16
H(10A)	4315	6740	9216	21
H(10B)	2884	6857	8613	21
H(15A)	5744	2999	7193	17
H(15B)	3578	3913	7112	17
H(17A)	896	2090	10932	31
H(17B)	1996	3521	10828	31
H(17C)	3027	1522	11087	31
H(18A)	3473	-59	9383	27
H(18B)	1592	-29	9858	27
H(18C)	3481	-621	10264	27
H(20)	-282	2919	8874	17
H(21)	-2009	1551	8321	18
H(23)	2460	-662	7043	19
H(24)	4185	704	7599	18

Table 5. Hydrogen coordinates ($x \ 10^4$) and isotropic displacement parameters (Å²x 10³) for **7f**.

Table 6. Torsion angles [°] for **7f**.

O(2)-S(1)-N(1)-C(8)	-170.73(12)	C(14)-C(13)-C(19)-C(20)	-137.52(17)
O(1)-S(1)-N(1)-C(8)	-42.23(14)	C(12)-C(13)-C(19)-C(20)	34.5(2)
C(1)-S(1)-N(1)-C(8)	73.73(14)	C(14)-C(13)-C(19)-C(24)	41.5(2)
O(2)-S(1)-N(1)-C(15)	55.02(14)	C(12)-C(13)-C(19)-C(24)	-146.48(16)
O(1)-S(1)-N(1)-C(15)	-176.49(12)	C(24)-C(19)-C(20)-C(21)	-1.6(2)
C(1)-S(1)-N(1)-C(15)	-60.52(14)	C(13)-C(19)-C(20)-C(21)	177.43(15)
O(2)-S(1)-C(1)-C(6)	156 51(13)	C(19)-C(20)-C(21)-C(22)	0 3(2)
O(1)-S(1)-C(1)-C(6)	$24\ 27(15)$	C(20)-C(21)-C(22)-C(23)	10(2)
N(1)-S(1)-C(1)-C(6)	-89 18(14)	C(20)-C(21)-C(22)-Br(1)	-178.66(12)
O(2)-S(1)-C(1)-C(2)	-2444(16)	C(21)-C(22)-C(23)-C(24)	-0.9(2)
O(1)- $S(1)$ - $C(1)$ - $C(2)$	-156 68(13)	Br(1)-C(22)-C(23)-C(24)	17874(12)
N(1) - S(1) - C(1) - C(2)	89.87(15)	C(22) C(23) C(24) C(24)	-0.5(2)
C(6) C(1) C(2) C(3)	1 4(2)	C(22) - C(23) - C(24) - C(13)	-0.3(2) 1.7(2)
S(1)-C(1)-C(2)-C(3)	-177.69(14)	C(20) - C(19) - C(24) - C(23)	-177.33(15)
C(1) C(2) C(3)	-177.09(14)	C(13) - C(13) - C(24) - C(23)	-177.55(15)
C(1)-C(2)-C(3)-C(4)	-0.4(3)		
C(2) - C(3) - C(4) - C(3)	-1.2(3) 170.20(17)		
C(2)-C(3)-C(4)-C(7)	1/9.39(1/)		
C(3)-C(4)-C(5)-C(6)	1.8(3)		
C(7)-C(4)-C(5)-C(6)	-1/8./3(16)		
C(4)-C(5)-C(6)-C(1)	-0.9(3)		
C(2)-C(1)-C(6)-C(5)	-0.7(2)		
S(1)-C(1)-C(6)-C(5)	1/8.33(13)		
C(15)-N(1)-C(8)-C(9)	-38.91(16)		
S(1)-N(1)-C(8)-C(9)	-177.49(11)		
N(1)-C(8)-C(9)-C(14)	22.49(16)		
N(1)-C(8)-C(9)-C(10)	-94.41(16)		
C(14)-C(9)-C(10)-C(11)	53.99(18)		
C(8)-C(9)-C(10)-C(11)	169.22(13)		
C(9)-C(10)-C(11)-O(3)	158.33(15)		
C(9)-C(10)-C(11)-C(12)	-21.4(2)		
O(3)-C(11)-C(12)-C(16)	-25.1(3)		
C(10)-C(11)-C(12)-C(16)	154.64(16)		
O(3)-C(11)-C(12)-C(13)	156.36(16)		
C(10)-C(11)-C(12)-C(13)	-23.9(2)		
C(16)-C(12)-C(13)-C(14)	-142.13(17)		
C(11)-C(12)-C(13)-C(14)	36.4(2)		
C(16)-C(12)-C(13)-C(19)	45.6(2)		
C(11)-C(12)-C(13)-C(19)	-135.88(15)		
C(19)-C(13)-C(14)-C(15)	7.6(3)		
C(12)-C(13)-C(14)-C(15)	-164.64(15)		
C(19)-C(13)-C(14)-C(9)	172.09(15)		
C(12)-C(13)-C(14)-C(9)	-0.1(2)		
C(10)- $C(9)$ - $C(14)$ - $C(13)$	-45 5(2)		
C(8)-C(9)-C(14)-C(13)	-167.01(15)		
C(10)- $C(9)$ - $C(14)$ - $C(15)$	121 72(14)		
C(8)- $C(9)$ - $C(14)$ - $C(15)$	0.23(17)		
C(8)-N(1)-C(15)-C(14)	38 37(16)		
S(1)-N(1)-C(15)-C(14)	177 12(11)		
C(13)-C(14)-C(15)-N(1)	1/7, 1/2(11) 1/3, 31(17)		
C(9) - C(14) - C(15) - N(1)	-27.66(17)		
C(13) - C(12) - C(16) - C(17)	$_{-177}^{-22.00(17)}$		
C(11) - C(12) - C(16) - C(17)	-1/7.21(10)		
C(12)-C(12)-C(10)-C(17)	4.4(<i>3</i>) <i>A</i> 1(2)		
C(13) - C(12) - C(10) - C(18)	4.1(3) 174.26(15)		
C(11)-C(12)-C(10)-C(18)	-1/4.30(15)		

Symmetry transformations used to generate equivalent atoms:

























































































































